



# Clinical considerations in early-onset cerebral amyloid angiopathy

Cerebral amyloid angiopathy (CAA) is an important cerebral small vessel disease associated with brain haemorrhage and cognitive change. The commonest form, sporadic amyloid- $\beta$  CAA, usually affects people in mid- to later life. However, early-onset forms, though uncommon, are increasingly recognized and may result from genetic or iatrogenic causes that warrant specific and focused investigation and management.

In this review, we firstly describe the causes of early-onset CAA, including monogenic causes of amyloid- $\beta$  CAA (APP missense mutations and copy number variants; mutations of PSEN1 and PSEN2) and non-amyloid- $\beta$  CAA (associated with ITM2B, CST3, GSN, PRNP and TTR mutations), and other unusual sporadic and acquired causes including the newly-recognized iatrogenic subtype. We then provide a structured approach for investigating early-onset CAA, and highlight important management considerations.

Improving awareness of these unusual forms of CAA amongst healthcare professionals is essential for facilitating their prompt diagnosis, and an understanding of their underlying pathophysiology may have implications for more common, late-onset, forms of the disease.

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#### Introduction

Descriptions of amyloid deposition within the cerebral vasculature have existed for more than a century<sup>1</sup>; these amyloids are fibrillary protein assemblies composed of stacks of monomers with a beta-

sheet structure. They demonstrate distinct ultra-structural properties and a characteristic cross- $\beta$  X-ray crystallographic diffraction pattern, as well as specific reactions to certain dyes, including Thioflavin T and Congo red.  $^2$  As of September 2020, the International Society of Amyloidosis has identified 36 amyloid

proteins that can cause human disease,<sup>2</sup> of which six can involve the cerebral vasculature, either exclusively or as part of a systemic amyloidosis. In recent years, however, cerebral amyloid angiopathy (CAA) has become near synonymous with the cerebrovascular deposition of one of these amyloids, amyloid-β, notable also for its frequent pathological presence in Alzheimer's disease. Sporadic amyloid-β CAA in this context refers to disease where a single explanatory cause has not been identified, and is usually age-related. As by far the commonest form of cerebrovascular amyloidosis,<sup>3</sup> sporadic amyloid-β CAA is primarily associated with brain haemorrhage (Fig. 1A and B) but also cognitive impairment, both of which can occur in association with, or independently of, Alzheimer's disease.4 Predominantly involving the small cortical and leptomeningeal vessels, amyloid- $\beta$  CAA can be identified in life by characteristic haemorrhagic structural imaging markers on bloodsensitive MRI sequences, namely cortical superficial siderosis and lobar cerebral microbleeds (Fig. 1C and D). These haemorrhagic imaging markers, together with more obviously symptomatic bleeding events, namely parenchymal intracerebral haemorrhage (ICH) and acute convexity subarachnoid haemorrhage, form the basis of the diagnostic Boston criteria. 7,8 Other non-haemorrhagic imaging features, such as cerebral atrophy, white matter hyperintensities and MRI visible perivascular spaces in the centrum semi-ovale (CSO-PVS), are also commonly observed in sporadic CAA<sup>8-10</sup>; whilst these are not necessarily specific for CAA, white matter hyperintensities in a 'multispot' pattern and CSO-PVS are included in the most recent Boston criteria (Version 2.0).8 There are also CT-based diagnostic criteria for CAA (the Edinburgh criteria<sup>11</sup>), but these require knowledge of APOE genotype in order to be applied fully; validation of using the imaging components alone is ongoing, but there is an early suggestion that they do have some diagnostic accuracy. 12-15

Pathologically, amyloid- $\beta$  can be immunohistochemically identified with specific antibodies, and descriptions include details of both which vessels (small arteries, arterioles, capillaries, venules, veins) and which locations (meningeal, cortical/parenchymal) and brain regions are involved.  $^{16}$  Classifications of pathological severity consider both the extent of amyloid deposition within the vessel, which begins in the abluminal layer of the tunica media and progresses to involve all layers of the vessel wall, and the degree of the associated vasculopathy, which can be characterized by fibrinoid necrosis, microaneurysm formation and concentric splitting of vessel wall ('double barrelling') at its most severe.  $^{17-19}$  Sporadic amyloid- $\beta$  CAA has a predominantly posterior distribution, with the occipital lobe most commonly affected.  $^{20}$ 

As MRI is more widely used for the clinical assessment of a range of neurological conditions, an increasing number of people are identified as having imaging features of CAA, which can be an incidental or unexpected finding. Although clinical context is essential for determining their importance, in younger patients presenting with ICH, cognitive symptoms or transient focal neurological episodes, the presence of these markers might indicate certain rare but important causes of CAA that warrant specific and focused investigation with implications for patients and their families. Young or early-onset in this context is often considered to be before the age of 50 years, when sporadic amyloid-β CAA would be unusual or unexpected,8 although strict age criteria are necessarily arbitrary. The amyloid-β protein has a central and defining role in both CAA and Alzheimer's disease, a role underpinned by the identification of mutations in genes involved in amyloid-β production.  $^{21-23}$  However, the existence of other, non-amyloid- $\beta$  forms of CAA also has important mechanistic implications. In this review,

our aim is firstly to describe the causes of early-onset CAA (summarized in Fig. 2), including rare monogenic causes and other unusual types of CAA that can affect younger people. Our second aim is to suggest a structured approach to the investigation and management of people who meet the diagnostic clinicoradiological criteria for CAA, and present with early-onset disease.

CAA can be mimicked by other conditions, and in young onset cases it is important to consider other monogenic disorders, including those associated with familial cerebral cavernous malformations (which can resemble cerebral microbleeds on imaging) and those which can cause ICH in the context of other (non-CAA) cerebral small vessel diseases, including COL4A1/COL4A2 haemorrhagic microangiopathy and CADASIL (cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy). 24,25 Other, non-genetic, diagnoses to consider include reversible cerebral vasoconstriction syndrome (RCVS), cerebral venous thrombosis, vascular malformations (e.g. dural arteriovenous fistulas), posterior reversible encephalopathy syndrome (PRES) and infective endocarditis<sup>26</sup>; these can all result in ICH, acute convexity subarachnoid haemorrhage and cerebral microbleeds. Cerebral microbleeds can also occur following cardiac procedures, 27-29 head trauma<sup>30,31</sup> and critical illness, for example due to severe COVID-19 infection. 32-34 Although none of these conditions typically mimic the strictly lobar microbleed distribution characteristic of CAA, in practice this distinction may not be clear-cut.

# Monogenic forms of cerebral amyloid angiopathy

#### Amyloid-β cerebral amyloid angiopathy

All monogenic forms of CAA are rare, although definitive data on their prevalence are lacking. Mutations relevant to amyloid-β CAA can be considered in two broad categories, on the basis of dominant clinical and pathological phenotype. The first are those where pathological vascular amyloid-β deposition and/or presentation with ICH are the dominant feature(s); these mutations nearly exclusively involve the amyloid-β coding domains of the amyloid precursor protein gene (APP). Mutations of APP outside of the amyloid-β coding domain that cause familial Alzheimer's disease can show significant pathological evidence of CAA, but the phenotype is cognitive rather than haemorrhagic; this includes London NM\_000484.4(APP):c.2149G>A (p.Val717Ile) mutation,<sup>35</sup> the Indiana NM\_000484.4(APP):c.2149G>A (p.Val717Phe) mutation<sup>36</sup> and Swedish NM\_000484.4(APP):c.2010\_2011inv (p.Lys670\_Met671delinsAsnLeu) double mutation. 37,38 The second group are mutations primarily associated with familial Alzheimer's disease, where CAA can be a significant feature. Of these, copy number variants of APP (including trisomy 21) are most frequently associated with clinical and radiological haemorrhagic features typical for sporadic CAA; mutations of PSEN1 and PSEN2 will also be considered. It is important to recognize that these groups are not necessarily distinct and often overlap; this clinical heterogeneity can result in different presentations (haemorrhagic, cognitive or both) even within a single family carrying a particular mutation.

In monogenic forms of CAA, the genetic mutation or duplication plays a causal role in the disease. Genetic variants that confer increased risk are also likely to make important contributions to early-onset CAA. This has been less studied in CAA compared with Alzheimer's disease, <sup>39,40</sup> but one well recognized risk gene is APOE. APOE encodes apolipoprotein E (ApoE), a glycoprotein with

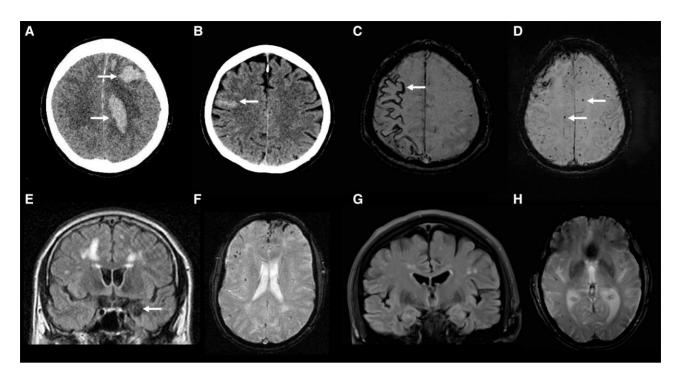


Figure 1 Imaging features of amyloid-β CAA. (A-D) Examples of brain imaging from individuals with sporadic amyloid-β CAA. CT images demonstrating (A) acute ICH and (B) acute convexity subarachnoid haemorrhage. Susceptibility-weighted MRI images demonstrating (C) disseminated cortical superficial siderosis of the right cerebral hemisphere and (D) lobar (cortical) cerebral microbleeds. B and C reprinted and adapted from Banerjee et al.4 (open access article distributed under the terms of the Creative Commons CC BY 4.0 license). (E-H) MRI from individuals with features of CAA accompanying familial Alzheimer's disease. An old amygdala haemorrhage (arrow) and extensive white matter hyperintensities on T2 FLAIR (E), with multiple microbleeds on T2\* imaging (F) in an APP duplication carrier. White matter hyperintensities on T2 FLAIR (G) and widespread lobar microbleeds on susceptibility-weighted imaging (H) in an individual with the PSEN1 R269H mutation. E and F reprinted and adapted from McNaughton et al. 5 G and H reprinted and adapted from Ryan et al.<sup>6</sup> by permission of Oxford University Press.

an important role in peripheral cholesterol metabolism, which also acts as the main lipid and cholesterol transporter within the CNS. 41 The commonest isoform of ApoE is ApoE3; carrying the APOEc4 genotype (encoding the ApoE4 isoform) is a major risk factor for Alzheimer's disease. 41 In CAA, associations with both APOE €2 and APOE $\epsilon$ 4 genotypes have been described, 39 with evidence most convincing for the latter<sup>42</sup>; both these risk alleles can be associated with earlier age of first ICH in CAA. 43,44 The relevance of other genetic risk factors identified in studies of Alzheimer's disease remains an area for active study; of these, there are initial data suggesting that ABCA7, CLU and CR1 might be of relevance to CAA45-48 and lobar ICH.49

#### Mutations with cerebral amyloid angiopathy as the dominant clinical or pathological phenotype

To date, all identified mutations that primarily result in amyloid-β CAA involve the amyloid precursor protein gene (APP); APP mutations associated with severe CAA are summarized in Table 1. There are six mutations with confirmed pathogenicity (Fig. 3) and several of uncertain pathogenicity, described in detail below. With the exception of the Dutch-type CAA, only a small number of cases have been reported for each mutation and therefore the full spectrum of presentations might be broader than that currently

The amyloid precursor protein can undergo cleavage via two alternative pathways, which are in competition with one another<sup>87</sup>: the first involves cleavage by  $\alpha$ -secretase, and does not result in amyloid-β production; the second requires sequential cleavage by BACE1 (β-site amyloid precursor protein cleaving enzyme 1, also known as  $\beta$ -secretase) and gamma-secretase enzymes, which produces amyloid- $\beta$  peptides.  $\gamma$ -Secretase, which contains either presenilin 1 or 2 as its catalytic subunit, carries out a series of successive cleavages generating shorter amyloid-β peptides. The amyloid-β peptides released can be between 37 and 49 amino acids in length but under physiological conditions the 40 amino acid (amyloid- $\beta_{1-40}$ , normally 80% to 90% of the amyloid- $\beta$  peptides) and 42 amino acid (amyloid- $\beta_{1-42}$ , up to 10%) fragments are most common.<sup>87</sup> The coding region for amyloid- $\beta$  within the APP gene can start at either codon 672 or codon 682, depending on BACE1 cleavage, which can occur at either site, and ends variably between codon 709 and 720 depending on  $\gamma$ -secretase cleavage (Fig. 3).87 Whilst APP mutations around the  $\gamma$ -secretase cleavage site typically cause familial Alzheimer's disease presenting with memory impairment, mutations that occur within the amyloid-β coding domain give rise to severe CAA, which can manifest with ICH, dementia or both.

#### Dutch mutation; NM\_000484.4(APP):c.2077G>C (p.Glu693Gln)

Dutch-type CAA (D-CAA; previously called hereditary cerebral haemorrhage with amyloidosis-Dutch type, HCHWA-D) is arguably the archetypal monogenic form of amyloid-β CAA, and remains the most studied clinically.<sup>22</sup> The clinical findings were first described in 1964 in two families originally from the seaside villages of Katwijk and Scheveningen in The Netherlands. 22,88 Although most people with the mutation still live in The Netherlands, there is a kindred now based in Western Australia, who are descendants from a branch of the family who emigrated from Katwijk some

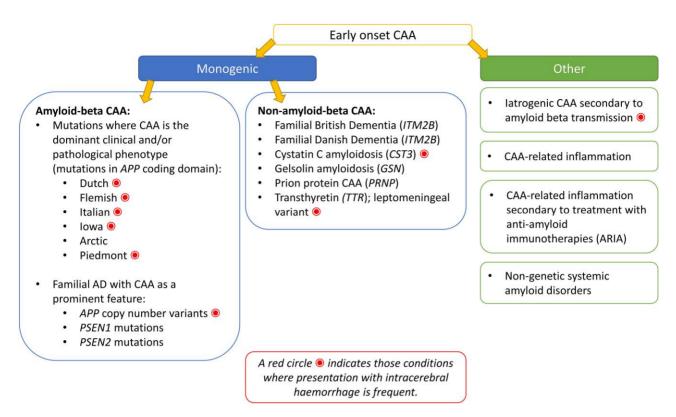


Figure 2 Potential causes of early-onset CAA. AD = Alzheimer's disease; ARIA = amyloid-related imaging abnormalities; CAA = cerebral amyloid angiopathy.

years previously.<sup>89</sup> D-CAA classically presents with recurrent ICH, 90,91 although migraine with aura can also be a presenting feature, 50 as noted in some of the earliest case descriptions. 51 ICH can be followed by seizures (observed in around half of ICH survivors), cognitive decline and dementia, although cases of cognitive impairment in the absence of ICH have also been described. 22,90,92,93 Pathologically, there is severe CAA with a relative paucity of neuritic plaques and neurofibrillary tangles rarely seen, although diffuse amyloid-β deposits may be found. 94,95

All haemorrhagic and non-haemorrhagic structural imaging markers observed in sporadic age-related amyloid-β CAA have been reported in symptomatic mutation carriers, in addition to a number of novel neuroimaging biomarkers (Table 1). Data from presymptomatic mutation carriers has provided important insights into possible biomarkers for early disease, including decreased levels of CSF<sup>62</sup> and plasma<sup>96</sup> amyloid- $\beta_{1-40}$  and amyloid- $\beta_{1-42}$ , increased retention of the PET agent Pittsburgh compound B (PiB) on amyloid imaging,61 reductions in occipital vascular reactivity when presented with visual stimuli, 58 non-haemorrhagic imaging markers (white matter hyperintensities, CSO-PVS and microinfarcts)52,97 and retinal changes on optical coherence tomography (OCT).64

#### Flemish mutation; NM\_000484.4(APP):c.2075C>G (p.Ala692Gly)

This mutation was first described in 1992 in a Dutch family, and of the six patients described, four presented with dementia and two with ICH.<sup>65</sup> Further review of this family<sup>66,98</sup> has confirmed that of the 20 suspected symptomatic mutation carriers, five presented with ICH and a further two family members had strokes (unspecified) during the course of their disease. The mutation was later described in a second unrelated British family, 67 in which six of seven

patients presented with dementia and one with ICH. This mutation has also been described in a French man of Portuguese descent.<sup>68</sup> MRI studies have shown extensive white matter hyperintensities, which are present to a lesser extent (but more than expected for age) in pre-symptomatic mutation carriers<sup>66,67,69</sup>; multiple cortical and juxtacortical microbleeds, cortical superficial siderosis and atrophy (cortical and hippocampal) have also been reported.<sup>68</sup> The pathology is characterized by severe CAA and 'dense-core' plaques, which are frequently centred around vessels and predominantly composed of amyloid- $\beta_{1-40}$ ; diffuse amyloid- $\beta$  deposits and neurofibrillary tangles are also observed. 98,99

#### Italian mutation; NM\_000484.4(APP):c.2077G>A (p.Glu693Lys)

Twenty symptomatic individuals from four families from Lombardy, Italy, with this mutation have been described, as well as a brother and sister investigated in France, 68 in which affected individuals can present with strokes (haemorrhagic and ischaemic) and progressive cognitive impairment, as well as headaches and seizures (which in some cases appear secondary to the incident haemorrhage).<sup>70</sup> MRI can show white matter hyperintensities and cerebral microbleeds, and cortical occipital calcifications.<sup>68,70</sup> Pathologically there is abundant CAA, and diffuse parenchymal deposition of amyloid-β, but no neuritic plaques or neurofibrillary tangles.70

#### Arctic mutation; NM\_000484.4(APP):c.2078A>G (p.Glu693Gly)

This mutation has been reported in a single family, originally from northern Sweden (hence 'Arctic', allowing differentiation from a different APP mutation known as the 'Swedish', which is located outside the amyloid- $\beta$  coding domain adjacent the  $\beta$ -secretase cleavage site). 37 The Arctic mutation causes progressive amnestic

Table 1 Pathogenic missense mutations in the APP gene associated with severe CAA

Name	Mutation	Αβ		Pathology			Clinical pı	Clinical presentation		Biomarkers	
		pepude	CAA	Amyloid plaques	Tau	Typical presentation and AAO, y	ICH, including cSAH	Dementia	Other	Imaging	Other
Confirmed p	Confirmed pathogenicity; NM_000484.4(APP) Dutch c.2077G>C + (p.Glu693Gln)		(dat\)	1	1	Recurrent ICH from ~50 (range 39-76) followed or preceded by cognitive impairment. <sup>22</sup>	+	+	Migraine with aura (can manifest 8-9 y prior to first ICH), <sup>50,51</sup> seizures.		Reduced CSF Aβ <sub>1-40</sub> and Aβ <sub>1-42</sub> . <sup>61,62</sup> Reduced plasma Aβ <sub>1-42</sub> . <sup>63</sup> Retinal changes on OCT. <sup>64</sup>
Flemish	c.2075G>G (p.Ala692Gly)	A21G	+	+	+	ICH, dementia or both. Mid-40s (range 35– 61) <sup>22,65-68</sup>	+	+	Seizures	PIB-PET positivity. **  Extensive WMH on MRI; WMH in pre-symptomatic mutation carriers. **  **cSS, microbleeds, cortical and hippocampal atrophy. **  Parieto-temporal hypometabolism on FPG-PET **	Intermediate CSF Aβ <sub>1-4-2</sub> ; normal CSF total-tau and phospho-tau. <sup>68</sup>
Italian	c.2077G>A (p.Glu693Lys)	E22K	+	I	1	Recurrent ICH mid-50s (range 44–63) <sup>68,70</sup>	+	+	Headache, seizures	WMH, cSS, microbleeds, occipital calcifications.	Reduced CSF A\(\beta_{1-42}\), increased total and
Arctic	c.2078A>G (p.Glu693Gly)	E22G	+	+	+	Memory-led cognitive impairment, 50s (range 52-62) <sup>71-73</sup>	T	+	Headache, fatigue, psychiatric symptoms, myoclonus, rigidity.	MRI findings variable. 73-75  WMH in some cases, usually mild-to-moderate, extensive in one case. Haemorrhagic imaging markers have not been reported to date.  Cortical atrophy (left precuneus, superior temporal and fusiform gyri). 40  Pil-PFT normal; reduced glucose	Purospino-tau. Reductions in CSF Aβ <sub>1-42</sub> , elevations in CSF total tau and phospho-tau. <sup>74,76</sup> Reduced plasma Aβ <sub>1-40</sub> and 1 Aβ <sub>1-42</sub> , also in pre-symptomatic carriers. <sup>72</sup>
Iowa	c.2080G>A (p.Asp694Asn)	D23N	+	+	+	ICH, dementia or both. Early 50s (range 38– 67). <sup>68,77-82</sup>	+	+	Seizures, expressive language dysfunction, personality change, myoclonus, gait	unetacousm (t.D.G.T.E.1). WMH, microbleeds, cortical calcifications. <sup>77-81,83</sup>	
Piedmont	c.2113C>G (p.Leu705Val)	L34V	+	ı	I	Recurrent ICH (range 45–72). ***85	+	ı	TFNE	Diffuse WMH. <sup>84</sup> Lobar microbleeds and cSS on <i>ex</i> vivo MRI. <sup>85</sup>	

siderosis; DTI = diffusion tensor imaging; ICH = intracerebral haemorrhage; OCT = optical coherence tomography; PiB = Pittsburgh Compound B; rCBF = regional cerebral blood flow; TFNE = transient focal neurological episodes ('amyloid spells'); WMH = white matter hyperintensities; y = years. A $\beta$  = amyloid- $\beta$ ; AAO = age at onset; CAA = cerebral amyloid angiopathy; cSAH = convexity subarachnoid haemorrhage; CSO-PVS = MRI visible perivascular spaces in the centrum semi-ovale; cSS = cortical superficial

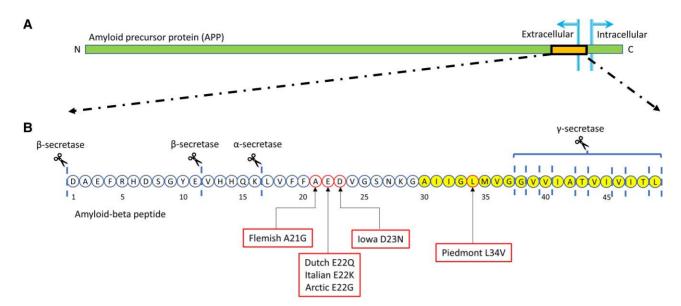


Figure 3 Schematics of the amyloid precursor protein (APP) and amyloid- $\beta$  peptide. (A) Schematic of the amyloid precursor protein (APP). APP is a transmembrane glycoprotein that can exist in multiple isoforms of differing lengths, the most common in the brain being 695 amino acids in length (APP695). This schematic of APP demonstrates its larger extracellular N-terminal end and smaller intracellular C-terminal end (both green). The amyloid- $\beta$  peptide (orange) starts in the extracellular domain and enters the transmembrane domain. (B) Schematic of the amyloid- $\beta$  peptide and its amino acid sequence. The  $\alpha$ -secretase cleavage site, in addition to those for  $\beta$ -secretase (two potential sites) and  $\gamma$ -secretase (multiple sites, resulting in amyloid- $\beta$  fragments between 37 and 49 amino acids in length) are shown. Amino acids coloured yellow fall within the transmembrane section of APP. Mutations affecting the amino acids outlined in red have confirmed pathogenicity and are particularly associated with CAA.

cognitive impairment.  $^{71,72}$  Pathologically, there is severe CAA, together with distinctive amyloid- $\beta$  plaques (lacking a dense core and described as 'ring-like' or 'targetoid') and neurofibrillary tangles.  $^{73,100,101}$  Intriguingly, very low PiB retention was observed on amyloid PET imaging of two Arctic mutation carriers, despite clearly pathological levels of CSF amyloid- $\beta$ , total tau and phosphotau  $^{74}$ ; given that PiB binds fibrillary amyloid,  $^{102}$  this highlights the potential importance of non-fibrillar forms of amyloid- $\beta$  in the disease process associated with this particular mutation.  $^{74}$ 

#### Iowa mutation; NM\_000484.4(APP):c.2080G>A (p.Asp694Asn)

First described in a family from Iowa of German descent, 77 this mutation has subsequently been described in a French family of Spanish descent, 78,83 and later in families of Austro-Hungarian, 68 Irish, <sup>79</sup> Polish <sup>80</sup> and Spanish (Basque) <sup>81</sup> descent. The clinical presentation can be either with ICH (as observed in the Austro-Hungarian, French-Spanish, Irish, Polish and Spanish-Basque kindreds), non-aneurysmal subarachnoid haemorrhage (Austro-Hungarian) or dementia (Austro-Hungarian, Iowa and Irish kindreds); other features include seizures, a short-stepped gait, expressive language dysfunction, personality change, and extracerebral vascular changes (external carotid artery dysplasia, thickening of basement membranes in skin capillaries). MRI can show white matter changes (which can be severe), cerebral microbleeds, and posterior 'tram-line' or gyriform cortical calcifications. 77-81,83 Pathologically, there is evidence of severe CAA, together with diffuse plaques and neurofibrillary tangles of varying severity. 77,79,81

#### Piedmont mutation; NM\_000484.4(APP):c.2113C>G (p.Leu705Val)

This was first described in an Italian family with affected members presenting with recurrent ICH. A subsequent, and apparently unrelated, family has since been described, although confirmatory genetic testing has only been completed in a single case. The clinical presentation is with ICH (cognitive impairment is described in

one case, but in the context of multiple ICH). Diffuse white matter hyperintensities have been reported<sup>84</sup>; lobar microbleeds and cortical superficial siderosis have also been described on *ex vivo* MRI.<sup>85</sup> The pathological findings demonstrate CAA without parenchymal plaques or tau pathology.<sup>84,85</sup>

#### Mutations of uncertain pathogenicity

The 'Greek' mutation, NM 000484.4(APP):c.2062T>G (p.Leu688Val), has been reported in two Greek families, in which individuals had been diagnosed clinically with Alzheimer's disease and vascular dementia. 103,104 The presentation is with mood and gait disturbances, dementia and ischaemic strokes. Although the MRI findings were similar to those observed in another monogenic cerebral small vessel disease, CADASIL, with extensive white matter hyperintensities with involvement of the anterior temporal lobes, no relevant mutations in the causative NOTCH3 gene were found. 103 Cerebral microbleeds and occipital calcifications were also observed on MRI. 103,104 The APP mutation identified is considered to be pathogenic and the CSF findings are also supportive of amyloid-β disease. The neuropathological findings associated with this mutation have not yet been reported, and will provide important confirmatory evidence for establishing whether this mutation is causative in these cases.

The NM\_000484.4(APP):c.2137G>A (p.Ala713Thr) mutation has also been described in the context of Alzheimer's disease, <sup>105-108</sup> but remains of uncertain significance as its frequency in the population suggests limited penetrance. <sup>109</sup> A single case with pathological evidence of severe CAA has been described <sup>108</sup>; this patient had a peri-procedural brain haemorrhage at the time of brain biopsy.

Three mutations in the 3' untranslated region (UTR) of the APP gene have been described, 110 one of which is potentially of pathogenic relevance (c.\*331\_\*332del); this patient had early onset CAA characterized by recurrent ICH, with presentation at 39 years.

However, this patient also had a history of childhood neurosurgery for spina bifida, and therefore iatrogenic CAA might be an alternative explanation for their presentation; the latency between surgery and clinical presentation would be in keeping with this latter hypothesis. 111

# Mutations resulting in familial Alzheimer's disease, in which cerebral amyloid angiopathy can be a prominent feature

#### APP copy number variants, including duplications and triplications

APP duplications are associated with early onset Alzheimer's disease and prominent CAA, 5,112-118 with ICH described in approximately one-third of cases. 119 Seizures are also commonly observed. 120 White matter hyperintensities and cerebral microbleeds have been reported in some of these patients.<sup>5,53,114,121</sup> In trisomy 21 (Down syndrome), where people carry three copies of the APP gene (given its location on chromosome 21), there is an increased risk of Alzheimer's disease, which also occurs at younger ages. 122 People with trisomy 21 show an increased prevalence of ICH compared with the general population, but not at the levels observed in APP duplications. 119,123 Pathologically (Fig. 4), moderate-to-severe CAA is frequently, but not always, observed, 119,124 and MRI features of CAA (white matter changes, cerebral microbleeds, MRI-visible perivascular spaces) have also been reported 122,125 (Fig.1E and F). More recently, two related cases of APP triplication have been described, 126 both of whom presented with early onset dementia; in one of these cases, seizures and recurrent ICH developed during the disease course. As with APP duplications, white matter hyperintensities and cerebral microbleeds were observed; in one case CSF findings were supportive of cerebral amyloid- $\beta$  deposition (low CSF amyloid- $\beta_{1-42}$ , elevated total-tau and phospho-tau), and brain biopsy of the other confirmed significant cerebral vascular amyloid- $\beta$  deposition.

#### **PSEN1** mutations

Presenilin-1, encoded by the PSEN1 gene, is an important component of the  $\gamma$ -secretase enzymatic complex (providing its catalytic site)<sup>127</sup>; mutations in this gene remain the commonest cause of autosomal dominantly inherited familial Alzheimer's disease, with over 300 mutations identified to date. <sup>128</sup> Clinical presentation is variable, and can include features such as seizures, myoclonus, spastic paraparesis, extrapyramidal and cerebellar signs. 23,129,130 Whilst individuals typically present with memory symptoms, nonamnestic cognitive presentations may also occur, for example behavioural or dysexecutive syndromes, or language impairment. 131,132 Age at onset is typically in the mid-40s, but can range from the third to eighth decade of life. 133 The presence of severe CAA on neuropathological examination has been described<sup>23,134,135</sup> in a number of PSEN1 mutations and is particularly, although not exclusively, associated with mutations beyond codon 200; an example of the neuropathological findings is shown in Fig. 4. However, despite these pathological findings, the occurrence of ICH is unusual, and has only been described in two families, who carry the NM\_000021.4(PSEN1):c.49976C>T (p.Pro264Leu) and NM\_000021.4(PSEN1):c.857T>C (p.Leu286Pro) mutations. 136,137 White matter hyperintensities on MRI are well recognized, 138 particularly in association with mutations beyond codon 200,69 and cerebral microbleeds have been reported in some cases<sup>6,53,139,140</sup> (Fig. 1G and H). As in sporadic disease, cortical and hippocampal atrophy are characteristic radiological features of familial Alzheimer's disease, although brain volumes may sometimes appear normal to visual inspection in the early stages of the disease. <sup>141</sup> Rates of atrophy, however, are pathologically increased several years prior to symptom onset, so serial MRI may be helpful in cases of diagnostic uncertainty. <sup>142</sup>

#### PSEN2 mutations

The PSEN2 gene encodes the presenilin-2 protein, a homologue of presenilin-1 that can also form the catalytic part of the γ-secretase enzymatic complex.<sup>127</sup> Mutations in this gene most commonly result in familial Alzheimer's disease with a phenotype that is typically amnestic but can include psychosis and seizures. Severe CAA with ICH has been reported in two people carrying the NM\_000447.3(PSEN2):c.422A>T (p.Asn141Ile) mutation<sup>143,144</sup>, this mutation, first identified in families of Volga German descent, accounts for the majority of reported PSEN2 cases.<sup>145</sup> PSEN2 mutations are much rarer than mutations in PSEN1 and APP, and typically present at a later age, on average in the mid-50s, but which can vary widely within families and be as late as the early 80 s.<sup>133,145</sup> Mutations in this gene should therefore be considered in individuals with early-onset CAA with a family history of late onset Alzheimer's disease.

#### Non-amyloid-β cerebral amyloid angiopathy

As noted in the 'Introduction' section, although CAA more recently has become near synonymous with amyloid- $\beta$  CAA, its definition describes any condition in which proteins with biochemical features of amyloid are deposited within the cerebral vasculature. Of the six amyloid proteins that can involve the CNS, four cause localized disease and two can cause CNS disease as part of systemic amyloidosis²; these are summarized in Table 2. As with monogenic forms of amyloid- $\beta$  CAA, obtaining recent and accurate figures for prevalence is challenging, but these are generally accepted to be very rare conditions. The number of reported cases is small and investigation in some cases is limited, and so current descriptions might not reflect the true extent of the clinical phenotype.

#### ITM2B: familial British and familial Danish dementias

Different mutations in the integral membrane protein 2B (ITM2B; previously called BRI2) gene result in ABri and ADan amyloidoses, associated with Familial British and Familial Danish Dementia (FBD and FDD), respectively. In both cases, the mutations result in an elongated version of the precursor protein (BRI2), and subsequent cleavage by furin and furin-like endoproteases results in the production of the pathogenic amyloid fibril (ABri and ADan). 146-148

FBD was first described by Worster-Drought in 1933, and is characterized by a progressive dementia, spasticity and cerebellar ataxia. 149 It is caused by a mutation at codon 267, NM\_021999.5 (ITM2B): c.799T>A (p.Ter267Arg), which extends the BRI2 protein to 277 amino acids. 150 Extracranial systemic vascular ABri deposition also occurs, and can involve the pancreas, adrenal glands, lungs, myocardium, liver, spleen and skeletal muscle. 151 'Stroke-like episodes' are a recognized feature (approximately a quarter of historical cases) $^{152,153}$  but their aetiology remains unclear, and in particular, whether they resemble the transient focal neurological episodes (TFNE) or 'amyloid spells' observed in sporadic amyloid-β CAA; this has been hypothesized to be the case in at least one patient. 154 ICH has been reported in two patients, 152,154 one of which primarily involved the anterior putamen and globus pallidus (i.e. non-cortical areas). 152 Two further cases of intracranial bleeding were due to subdural haemorrhage and intracranial saccular 3998 | BRAIN 2023: 146; 3991–4014 G. Banerjee et al.

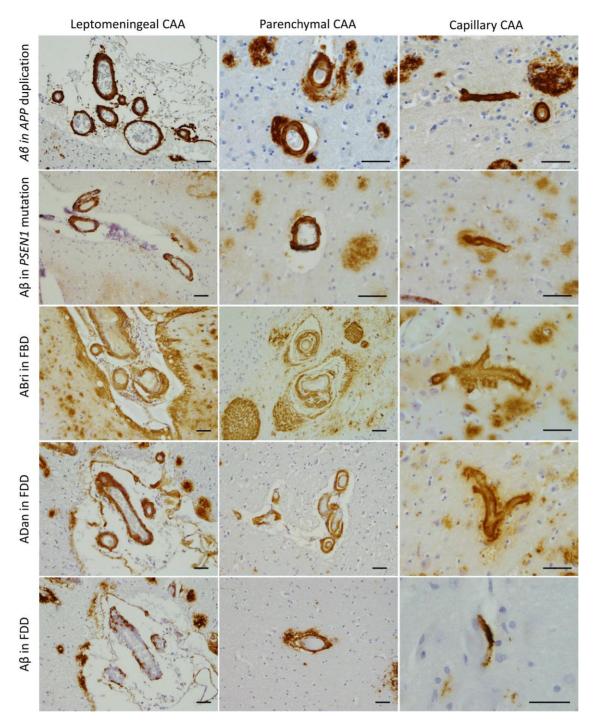


Figure 4 Immunohistochemical staining of early-onset cerebral amyloidosis. Amyloid- $\beta$  (A $\beta$ ) immunohistochemistry in an APP duplication and PSEN1 post-codon mutation case showing severe cerebral amyloid angiopathy (CAA) in both leptomeningeal and parenchymal blood vessels; there is also evidence of amyloid- $\beta$  deposition in the capillaries. In familial British dementia (FBD) the amyloidogenic protein ABri is deposited in leptomeningeal and parenchymal blood vessels, as well as capillaries. In familial Danish dementia (FDD), both ADan and amyloid- $\beta$  are found deposited as amyloid. ADan and amyloid- $\beta$  are found within the same blood vessels shown in the leptomeningeal vessels on sequential sections. ADan and amyloid- $\beta$  are also found in parenchymal vessels and capillaries. Scale bars = 50  $\mu$ m.

aneurysm rupture, and details for a third potential intracranial haemorrhage are unconfirmed. <sup>152,153</sup> MRI brain findings include white matter hyperintensities, which can be extensive and involve the corpus callosum, the presence of lacunar infarcts and cerebral microbleeds. <sup>152-154</sup> Pathological observations (Fig. 4) include vascular amyloid deposition in the brain, spinal cord and leptomeninges, parenchymal (most frequently involving the hippocampus and

cerebellum) and perivascular plaques of varying morphology, and diffuse non-fibrillar parenchymal deposition, as well as neurofibrillary tangles (particularly involving limbic areas) and ischaemic white matter changes.  $^{146,152,153,155}$ 

FDD, originally called heredopathia ophthalmo-oto-encephalica, was first described by Strömgren and colleagues in 1970,<sup>156</sup> and is characterized clinically by cataracts, deafness, progressive ataxia

Table 2 Causes of non-amyloid-β CAA

Name	Gene		Protein		Clinical features	Imaging
		Fibril	Precursor	Those shared with sporadic Aβ CAA	Others	
Familial British Dementia	ITM2B	ABri	ABriPP	'Stroke-like episodes' (possibly TFNE), ICH (rare), dementia	Spasticity, cerebellar ataxia	Extensive white matter hyperintensities Lacunar infarcts Cerebral mirrohleeds
Familial Danish Dementia Heredonathia onhthalmo-oto-encenhalica	ITM2B	ADan	ADanPP	Dementia	Cataracts, deafness, progressive ataxia, paranoid nswrhosis (as nart of dementia sundrome)	White matter
Cystatin C amyloidosis 'Icelandic type', HCHWA-1, HCCAA	CST3	ACys	Cystatin C	Recurrent ICH, progressive cognitive impairment	None	White matter changes
Gelsolin amyloidosis 'Finnish type', FAF, HGA, Meretoja syndrome	GSN	AGel	Gelsolin	None	Comeal lattice dystrophy, progressive cranial neuropathy, peripheral neuropathy (predominantly sensory), cutis laxa, myokymia, autonomic dysfunction onit attaxia cardiac and renal disease	White matter changes Cerebral microbleeds
PrP-CAA	PRNP	APrP	PrP	Dementia (Y145X, Q160X, Y163X)	Diarrhoea, autonomic dysfunction, peripheral neuropathy (Q160X, Y163X)	Non-specific white matter changes
Leptomeningeal variants of hereditary transthyretin ATTR amyloidosis De novo leptomeningeal involvement following liver transplantation in other variants of hereditary TTR amyloidosis	TTR	ATTR	Transthyretin	ICH, SAH (can be recurrent), TFNE, progressive cognitive impairment	Headache, ataxia, myelopathy, seizures, psychosis.	Superficial siderosis (supra- and infratentorial) Cerebral microbleeds Meningeal

Aβ = amyloid-β; CAA = cerebral amyloid angiopathy; CMB = cerebral microbleeds; FAF = familial amyloidosis of Finnish type; HCCAA = hereditary cystatin C amyloid angiopathy; HCHWA-1 = hereditary cerebral haemorrhage with amyloidosis-Icelandic type; HCA = hereditary gelsolin amyloidosis; ICH = intracerebral haemorrhage, PP = precursor protein; PPP = prion protein; SAH = subarachnoid haemorrhage; TFNE = transient focal neurological episodes.

and dementia, with paranoid psychosis a common feature. 157 It is caused by a decamer duplication of the nucleotides 786 and 795, NM\_021999.5(ITM2B):c.787\_796dup (p.Ser266fs), thereby shifting the reading frame and increasing the BRI2 length to 277 amino acids (the same length observed in FBD, although the C-terminal amino acid sequence is different). 157 Whilst ICH has not been reported, there has been a recorded death due to cerebrovascular disease, 156 and ischaemic stroke has also been reported. 158 Brain MRI can show white matter hyperintensities; evidence of cSS or cerebral microbleeds have not been reported. The neuropathological findings (Fig. 4) include widespread amyloid angiopathy of the neocortex, choroid plexus, cerebellum, spinal cord and retina, as well as parenchymal and leptomeningeal deposits and extensive tau pathology. 146,158 The neocortex and retina are more severely affected pathologically in FDD than FBD; another important difference is the presence of parenchymal and vascular amyloid-β in FDD that is not present in FBD. The amyloid- $\beta$  can be found in association with ADan deposits as well as independently. 158

More recently, a new mutation in ITM2B, NM\_021999.5(ITM2B): c.800G>T (p.\*267Leuext\*11), associated with dementia, ataxia, deafness and paraplegia has been described (familial Chinese dementia). 159 This mutation also results in an abnormal extension of the BRI2 protein. Brain MRI shows white matter hyperintensities but no cerebral microbleeds; confirmation of any neuropathological findings and the nature of the amyloidogenic protein is pending.

#### Cystatin C amyloidosis

A hereditary disease affecting certain families in Iceland and causing fatal ICH in young people was first described in 1935; the association with cerebrovascular amyloid deposition was made in 1972, with the causative protein and then gene mutation in the CST3 gene, NM\_000099.4(CST3):c.281T>A (p.Leu94Gln), identified in the early and mid-1980s, respectively. 160-162 This condition is now recognized as hereditary cystatin C amyloid angiopathy (HCCAA), also called hereditary cerebral haemorrhage with amyloidosis-Icelandic type (HCHWA-I); cystatin C is an inhibitor of extracellular cysteine proteinases and is found in all body fluids. 163 This presents with ICH in the 20s, with first ICH nearly always before the age of 40,164 and progressive neurological sequelae as a consequence of multiple strokes<sup>165</sup>; average life expectancy is ~30 years. 166,167 Pathologically there is significant ACys deposition within the small arteries and arterioles of the leptomeninges and brain; deposits are also found within the skin, lymph nodes, spleen, salivary glands and testes. 160,168,169 White matter changes 170 have been reported; it is not clear whether structural haemorrhagic features (cerebral microbleeds, cortical superficial siderosis) are also present.

#### Gelsolin amyloidosis

AGel or gelsolin amyloidosis, also referred to as familial amyloidosis of Finnish type (FAF), hereditary gelsolin amyloidosis (HGA) and Meretoja syndrome (after Jouko Meretoja, who in 1969 first described the syndrome in three Finnish families 171,172), occurs due to mutations in the GSN gene. GSN encodes the gelsolin protein, which binds actin and is involved in cytoskeletal remodelling. Two mutations at the same locus, NM\_198252.3(GSN):c.487G>A (p.Asp163Asn) and NM\_198252.3(GSN):c.487G>T (p.Asp163Tyr), cause the classical syndrome, although other mutations have more recently been described. 171-175 Onset is typically in the fourth or fifth decade of life, and corneal lattice dystrophy is often the first clinical sign. Stroke, and in particular ICH, is unusual, 172,176,177 as is dementia,177 although subtle neuropsychological deficits have

been reported. 176 Brain MRI features can include white matter changes<sup>178</sup> (although not necessarily in excess of that expected for age<sup>176,179</sup>) and a small number of microhaemorrhages.<sup>176</sup> Classical features include a progressive cranial neuropathy (typically starting with the facial nerve and progressing to involve other lower cranial nerves, thereby compromising speech and swallowing), a mild predominantly sensory peripheral neuropathy, and dermatological involvement, particularly cutis laxa. 172,180 Other features can include myokymia, autonomic dysfunction, gait ataxia and the consequences of cardiac and renal involvement. 172,180 Pathologically, there is deposition of AGel throughout in multiple organs and particularly involving the basement membranes of epithelial, smooth and striated muscle cells<sup>177,181</sup>; in the CNS deposition is predominantly vascular, and involves the grey and white matter of the brain, 182,183 spinal cord and meninges. 182

#### Prion protein cerebral amyloid angiopathy

Prion diseases are associated with the propagation of diseaserelated assemblies of misfolded prion protein (PrP) or prions and can be sporadic, acquired or inherited in aetiology<sup>184</sup>; they have wide phenotypic diversity which can readily mimic other neurodegenerative diseases with the classical syndromes being Creutzfeldt-Jakob disease (CJD), Gerstmann-Sträussler-Scheinker syndrome (GSS), fatal familial insomnia (FFI) and kuru. 184 Whilst the common pathological findings in most cases, regardless of aetiology, are spongiform degeneration, gliosis, neuronal loss and abnormal PrP immunoreactivity (synaptic deposits and/or plaques of multiple morphologies<sup>21,185,186</sup>), amyloid angiopathy can occur. This appears to be an exclusive feature of certain inherited forms of PrP disease, namely stop codon mutations in the PRNP gene which result in truncated forms of PrP with loss of the glycosylphosphatidylinositol (GPI) anchor (which allows PrP to attach to the cell membrane), and some octapeptide repeat insertional mutations. 21,169,186-188

Mutations associated neuropathologically with PrP-CAA include NM\_000311.5(PRNP):c.435T>G (p.Tyr145Ter), 187,189-191 NM\_ 000311.5(PRNP):c.478C>T (p.Gln160Ter), 189,191-194 NM\_000311.5 (PRNP):c.489C>G (p.Tyr163Ter), 195,196 NM\_000311.5(PRNP):c.678C>A (p.Tyr226Ter)<sup>197</sup> and NM\_000311.5(PRNP):c.534\_535del (p.Asp178fs)<sup>188,198,199</sup> The clinical phenotype of these mutations includes dementia, autonomic dysfunction, chronic diarrhoea and peripheral neuropathy and some have been associated with nonspecific white matter hyperintensities on MRI (Table 2). To date, other clinical or neuroimaging features associated with sporadic amyloid-β CAA have not been described. Three further stop codon mutations have been described, 196,197,199 but neuropathological data are not available and therefore the presence of PrP-CAA is unknown; imaging data, where reported, have not been typical for CAA.

#### Transthyretin

Transthyretin (TTR) is a transport protein synthesized by the liver, choroid plexus and retinal pigment epithelium, 200,201 and in humans it can result in amyloidosis in one of two ways. The first is amyloidosis of wild-type TTR, which results in senile systemic amyloidosis (also called wild-type ATTR or ATTRwt amyloidosis) characterized by cardiomyopathy in the elderly (over the age of 80).  $^{201,202}$  The second is amyloidosis due to variant forms of the TTR protein, caused by mutations in the TTR gene and resulting in hereditary ATTR amyloidosis.<sup>201</sup> Hereditary ATTR amyloidosis usually presents with one of three main phenotypes: familial amyloid polyneuropathy (FAP), familial amyloid cardiomyopathy and familial leptomeningeal (sometimes meningovascular, or oculoleptomeningeal/

oculomeningovascular, if there is eye involvement) amyloidosis. This latter presentation is associated with widespread amyloid deposition within the small and medium arteries, arterioles and veins of the leptomeninges, and more variably can involve the brain and eyes. <sup>201,203-208</sup> Leptomeningeal involvement can also occur in people with FAP. <sup>206,209</sup> Several mutations associated with familial leptomeningeal amyloidosis have been identified, <sup>204</sup> and this syndrome can rarely develop *de novo* in other phenotypes after liver transplantation, which is the usual treatment for ATTR amyloidosis (as it removes the main source of variant TTR) due to ongoing TTR synthesis by the choroid plexus and retinal pigment epithelium. <sup>202,210-213</sup> Improved treatment of the systemic amyloid component of ATTR amyloidosis (without effective treatment of CNS TTR synthesis) is leading to increased recognition of the leptomeningeal manifestations, creating an unmet need for treatment.

Clinically, leptomeningeal involvement can present with intracerebral and subarachnoid haemorrhage, which can be recurrent<sup>204,214-218</sup>; progressive cognitive impairment<sup>201,219</sup>; transient focal neurological episodes<sup>210,212</sup> can occur, but not always with radiological evidence of leptomeningeal disease; they may be an early clinical clue to leptomeningeal involvement. Other features can include headache, ataxia, myelopathy, seizures and psychosis.<sup>201,204</sup> Brain MRI can show superficial siderosis (supra- and infratentorial, with the latter resulting in a clinical syndrome analogous to classical superficial siderosis of the CNS) and cerebral microbleeds, <sup>212,215,220,221</sup> in addition to meningeal enhancement with contrast.<sup>204-207,222</sup> White matter hyperintensities do not appear to be a prominent feature.

# Other causes of early-onset cerebral amyloid angiopathy

#### Iatrogenic cerebral amyloid angiopathy

Whilst many proteins associated with neurodegeneration have been shown experimentally to have 'prion-like' properties, there has been no clear evidence for their iatrogenic transmission until recently with the recognition of human transmission of amyloid-\$\beta\$ pathology following treatment with human cadaverderived pituitary growth hormone contaminated with amyloid-\u03b4 seeds<sup>223</sup> and causing iatrogenic CAA by a prion-like process.<sup>224</sup> This iatrogenic form of CAA has only been recognized relatively recently,<sup>223</sup> and several cases have now been reported.<sup>225,226</sup> Clinical presentation is typically with ICH, occurring after a latency of between two and four decades after exposure to amyloid-β, often in childhood; cases presenting with cognitive impairment and seizures have also been described.<sup>226</sup> Associated medical interventions include neurosurgery, and procedures or treatments involving cadaveric human material, for example dura mater (used either as a surgical material or for embolization of vascular malformations) and pituitary-derived (cadaveric) human growth hormone.<sup>223,225</sup> The MRI and other investigative findings (amyloid-PET, CSF) are similar to those observed in sporadic amyloid-β CAA.<sup>226-238</sup> Pathologically, there is evidence of vascular amyloid-β deposition, but findings can also include parenchymal amyloid-β plaques and tau pathology, which are more commonly observed in Alzheimer's disease. 111

#### Inflammatory cerebral amyloid angiopathy

Inflammatory forms of CAA, a spectrum of disease ranging from CAA-related inflammation (CAA-ri) to amyloid- $\beta$  related angiitis

(ABRA), describe forms of CAA where pathologically there is an inflammatory response to vascular amyloid- $\beta$ . <sup>239,240</sup> Although the vast majority of cases present at an older age (over 65 years <sup>239</sup>; mean age 72.9 years in a recent prospective cohort of 113 patients <sup>241</sup>), younger onset cases have been described, including cases in people with symptomatic and presymptomatic familial Alzheimer's disease. <sup>242,243</sup> The current clinico-radiological diagnostic criteria allow the diagnosis to be made from the age of 40 onwards, though the spectrum of disease is widening. <sup>239,244,245</sup>

CAA-related inflammation is thought to occur as a result of the spontaneous generation of auto-antibodies against amyloid- $\beta$ , supported by the identification by one group of such antibodies in the CSF of some patients with inflammatory CAA,  $^{246\text{-}248}$  and the observation that similar imaging appearances sometimes occur following the administration of anti- amyloid- $\beta$  antibodies as a treatment for Alzheimer's disease (amyloid-related imaging abnormalities, ARIA).  $^{249}$  Clinically, CAA-related inflammation classically presents with seizures, altered consciousness, cognitive decline (often rapidly progressive) and focal neurological deficits due to stroke, which are frequently but not exclusively haemorrhagic.  $^{239}$  Milder forms in which there is a mismatch between the severity of clinical and imaging features have also been reported.  $^{250}$  This form of CAA can also manifest in people with established cognitive impairment.  $^{251}$ 

The imaging criteria  $^{245}$  for CAA-related inflammation require the presence of one or more cortico-subcortical haemorrhagic lesions (ICH, acute convexity subarachnoid haemorrhage, cortical superficial siderosis, cerebral microbleeds) and characteristic white matter lesions that extend to the immediately subcortical white matter; these can be unifocal or multifocal, cortico-subcortical or deep, and are usually asymmetric. Some cases seem to respond well to immunosuppression, but there are also cases of spontaneous improvement and treatment failure (either failure to respond, or relapse following an initial response), highlighting the paucity of natural history data for this condition.  $^{239,241,252}$  The APOE  $\epsilon 4$  genotype appears to be a risk factor for both inflammatory CAA and ARIA.  $^{239,253-257}$ 

#### Non-genetic systemic amyloid disorders

There are pathological descriptions of cerebrovascular involvement in patients with light chain (AL) and serum amyloid A (AA) systemic amyloidoses.<sup>258</sup> ICH in the context of AL amyloidosis has been reported,<sup>259</sup> but appears to be a rare complication of this condition.

#### Aluminium toxicity

Two pathological cases of CAA occurring in the context of aluminium toxicity have been reported, one of which presented clinically at the age of 49 years, <sup>260</sup> following a water pollution incident in 1988 in North Cornwall, UK. <sup>260-262</sup> In this accident, high concentrations of aluminium sulphate were discharged into the drinking water supply over a period of weeks. <sup>261</sup> However, genetic testing in these cases was limited (reflecting the availability of testing at the time) and there have been no further reports of cases following this exposure; given the small number of cases, it is difficult to draw firm conclusions regarding causality. Further details regarding the clinical presentation (particularly details of past medical history) would help to exclude other potentially relevant exposures (particularly iatrogenic ones).

### Making the diagnosis and what to do

#### Approach to investigation

Whilst there is no consensus on how patients with early-onset CAA should be investigated (perhaps unsurprisingly, given the relative rarity of cases), we recommend the approach outlined in Fig. 5. Some of the recommended tests might only be available in centres with a specialist interest in CAA, and therefore onward referral should be considered.

Early-onset cases can present via acute stroke services (after ICH, transient focal neurological episodes or symptoms of CAA-related inflammation), to memory clinics and other cognitive services, or following identification of imaging features of CAA whilst undergoing investigation for other neurological symptoms (e.g. headache). For those presenting with ICH, causes which are particularly associated with haemorrhagic manifestations of CAA should be initially considered (Fig. 2). Familial Alzheimer's disease associated with CAA might present to cognitive services without clinical or imaging features of haemorrhage8; in such cases hippocampal atrophy and white matter changes (Fig. 1E and G) might be important, as is a family history of dementia or stroke (early or late onset). Such patients should all have MRI with blood-sensitive sequences to review for clinically silent markers of CAA (particularly cerebral microbleeds; Fig. 1D, F and H), in addition to standard volumetric and FLAIR sequences.

In all cases, details of other potentially relevant neurological (including migraine and the nature of any aura, transient neurological disturbances) and non-neurological symptoms should be recorded;

CAA-related TFNE are stereotyped episodes of cortical disturbance, usually lasting for less than 30 min and classically with a 'spreading' onset and progression.<sup>263</sup> In addition, it is important to establish whether there is history of potential iatrogenic amyloid- $\beta$ exposure, particularly treatment or operations involving cadaveric pituitary hormones or dura mater and any other previous medical or surgical procedures involving the brain, spinal cord or posterior eye. This may require a diligent search of all available previous medical records, including operation notes from decades earlier. In addition to obtaining a detailed family history of neurological disease, we would advise confirming the patient's ethnic background and family origins.

In cases where there is clinical and radiological evidence of early-onset CAA, with or without a supporting family history, we would recommend genetic testing, either via a neurodegenerative panel, whole exome or whole genome sequencing (WES and WGS, respectively); copy number variants may need to be requested separately, depending on the method used. In centres where only individual gene testing is available, the presence or absence of cognitive impairment in addition to haemorrhagic markers of CAA can be useful in establishing which genes to prioritize. In people with haemorrhagic markers of CAA but no cognitive symptoms, and no family history of dementia, focused initial testing of APP (to include both missense and copy number variants) is likely to have the highest yield. In people with cognitive symptoms as well as haemorrhagic CAA, or cases where there is a family history, testing of APP (missense mutations and copy number variants), PSEN1 and PSEN2 is needed. In individuals with a later age at symptom onset but strong family history, genetic testing may also be considered.

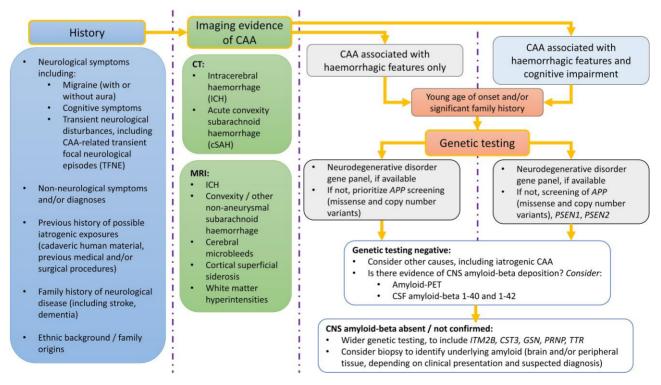


Figure 5 An outline approach to investigating early-onset CAA. The order of investigations will necessarily depend on the clinical context for an individual patient and the investigations available at a particular centre. In cases where gene panel testing is requested, it is important to ensure that the method allows duplications (and other copy number variants) to be identified, as this is not universally the case. CAA = cerebral amyloid angiopathy.

Should this genetic testing be negative, other diagnoses (such as iatrogenic CAA) should be explored. Further investigations to evaluate the presence (or absence) of amyloid- $\beta$  can be considered, as this potentially allows exclusion of the rarer non-amyloid-\u03b3 forms of CAA. This can be achieved by CSF measurement of amyloid-β markers, although validated thresholds for the clinical diagnosis of CAA have not been defined. The presence of CSF amyloid- $\beta_{1-42}$  levels, or a CSF amyloid- $\beta_{1-42/1-40}$  ratio consistent Alzheimer's disease would support beta-amyloidosis<sup>264,265</sup>; plasma measurement of these markers might also be possible clinically in the future.<sup>266</sup> Amyloid-PET imaging can also be helpful, although this imaging modality might not be available outside non-specialist centres and it is important to recognize that tracers can bind other amyloids and are therefore not specific for amyloid- $\beta$ . <sup>154,267,268</sup> As with CSF measures, thresholds for CAA diagnosis have not been established, 269 but amyloid-PET imaging consistent with Alzheimer's disease may support the presence of cerebral amyloid-β deposition.<sup>265</sup> In cases where amyloid-β deposition has not been confirmed, broader genetic testing to include other causes of cerebrovascular amyloidosis should be considered.

Rarely, and particularly when other treatable conditions are in the differential diagnosis (e.g. cerebral vasculitis or intravascular lymphoma), it may be appropriate to consider a brain biopsy, with CJD precautions where appropriate. Brain biopsy and other relevant ancillary tests (such as digital subtraction angiography) might be considered in cases with an unusual clinical presentation (for example, rapidly progressive symptoms, or those where encephalopathy or seizures without ICH are a dominant feature), atypical imaging findings (for example, prominent cortical swelling, acute ischaemic lesions, isolated white matter lesions with an inflammatory appearance) and inflammatory changes in the CSF. A previous series of cerebral biopsies undertaken to investigate patients with dementia found that a raised CSF white cell count was the best predictor for identifying a potential treatable, inflammatory process. 270,271

The nature and provision of genetic testing is changing, and the increasing availability of next generation sequencing techniques including WES and WGS has the potential to change the diagnostic approach in early onset-CAA.<sup>272</sup> In particular 'neurodegenerative disorder' gene panels, which contain APP, PSEN1 and PSEN2 as well as other genes of interest, can be a more practical option in those with a strong family history, and can be more readily accessible than investigations such as amyloid-PET or CSF amyloid-β measures. Access and approaches to genetic testing may vary between countries. In the UK, the National Health Service (NHS) Genomic Medicine Service aims to provide consistent and equitable access to genomic medicine and has developed the publicly available, annually updated, National Genomic Test Directory (https://www.england.nhs.uk/genomics/the-national-genomic-tes t-directory/), which lists the available tests, their indications, who can access them and the testing methodologies by which they should be delivered. WGS is being introduced for neurodegenerative disorder panel testing, which brings increased diagnostic capabilities including the ability to detect copy number variants, in contrast to WES. Prior to the introduction of WGS, testing for APP duplications had to be requested and performed separately.<sup>272</sup> For both whole exome and whole genome approaches, careful pre-test discussions regarding variants of uncertain significance and secondary findings (mutations unrelated to the condition being investigated, which could have implications for future health) should be included as part of the informed consent process.<sup>272</sup>

#### Management considerations

#### Anti-platelet and anticoagulant medications

Given that CAA is clinically associated with intracranial bleeding, there is understandable anxiety when people with CAA require treatment with medications that increase that bleeding risk, namely anti-platelet and anticoagulant medications. An MRI can be helpful in this context for confirming the diagnosis of CAA, particularly in people presenting with a single lobar ICH. Whilst younger patients (particularly those without systemic amyloidosis) are less likely to have age-associated comorbid conditions that might require the use of such medications (e.g. atrial fibrillation, ischaemic heart disease, peripheral vascular disease), the risk of recurrent haemorrhage in individuals with familial CAA presenting with or without ICH might be significant, although definitive natural history data are lacking.

In the absence of good quality trial data in these specific youngonset cohorts, we base our approach on that used for patients with sporadic CAA. In people who have had one or more ICH, evidence from the RESTART trial<sup>273</sup> suggests no difference in outcomes between those that do and do not restart anti-platelet therapy for the secondary prevention of occlusive vascular disease, even in those with lobar ICH (which has a higher recurrence rate and is associated with CAA). However, this trial included few individuals with CAA. In view of this, our practice is to reserve anti-platelet therapy only for those with strong indications for use, such as significant ischaemic heart disease or previous ischaemic stroke. There are fewer data regarding anticoagulant medications after ICH, and no randomized data specific to CAA; this creates a management challenge for those with long-term or lifelong indications for use (atrial fibrillation, recurrent venous thromboembolism). The APACHE-AF trial of apixaban in survivors of an anticoagulant associated ICH with co-existent atrial fibrillation found that the risk of non-fatal stroke or vascular death was similar regardless of whether a participant received apixaban or not.<sup>274</sup> However, the event rate was low, precluding analysis by original haemorrhage location or presence of CAA. SoSTART, <sup>275</sup> a randomized open-label non-inferiority trial investigating starting versus stopping oral anticoagulation in survivors of intracranial haemorrhage with atrial fibrillation and which included twice as many participants as APACHE-AF, was also inconclusive. In both SoSTART and APACHE-AF, the majority of recurrent ICH events occurred in people randomized to an anticoagulant group. A recent Cochrane synthesis review,<sup>276</sup> which included the APACHE-AF, ReSTART and SoSTART studies (and others) similarly concluded that there is limited evidence to support either benefit or harm when treating with anti-platelets or anticoagulants after ICH. For these reasons, in people with CAA and a long-term or lifelong indication for anticoagulation, a direct oral anticoagulant (DOAC) is preferred over vitamin K antagonists like warfarin due to the ~50% lower risk of intracranial haemorrhage. Alternative approaches that limit the duration of anticoagulation (e.g. left atrial appendage closure) can also be considered, although as yet there are no data for these approaches in early-onset CAA. In people with haemorrhagic MRI markers of CAA (cortical superficial siderosis and cerebral microbleeds; Fig. 1C and D) but no history of symptomatic haemorrhage, we would recommend a similar approach, but one that considers other symptoms. Transient focal neurological episodes and cortical superficial siderosis are both likely to result from episodes of acute convexity subarachnoid haemorrhage (Fig. 1B), which is associated with a high risk of later ICH<sup>277,278</sup>; in view of this, we consider these

patients in the same way as those with a history of symptomatic ICH. For those with cerebral microbleeds only, our approach is more nuanced; the Microbleeds International Collaborative Network (MICON) ischaemic stroke and ICH scores (MICON-IS and MICON-ICH, respectively)<sup>279</sup> provide one approach for evaluating the impact of microbleed presence on ischaemic stroke and ICH risk, but the scores have not been validated outside patients with ischaemic stroke or transient ischaemic attack (TIA).

#### Vascular risk factors including blood pressure

As with anti-platelet and anticoagulant medications, specific trial evidence for early-onset CAA is lacking, and therefore our practice is based on our approach for people with sporadic CAA. Good blood pressure control in people with prior ICH reduces subsequent haemorrhage risk, 280 including those with ICH due to sporadic CAA, 281 as demonstrated in the PROGRESS trial. There are also observational data that support stricter blood pressure targets in people with ICH (including lobar ICH) to reduce future haemorrhage recurrrence.<sup>282</sup> Although evidence for a specific target blood pressure for secondary prevention in CAA is limited (due to methodological differences in the relevant trials),<sup>283</sup> we aim for a blood pressure target of 130/80 mmHg or lower (as tolerated) in people with sporadic CAA, with or without a prior history of ICH; this target is recommended for all people with ICH in recently published guidelines from the American Heart Association (AHA) and American Stroke Association (ASA). 284 In early-onset CAA, we regularly monitor blood pressure and aim for a similar blood pressure target, particularly in those with prior ICH.

Statin use in people with CAA remains controversial. Two large, randomized placebo-controlled trials (SPARCL, <sup>285</sup> HPS<sup>286</sup>) both suggested an increased risk of ICH with statin use, and there have been subsequent recommendations that statin use should be avoided in those with lobar ICH. <sup>287</sup> Meta-analysis of these and other statin trials did not find an increased risk of ICH, but did note a positive impact of statin use on mortality and functional outcome, although this work did not consider CAA specifically. <sup>288</sup> The recent AHA/ASA guidelines<sup>284</sup> recommend that statin use following ICH should be considered on an individual basis.

The impact of other vascular risk factors for early-onset CAA, as for other types of CAA, is less clear. There is a suggestion that life expectancy of people with cystatin C amyloidosis dramatically reduced during the 19th century, which was thought to reflect environmental changes and specifically an increased consumption of dietary salt and imported foods high in carbohydrates. <sup>167</sup> Whilst this might simply relate to higher rates of hypertension, it does raise a question of whether aggressive management of other cardiovascular risk factors in people with genetic forms of CAA might have prognostic benefits. Our approach is to screen for cardiovascular risk (diabetes, smoking, lipid profile) and treat only where abnormalities are identified. We also advise regular exercise, a healthy diet low in saturated fats, avoidance of alcohol and smoking cessation.

#### Symptomatic treatments for Alzheimer's disease

Pharmacological treatments such as acetylcholinesterase inhibitors (donepezil, rivastigmine, galantamine) and the NMDA receptor antagonist memantine<sup>289</sup> are widely licensed for symptomatic management in Alzheimer's disease and should be offered to individuals with familial Alzheimer's disease. Whilst these medications are not routinely offered to individuals diagnosed with CAA and/or vascular dementia alone (trials to date of these agents in

vascular dementia show modest cognitive benefits, but with effect sizes felt unlikely to be clinically important and no real change for functional outcomes<sup>290,291</sup>), there are case reports suggesting that acetylcholinesterase inhibitors might provide some benefit in those thought to have Alzheimer's disease co-pathology contributing to cognitive impairment in addition to CAA.<sup>292</sup> Given their relatively benign side effect profile, treatment with these agents can be considered in early-onset cases suspected to have both diagnoses.

#### Other considerations

The diagnosis of a monogenic form of CAA has implications for the patient but also for their families, who may benefit from early involvement of a clinical genetics team. Related individuals should, if they wish, be given access to genetic counselling to explore their choices in a variety of areas including predictive (pre-symptomatic) or diagnostic genetic testing, and reproductive options such as preimplantation genetic diagnosis. They may wish to explore opportunities to connect with other similarly affected families, and to participate in research including pre-symptomatic trials of therapies with the potential for disease modification.<sup>293</sup>

Inflammatory forms of CAA are important to recognize as they can respond to immunosuppressive treatment (usually steroids in the first instance). Although clinically and radiologically relapses following treatment withdrawal are now increasingly reported, <sup>239,252</sup> findings from a series of patients with CAA-related inflammation suggest that early immunosuppressive treatment may both improve the initial disease course and reduce the risk of recurrence. <sup>252</sup>

Finally, there are data suggesting that selective serotonin reuptake inhibitors (SSRI) can be associated with increased recurrent ICH risk, <sup>294-296</sup> although the relationship may reflect confounding influences rather than causation and data specifically for CAA and for younger people are lacking. AHA and ASA ICH guidance recommends reserving use of SSRIs for people with moderate-to-severe depression following ICH, but recognizes there is a paucity of data on risk of ICH for specific SSRI medications and on distinguishing risk profiles between SSRIs and other antidepressants including serotonin-noradrenaline reuptake inhibitors (SNRI). <sup>284</sup>

#### **Discussion**

In this review, we provide an overview of diagnoses to consider when assessing patients with early-onset CAA, and suggest an approach to their investigation and management. These unusual forms of CAA provide potential insights into the pathophysiology of vascular amyloid deposition. The observation that a range of different amyloidogenic proteins can be deposited in the vasculature has led to the protein elimination failure hypothesis, which proposes that deposition of insoluble amyloid proteins within the walls of blood vessels is associated with failure of normal perivascular clearance mechanisms.<sup>297</sup> The effectiveness of these systems is thought to be influenced by age and APOE genotype and might be additionally challenged by certain protein structures particularly prone to vascular aggregation. 297,298 Examples include: mutations associated with PrP-CAA, which all result in 'anchorless' PrP<sup>297</sup>; ITM2B mutations in FBD and FDD, which both produce elongated forms of BRI2 protein of identical length; and APP missense mutations associated with severe CAA, most of which fall within a narrow region of the amyloid- $\!\beta$  coding domain, between codons 692 and 694.<sup>22</sup> Monogenic forms of CAA might also allow the identification of elusive early disease biomarkers, particularly in pre-symptomatic

mutation carriers, as has been the case in D-CAA. The hypothesized prion mechanism of seeding and spread of iatrogenic CAA may have relevance for the pathogenesis of sporadic amyloid- $\beta$  CAA, for example in understanding the pre-clinical latency period (analogous to that hypothesized in Alzheimer's disease) and some features of sporadic CAA including frequent spatial clustering of siderosis, microbleeds and ICH. Therefore, despite being rare, improving our understanding of disease processes in early-onset CAA subtypes might provide important mechanistic information for other, more commonly encountered, forms of CAA.

The monogenic forms of CAA also serve to demonstrate the range of clinical phenotypes associated with pathological evidence of CAA; it is striking that only a handful of the genetic forms are associated with ICH, and how this does not always correlate with pathological severity. Of the non-amyloid-β CAAs, only cystatin C and leptomeningeal TTR amyloidosis are clearly associated with ICH, with the latter bearing the closest semblance to the clinical phenotype typically associated with sporadic amyloid-β CAA with prominent (and sometimes exclusive) leptomeningeal involvement. One hypothesis might be that significant anatomical involvement of larger leptomeningeal vessels might predispose to a 'haemorrhagic' phenotype, characterized by symptomatic ICH, convexity subarachnoid haemorrhage, cortical superficial siderosis and transient focal neurological episodes, whereas predominantly cortical involvement might have a more 'cognitive' phenotype, where cerebral microbleeds and white matter changes might be present, but other haemorrhagic features are rare. This is supported by data suggesting that the APOE €2 allele is associated with leptomeningeal and haemorrhagic phenotypes, whereas the €4 allele is associated with capillary CAA, cognitive phenotypes and Alzheimer's disease pathology. 42,299-304 However, significant and dominant leptomeningeal involvement can be a feature of APP and PSEN1 mutations, 305 and in other non-amyloid-β disorders including those caused by ITM2B mutations. An alternative hypothesis is that certain mutations render the resulting protein more toxic to the cellular components of the cerebral (cortical and leptomeningeal) vasculature, due to structural changes. 306,307 The concept of strains, where proteins with the same amino acid sequence can form multimeric assemblies or seeds with distinct aberrant folds of the monomer subunits, is well recognized in prion biology, 308 and there is some evidence that a similar conformational effect might have relevance to phenotype for other proteins including amyloid- $\beta^{309,310}$ ; such that structurally distinct amyloid- $\beta$ seeds or strains might provide an explanation for the clinical heterogeneity observed within families with the same mutation. Finally, it is important to consider the potential impact of these mutations on other non-vascular functions of a given protein. As an example, amyloid-β might have a physiological role in clotting<sup>311</sup>; it is produced by platelets and can induce their aggregation,87 and further influences clotting via its interactions with fibrinogen.<sup>312</sup> There is evidence that the Dutch and Iowa APP mutations result in altered amyloid-β interactions with fibrinogen and subsequent alterations in clot structure. 313 It is therefore possible that haemorrhage in amyloid-β CAA is a 'two-hit' phenomenon, where vascular amyloid deposition (affecting vessel structural integrity) and clotting perturbation are both necessary. Further work will be needed to explore this and related hypotheses.

Early-onset CAA is an important diagnosis to recognize; these rare forms of CAA require focused investigation and management, and have significant implications for both the affected patient and their families. Improved awareness of these unusual forms of CAA amongst healthcare professionals is essential for facilitating their

prompt diagnosis, and understanding their underlying pathophysiology is likely to have implications for our understanding of more common, late-onset, forms of the disease.

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#### **Competing interests**

The authors report no competing interests.

#### References

- Biffi A, Greenberg SM. Cerebral amyloid angiopathy: a systematic review. J Clin Neurol. 2011;7:1-9.
- Benson MD, Buxbaum JN, Eisenberg DS, et al. Amyloid nomenclature 2020: update and recommendations by the International Society of Amyloidosis (ISA) nomenclature committee. Amyloid. 2020;27:217-222.
- Jakel L, De Kort AM, Klijn CJM, Schreuder F, Verbeek MM. Prevalence of cerebral amyloid angiopathy: A systematic review and meta-analysis. Alzheimers Dement. 2022;18:10-28.
- Banerjee G, Carare R, Cordonnier C, et al. The increasing impact of cerebral amyloid angiopathy: Essential new insights for clinical practice. J Neurol Neurosurg Psychiatr. 2017;88:982-994.
- McNaughton D, Knight W, Guerreiro R, et al. Duplication of amyloid precursor protein (APP), but not prion protein (PRNP) gene is a significant cause of early onset dementia in a large UK series. Neurobiol Aging. 2012;33:426 e13-21.
- Ryan NS, Bastos-Leite AJ, Rohrer JD, et al. Cerebral microbleeds in familial Alzheimer's disease. Brain. 2012;135(Pt 1):e201.
- Linn J, Halpin A, Demaerel P, et al. Prevalence of superficial siderosis in patients with cerebral amyloid angiopathy. Neurology. 2010;74:1346-1350.
- Charidimou A, Boulouis G, Frosch MP, et al. The Boston criteria version 2.0 for cerebral amyloid angiopathy: a multicentre, retrospective, MRI-neuropathology diagnostic accuracy study. Lancet Neurol. 2022;21:714-725.
- Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. Lancet Neurol. 2013;12: 822-838.
- Malhotra K, Theodorou A, Katsanos AH, et al. Prevalence of clinical and neuroimaging markers in cerebral amyloid

- angiopathy: a systematic review and meta-analysis. Stroke. 2022;53:1944-1953.
- 11. Rodrigues MA, Samarasekera N, Lerpiniere C, et al. The Edinburgh CT and genetic diagnostic criteria for lobar intracerebral haemorrhage associated with cerebral amyloid angiopathy: model development and diagnostic test accuracy study. Lancet Neurol. 2018;17:232-240.
- 12. van Etten ES, Kaushik K, van Zwet EW, et al. Sensitivity of the Edinburgh criteria for lobar intracerebral hemorrhage in hereditary cerebral amyloid angiopathy. Stroke. 2020;51:3608-3612.
- 13. Sembill JA, Knott M, Xu M, et al. Simplified Edinburgh CT criteria for identification of lobar intracerebral hemorrhage associated with cerebral amyloid angiopathy. Neurology. 2022;98: e1997-e2004.
- 14. Baron JC, Boulouis G, Benzakoun J, et al. Cerebral amyloid angiopathy-related acute lobar intra-cerebral hemorrhage: Diagnostic value of plain CT. J Neurol. 2022;269:2126-2132.
- 15. Schwarz G, Banerjee G, Hostettler IC, et al. MRI And CT imaging biomarkers of cerebral amyloid angiopathy in lobar intracerebral hemorrhage. Int J Stroke. 2023;18:85-94.
- 16. Fisher RA, Miners JS, Love S. Pathological changes within the cerebral vasculature in Alzheimer's disease: New perspectives. Brain Pathol. 2022;32:e13061.
- 17. Vonsattel JP, Myers RH, Hedley-Whyte ET, Ropper AH, Bird ED, Richardson EP Jr. Cerebral amyloid angiopathy without and with cerebral hemorrhages: A comparative histological study. Ann Neurol. 1991;30:637-649.
- 18. Love S, Chalmers K, Ince P, et al. Development, appraisal, validation and implementation of a consensus protocol for the assessment of cerebral amyloid angiopathy in post-mortem brain tissue. Am J Neurodegener Dis. 2014;3:19-32.
- 19. Olichney JM, Hansen LA, Hofstetter CR, Grundman M, Katzman R, Thal LJ. Cerebral infarction in Alzheimer's disease is associated with severe amyloid angiopathy and hypertension. Arch Neurol. 1995;52:702-708.
- 20. Kovari E, Herrmann FR, Hof PR, Bouras C. The relationship between cerebral amyloid angiopathy and cortical microinfarcts in brain ageing and Alzheimer's disease. Neuropathol Appl Neurobiol. 2013;39:498-509.
- 21. Revesz T, Holton JL, Lashley T, et al. Genetics and molecular pathogenesis of sporadic and hereditary cerebral amyloid angiopathies. Acta Neuropathol. 2009;118:115-130.
- 22. Zhang-Nunes SX, Maat-Schieman ML, van Duinen SG, Roos RA, Frosch MP, Greenberg SM. The cerebral beta-amyloid angiopathies: Hereditary and sporadic. Brain Pathol. 2006;16:30-39.
- 23. Ryan NS, Rossor MN. Correlating familial Alzheimer's disease gene mutations with clinical phenotype. Biomark Med. 2010;4: 99-112.
- 24. Mancuso M, Arnold M, Bersano A, et al. Monogenic cerebral small-vessel diseases: Diagnosis and therapy. Consensus recommendations of the European academy of neurology. Eur J Neurol. 2020;27:909-927.
- 25. Guey S, Lesnik Oberstein SAJ, Tournier-Lasserve E, Chabriat H. Hereditary cerebral small vessel diseases and stroke: A guide for diagnosis and management. Stroke. 2021;52:3025-3032.
- 26. Forman R, Conners JJ, Song SY, et al. The Spectrum of nontraumatic convexity subarachnoid hemorrhage. Journal of stroke and cerebrovascular diseases: the official journal of National Stroke Association. 2019;28:104473.
- 27. Patel N, Banahan C, Janus J, et al. Perioperative cerebral microbleeds after adult cardiac surgery. Stroke. 2019;50:336-343.
- 28. De Sciscio M, De Sciscio P, Vallat W, Kleinig T. Cerebral microbleed distribution following cardiac surgery can mimic cerebral amyloid angiopathy. BMJ Neurol Open. 2021;3:e000166.

- 29. Van Belle E, Debry N, Vincent F, et al. Cerebral microbleeds during transcatheter aortic valve replacement: A prospective magnetic resonance imaging cohort. Circulation. 2022;146: 383-397.
- Scheid R, Preul C, Gruber O, Wiggins C, von Cramon DY. Diffuse axonal injury associated with chronic traumatic brain injury: Evidence from T2\*-weighted gradient-echo imaging at 3 T. AJNR Am J Neuroradiol. 2003;24:1049-1056.
- 31. Koschmieder K, Paul MM, van den Heuvel TLA, van der Eerden AW, van Ginneken B, Manniesing R. Automated detection of cerebral microbleeds via segmentation in susceptibilityweighted images of patients with traumatic brain injury. Neuroimage Clin. 2022;35:103027.
- Fanou EM, Coutinho JM, Shannon P, et al. Critical illnessassociated cerebral microbleeds. Stroke. 2017;48:1085-1087.
- Backman L, Möller MC, Thelin EP, et al. Monthlong intubated patient with life-threatening COVID-19 and cerebral microbleeds suffers only mild cognitive sequelae at 8-month follow-up: A case report. Arch Clin Neuropsychol. 2022;37: 531-543.
- Agarwal S, Jain R, Dogra S, et al. Cerebral microbleeds and leukoencephalopathy in critically ill patients with COVID-19. Stroke. 2020;51:2649-2655.
- Lloyd GM, Trejo-Lopez JA, Xia Y, et al. Prominent amyloid plaque pathology and cerebral amyloid angiopathy in APP V717I (London) carrier—Phenotypic variability in autosomal dominant Alzheimer's disease. Acta Neuropathol Commun. 2020;8:31.
- 36. Murrell J, Farlow M, Ghetti B, Benson MD. A mutation in the amyloid precursor protein associated with hereditary Alzheimer's disease. Science. 1991;254:97-99.
- 37. Mullan M, Crawford F, Axelman K, et al. A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N-terminus of beta-amyloid. Nat Genet. 1992;1:345-347.
- 38. Lannfelt L, Bogdanovic N, Appelgren H, et al. Amyloid precursor protein mutation causes Alzheimer's disease in a Swedish family. Neurosci Lett. 1994;168(1-2):254-256.
- Markus HS, Schmidt R. Genetics of vascular cognitive impairment. Stroke. 2019;50:765-772.
- Carpenter AM, Singh IP, Gandhi CD, Prestigiacomo CJ. Genetic risk factors for spontaneous intracerebral haemorrhage. Nat Rev Neurol. 2016;12:40-49.
- 41. Chen Y, Strickland MR, Soranno A, Holtzman DM. Apolipoprotein E: Structural insights and links to Alzheimer disease pathogenesis. Neuron. 2021; 109:205-221. doi:10.1016/ j.neuron.2020.10.008
- 42. Rannikmae K, Samarasekera N, Martinez-Gonzalez NA, Al-Shahi Salman R, Sudlow CL. Genetics of cerebral amyloid angiopathy: systematic review and meta-analysis. J Neurol Neurosurg Psychiatr. 2013;84:901-908.
- 43. Greenberg SM, Briggs ME, Hyman BT, et al. Apolipoprotein E epsilon 4 is associated with the presence and earlier onset of hemorrhage in cerebral amyloid angiopathy. Stroke. 1996;27:
- 44. Greenberg SM, Vonsattel JP, Segal AZ, et al. Association of apolipoprotein E epsilon2 and vasculopathy in cerebral amyloid angiopathy. Neurology. 1998;50:961-965.
- 45. Hens E, Bossaerts L, Van den Bossche T, et al. In a large Belgian AD cohort loss of ABCA7 mutations are associated with Alzheimer's disease and cerebral amyloid angiopathy. (1779). Neurology. 2020;94(15 Supplement):1779.
- 46. Hens E, Bossaerts L, Sieben A, et al. Belgian Carriers of rare ABCA7 mutations present with pronounced cerebral amyloid angiopathy and Alzheimer's disease (S15.004). Neurology. 2022;98(18 Supplement):2645.

- 47. Bossaerts L, Van de Craen EH, Cacace R, Asselbergh B, Van Broeckhoven C. Rare missense mutations in ABCA7 might increase Alzheimer's disease risk by plasma membrane exclusion. Acta Neuropathol Commun. 2022;10:43.
- 48. Biffi A, Shulman JM, Jagiella JM, et al. Genetic variation at CR1 increases risk of cerebral amyloid angiopathy. Neurology. 2012;78:334-341.
- 49. Sawyer RP, Demel SL, Comeau ME, et al. Alzheimer's disease related single nucleotide polymorphisms and correlation with intracerebral hemorrhage incidence. Medicine (Baltimore). 2022;101:e30782.
- 50. Koemans EA, Voigt S, Rasing I, et al. Migraine with aura as early disease marker in hereditary Dutch-type cerebral amyloid angiopathy. Stroke. 2020;51:1094-1099.
- 51. Wattendorff AR, Bots GT, Went LN, Endtz LJ. Familial cerebral amyloid angiopathy presenting as recurrent cerebral haemorrhage. J Neurol Sci. 1982;55:121-135.
- 52. van Rooden S, van Opstal AM, Labadie G, et al. Early magnetic resonance imaging and cognitive markers of hereditary cerebral amyloid angiopathy. Stroke. 2016;47:3041-3044.
- 53. Joseph-Mathurin N, Wang G, Kantarci K, et al. Longitudinal accumulation of cerebral microhemorrhages in dominantly inherited Alzheimer disease. Neurology. 2021;96:e1632-e1645.
- 54. van Rooden S, van der Grond J, van den Boom R, et al. Descriptive analysis of the Boston criteria applied to a Dutch-type cerebral amyloid angiopathy population. Stroke. 2009;40:3022-3027.
- 55. van den Boom R, Bornebroek M, Behloul F, van den Berg-Huysmans AA, Haan J, van Buchem MA. Microbleeds in hereditary cerebral hemorrhage with amyloidosis-Dutch type. Neurology. 2005;64:1288-1289.
- 56. Fotiadis P, van Rooden S, van der Grond J, et al. Cortical atrophy in patients with cerebral amyloid angiopathy: A case-control study. Lancet Neurol. 2016;15:811-819.
- 57. Schouten TM, de Vos F, van Rooden S, et al. Multiple approaches to diffusion magnetic resonance imaging in hereditary cerebral amyloid angiopathy mutation carriers. J Am Heart Assoc. 2019;8:e011288.
- 58. van Opstal AM, van Rooden S, van Harten T, et al. Cerebrovascular function in presymptomatic and symptomatic individuals with hereditary cerebral amyloid angiopathy: A case-control study. Lancet Neurol. 2017;16:115-122.
- 59. Koemans EA, van Etten ES, van Opstal AM, et al. Innovative magnetic resonance imaging markers of hereditary cerebral amyloid angiopathy at 7 tesla. Stroke. 2018;49: 1518-1520.
- 60. Bulk M, Moursel LG, van der Graaf LM, et al. Cerebral amyloid angiopathy with vascular iron accumulation and calcification. Stroke. 2018;49:2081-2087.
- 61. Schultz AP, Kloet RW, Sohrabi HR, et al. Amyloid imaging of Dutch-type hereditary cerebral amyloid angiopathy carriers. Ann Neurol. 2019;86:616-625.
- 62. van Etten ES, Verbeek MM, van der Grond J, et al. beta-Amyloid in CSF: Biomarker for preclinical cerebral amyloid angiopathy. Neurology. 2017;88:169-176.
- 63. Bornebroek M, De Jonghe C, Haan J, et al. Hereditary cerebral hemorrhage with amyloidosis Dutch type (AbetaPP 693): Decreased plasma amyloid-beta 42 concentration. Neurobiol Dis. 2003;14:619-623.
- 64. van Etten ES, de Boer I, Steenmeijer SR, et al. Optical coherence tomography detects retinal changes in hereditary cerebral amyloid angiopathy. Eur J Neurol. 2020;27:2635-2640.
- 65. Hendriks L, van Duijn CM, Cras P, et al. Presenile dementia and cerebral haemorrhage linked to a mutation at codon 692 of the

- beta-amyloid precursor protein gene. Nat Genet. 1992;1: 218-221.
- Roks G, Van Harskamp F, De Koning I, et al. Presentation of amyloidosis in carriers of the codon 692 mutation in the amyloid precursor protein gene (APP692). Brain. 2000;123(Pt 10): 2130-2140.
- 67. Brooks WS, Kwok JB, Halliday GM, et al. Hemorrhage is uncommon in new Alzheimer family with flemish amyloid precursor protein mutation. Neurology. 2004;63:1613-1617.
- Sellal F, Wallon D, Martinez-Almoyna L, et al. APP Mutations in cerebral amyloid angiopathy with or without cortical calcifications: Report of three families and a literature review. J Alzheimer's Dis: JAD. 2017;56:37-46.
- 69. Ryan NS, Biessels GJ, Kim L, et al. Genetic determinants of white matter hyperintensities and amyloid angiopathy in familial Alzheimer's disease. Neurobiol Aging. 2015;36:3140-3151.
- Bugiani O, Giaccone G, Rossi G, et al. Hereditary cerebral hemorrhage with amyloidosis associated with the E693K mutation of APP. Arch Neurol. 2010;67:987-995.
- 71. Kamino K, Orr HT, Payami H, et al. Linkage and mutational analysis of familial Alzheimer disease kindreds for the APP gene region. Am J Hum Genet. 1992;51:998-1014.
- 72. Nilsberth C, Westlind-Danielsson A, Eckman CB, et al. The 'Arctic' APP mutation (E693G) causes Alzheimer's disease by enhanced abeta protofibril formation. Nat Neurosci. 2001;4: 887-893.
- 73. Basun H, Bogdanovic N, Ingelsson M, et al. Clinical and neuropathological features of the Arctic APP gene mutation causing early-onset Alzheimer disease. Arch Neurol. 2008;65:499-505.
- Scholl M, Wall A, Thordardottir S, et al. Low PiB PET retention in presence of pathologic CSF biomarkers in Arctic APP mutation carriers. Neurology. 2012;79:229-236.
- Almkvist O, Axelman K, Basun H, et al. Clinical findings in nondemented mutation carriers predisposed to Alzheimer's disease: A model of mild cognitive impairment. Acta Neurol Scand Suppl. 2003;179:77-82.
- Thordardottir S, Stahlbom AK, Ferreira D, et al. Preclinical cerebrospinal fluid and volumetric magnetic resonance imaging biomarkers in Swedish familial Alzheimer's disease. J Alzheimer's Dis: JAD. 2015;43:1393-1402.
- 77. Grabowski TJ, Cho HS, Vonsattel JP, Rebeck GW, Greenberg SM. Novel amyloid precursor protein mutation in an Iowa family with dementia and severe cerebral amyloid angiopathy. Ann Neurol. 2001;49:697-705.
- 78. Greenberg SM, Shin Y, Grabowski TJ, et al. Hemorrhagic stroke associated with the Iowa amyloid precursor protein mutation. Neurology. 2003;60:1020-1022.
- 79. Mok T, Chalissery AJ, Byrne S, et al. Familial cerebral amyloid angiopathy due to the Iowa mutation in an Irish family. Can J Neurol Sci. 2014;41:512-517.
- Iwanowski P, Kozubski W, Losy J. Iowa-type hereditary cerebral amyloid angiopathy in a Polish family. J Neurol Sci. 2015; 356(1-2):202-204.
- 81. Zarranz JJ, Fernandez-Martinez M, Rodriguez O, Mateos B, Iglesias S, Baron JC. Iowa APP mutation-related hereditary Cerebral Amyloid Angiopathy (CAA): A new family from Spain. J Neurol Sci. 2016;363:55-56.
- 82. Bermejo-Guerrero L, Sánchez-Tejerina D, Sánchez-Tornero M, et al. Low amyloid-PET uptake in Iowa-type cerebral amyloid angiopathy with cerebral venous thrombosis. J Alzheimer's Dis: JAD. 2019;72:677-681.
- 83. Iglesias S, Chapon F, Baron JC. Familial occipital calcifications, hemorrhagic strokes, leukoencephalopathy, dementia, and external carotid dysplasia. Neurology. 2000;55:1661-1667.

- 84. Obici L, Demarchi A, de Rosa G, et al. A novel AbetaPP mutation exclusively associated with cerebral amyloid angiopathy. Ann Neurol. 2005;58:639-644.
- 85. Kozberg MG, van Veluw SJ, Frosch MP, Greenberg SM. Hereditary cerebral amyloid angiopathy, Piedmont-type mutation. Neurol Genet. 2020;6:e411.
- 86. Guerreiro RJ, Gustafson DR, Hardy J. The genetic architecture of Alzheimer's disease: Beyond APP, PSENs and APOE. Neurobiol Aging. 2012;33:437-456.
- 87. Kent SA, Spires-Jones TL, Durrant CS. The physiological roles of tau and Abeta: Implications for Alzheimer's disease pathology and therapeutics. Acta Neuropathol. 2020;140:417-447.
- Maat-Schieman M, Roos R, van Duinen S. Hereditary cerebral hemorrhage with amyloidosis-Dutch type. Neuropathology. 2005;25:288-297.
- 89. Panegyres PK, Kwok JB, Schofield PR, Blumbergs PC. A western Australian kindred with Dutch cerebral amyloid angiopathy. J Neurol Sci. 2005;239:75-80.
- 90. Bornebroek M, Haan J, Maat-Schieman ML, Van Duinen SG, Roos RA. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): I-A review of clinical, radiologic and genetic aspects. Brain Pathol. 1996;6:111-114.
- 91. van Etten ES, Gurol ME, van der Grond J, et al. Recurrent hemorrhage risk and mortality in hereditary and sporadic cerebral amyloid angiopathy. Neurology. 2016;87:1482-1487.
- 92. Haan J, Bakker E, Jennekens-Schinkel A, Roos RA. Progressive dementia, without cerebral hemorrhage, in a patient with hereditary cerebral amyloid angiopathy. Clin Neurol Neurosurg. 1992;94:317-318.
- 93. Bornebroek M, Haan J, Roos RA. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): a review of the variety in phenotypic expression. Amyloid. 1999;6:215-224.
- 94. Natte R, Maat-Schieman ML, Haan J, Bornebroek M, Roos RA, van Duinen SG. Dementia in hereditary cerebral hemorrhage with amyloidosis-Dutch type is associated with cerebral amyloid angiopathy but is independent of plaques and neurofibrillary tangles. Ann Neurol. 2001;50:765-772.
- 95. Maat-Schieman ML, van Duinen SG, Bornebroek M, Haan J, Roos RA. Hereditary cerebral hemorrhage with amyloidosis-Dutch type (HCHWA-D): II-A review of histopathological aspects. Brain Pathol. 1996;6:115-120.
- 96. Chatterjee P, Tegg M, Pedrini S, et al. Plasma amyloid-Beta levels in a Pre-symptomatic Dutch-type hereditary cerebral amyloid angiopathy pedigree: A cross-sectional and longitudinal investigation. Int J Mol Sci. 2021;22:2931.
- 97. Martinez-Ramirez S, van Rooden S, Charidimou A, et al. Perivascular spaces volume in sporadic and hereditary (Dutch-type) cerebral amyloid angiopathy. Stroke. 2018;49:
- 98. Kumar-Singh S, Cras P, Wang R, et al. Dense-core senile plaques in the flemish variant of Alzheimer's disease are vasocentric. Am J Pathol. 2002;161:507-520.
- 99. Cras P, van Harskamp F, Hendriks L, et al. Presenile Alzheimer dementia characterized by amyloid angiopathy and large amyloid core type senile plaques in the APP 692Ala->Gly mutation. Acta Neuropathol. 1998;96:253-260.
- 100. Kalimo H, Lalowski M, Bogdanovic N, et al. The Arctic AbetaPP mutation leads to Alzheimer's disease pathology with highly variable topographic deposition of differentially truncated abeta. Acta Neuropathol Commun. 2013;1:60.
- 101. Philipson O, Lord A, Lalowski M, et al. The Arctic amyloid-beta precursor protein (AbetaPP) mutation results in distinct plaques and accumulation of N- and C-truncated Abeta. Neurobiol Aging. 2012;33:1010 e1-13.

- 102. Nordberg A, Rinne JO, Kadir A, Langstrom B. The use of PET in Alzheimer disease. Nat Rev Neurol. 2010;6:78-87.
- 103. Psychogios K, Xiromerisiou G, Kargiotis O, et al. Hereditary cerebral amyloid angiopathy mimicking CADASIL syndrome. Eur J Neurol. 2021;28:3866-3869.
- 104. Kalampokini S, Georgouli D, Patrikiou E, et al. Tauhe Greek variant in APP gene: The phenotypic Spectrum of APP mutations. Int J Mol Sci. 2021;22:12355.
- 105. Conidi ME, Bernardi L, Puccio G, et al. Homozygous carriers of APP A713T mutation in an autosomal dominant Alzheimer disease family. Neurology. 2015;84:2266-2273.
- 106. Bernardi L, Geracitano S, Colao R, et al. AbetaPP A713T mutation in late onset Alzheimer's disease with cerebrovascular lesions. J Alzheimer's Dis: JAD. 2009;17:383-389.
- 107. Carter DA, Desmarais E, Bellis M, et al. More missense in amyloid gene. Nat Genet. 1992;2:255-256.
- 108. Rossi G, Giaccone G, Maletta R, et al. A family with Alzheimer disease and strokes associated with A713T mutation of the APP gene. Neurology. Sep. 14 2004;63:910-912.
- 109. Koriath C, Kenny J, Adamson G, et al. Predictors for a dementia gene mutation based on gene-panel next-generation sequencing of a large dementia referral series. Mol Psychiatry. 2020;
- 110. Nicolas G, Wallon D, Goupil C, et al. Mutation in the 3'untranslated region of APP as a genetic determinant of cerebral amyloid angiopathy. Eur J Hum Genet. 2016;24:92-98.
- 111. Jaunmuktane Z, Banerjee G, Paine S, et al. Alzheimer's disease neuropathological change three decades after iatrogenic amyloid-beta transmission. Acta Neuropathol. 2021;142: 211-215.
- 112. Rovelet-Lecrux A, Hannequin D, Raux G, et al. APP Locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. Nat Genet. 2006;38: 24-26.
- 113. Sleegers K, Brouwers N, Gijselinck I, et al. APP Duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy. Brain. 2006;129(Pt 11):2977-2983.
- 114. Cabrejo L, Guyant-Marechal L, Laquerriere A, et al. Phenotype associated with APP duplication in five families. Brain. 2006; 129(Pt 11):2966-2976.
- 115. Rovelet-Lecrux A, Frebourg T, Tuominen H, Majamaa K, Campion D, Remes AM. APP Locus duplication in a Finnish family with dementia and intracerebral haemorrhage. J Neurol Neurosurg Psychiatr. 2007;78:1158-1159.
- 116. Llado A, Grau-Rivera O, Sanchez-Valle R, et al. Large APP locus duplication in a sporadic case of cerebral haemorrhage. Neurogenetics. 2014;15:145-149.
- 117. Kasuga K, Shimohata T, Nishimura A, et al. Identification of independent APP locus duplication in Japanese patients with early-onset Alzheimer disease. J Neurol Neurosurg Psychiatr. 2009;80:1050-1052.
- 118. Thonberg H, Fallstrom M, Bjorkstrom J, Schoumans J, Nennesmo I, Graff C. Mutation screening of patients with Alzheimer disease identifies APP locus duplication in a Swedish patient. BMC Res Notes. 2011;4:476.
- 119. Buss L, Fisher E, Hardy J, et al. Intracerebral haemorrhage in down syndrome: Protected or predisposed? F1000Res. 2016;5: F1000 Faculty Rev-876.
- 120. Zarea A, Charbonnier C, Rovelet-Lecrux A, et al. Seizures in dominantly inherited Alzheimer disease. Neurology. 2016;87:
- 121. Guyant-Marechal I, Berger E, Laquerriere A, et al. Intrafamilial diversity of phenotype associated with app duplication. Neurology. 2008;71:1925-1926.

- 122. Neale N, Padilla C, Fonseca LM, Holland T, Zaman S. Neuroimaging and other modalities to assess Alzheimer's disease in down syndrome. *Neuroimage Clin*. 2018;17:263-271.
- 123. Sobey CG, Judkins CP, Sundararajan V, Phan TG, Drummond GR, Srikanth VK. Risk of Major cardiovascular events in people with down syndrome. PLoS One. 2015;10:e0137093.
- 124. Mann DMA, Davidson YS, Robinson AC, et al. Patterns and severity of vascular amyloid in Alzheimer's disease associated with duplications and missense mutations in APP gene, down syndrome and sporadic Alzheimer's disease. Acta Neuropathol. 2018;136:569-587.
- 125. Lao PJ, Gutierrez J, Keator D, et al. Alzheimer-Related Cerebrovascular disease in down syndrome. Ann Neurol. 2020;88:1165-1177.
- 126. Grangeon L, Cassinari K, Rousseau S, et al. Early-Onset cerebral amyloid angiopathy and Alzheimer disease related to an APP locus triplication. *Neurol Genet*. 2021;7:e609.
- 127. Selkoe DJ, Wolfe MS. Presenilin: Running with scissors in the membrane. *Cell*. 2007;131:215-221.
- 128. AlzForum Alzheimer's Disease Mutation Database. Accessed 14 June 2021. https://www.alzforum.org/mutations
- 129. Tang M, Ryman DC, McDade E, et al. Neurological manifestations of autosomal dominant familial Alzheimer's disease: a comparison of the published literature with the Dominantly Inherited Alzheimer Network observational study (DIAN-OBS). Lancet Neurol. 2016;15:1317-1325.
- 130. Voglein J, Paumier K, Jucker M, et al. Clinical, pathophysiological and genetic features of motor symptoms in autosomal dominant Alzheimer's disease. Brain. 2019;142:1429-1440.
- 131. Ryan NS, Nicholas JM, Weston PSJ, et al. Clinical phenotype and genetic associations in autosomal dominant familial Alzheimer's disease: A case series. Lancet Neurol. 2016;15: 1326-1335.
- 132. Wallon D, Rousseau S, Rovelet-Lecrux A, et al. The French series of autosomal dominant early onset Alzheimer's disease cases: mutation spectrum and cerebrospinal fluid biomarkers. *J Alzheimer's* Dis. 2012;30:847-856.
- 133. Ryman DC, Acosta-Baena N, Aisen PS, et al. Symptom onset in autosomal dominant Alzheimer disease: a systematic review and meta-analysis. Neurology. 2014;83:253-260.
- 134. Mann DM, Pickering-Brown SM, Takeuchi A, Iwatsubo T. Members of the familial Alzheimer's Disease Pathology Study G. Amyloid angiopathy and variability in amyloid beta deposition is determined by mutation position in presenilin-1-linked Alzheimer's disease. *Am J Pathol.* 2001; 158:2165-2175.
- 135. Ringman JM, Monsell S, Ng DW, et al. Neuropathology of autosomal dominant Alzheimer disease in the national Alzheimer coordinating center database. J Neuropathol Exp Neurol. 2016;75: 284-290.
- 136. Dumanchin C, Tournier I, Martin C, et al. Biological effects of four PSEN1 gene mutations causing Alzheimer disease with spastic paraparesis and cotton wool plaques. Hum Mutat. 2006;27:1063.
- 137. Sanchez-Valle R, Llado A, Ezquerra M, Rey MJ, Rami L, Molinuevo JL. A novel mutation in the PSEN1 gene (L286P) associated with familial early-onset dementia of Alzheimer type and lobar haematomas. Eur J Neurol. 2007;14:1409-1412.
- 138. Lee S, Viqar F, Zimmerman ME, et al. White matter hyperintensities are a core feature of Alzheimer's disease: Evidence from the dominantly inherited Alzheimer network. Ann Neurol. 2016;79:929-939.
- 139. Lee S, Zimmerman ME, Narkhede A, et al. White matter hyperintensities and the mediating role of cerebral amyloid

- angiopathy in dominantly-inherited Alzheimer's disease. PLoS One. 2018;13:e0195838.
- 140. Floris G, Di Stefano F, Cherchi MV, Costa G, Marrosu F, Marrosu MG. Multiple spontaneous cerebral microbleeds and leukoencephalopathy in PSEN1-associated familial Alzheimer's disease: Mirror of cerebral amyloid angiopathy? *J Alzheimer's Dis: JAD.* 2015;47:535-538.
- 141. Ringman JM, Pope W, Salamon N. Insensitivity of visual assessment of hippocampal atrophy in familial Alzheimer's disease. *J Neurol.* 2010;257:839-842.
- 142. Ridha BH, Barnes J, Bartlett JW, et al. Tracking atrophy progression in familial Alzheimer's disease: a serial MRI study. *Lancet Neurol.* 2006;5:828-834.
- 143. Jayadev S, Leverenz JB, Steinbart E, et al. Alzheimer's disease phenotypes and genotypes associated with mutations in presenilin 2. *Brain*. 2010;133(Pt 4):1143-1154.
- 144. Nochlin D, Bird TD, Nemens EJ, Ball MJ, Sumi SM. Amyloid angiopathy in a Volga German family with Alzheimer's disease and a presentilin-2 mutation (N141I). Ann Neurol. 1998;43: 131-135.
- 145. Canevelli M, Piscopo P, Talarico G, et al. Familial Alzheimer's disease sustained by presenilin 2 mutations: Systematic review of literature and genotype-phenotype correlation. Neurosci Biobehav Rev. 2014;42:170-179.
- 146. Rostagno A, Tomidokoro Y, Lashley T, et al. Chromosome 13 dementias. Cell Mol Life Sci. 2005;62:1814-1825.
- 147. Kim SH, Wang R, Gordon DJ, et al. Furin mediates enhanced production of fibrillogenic ABri peptides in familial British dementia. Nat Neurosci. 1999;2:984-988.
- 148. Kim SH, Wang R, Gordon DJ, et al. Familial British dementia: Expression and metabolism of BRI. Ann N Y Acad Sci. 2000; 920:93-99.
- 149. Worster-Drought C, Hill TR, McMenemey WH. Familial presenile dementia with spastic paralysis. *J Neurol Psychopathol*. 1933;14:27-34.
- 150. Vidal R, Frangione B, Rostagno A, et al. A stop-codon mutation in the BRI gene associated with familial British dementia. *Nature*. 1999;399:776-781.
- 151. Ghiso JA, Holton J, Miravalle L, et al. Systemic amyloid deposits in familial British dementia. J Biol Chem. 2001;276:43909-43914.
- 152. Mead S, James-Galton M, Revesz T, et al. Familial British dementia with amyloid angiopathy: Early clinical, neuropsychological and imaging findings. Brain. 2000;123(Pt 5):975-991.
- 153. Plant GT, Revesz T, Barnard RO, Harding AE, Gautier-Smith PC. Familial cerebral amyloid angiopathy with nonneuritic amyloid plaque formation. Brain. 1990;113(Pt 3):721-747.
- 154. Harris MJ, Lane CA, Coath W, et al. Familial British dementia: a clinical and multi-modal imaging case study. *J Neurol.* 2022; 269:3926-3930.
- 155. Holton JL, Ghiso J, Lashley T, et al. Regional distribution of amyloid-Bri deposition and its association with neurofibrillary degeneration in familial British dementia. Am J Pathol. 2001; 158:515-526
- 156. Stromgren E, Dalby A, Dalby MA, Ranheim B. Cataract, deafness, cerebellar ataxia, psychosis and dementia–a new syndrome. Acta Neurol Scand. 1970;46(S43):261-262.
- 157. Vidal R, Revesz T, Rostagno A, et al. A decamer duplication in the 3' region of the BRI gene originates an amyloid peptide that is associated with dementia in a danish kindred. Proc Natl Acad Sci U S A. 2000;97:4920-4925.
- 158. Holton JL, Lashley T, Ghiso J, et al. Familial danish dementia: A novel form of cerebral amyloidosis associated with deposition of both amyloid-Dan and amyloid-beta. J Neuropathol Exp Neurol. 2002;61:254-267.

- 159. Liu X, Chen KL, Wang Y, et al. A novel ITM2B mutation associated with familial Chinese dementia. J Alzheimer's Dis: JAD. 2021:81:499-505
- 160. Gudmundsson G, Hallgrimsson J, Jonasson TA, Bjarnason O. Hereditary cerebral haemorrhage with amyloidosis. Brain. 1972:95:387-404.
- 161. Jensson O, Gudmundsson G, Arnason A, et al. Hereditary cystatin C (gamma-trace) amyloid angiopathy of the CNS causing cerebral hemorrhage. Acta Neurol Scand. 1987;76:102-114.
- 162. Palsdottir A, Abrahamson M, Thorsteinsson L, et al. Mutation in cystatin C gene causes hereditary brain haemorrhage. Lancet. 1988;2:603-604.
- 163. Palsdottir A, Snorradottir AO, Thorsteinsson L. Hereditary cystatin C amyloid angiopathy: Genetic, clinical, and pathological aspects. Brain Pathol. 2006;16:55-59.
- 164. Olafsson I, Grubb A. Hereditary cystatin C amyloid angiopathy. Amyloid. 2000;7:70-79.
- 165. Blondal H, Guomundsson G, Benedikz E, Johannesson G. Dementia in hereditary cystatin C amyloidosis. Prog Clin Biol Res. 1989;317:157-164.
- 166. March ME, Gutierrez-Uzquiza A, Snorradottir AO, et al. NAC Blocks cystatin C amyloid complex aggregation in a cell system and in skin of HCCAA patients. Nat Commun. 2021;12:1827.
- 167. Palsdottir A, Helgason A, Palsson S, et al. A drastic reduction in the life span of cystatin C L68Q carriers due to life-style changes during the last two centuries. PLoS Genet. 2008;4:e1000099.
- 168. Jensson O, Palsdottir A, Thorsteinsson L, Arnason A. The saga of cystatin C gene mutation causing amyloid angiopathy and brain hemorrhage-clinical genetics in Iceland. Clin Genet. 1989;36:368-377.
- 169. Revesz T, Holton JL, Lashley T, et al. Sporadic and familial cerebral amyloid angiopathies. Brain Pathol. 2002;12:343-357.
- 170. Sveinbjornsdottir S, Blondal H, Gudmundsson G, Kjartansson O, Jonsdottir S, Gudmundsson G. Progressive dementia and leucoencephalopathy as the initial presentation of late onset hereditary cystatin-C amyloidosis. Clinicopathological presentation of two cases. J Neurol Sci. 1996;140(1-2):101-108.
- 171. Solomon JP, Page LJ, Balch WE, Kelly JW. Gelsolin amyloidosis: genetics, biochemistry, pathology and possible strategies for therapeutic intervention. Crit Rev Biochem Mol Biol. 2012;47:282-296.
- 172. Schmidt EK, Mustonen T, Kiuru-Enari S, Kivela TT, Atula S. Finnish Gelsolin amyloidosis causes significant disease burden but does not affect survival: FIN-GAR phase II study. Orphanet J Rare Dis. 2020;15:19.
- 173. Cabral-Macias J, Garcia-Montano LA, Perezpena-Diazconti M, et al. Clinical, histopathological, and in silico pathogenicity analyses in a pedigree with familial amyloidosis of the Finnish type (meretoja syndrome) caused by a novel gelsolin mutation. Mol Vis. 2020;26:345-354.
- 174. Mullany S, Souzeau E, Klebe S, et al. A novel GSN variant outside the G2 calcium-binding domain associated with Amyloidosis of the Finnish type. Hum Mutat. 2021;42:818-826.
- 175. Potrc M, Volk M, de Rosa M, et al. Clinical and histopathological features of gelsolin amyloidosis associated with a novel GSN variant p.Glu580Lys. Int J Mol Sci. 2021;22:1084.
- 176. Kantanen M, Kiuru-Enari S, Salonen O, Kaipainen M, Hokkanen L. Subtle neuropsychiatric and neurocognitive changes in hereditary gelsolin amyloidosis (AGel amyloidosis). PeerJ. 2014;2:e493.
- 177. Kiuru-Enari S, Haltia M. Hereditary gelsolin amyloidosis. Handb Clin Neurol. 2013;115:659-681.
- 178. Kiuru S, Seppäläinen A-M, Salonen O, Hokkanen L, Somer H, Palo J. CNS Abnormalities in patients with familial amyloidosis, Finnish type (FAF). Amyloid. 2009;2:22-30.

- 179. Cheong EN, Paik W, Choi YC, et al. Clinical features and brain MRI findings in Korean patients with AGel amyloidosis. Yonsei Med J. 2021;62:431-438.
- 180. Kiuru S. Gelsolin-related familial amyloidosis, Finnish type (FAF), and its variants found worldwide. Amyloid. 1998;5:55-66.
- 181. Koskelainen S, Pihlamaa T, Suominen S, et al. Gelsolin amyloid angiopathy causes severe disruption of the arterial wall. APMIS. 2016;124:639-648.
- 182. Kiuru S, Salonen O, Haltia M. Gelsolin-related spinal and cerebral amyloid angiopathy. Ann Neurol. 1999;45:305-311.
- 183. Makishita H, Ikeda S-I, Yazaki M, et al. Postmortem pathological findings in a Japanese patient with familial amyloidosis, Finnish type (FAF). Amyloid. 2009;3:134-139.
- 184. Mead S, Lloyd S, Collinge J. Genetic factors in mammalian prion diseases. Annu Rev Genet. 2019;53:117-147.
- 185. Appleby BS, Rhoads DD, Mente K, Cohen ML. A practical primer on prion pathology. J Neuropathol Exp Neurol. 2018;77: 346-352.
- 186. Rostagno A, Holton JL, Lashley T, Revesz T, Ghiso J. Cerebral amyloidosis: Amyloid subunits, mutants and phenotypes. Cell Mol Life Sci. 2010;67:581-600.
- 187. Revesz T, Ghiso J, Lashley T, et al. Cerebral amyloid angiopathies: A pathologic, biochemical, and genetic view. J Neuropathol Exp Neurol. 2003;62:885-898.
- 188. Honda H, Matsuzono K, Fushimi S, et al. C-Terminal-Deleted prion protein fragment is a Major accumulated component of systemic PrP deposits in hereditary prion disease with a 2-bp (CT) deletion in PRNP Codon 178. J Neuropathol Exp Neurol. 2016;75:1008-1019.
- 189. Ghetti B, Piccardo P, Spillantini MG, et al. Vascular variant of prion protein cerebral amyloidosis with tau-positive neurofibrillary tangles: the phenotype of the stop codon 145 mutation in PRNP. Proc Natl Acad Sci U S A. 1996;93:744-748.
- 190. Kitamoto T, Iizuka R, Tateishi J. An amber mutation of prion protein in gerstmann-straussler syndrome with mutant PrP plaques. Biochem Biophys Res Commun. 1993;192:525-531.
- 191. Ghetti B, Piccardo P, Frangione B, et al. Prion protein amyloidosis. Brain Pathol. 1996;6:127-145.
- 192. Finckh U, Muller-Thomsen T, Mann U, et al. High frequency of mutations in four different disease genes in early-onset dementia. Ann N Y Acad Sci. 2000;920:100-106.
- 193. Jayadev S, Nochlin D, Poorkaj P, et al. Familial prion disease with Alzheimer disease-like tau pathology and clinical phenotype. Ann Neurol. 2011;69:712-720.
- 194. Guerreiro R, Bras J, Wojtas A, Rademakers R, Hardy J, Graff-Radford N. Nonsense mutation in PRNP associated with clinical Alzheimer's disease. Neurobiol Aging. 2014;35:2656 e13-2656 e16.
- 195. Mead S, Gandhi S, Beck J, et al. A novel prion disease associated with diarrhea and autonomic neuropathy. N Engl J Med. 2013;
- 196. Capellari S, Baiardi S, Rinaldi R, et al. Two novel PRNP truncating mutations broaden the spectrum of prion amyloidosis. Ann Clin Transl Neurol. 2018;5:777-783.
- 197. Jansen C, Parchi P, Capellari S, et al. Prion protein amyloidosis with divergent phenotype associated with two novel nonsense mutations in PRNP. Acta Neuropathol. 2010;119:189-197.
- 198. Matsuzono K, Honda H, Sato K, et al. 'Prp systemic deposition disease': clinical and pathological characteristics of novel familial prion disease with 2-bp deletion in codon 178. Eur J Neurol. 2016;23:196-200.
- 199. Matsuzono K, Ikeda Y, Liu W, et al. A novel familial prion disease causing pan-autonomic-sensory neuropathy and cognitive impairment. Eur J Neurol. 2013;20:e67-e69.

- 200. Liz MA, Coelho T, Bellotti V, Fernandez-Arias MI, Mallaina P, Obici L. A narrative review of the role of transthyretin in health and disease. *Neurol Ther.* 2020;9:395-402.
- 201. Sekijima Y. Transthyretin (ATTR) amyloidosis: clinical spectrum, molecular pathogenesis and disease-modifying treatments. *J Neurol Neurosurq Psychiatr*. 2015;86:1036-1043.
- 202. Pinheiro F, Varejao N, Esperante S, et al. Tolcapone, a potent aggregation inhibitor for the treatment of familial leptomeningeal amyloidosis. FEBS J. 2021;288:310-324.
- 203. Goren H, Steinberg MC, Farboody GH. Familial oculoleptomeningeal amyloidosis. *Brain*. 1980;103:473-495.
- 204. Sekijima Y. Hereditary transthyretin amyloidosis. In: Adam MP, Everman DB, Mirzaa GM, et al. eds. Genereviews((R)). University of Washington; 1993.
- 205. Garzuly F, Vidal R, Wisniewski T, Brittig F, Budka H. Familial meningocerebrovascular amyloidosis, Hungarian type, with mutant transthyretin (TTR Asp18Gly). Neurology. 1996;47: 1562-1567.
- 206. Herrick MK, DeBruyne K, Horoupian DS, Skare J, Vanefsky MA, Ong T. Massive leptomeningeal amyloidosis associated with a Val30Met transthyretin gene. *Neurology*. 1996;47:988-992.
- 207. McColgan P, Viegas S, Gandhi S, et al. Oculoleptomeningeal amyloidosis associated with transthyretin Leu12Pro in an African patient. *J Neurol*. 2015;262:228-234.
- 208. Martin SE, Benson MD, Hattab EM. The pathologic spectrum of oculoleptomeningeal amyloidosis with Val30Gly transthyretin gene mutation in a postmortem case. *Hum Pathol.* 2014;45: 1105-1108.
- 209. Brett M, Persey MR, Reilly MM, et al. Transthyretin Leu12Pro is associated with systemic, neuropathic and leptomeningeal amyloidosis. Brain. 1999;122(Pt 2):183-190.
- Sekijima Y, Yazaki M, Oguchi K, et al. Cerebral amyloid angiopathy in posttransplant patients with hereditary ATTR amyloidosis. Neurology. 2016;87:773-781.
- 211. Suhr OB, Larsson M, Ericzon BG, Wilczek HE, investigators FAs. Survival after transplantation in patients with mutations other than Val30Met: Extracts from the FAP world transplant registry. Transplantation. 2016;100:373-381.
- 212. Salvi F, Pastorelli F, Plasmati R, et al. Brain microbleeds 12 years after orthotopic liver transplantation in Val30Met amyloidosis. *J Stroke cerebrovascr Dis.* 2015;24:e149-e151.
- 213. De Carolis P, Galeotti M, Ficarra G, Masetti M, Grimaldi D, Cortelli P. Fatal cerebral haemorrhage after liver transplantation in a patient with transthyretin variant (gly53glu) amyloidosis. Neurol Sci. 2006;27:352-354.
- 214. Uitti RJ, Donat JR, Rozdilsky B, Schneider RJ, Koeppen AH. Familial oculoleptomeningeal amyloidosis. Report of a new family with unusual features. Arch Neurol. 1988;45:1118-1122.
- 215. Mascalchi M, Salvi F, Pirini MG, et al. Transthyretin amyloidosis and superficial siderosis of the CNS. Neurology. 1999;53: 1498-1503.
- 216. Kleefeld F, Knebel F, Eurich D, et al. Familial oculoleptomeningeal transthyretin amyloidosis caused by Leu55Arg mutation. *J Neuromuscul Dis.* 2020;7:515-519.
- 217. Mathieu F, Morgan E, So J, Munoz DG, Mason W, Kongkham P. Oculoleptomeningeal amyloidosis secondary to the rare transthyretin c.381T > G (p.Ile127Met) mutation. World Neurosurg. 2018;111:190-193.
- 218. Bevers MB, McGuone D, Jerath NU, Musolino PL. Leptomeningeal transthyretin-type amyloidosis presenting as acute hydrocephalus and subarachnoid hemorrhage. *J Clin Neurosci.* 2016;29:203-205.
- 219. Ziskin JL, Greicius MD, Zhu W, Okumu AN, Adams CM, Plowey ED. Neuropathologic analysis of Tyr69His TTR variant

- meningovascular amyloidosis with dementia. Acta Neuropathol Commun. 2015;3:43.
- 220. Salvi F, Volpe R, Pastorelli F, et al. Failure of tafamidis to halt progression of Ala36Pro TTR oculomeningovascular amyloidosis. J Stroke Cerebrovascr Dis. 2018;27:e212-e214.
- 221. Jin K, Sato S, Takahashi T, et al. Familial leptomeningeal amyloidosis with a transthyretin variant Asp18Gly representing repeated subarachnoid haemorrhages with superficial siderosis. *J Neurol Neurosurg Psychiatr.* 2004;75:1463-1466.
- 222. Urban PP, Hertkorn C, Schattenberg JM, et al. Leptomeningeal familial amyloidosis: A rare differential diagnosis of leptomeningeal enhancement in MRI. J Neurol. 2006;253:1238-1240.
- 223. Purro SA, Farrow MA, Linehan J, et al. Transmission of amyloidbeta protein pathology from cadaveric pituitary growth hormone. Nature. 2018;564:415-419.
- 224. Collinge J. Mammalian prions and their wider relevance in neurodegenerative diseases. *Nature*. 2016;539:217-226.
- 225. Lauwers E, Lalli G, Brandner S, et al. Potential human transmission of amyloid beta pathology: Surveillance and risks. *Lancet Neurol.* 2020;19:872-878.
- 226. Banerjee G, Samra K, Adams ME, et al. Iatrogenic cerebral amyloid angiopathy: An emerging clinical phenomenon. *J Neurol Neurosurg Psychiatr*. 2022;93:693-700.
- 227. Ehling R, Helbok R, Beer R, et al. Recurrent intracerebral haemorrhage after coitus: A case report of sporadic cerebral amyloid angiopathy in a younger patient. Lett Eur J Neurol. 2012;19: e29-e31.
- 228. Nakayama Y, Mineharu Y, Arawaka Y, et al. Cerebral amyloid angiopathy in a young man with a history of traumatic brain injury: A case report and review of the literature. Acta Neurochir (Wien). 2017;159:15-18.
- 229. Herve D, Porche M, Cabrejo L, et al. Fatal Abeta cerebral amyloid angiopathy 4 decades after a dural graft at the age of 2 years. Acta Neuropathol. 2018;135:801-803.
- 230. Jaunmuktane Z, Quaegebeur A, Taipa R, et al. Evidence of amyloid-beta cerebral amyloid angiopathy transmission through neurosurgery. Acta Neuropathol. 2018;135:671-679.
- 231. Banerjee G, Adams ME, Jaunmuktane Z, et al. Early onset cerebral amyloid angiopathy following childhood exposure to cadaveric dura. Ann Neurol. 2019;85:284-290.
- 232. Giaccone G, Maderna E, Marucci G, et al. Iatrogenic early onset cerebral amyloid angiopathy 30 years after cerebral trauma with neurosurgery: Vascular amyloid deposits are made up of both Abeta40 and Abeta42. Acta Neuropathol Commun. 2019;7:70.
- 233. Hamaguchi T, Komatsu J, Sakai K, *et al*. Cerebral hemorrhagic stroke associated with cerebral amyloid angiopathy in young adults about 3 decades after neurosurgeries in their infancy. *J Neurol Sci.* 2019;399:3-5.
- 234. Caroppo P, Marucci G, Maccagnano E, et al. Cerebral amyloid angiopathy in a 51-year-old patient with embolization by dura mater extract and surgery for nasopharyngeal angiofibroma at age 17. Amyloid. 2021;28:142-143.
- 235. Raposo N, Planton M, Siegfried A, et al. Amyloid-beta transmission through cardiac surgery using cadaveric dura mater patch. J Neurol Neurosurg Psychiatr. 2020;91:440-441.
- 236. Tachiyama K, Nakamori M, Hayashi Y, et al. Infant critical head injury could be a remote cause of middle-aged cerebral amyloid angiopathy. *Interdiscip Neurosurg*. 2020;22:100794.
- 237. Yoshiki K, Hirose G, Kumahashi K, et al. Follow-up study of a patient with early onset cerebral amyloid angiopathy following childhood cadaveric dural graft. Acta Neurochir (Wien). 2021;163:1451-1455.
- 238. Michiels L, Van Weehaeghe D, Vandenberghe R, Demeestere J, Van Laere K, Lemmens R. The role of amyloid PET in

- diagnosing possible transmissible cerebral amyloid angiopathy in young adults with a history of neurosurgery: a case series. Cerebrovasc Dis. 2021;50:356-360.
- 239. Corovic A, Kelly S, Markus HS. Cerebral amyloid angiopathy associated with inflammation: A systematic review of clinical and imaging features and outcome. Int J Stroke. 2018;13: 257-267.
- 240. Chwalisz BK. Cerebral amyloid angiopathy and related inflammatory disorders. J Neurol Sci. 2021;424:117425.
- 241. Antolini L, DiFrancesco JC, Zedde M, et al. Spontaneous ARIA-like events in cerebral amyloid angiopathy-related inflammation: A multicenter prospective longitudinal cohort study. Neurology. 2021;97:e1809-e1822.
- 242. Ryan NS, Lashley T, Revesz T, Dantu K, Fox NC, Morris HR. Spontaneous ARIA (amyloid-related imaging abnormalities) and cerebral amyloid angiopathy related inflammation in presenilin 1-associated familial Alzheimer's disease. J Alzheimer's Dis: JAD. 2015;44:1069-1074.
- 243. Chamard L, Wallon D, Pijoff A, et al. Amyloid-related imaging abnormalities in AbetaPP duplication carriers. J Alzheimer's Dis: JAD. 2013;37:789-793.
- 244. Chung KK, Anderson NE, Hutchinson D, Synek B, Barber PA. Cerebral amyloid angiopathy related inflammation: Three case reports and a review. J Neurol Neurosurg Psychiatr. 2011;
- 245. Auriel E, Charidimou A, Gurol ME, et al. Validation of clinicoradiological criteria for the diagnosis of cerebral amyloid angiopathy-related inflammation. JAMA Neurol. 2016;73:
- 246. Piazza F, Greenberg SM, Savoiardo M, et al. Anti-amyloid beta autoantibodies in cerebral amyloid angiopathy-related inflammation: Implications for amyloid-modifying therapies. Ann Neurol. 2013;73:449-458.
- 247. Boncoraglio GB, Piazza F, Savoiardo M, et al. Prodromal Alzheimer's disease presenting as cerebral amyloid angiopathy-related inflammation with spontaneous amyloid-related imaging abnormalities and high cerebrospinal fluid anti-abeta autoantibodies. J Alzheimer's Dis: JAD. 2015;45:363-367.
- 248. Carmona-Iragui M, Fernandez-Arcos A, Alcolea D, et al. Cerebrospinal fluid anti-amyloid-beta autoantibodies and amyloid PET in cerebral amyloid angiopathy-related inflammation. J Alzheimer's Dis: JAD. 2016;50:1-7.
- 249. Werring DJ, Sperling R. Inflammatory cerebral amyloid angiopathy and amyloid-modifying therapies: Variations on the same ARIA? Ann Neurol. 2013;73:439-441.
- 250. Banerjee G, Alvares D, Bowen J, Adams ME, Werring DJ. Minimally symptomatic cerebral amyloid angiopathy-related inflammation: Three descriptive case reports. J Neurol Neurosurg Psychiatr. 2019;90:113-115.
- 251. Plotzker AS, Henson RL, Fagan AM, Morris JC, Day GS. Clinical and paraclinical measures associated with outcome in cerebral amyloid angiopathy with related inflammation. J Alzheimer's Dis: JAD. 2021;80:133-142.
- 252. Regenhardt RW, Thon JM, Das AS, et al. Association between immunosuppressive treatment and outcomes of cerebral amyloid angiopathy-related inflammation. JAMA Neurol. 2020;77:1261-1269.
- 253. Sperling RA, Jack CR Jr, Black SE, et al. Amyloid-related imaging abnormalities in amyloid-modifying therapeutic trials: Recommendations from the Alzheimer's association research roundtable workgroup. AlzheimersDement. 2011;7:367-385.
- 254. Sperling R, Salloway S, Brooks DJ, et al. Amyloid-related imaging abnormalities in patients with Alzheimer's disease

- treated with bapineuzumab: A retrospective analysis. Lancet Neurol. 2012;11:241-249.
- 255. Mintun MA, Lo AC, Duggan Evans C, et al. Donanemab in early Alzheimer's disease. N Engl J Med. 2021;384:1691-1704.
- 256. Sevigny J, Chiao P, Bussiere T, et al. The antibody aducanumab reduces abeta plaques in Alzheimer's disease. Nature. 2016; 537:50-56.
- 257. Ostrowitzki S, Lasser RA, Dorflinger E, et al. A phase III randomized trial of gantenerumab in prodromal Alzheimer's disease. Alzheimers Res Ther. 2017;9:95.
- 258. Schroder R, Linke RP. Cerebrovascular involvement in systemic AA and AL amyloidosis: a clear haematogenic pattern. Virchows Arch. 1999;434:551-560.
- 259. Mawet J, Adam J, Errera MH, et al. Cerebral immunoglobulin light chain amyloid angiopathy-related hemorrhages. Rev Neurol (Paris). 2009;165:583-587.
- 260. King A, Troakes C, Aizpurua M, et al. Unusual neuropathological features and increased brain aluminium in a resident of Camelford, UK. Neuropathol Appl Neurobiol. 2017;43:537-541.
- 261. Exley C, Esiri MM. Severe cerebral congophilic angiopathy coincident with increased brain aluminium in a resident of camelford, Cornwall, UK. J Neurol Neurosurg Psychiatr. 2006;77:
- 262. Mold M, Cottle J, King A, Exley C. Intracellular aluminium in inflammatory and glial cells in cerebral amyloid angiopathy: a case report. Int J Environ Res Public Health. 2019;16:1459.
- 263. Smith EE, Charidimou A, Ayata C, Werring DJ, Greenberg SM. Cerebral amyloid angiopathy-related transient focal neurologic episodes. Neurology. 2021;97:231-238.
- 264. Keshavan A, Wellington H, Chen Z, et al. Concordance of CSF measures of Alzheimer's pathology with amyloid PET status in a preclinical cohort: A comparison of lumipulse and established immunoassays. Alzheimers Dement (Amst). 2021;13:e12131.
- 265. Jack CR J, Bennett DA, Blennow K, et al. NIA-AA Research framework: Toward a biological definition of Alzheimer's disease. Alzheimers Dement. 2018;14:535-562.
- 266. Zetterberg H, Schott JM. Blood biomarkers for Alzheimer's disease and related disorders. Acta Neurol Scand. 2022;146:51-55.
- 267. Hellstrom-Lindahl E, Westermark P, Antoni G, Estrada S. In vitro binding of [(3)H]PIB to human amyloid deposits of different types. Amyloid. 2014;21:21-27.
- 268. Pilebro B, Arvidsson S, Lindqvist P, et al. Positron emission tomography (PET) utilizing Pittsburgh compound B (PIB) for detection of amyloid heart deposits in hereditary transthyretin amyloidosis (ATTR). J Nucl Cardiol. 2018;25:240-248.
- 269. Charidimou A, Farid K, Baron JC. Amyloid-PET in sporadic cerebral amyloid angiopathy: A diagnostic accuracy meta-analysis. Neurology. 2017;89:1490-1498.
- 270. Schott JM, Reiniger L, Thom M, et al. Brain biopsy in dementia: Clinical indications and diagnostic approach. Acta Neuropathol. 2010;120:327-341.
- 271. Warren JD, Schott JM, Fox NC, et al. Brain biopsy in dementia. Brain. 2005;128(Pt 9):2016-2025.
- 272. Koriath CAM, Kenny J, Ryan NS, et al. Genetic testing in dementia—Utility and clinical strategies. Nat Rev Neurol. 2021;17:
- 273. Al-Shahi Salman R, Minks DP, Mitra D, et al. Effects of antiplatelet therapy on stroke risk by brain imaging features of intracerebral haemorrhage and cerebral small vessel diseases: Subgroup analyses of the RESTART randomised, open-label trial. Lancet Neurol. 2019;18:643-652.
- 274. Schreuder F, van Nieuwenhuizen KM, Hofmeijer J, et al. Apixaban versus no anticoagulation after anticoagulationassociated intracerebral haemorrhage in patients with atrial

- fibrillation in The Netherlands (APACHE-AF): A randomised, open-label, phase 2 trial. Lancet Neurol. 2021;20:907-916.
- 275. So SC. Effects of oral anticoagulation for atrial fibrillation after spontaneous intracranial haemorrhage in the UK: A randomised, open-label, assessor-masked, pilot-phase, noninferiority trial. Lancet Neurol. 2021:20:842-853.
- 276. Cochrane A, Chen C, Stephen J, et al. Antithrombotic treatment after stroke due to intracerebral haemorrhage. Cochrane Database Syst Rev. 2023;1:CD012144.
- 277. Hostettler IC, Wilson D, Fiebelkorn CA, et al. Risk of intracranial haemorrhage and ischaemic stroke after convexity subarachnoid haemorrhage in cerebral amyloid angiopathy: International individual patient data pooled analysis. J Neurol. 2022;269:1427-1438.
- 278. Sanchez-Caro JM, de Lorenzo Martinez de Ubago I, de Celis Ruiz E, et al. Transient focal neurological events in cerebral amyloid angiopathy and the long-term risk of intracerebral hemorrhage and death: A systematic review and meta-analysis. JAMA Neurol. 2022;79:38-47.
- 279. Best JG, Ambler G, Wilson D, et al. Development of imaging-based risk scores for prediction of intracranial haemorrhage and ischaemic stroke in patients taking antithrombotic therapy after ischaemic stroke or transient ischaemic attack: A pooled analysis of individual patient data from cohort studies. Lancet Neurol. 2021;20:294-303.
- 280. PROGRESS Collaborative Group. Randomised trial of a perindopril-based blood-pressure-lowering regimen among 6,105 individuals with previous stroke or transient ischaemic attack. Lancet. 2001;358:1033-1041.
- 281. Arima H, Tzourio C, Anderson C, et al. Effects of perindoprilbased lowering of blood pressure on intracerebral hemorrhage related to amyloid angiopathy: The PROGRESS trial. Stroke. 2010;41:394-396.
- 282. Biffi A, Anderson CD, Battey TWK, et al. Association between blood pressure control and risk of recurrent intracerebral hemorrhage. Jama. 2015;314:904-912.
- 283. Steiner T, Al-Shahi Salman R, Beer R, et al. European Stroke Organisation (ESO) guidelines for the management of spontaneous intracerebral hemorrhage. Int J Stroke. 2014;9: 840-855.
- 284. Greenberg SM, Ziai WC, Cordonnier C, et al. 2022 Guideline for the management of patients with spontaneous intracerebral hemorrhage: A guideline from the American heart association/American stroke association. Stroke. 2022;53:e282-e361.
- 285. Amarenco P, Bogousslavsky J, Callahan A 3rd, et al. High-dose atorvastatin after stroke or transient ischemic attack. N Engl J Med. 2006;355:549-559.
- 286. Heart Protection Study Collaborative Group. MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in  $20,\!536\ high-risk\ individuals:\ a\ randomised\ placebo-controlled$ trial. Lancet. 2002;360:7-22.
- 287. Westover MB, Bianchi MT, Eckman MH, Greenberg SM. Statin use following intracerebral hemorrhage: A decision analysis. Arch Neurol. 2011;68:573-579.
- 288. Ziff OJ, Banerjee G, Ambler G, Werring DJ. Statins and the risk of intracerebral haemorrhage in patients with stroke: Systematic review and meta-analysis. J Neurol Neurosurg Psychiatr. 2019;90:75-83.
- 289. Arvanitakis Z, Shah RC, Bennett DA. Diagnosis and management of dementia: Review. Jama. 2019;322:1589-1599.
- 290. Battle CE, Abdul-Rahim AH, Shenkin SD, Hewitt J, Quinn TJ. Cholinesterase inhibitors for vascular dementia and other vascular cognitive impairments: A network meta-analysis. Cochrane Database Syst Rev. 2021;2:CD013306.

- 291. Smith EE, Cieslak A, Barber P, et al. Therapeutic strategies and drug development for vascular cognitive impairment. J Am Heart Assoc. 2017;6:e005568.
- 292. Paterson RW, Abdi Z, Haines A, Schott JM. Significant cognitive improvement with cholinesterase inhibition in AD with cerebral amyloid angiopathy. Clin Neurol Neurosurg. 2016;144:64-66.
- 293. Bateman RJ, Benzinger TL, Berry S, et al. The DIAN-TU next generation Alzheimer's prevention trial: Adaptive design and disease progression model. Alzheimers Dement. 2017;13:8-19.
- 294. Jensen MP, Ziff OJ, Banerjee G, Ambler G, Werring DJ. The impact of selective serotonin reuptake inhibitors on the risk of intracranial haemorrhage: A systematic review and meta-analysis. Eur Stroke J. 2019;4:144-152.
- 295. Liu L, Fuller M, Behymer TP, et al. Selective serotonin reuptake inhibitors and intracerebral hemorrhage risk and outcome. Stroke. 2020;51:1135-1141.
- 296. Kubiszewski P, Sugita L, Kourkoulis C, et al. Association of selective serotonin reuptake inhibitor use after intracerebral hemorrhage with hemorrhage recurrence and depression severity. JAMA Neurol. 2020;78:1-8.
- 297. Carare RO, Hawkes CA, Jeffrey M, Kalaria RN, Weller RO. Review: Cerebral amyloid angiopathy, prion angiopathy, CADASIL and the spectrum of Protein Elimination Failure Angiopathies (PEFA) in neurodegenerative disease with a focus on therapy. Neuropathol Appl Neurobiol. 2013;39:593-611.
- 298. Carare RO, Aldea R, Agarwal N, et al. Clearance of interstitial fluid (ISF) and CSF (CLIC) group-part of Vascular Professional Interest Area (PIA): Cerebrovascular disease and the failure of elimination of Amyloid-beta from the brain and retina with age and Alzheimer's disease-Opportunities for Therapy. Alzheimers Dement (Amst). 2020;12:e12053.
- 299. Rannikmae K, Kalaria RN, Greenberg SM, et al. APOE Associations with severe CAA-associated vasculopathic changes: Collaborative meta-analysis. J Neurol Neurosurg Psychiatr. 2014;85:300-305.
- 300. Ye X, Li G, Liu X, et al. Apolipoprotein E genotype predicts subarachnoid extension in spontaneous intracerebral haemorrhage. Eur J Neurol. 2021;28:1992-1999.
- 301. Goldberg TE, Huey ED, Devanand DP. Associations of APOE e2 genotype with cerebrovascular pathology: A postmortem study of 1275 brains. J Neurol Neurosurg Psychiatry. 2021;92:7-11.
- 302. Charidimou A, Zonneveld HI, Shams S, et al. APOE And cortical superficial siderosis in CAA: Meta-analysis and potential mechanisms. Neurology. 2019;93:e358-e371.
- 303. Thal DR, Ghebremedhin E, Rub U, Yamaguchi H, Del Tredici K, Braak H. Two types of sporadic cerebral amyloid angiopathy. J Neuropathol Exp Neurol. 2002;61:282-293.
- 304. Attems J, Jellinger KA. Only cerebral capillary amyloid angiopathy correlates with Alzheimer pathology-a pilot study. Acta Neuropathol. 2004;107:83-90.
- 305. Willumsen N, Poole T, Nicholas JM, Fox NC, Ryan NS, Lashley T. Variability in the type and layer distribution of cortical abeta pathology in familial Alzheimer's disease. Brain Pathol. 2022; 32:e13009.
- 306. Hatami A, Monjazeb S, Milton S, Glabe CG. Familial Alzheimer's disease mutations within the amyloid precursor protein Alter the aggregation and conformation of the amyloid-beta peptide. J Biol Chem. 2017;292:3172-3185.
- 307. Miravalle L, Tokuda T, Chiarle R, et al. Substitutions at codon 22 of Alzheimer's abeta peptide induce diverse conformational changes and apoptotic effects in human cerebral endothelial cells. J Biol Chem. 2000;275:27110-6.
- 308. Collinge J, Clarke AR. A general model of prion strains and their pathogenicity. Science. 2007;318:930-936.

- 309. Qiang W, Yau WM, Lu JX, Collinge J, Tycko R. Structural variation in amyloid-beta fibrils from Alzheimer's disease clinical subtypes. *Nature*. 2017;541:217-221.
- 310. Yang Y, Arseni D, Zhang W, et al. Cryo-EM structures of amyloid-beta 42 filaments from human brains. Science. 2022; 375:167-172.
- 311. Mazinani N, Strilchuk AW, Baylis JR, Hur WS, Jefferies WA, Kastrup CJ. Bleeding is increased in amyloid precursor protein knockout mouse. Res Pract Thromb Haemost. 2020;4:823-828.
- 312. Cortes-Canteli M, Paul J, Norris EH, et al. Fibrinogen and beta-amyloid association alters thrombosis and fibrinolysis: a possible contributing factor to Alzheimer's disease. *Neuron*. 2010;66:695-709.
- 313. Cajamarca SA, Norris EH, van der Weerd L, Strickland S, Ahn HJ. Cerebral amyloid angiopathy-linked beta-amyloid mutations promote cerebral fibrin deposits via increased binding affinity for fibrinogen. Proc Natl Acad Sci U S A. 2020;117: 14482-14492.