REVIEW

Cannabis and Cannabinoid Research

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Endocannabinoids and Immunity

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Abstract

Over the last 10 years, a great boost of knowledge accumulated on the immunomodulatory and anti-inflammatory properties of endocannabinoids (eCBs). In this scenario, these bioactive lipids, which are produced by most immune cells along with a set of receptors and enzymes that regulate their synthesis and degradation, act as secondary modulators and increase or decrease a plethora of immune functions. In this review, the manifold immunomodulatory effects of the main eCBs in different compartments of innate and adaptive immunity will be discussed, suggesting that they could be considered as master regulators of innate-adaptive immune axis and as potent immunoresolvents.

Key words: endocannabinoid; immunology; immunosuppression

Endocannabinoids and Their Metabolism in the Immune System

Endocannabinoids (eCBs) include a group of lipid mediators, the best characterized of which are Narachidonoylethanolamine (mostly known as anandamide, AEA) and 2-arachidonovlglycerol (2-AG).^{1,2} Some other compounds have been proposed to belong to the eCB family, including 2-AG-ether (noladin ether) and O-arachidonoylethanolamine (virodhamine), as well as two additional N-acylethanolamines (NAEs), namely Npalmitoylethanolamine (PEA) and N-oleoylethanolamine (OEA).^{1,2} eCBs are synthesized and released on demand (if and when needed) from membrane phospholipids in response to physiological or pathological stimuli, although recent evidence has shown that they could be accumulated and stored by intracellular transporters and storage organelles/pools.³ It is now well established that all body districts and tissues produce eCBs as part of a homeostatic system that acts at almost every level of biological life, with the aim of controlling several physiopathological states and maintaining human health. Their role in the regulation of the immune system is probably the most flourishing and promising research field due to the increasing recognition of the eCBs signaling in several

chronic inflammatory diseases.^{1,4} In particular, AEA and other NAEs are synthesized from phosphatidylethanolamine through a series of enzymatic steps that involve mainly the enzyme N-acyl-phosphatidylethanolaminehydrolyzing phospholipase D (NAPE-PLD) as well as, alternatively, phospholipase A and lyso-PLD, α/β hydrolase 4 (ABDH4) and glycerophosphodiesterase 1, or phosholipase C and protein tyrosine phosphatase type-22.² On the contrary, 2-AG is synthesized from sn-1-acyl-2-arachidonoylglycerols (DAGs) through two Ca^{2+} -sensitive *sn*-2-selective DAG lipases (DAGL- α and DAGL- β) or more rarely from 2-arachidonoylglycerol-3-phosphate (2-AG-3P).² Once synthesized, eCBs bind to and functionally activate their target receptors, triggering various signaling pathways and causing several biological effects on different tissues. The main receptor targets for eCBs are type-1 (CB₁) and type-2 (CB₂) G protein-coupled cannabinoid receptors. CB₁ is widely expressed in the nervous system mainly at the terminal ends of central and peripheral neurons and its presence has also been widely documented in periphery. CB₂ is mainly expressed by cells of the immune system, where it is commonly associated with the regulation of different immune functions.^{1,4} Yet, CB₂ can also be found in

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neurons and activated microglial cells and astrocytes in response to various insults, particularly in association with chronic inflammation of the nervous system.⁵ However, eCBs can bind to and activate other receptors, including the transient receptor potential vanilloid 1 (TRPV1) channel, peroxisome proliferatoractivated receptor (PPAR) α and γ , and the orphan G protein-coupled receptor GPR55, all of them being also widely expressed in immune cells.^{1,5,6} For instance, PPAR α and PPAR γ mediate part of the eCB-induced immunomodulatory effects on several immune cells,^{7,8} and GPR55 was recently found to be specifically expressed on monocytes and natural killer (NK) cells and to mediate activatory properties.⁹ The inactivation of eCBs involves cellular uptake through a purported and still unidentified endocannabinoid membrane transporter and subsequent intracellular hydrolysis. AEA is mainly cleaved not only by fatty acid amide hydrolase (FAAH) into arachidonic acid and ethanolamine but also by another enzyme, N-acylethanolamine-hydrolyzing acid amidase (NAAA), which is mainly involved in the hydrolysis of PEA.² 2-AG, instead, is cleaved into glycerol and arachidonic acid not only mainly by monoacylglycerol lipase (MAGL) but, in part, also by FAAH, α/β -hydrolase domain-containing protein 6 (ABHD6), and 12 (ABHD12).² Furthermore, AEA and 2-AG can also be metabolized by cyclooxygenase-2 (COX-2), lipoxygenase (LOX) isozymes, and cytochrome P450, generating several oxidized compounds such as prostaglandin-ethanolamides and glyceryl esters, hydroxy-anandamides, and hydroxyeicosatetraenoyl-glycerols.¹⁰ As it will be discussed in detail in the next session, it is no surprise that the immunosuppressive effects of eCBs on immune cells are primarily mediated through CB₂, whose expression is indeed higher than that of CB₁.^{1,4}

Innate Immunity

Monocytes/macrophages

Macrophages (and monocytes, their precursor cells) play an important role in innate immunity since they do not only clear apoptotic cells and pathogens but they also instruct other immune cells (Fig. 1). Monocytes/macrophages are highly plastic (they can change their functional phenotype depending on environmental cues) and reside in every tissue of the body, where they bear different names (i.e., Kupffer cells in the liver or microglia in the central nervous system),¹¹

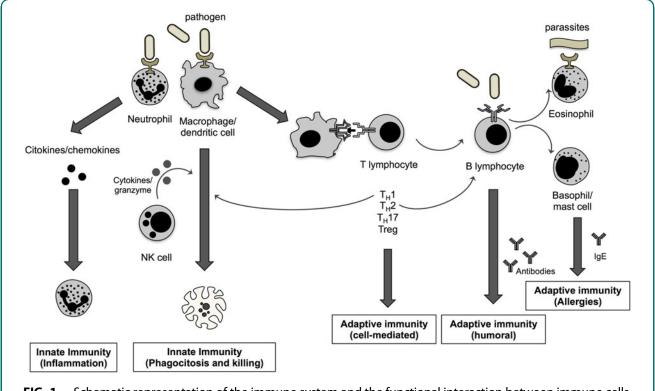


FIG. 1. Schematic representation of the immune system and the functional interaction between immune cells.

CB₁ and CB₂ receptors are highly expressed in both murine and human monocytes/macrophages and microglial cells regardless of cellular models.^{1,12,13}

Dendritic cells

Dendritic cells (DCs) are the most professional antigen-presenting cells, crucial in the development of antigen-specific T-cell responses (Fig. 1). They are present in those tissues that are in contact with the external environment, such as the skin (i.e., Langerhans cells) and the inner lining of several organs; they can also be found in peripheral blood (i.e., myeloid and plasmacytoid DCs).¹⁴ Despite their role in shaping the type and quality of immune responses due to their position at the crossroads between innate and adaptive immunity, very few studies have investigated eCB signaling in these cells, especially in humans.

Neutrophils and NK cells

Neutrophils and NK cells are crucial elements of innate immunity and are both involved in host defense against cancer and antimicrobial responses (Fig. 1). Neutrophils are the first inflammatory cells to be recruited at the site of inflammation/injury and are the hallmark of acute inflammation. Although, neutrophils express very low levels of cannabinoid receptors, a great deal of information has been accumulated on the role of both AEA and 2-AG in human neutrophils, maybe due to their abundance in peripheral blood. On the contrary, knowledge on eCB signaling in NK cells, which are a type of cytotoxic lymphocytes that provide rapid responses against virally infected cells and cancer cells,^{15,16} is almost null, although these cells have been shown to express CB₁, CB₂, and GPR55, as well as to release high levels of AEA and 2-AG.¹

Eosinophils, basophils, and mast cells

These rare cell populations share similar appearance and function and are involved in allergy and anaphylaxis, as well as in wound healing and in defense against pathogens (Fig. 1). However, they differ, in that they arise from different cell lines and in that eosinophils and basophils are found in the blood, whereas mast cells are tissue resident (i.e., connective and mucosal tissue, nervous system).^{17,18} Furthermore, eosinophils play a major role in dealing with elimination of large parasites.¹⁸ As yet, no evidence has been reported on eCB signaling either on murine or on human basophils. Very few reports addressed eosinophil response to eCBs and in particular to 2-AG. Concerning mast cells, PEA is the most extensively investigated eCB (especially in wild-type rats) also due to the fact that these cells produce high levels of PEA and express both CB_1 and CB_2 .^{1,19}

AEA and innate immunity. The first evidence for an immunoregulatory role of eCBs on monocytes/macrophages came from a study on mouse alveolar macrophages, where AEA inhibited macrophage-mediated killing of tumor necrosis factor (TNF)-sensitive cells.²⁰ Later evidence supported the anti-inflammatory activity of AEA, whereby it inhibited expression of proinflammatory mediators such as nitric oxide and interleukins, IL-6, IL-12, and IL23, and enhanced anti-inflammatory mediators such as IL-10 and CD200R.^{1,21} These immunosuppressive effects were mostly mediated by CB₂. The presence of the endocannabinoid system (ECS) on DCs was demonstrated for the first time by Di Marzo and coworkers (AEA, 2-AG, PEA, CB₁, CB₂, and FAAH), with 2-AG levels increasing significantly on activated human DCs.²² At the same time, it was found that high (micromolar) doses of AEA induce apoptosis in murine bone marrow-derived DCs, through both CB1 and CB2 receptors, providing a potential mechanism for eCB-mediated immunosuppression of immune cells.²³ Interestingly, the efficacy of AEA depended on its rapid hydrolysis by FAAH, whose pharmacological inhibition led to reduced resistance to apoptosis. The involvement of CB1 and CB2 in determining DC responses was clearly elucidated by analyzing the phenotypic and functional profile of murine bone marrow-derived DCs from $CB_1^{-/-}CB_2^{-/-}$ mice. Indeed, deletion of both receptors exacerbated DC function by increasing their activation markers, MHC-I/II, CD80, and CD86, and by eliciting a more robust T-cell response.²⁴ In contrast, another work reported that nanomolar and low micromolar doses of AEA before sensitization increased both the expression of murine DC costimulatory molecules CD80/ CD86 and IL-12/IL-23 production ex vivo.²⁵ Moreover, our group investigated more deeply the two subsets of DCs, that is, myeloid and plasmacytoid DCs. Notably, we found that only AEA was able to inhibit $TNF-\alpha$, IL-12p40, and IL-6 from activated myeloid as well as TNF- α and IFN- α release from plasmacytoid DCs, both in a CB₂-dependent mechanism.²⁶ Furthermore, AEAmediated immunosuppression of both DC subsets was also paralleled by a reduced ability of myeloid and plasmacytoid DCs to polarize naïve CD4 T cells into Th1 and Th17 lineages. AEA has been shown to inhibit neutrophil migration, and its levels positively correlated with their phagocytic capabilities^{1,27}; yet, many studies consistently reported a failure of AEA to effectively inhibit superoxide and hydrogen peroxide production, thus being almost ineffective in regulating microbial neutrophil burst reaction. Interestingly, these effects seemed to be independent of cannabinoid receptors. In addition, AEA has been shown to limit excessive mast cell maturation and activation in a CB₁-dependent mechanism in a human hair follicle organ culture model, suggesting that normal skin mast cells are indeed modulated by the ECS.²⁸ The involvement of AEA and CB₁ in modulating human mast cell functions was further confirmed by the observation that in human airway mucosal mast cells, maturation and excessive activation were inhibited by the eCB tone through CB₁ stimulation.²⁹ A very recent and interesting work further unraveled the biological implication of AEA-CB1-mediated mast cell modulation in mast cell-deficient mice, showing that AEA activation of CB₁ in mast cells induced MCP-1-mediated recruitment of monocytic and anti-inflammatory myeloidderived suppressor cells.³⁰

2-AG in innate immunity. Controversial data exist on the role of 2-AG in the modulation of macrophages/ microglia responses. On the one hand, 2-AG inhibits TNF-α and IL-6 production and promotes phagocytosis of opsonized zymosan and alternatively activated and anti-inflammatory M2 macrophages.¹ On the other hand, it increases iNOS-dependent nitric oxide and chemokine production, as well as migration and cell adhesion.¹ In some cases, it was not entirely clear whether the effects of 2-AG were actually mediated through CB₂ receptors. Additionally, it has been suggested that discrepancies on the effects of 2-AG can be due to their conversion into bioactive COX-2 metabolites.¹⁰ On DCs, 2-AG acts as a chemoattractant for both immature and mature bone marrow-derived mouse DCs, also shifting the response toward the Th1 type.³¹ In parallel, 2-AG seems to be also an activator of human neutrophils by stimulating myeloperoxidase release, leukotriene B4 biosynthesis, kinase activation, and calcium mobilization.³² It also induces increased levels of antimicrobial effectors, thereby being a potent regulator of host defense in vivo. As expected, these effects on neutrophil activation were not mediated by CB₂, also due to the very low levels of its expression in these cells, but were rather due to its hydrolysis and subsequent metabolism into LTB₄, with activation of BLT₁ receptors. Additional data supported a role for 2-AG in controlling RhoA activation, thereby suppressing neutrophil migration.³³ Almost no evidence exists on the effects of eCBs on NK cells, where this compound induced the migration of NKdifferentiated human HL-60 cells through CB2.1,34 2-AG induced the migration of human eosinophils in a CB₂-dependent manner, where such receptor was particularly expressed, although less potently than typical, strong eosinophil chemoattractants such as plateletactivating factor, RANTES, and eotaxin.³⁵ These studies suggest that CB₂ and its endogenous ligand 2-AG may be potentially involved in allergic inflammation, accompanied by eosinophil infiltration, and this was demonstrated in a mouse model of contact dermatitis. A very recent article investigated the mechanisms of 2-AG-induced migration of human eosinophils, confirming that this eCB in combination with IL-5 has the ability to activate and modulate eosinophil functional responses and that the 15-LOX pathway is very likely involved in the regulation of these activities.³⁶

PEA in innate immunity. The immunomodulatory role of PEA has been, so far, investigated only on monocytes/ macrophages and on mast cells. In particular, PEA exerts anti-inflammatory properties on the macrophages of the brain (i.e., microglia), mainly by stimulating phagocytosis and clearance of pathogens and by increasing resistance to infection and microglial cell motility.¹ On mast cells, PEA is a strong inhibitor of mast cell degranulation and activation, also contributing to reduce the severity of spinal cord trauma.¹⁹ Interestingly, a recent work hypothesized that the anti-nociceptive role of PEA in inducing relief in neuropathic pain correlates with its ability to modulate these cells.³⁷

Adaptive Immunity

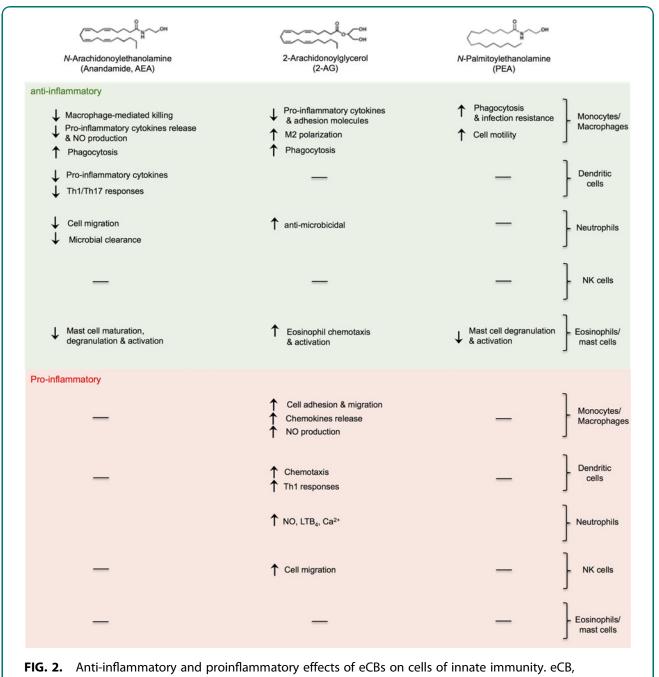
T Lymphocytes

T lymphocytes (or T cells) play a central role in cellmediated immunity and comprise several subsets (Fig. 1), each with a distinct function, including CD4+ T helper cells (Th), CD8+ cytotoxic T cells, memory T cells, regulatory T cells, $\gamma\delta$ T cells, and mucosal-associated invariant T cells (MAIT).³⁸ Although, it is not known whether these specific cell subsets are capable to produce eCBs, their expression of cannabinoid receptors has been extensively investigated and it seems that T cells usually bear very low levels of both CB₁ and CB₂.^{1,39} However, we have demonstrated that CB₂ significantly increases CD4+ and CD8+ human T cells when activated,³⁹ supporting the view that these cells are indeed responsive to the effects of eCBs.

B Lymphocytes

B lymphocytes (or B cells) are not only involved in the production of antibodies against antigens (humoral immunity) but they are also capable of acting as antigen-presenting cells (Fig. 1).⁴⁰ Antibody-producing plasma cells are among the immune cells that express the highest levels of CB₂, with human B cells expressing

one transcript and mouse B cells expressing three transcripts, specifically selected during B-cell activation by lipopolysaccharide. However, most of the research has focused only on the use of phytocannabinoids and syntho-cannabinoids, rather than on eCBs, trying to understand the functional role of this receptor in B cells. Indeed, CB₂ was identified as a crucial receptor

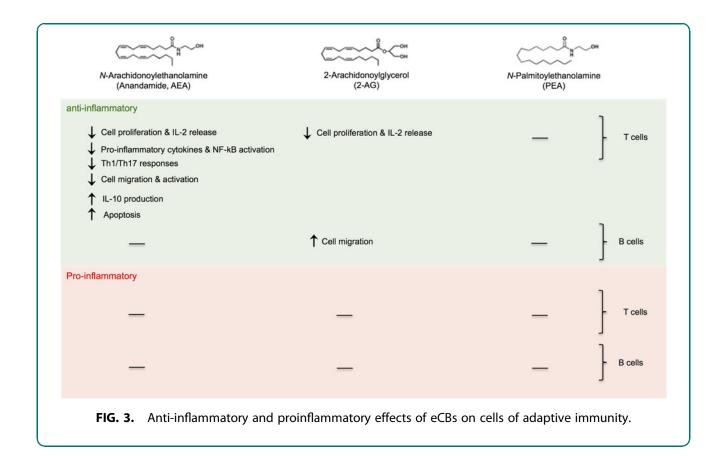


endocannabinoid.

for mouse B-cell differentiation since the end of the 90s as it was markedly expressed in mantle zones of secondary follicles and less in germinal centers and its expression was downregulated during B-cell differentiation. Furthermore, CB_2 was found to be essential also for mouse B-cell subset formation and for retention of immature B cells in bone marrow sinusoids and in the splenic marginal zone. CB_2 was also reported to mediate immunoglobulin class switching from IgM to IgE, suggesting that this CB receptor could have a crucial role in the generation of B-cell repertoire and the regulation of Th2-type humoral responses.¹

AEA in adaptive immunity. The first evidence for an immunosuppressive role of eCBs on T cells came just 2 years after the isolation and purification of AEA, demonstrating its dose-dependent antiproliferative effects on human T cells. Indeed, micromolar doses of AEA rapidly inhibited mitogen-induced DNA synthesis and this was associated with induction of apoptotic cell death.¹ Since then, most of the literature focused only on phytocannabinoids and synthetic agonists/antagonists selective for CB₁ or CB₂. It is now well ac-

cepted that AEA is a potent immunosuppressor of T-cell proliferation and cytokine release, acting mainly through CB_2 and PPAR- γ and most likely through NFkB inhibition. This pathway has been largely investigated in mouse and human T cells.^{1,41,42} Our group was the first to demonstrate the antiproliferative effect of AEA on both CD4 and CD8 T-cell subsets, without any effect on cell viability.³⁹ In addition, we disclosed its inhibitory effect on IFN-y-producing Th1 and IL-17-producing Th17. This effect of AEA on Th17 has been recently reproduced in a mouse model of hypersensitivity, where it was also shown to be mediated by IL-10 and miRNA induction.⁴³ Interestingly, cytokines have been shown to directly influence other elements of the eCB system in T lymphocytes since Th2 cytokines, IL-4 or IL-10, had a stimulatory effect on FAAH, whereas the Th1 cytokines, IL-12 and IFN- γ , reduced FAAH activity and protein expression, overall suggesting an eCBtriggered self-sustaining anti-inflammatory loop. The strong involvement of CB₂ in mediating AEA antiinflammatory effects is supported by a reduction of eCB immune modulation of T cells from a common CB₂ polymorphism and the evidence that formation of T cells



requires this receptor.¹ Surprisingly, no evidence of AEA has been gathered so far in either mouse or human B cells.

2-AG in adaptive immunity. Of note, only two works of Rockwell et al. demonstrated directly the effect of 2-AG on T lymphocytes, where it induced a significant suppression of IL-2 expression⁴⁴ and such antiinflammatory effect was independent of CB receptors, but it was rather mediated by a COX-2 metabolite of 2-AG, probably by activating the PPAR- γ .⁴⁵ As for B lymphocytes, this bioactive lipid induced migration of B220+CD19+ B cells, preferentially by attracting unstimulated naïve B cells rather than activated and/ or class-switched germinal center B cells in a CB₂dependent manner.⁴⁶ It has been postulated that these effects might be indirect since they could involve other immune cells (such as T cells and macrophages) that are required for B-cell activation.

Conclusions

The majority of scientific studies on the immunoregulatory role of eCBs concentrated on whole immune cells, either on peripheral blood mononuclear cells or on mouse splenocytes. When investigating the specific immunoregulatory role of eCBs on each specific immune cell subset of both innate (Fig. 2) and adaptive (Fig. 3) immunity, most of the research has focused on monocytes/macrophages and T cells and mainly on AEA. Of note, AEA is the most potent anti-inflammatory eCB and it practically acts on all cell subsets of either innate or adaptive immunity (except for NK and B cells). Instead, 2-AG exerts both pro- and anti-inflammatory effects and it seems that its effects are strictly dependent on cell type. Although some specific and rarely represented immune cells (i.e., regulatory T cells, $\gamma \delta$ T cells, or MAIT cells) were never investigated, neither were the subpopulations of each innate immune cell type, it can be generally stated that eCBs, particularly AEA and PEA, could be considered as master regulators of the innateadaptive immune axis and as valuable immunoresolvents. Indeed, the fact that most of these molecules as well as several elements of their metabolism and signaling (i.e., enzymes and receptors) are dysregulated in many pathological states where the immune system is a crucial factor suggests that their exploitation in treating several chronic inflammatory diseases could just be around the corner, providing that their role will be also confirmed in vivo and that their underlying molecular mechanism elucidated.

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Author Disclosure Statement

No competing financial interests exist.

References

- Chiurchiù V, Battistini L, Maccarrone M. Endocannabinoid signaling in innate and adaptive immunity. Immunology 2015. [Epub ahead of print]; DOI: 10.1111/imm.12441.
- Fezza F, Bari M, Florio R, et al. Endocannabinoids, related compounds and their metabolic routes. Molecules 2014;19:17078–17106.
- Maccarrone M, Dainese E, Oddi S. Intracellular trafficking of anandamide: new concepts for signaling. Trends Biochem Sci. 2010;35:601–608.
- Chiurchiù V, Leuti A, Maccarrone M. Cannabinoid signaling and neuroinflammatory diseases: a melting pot for the regulation of brain immune responses. J Neuroimmune Pharmacol. 2015;10:268–280.
- Galve-Roperh I, Chiurchiù V, Díaz-Alonso J, et al. Cannabinoid receptor signaling in progenitor/stem cell proliferation and differentiation. Prog Lipid Res 2013;52:633–650.
- Pistis M, Melis M. From surface to nuclear receptors: the endocannabinoid family extends its assets. Curr Med Chem. 2010;17:1450–1467.
- Fidaleo M, Fanelli F, Ceru MP, et al. Neuroprotective properties of peroxisome proliferator-activated receptor alpha (PPARα) and its lipid ligands. Curr Med Chem. 2014;21:2803–2821.
- Pontis S, Ribeiro A, Sasso O, et al. Macrophage-derived lipid agonists of PPAR-α as intrinsic controllers of inflammation. Crit Rev Biochem Mol Biol. 2015:1–8. [Epub ahead of print].
- Chiurchiù V, Lanuti M, De Bardi M, et al. The differential characterization of GPR55 receptor in human peripheral blood reveals a distinctive expression in monocytes and NK cells and a proinflammatory role in these innate cells. Int Immunol. 2015;27:153–160.
- Alhouayek M, Muccioli GG. COX-2-derived endocannabinoid metabolites as novel inflammatory mediators. Trends Pharmacol Sci. 2014;35:284–292.
- 11. Ginhoux F, Jung S. Monocytes and macrophages: developmental pathways and tissue homeostasis. Nat Rev Immunol. 2014;14:392–404.
- Maresz K, Carrier EJ, Ponomarev ED, et al. Modulation of the cannabinoid CB2 receptor in microglial cells in response to inflammatory stimuli. J Neurochem. 2005;95:437–445.
- Chiurchiù V, Lanuti M, Catanzaro G, et al. Detailed characterization of the endocannabinoid system in human macrophages and foam cells, and anti-inflammatory role of type-2 cannabinoid receptor. Atherosclerosis 2014;233:55–63.
- 14. Merad M, Sathe P, Helft J, et al. The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. Annu Rev Immunol. 2013;31:563–604.
- Mócsai A. Diverse novel functions of neutrophils in immunity, inflammation, and beyond. J Exp Med. 2013;210:1283–1299.
- Vivier E, Raulet DH, Moretta A, et al. Innate or adaptive immunity? The example of natural killer cells. Science 2011;331:44–49.
- Voehringer D. Protective and pathological roles of mast cells and basophils. Nat Rev Immunol. 2013;13:362–375.
- Jacobsen EA, Helmers RA, Lee JJ, et al. The expanding role(s) of eosinophils in health and disease. Blood 2012;120:3882–3890.
- De Filippis D, Negro L, Vaia M, et al. New insights in mast cell modulation by palmitoylethanolamide. CNS Neurol Disord Drug Targets 2013;12:78–83.
- Cabral GA, Toney DM, Fischer-Stenger K, et al. Anandamide inhibits macrophage-mediated killing of tumor necrosis factor-sensitive cells. Life Sci. 1995;56:2065–2072.
- Hernangómez M, Mestre L, Correa FG, et al. CD200-CD200R1 interaction contributes to neuroprotective effects of anandamide on experimentally induced inflammation. Glia 2012;60:1437–1450.
- 22. Matias I, Pochard P, Orlando P, et al. Presence and regulation of the endocannabinoid system in human dendritic cells. Eur J Biochem. 2002;269:3771–3778.

- Do Y, McKallip RJ, Nagarkatti M, et al. Activation through cannabinoid receptors 1 and 2 on dendritic cells triggers NF-kappaB-dependent apoptosis: novel role for endogenous and exogenous cannabinoids in immunoregulation. J Immunol. 2004;173:2373–2382.
- Karmaus PW, Chen W, Crawford RB, et al. Deletion of cannabinoid receptors 1 and 2 exacerbates APC function to increase inflammation and cellular immunity during influenza infection. J Leukoc Biol. 2011;90:983–995.
- Ribeiro A, Ferraz-de-Paula V, Pinheiro ML, et al. Anandamide prior to sensitization increases cell-mediated immunity in mice. Int Immunopharmacol. 2010;10:431–439.
- Chiurchiù V, Cencioni MT, Bisicchia E, et al. Distinct modulation of human myeloid and plasmacytoid dendritic cells by anandamide in multiple sclerosis. Ann Neurol. 2013;73:626–636.
- McHugh D, Tanner C, Mechoulam R, et al. Inhibition of human neutrophil chemotaxis by endogenous cannabinoids and phytocannabinoids: evidence for a site distinct from CB1 and CB2. Mol Pharmacol. 2008;73:441–450.
- Sugawara K, Bíró T, Tsuruta D, et al. Endocannabinoids limit excessive mast cell maturation and activation in human skin. J Allergy Clin Immunol. 2012;129:726–738.e8.
- Sugawara K, Zákány N, Hundt T, et al. Cannabinoid receptor 1 controls human mucosal-type mast cell degranulation and maturation in situ. J Allergy Clin Immunol. 2013;132:182–193.
- Jackson AR, Hegde VL, Nagarkatti PS, et al. Characterization of endocannabinoid-mediated induction of myeloid-derived suppressor cells involving mast cells and MCP-1. J Leukoc Biol. 2014;95:609–619.
- Maestroni GJ. The endogenous cannabinoid 2-arachidonoyl glycerol as in vivo chemoattractant for dendritic cells and adjuvant for Th1 response to a soluble protein. FASEB J. 2004;18:1914–1916.
- Chouinard F, Lefebvre JS, Navarro P, et al. The endocannabinoid 2arachidonoyl-glycerol activates human neutrophils: critical role of its hydrolysis and de novo leukotriene B4 biosynthesis. J Immunol. 2011;186:3188–3196.
- 33. Kurihara R, Tohyama Y, Matsusaka S, et al. Effects of peripheral cannabinoid receptor ligands on motility and polarization in neutrophil-like HL60 cells and human neutrophils. J Biol Chem. 2006;281:12908–12918.
- Sugiura T, Oka S, Gokoh M, et al. New perspectives in the studies on endocannabinoid and cannabis: 2-arachidonoylglycerol as a possible novel mediator of inflammation. J Pharmacol Sci. 2004;96:367–375.
- Kishimoto S, Oka S, Gokoh M, et al. Chemotaxis of human peripheral blood eosinophils to 2-arachidonoylglycerol: comparison with other eosinophil chemoattractants. Int Arch Allergy Immunol. 2006;140:3–7.
- Larose MC, Turcotte C, Chouinard F, et al. Mechanisms of human eosinophil migration induced by the combination of IL-5 and the endocannabinoid 2-arachidonoyl-glycerol. J Allergy Clin Immunol. 2014;133:1480–1482.
- Bettoni I, Comelli F, Colombo A, et al. Non-neuronal cell modulation relieves neuropathic pain: efficacy of the endogenous lipid palmitoylethanolamide. CNS Neurol Disord Drug Targets 2013;12:34–44.
- Bluestone JA, Mackay CR, O'Shea JJ, et al. The functional plasticity of T cell subsets. Nat Rev Immunol. 2009;9:811–816.
- Cencioni MT, Chiurchiù V, Catanzaro G, et al. Anandamide suppresses proliferation and cytokine release from primary human T-lymphocytes mainly via CB2 receptors. PLoS One 2010;5:e8688.

- Pieper K, Grimbacher B, Eibel H. B-cell biology and development. J Allergy Clin Immunol. 2013;131:959–971.
- 41. Sancho R, Calzado MA, Di Marzo V, et al. Anandamide inhibits nuclear factor-kappaB activation through a cannabinoid receptor-independent pathway. Mol Pharmacol. 2003;63:429–438.
- Joseph J, Niggemann B, Zaenker KS, et al. Anandamide is an endogenous inhibitor for the migration of tumor cells and T lymphocytes. Cancer Immunol Immunother. 2004;53:723–728.
- Jackson AR, Nagarkatti P, Nagarkatti M. Anandamide attenuates Th-17 cell-mediated delayed-type hypersensitivity response by triggering IL-10 production and consequent microRNA induction. PLoS One 2014;9:e93954.
- Rockwell CE, Raman P, Kaplan BL, et al. A COX-2 metabolite of the endogenous cannabinoid, 2-arachidonyl glycerol, mediates suppression of IL-2 secretion in activated Jurkat T cells. Biochem Pharmacol. 2008;76:353–361.
- Rockwell CE, Snider NT, Thompson JT, et al. Interleukin-2 suppression by 2-arachidonyl glycerol is mediated through peroxisome proliferatoractivated receptor gamma independently of cannabinoid receptors 1 and 2. Mol Pharmacol. 2006;70:101–111.
- Tanikawa T, Kurohane K, Imai Y. Induction of preferential chemotaxis of unstimulated B-lymphocytes by 2-arachidonoylglycerol in immunized mice. Microbiol Immunol. 2007;51:1013–1019.

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Abbreviations Used

- 2-AG = 2-arachidonoylglycerol
- COX-2 = cyclooxygenase-2
- DC = dendritic cell
- eCB = endocannabinoid
- ECS = endocannabinoid system
- FAAH = fatty acid amide hydrolase ABDH4 = α/β -hydrolase 4
- LOX = lipoxygenase
- MAIT = mucosal-associated invariant T cells
- NAE = N-acylethanolamine
- NAPE-PLD = N-acyl-phosphatidylethanolamine-hydrolyzing
 - phospholipase D

Cannabis and Cannabinoid

Research

- NK = natural killer cells
- OEA = N-oleoylethanolamine
- PEA = N-palmitoylethanolamine
- PPAR = peroxisome proliferator-activated receptor
- TNF = tumor necrosis factor

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