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Low diagnostic accuracy of Xpert MTB/RIF assay for extrapulmonary tuberculosis: A multicenter surveillance

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Diagnostic accuracy of Xpert MTB/RIF assay for pulmonary tuberculosis (PTB) and extrapulmonary TB (EPTB) has not been investigated in Iran. This study was aimed to assess the diagnostic accuracy of Xpert MTB/RIF assay for both PTB and EPTB. A total of 2111 clinical samples (1218 pulmonary and 838 extra-pulmonary) were collected from 16 medical centers during the study period and were analyzed for detection of PTB and EPTB by both Xpert MTB/RIF assay and standard conventional methods (culture and direct smear microscopy). The sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) of Xpert MTB/RIF assay for PTB were found to be 95.5%, 96.7%, 83.8%, and 99.1% respectively. For EPTB, the sensitivity, specificity, PPV and NPV of Xpert MTB/RIF assay counted for 76.5%, 95.9%, 62%, and 97.9% respectively. Xpert MTB/RIF assay found to be highly sensitive, specific and comparable to standard conventional methods for the diagnosis of PTB. However, the sensitivity and specificity of Xpert MTB/RIF for EPTB specimens were highly variable; thus, Xpert MTB/RIF cannot be recommended to replace standard conventional tests for diagnosis of EPTB.

Tuberculosis (TB) still remains a public health problem with an increasing death rate worldwide, especially in developing countries¹. Early detection of bacilli in clinical samples and starting sufficient treatment is extremely important to reduce the death rate².³. Acid-fast bacilli microscopy and culture are still the cornerstones of TB diagnosis⁴. Although culture is a gold standard technique but is cumbersome and time-consuming⁵.⁶. Likewise, the microscopy, although rapid and inexpensive, but its sensitivity is variable (20–80%)⁶. In the context of these limitations, Xpert MTB/RIF assay, a fully automated real-time semi-nested PCR system was endorsed by World Health Organization (WHO) as the most rapid test for diagnosis of pulmonary TB (PTB)⁷. Extrapulmonary TB (EPTB) accounts for more than 20% of all TB cases, and even higher percentages in HIV-infected persons¹.⅙. Differently from PTB, the diagnosis of EPTB is still a serious problem and existing tests are limited in accuracy⁶. Given the limitations of tests for EPTB detection, Xpert MTB/RIF has been evaluated in several studies⁶-1⁵. Although Xpert MTB/RIF assay has been validated for TB detection in sputum by authors in different countries, the efficacy of this automated molecular platform for TB detection in nonrespiratory specimens was highly variable, as its sensitivity ranged from 30% to 100%¹6-2⁶. Furthermore, the efficacy of Xpert MTB/RIF has not been yet investigated in Iran. Thus, the current study was aimed to comprehensively analyze the diagnostic accuracy of Xpert MTB/RIF assay for PTB and EPTB in Iran.

Methods

Setting and sampling. This cross-sectional study performed from Sep 2015 to Jan 2018 in regional reference laboratory of TB in Tehran, Iran. This center is well-equipped biosafety level III laboratory facilities, and standard biosafety precautions were followed for specimen processing, inoculation, and drug susceptibility testing. The Swedish Institute for Infectious Disease Control monitored and supervised the laboratories' quality. A total of 2111 specimens from TB suspected cases from 6 public hospitals and 10 medical universities from

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Category	Specimen type	Number (%)	Xpert MTB/RIF Error& Invalid	True negative	False negative	True positive	False positive	Xpert Sensitivity (%) with 95% CI	Xpert Specificity (%) with 95% CI	Xpert Positive Predictive Value (%) with 95% CI	Xpert Negative Predictive Value (%) with 95% CI
Pulmonary	BAL	340 (16.5)	4	327	0	3	6	100 (29.2–100)	98.2 (29.2–100)	33.3 (18.4-52.4)	100
	Tracheal	30 (1.4)	0	29	0	1	0	100 (25-100)	100 (88-100)	100	100
	Sputum	848 (41.2)	20	626	8	175	19	95.6 (91.5–98)	97 (95.4–98.2)	90.2 (85.5-93.4)	98.7 (97.5-99.3)
Extrapulmonary	Urine	44 (2.1)	2	40	0	2	1	100 (15.8–100)	97.5 (86.8–99.9)	66.6 (22.4-93.2)	100
	Abscess	49 (2.4)	1	36	0	10	2	100 (69.1–100)	94.7 (82.2–99.3)	83.3 (56.4-95)	100
	Osteoarticular	80 (3.4)	2	66	2	3	7	60 (14.6-94.7)	90.4 (81.2-96)	30 (13.5-53.9)	97 (91.8-98.9)
	Biopsy	162 (7.8)	9	119	5	13	16	72.2 (46.5–90.3)	88.1 (81.4-93)	44.8 (32-58.2)	95.9 (91.8-98)
	Pericardium	120 (5.8)	2	112	3	2	1	40 (5.2-85.3)	99.1 (95.1–99.9)	66.6 (17.7-94.8)	97.3 (94.8-98.7)
	Cerebrospinal fluid (CSF)	44 (2.1)	1	39	0	2	1	100 (15.8–100)	97.5 (86.8–99.9)	66.6 (22.4–93.2)	100
	Gastric Lavage	111 (5.4)	2	103	1	4	1	80 (28.3-99.4)	99 (94.7-99.9)	80 (35.1-96.7)	99 (94.6–99.8)
	Blood	15 (0.7)	2	13	0	0	0	_	100 (75.2–100)	_	100
	Plural fluid	175 (8.4)	7	153	4	7	4	63.6 (30.7-89)	97.4 (93.6–99.3)	63.6 (37.6-83.5)	97.4 (94.5-98.8)
	Ascites	38 (1.8)	2	33	0	3	0	100 (29.2–100)	100 (89.4–100)	100	100
Pulmonary		1218 (59.2)	24	982	8	179	25	95.7 (91.7–98.1)	97.5 (96.3–98.3)	87.7 (82.9-91.3)	99.1 (98.4-99.5)
Extrapulmonary		838 (40.7)	31	713	15	49	30	76.5 (64.3–86.2)	95.9 (94.2–97.2)	62 (52.8-70.4)	97.9 (96.8–98.6)
Total Number		2056	55	_	_	_	_	_	_	_	_

Table 1. Xpert MTB/RIF assay performance for pulmonary and extrapulmonary specimens.

different provinces of Iran were included in this study. From the collected samples, 1218 (59.2 percent) were pulmonary; bronchoalveolar lavage fluid (BAL), tracheal, sputum and 838 (40.7 percent) were extrapulmonary; urine, abscess, osteoarticular, biopsy, pericardium, cerebrospinal fluid (CSF), gastric lavage, blood, real-time fluid, and ascites.

Specimens were collected from both outpatients and inpatients with a history of cough, unintentional weight loss, and fever for more than two weeks. Specimens were either from new cases or from patients with treatment failure or relapse.

The Ethics Committee of Shahid Beheshti University of Medical Sciences, Tehran, Iran approved the study and all research was performed in accordance with the relevant guidelines. All the patients and/or their legal guardians have signed informed consent.

Microscopy examination and culture. Specimens (2.5 to 10 mL) were decontaminated using 4% sodium hydroxide (NaOH) (Petroff method)²⁷. Sediments of each treated sample were used to prepare a smear for Ziehl-Neelsen and Auramine fluorochrome staining and were cultured in Löwenstein-Jensen medium^{27,28}. Only one culture isolated per study subject was considered for further analysis. Each isolate was examined regarding morphology, pigmentation, and date of growth. Bacterial isolates identified as *M. tuberculosis* using standard biochemical tests, including production of niacin, nitrate reduction and catalase²⁷.

Xpert MTB/RIF. One mL unconcentrated specimens used (without centrifuge) for Xpert MTB/RIF assay. Specimens, firstly, homogenized with a 5% acid-alcohol buffer, a 2:1 ratio as the Cepheid Company (USA) prepared in sample reagent (SR) and then added to the Xpert MTB/RIF cartridge to do the assay in Genexpert instrument. Xpert MTB/RIF, also, uses five overlapping probes (A, B, C, D, and E) to detect five mutations in the *rpoB* gene that accord rifampicin resistance².

Data analysis. The sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) with 95% confidence intervals were calculated for the Xpert MTB/RIF, using culture and direct smear microscopy as the gold standard. The summary receiver operating characteristic (SROC) curve was constructed based on a bivariate regression approach to show the level of accuracy of the Xpert MTB/RIF. The multiple regression analysis was used to shows any relationship between semi-quantitative Xpert MTB/RIF results and direct smear and culture results. All statistical analyses were performed with MedCalc (version 14.8.1, Medcalc Software).

Results

Microbiological findings. As shown in Table 1, of 1218 pulmonary specimens, 171 were positive and 982 were negative by standard conventional methods. Likewise, of 838 extrapulmonary specimens, 49 positive and 713 were negative by standard methods. Biopsy (26.5%), abscess (20.4%) and pleural fluid (14.2%) were the most commonly involved organs for EPTB.

Xpert MTB/RIF results. *M. tuberculosis* was present in 204 (13.4%) of pulmonary specimens and 79 of extrapulmonary specimens (Table 1). "Invalid," "error," and "no result" of Xpert MTB/RIF were excluded from the study. The proportion of Xpert MTB/RIF invalid and error cases in extrapulmonary specimens was significantly

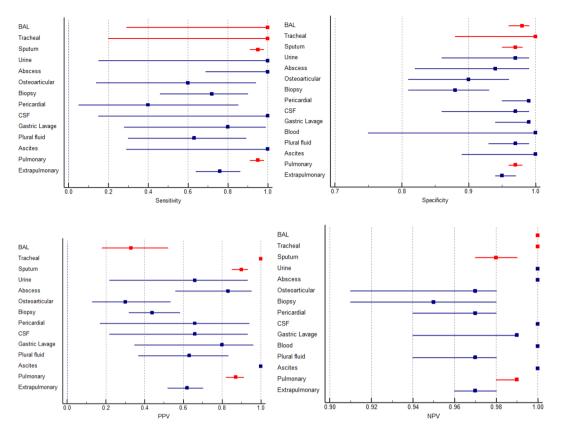


Figure 1. Forest plots give details of sensitivity, specificity, PPV, and NPV. BAL: bronchoalveolar lavage fluid, CSF: cerebrospinal fluid (Red colors are representative for pulmonary samples).

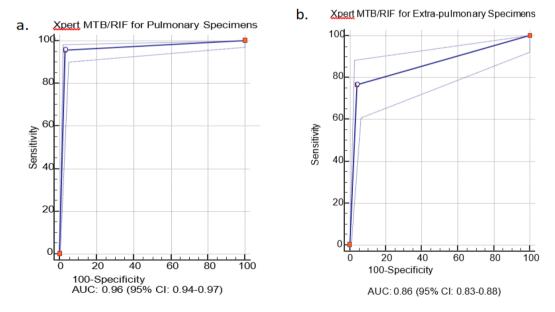


Figure 2. The summary receiver operating characteristic (SROC) curves for Xpert MTB/RIF assay. The SROC plot shows a summary of test performance, visual assessment of threshold effect, and heterogeneity of data in SROC space between sensitivity and specificity. The dashed blue line that is around the point estimate (blue line) shows 95% confidence region. The area under the curve (AUC), acts as an overall measure for test performance. Particularly, when AUC would be between 0.8 and 1, the accuracy is relatively high. As a matter of fact, AUC was 0.96 for pulmonary specimens in this report which represented a high level of accuracy. If SROC curve was in the upper left corner it would show the best combination of sensitivity and specificity for the diagnostic test. Part "a" and "b" are representative for pulmonary and extra-pulmonary specimens, respectively.

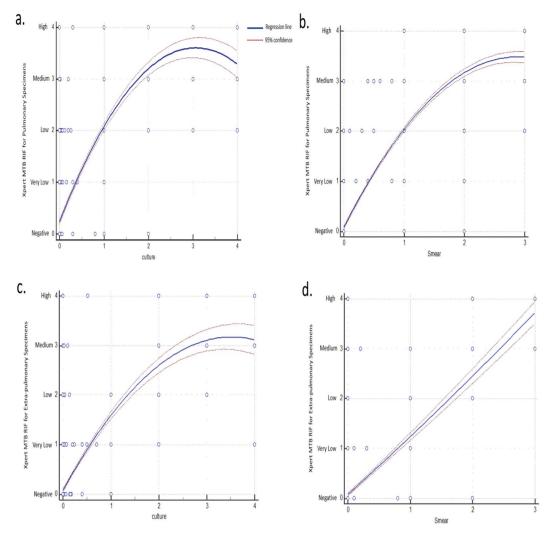


Figure 3. Quadratic regression plot to compare the Xpert MTB/RIF results with culture and smear microscopy. On Xpert MTB/RIF axis, results are categorized as 0, 1, 2, 3 and 4 which are indicated as negative, very low, low, medium and high scales, respectively. On the smear axis, results are categorized as 0, 1, 2 and 3 which are indicated as negative, 1+, 2+ and 3+ grading, respectively. On culture axis, results are categorized as 0, 1, 2, 3 and 4 which are indicated as negative, 1+, 2+, 3+ and 4+ grading, respectively. Part "a" and "b" are representative for pulmonary specimens and part "c" and "d" are representative for extra-pulmonary specimens. Regression lines showed the correlations between Xpert MTB/RIF results and culture/direct smear methods results.

more than pulmonary specimens with Chi-squared 5.62 (p < 0.02). Comparison of false positives in pulmonary and extrapulmonary specimens have not shown meaningful difference whilst false-negative results illustrated a significant difference with Chi-squared 5.93 for two groups (p < 0.02). The percentage result for each subgroup specimen of pulmonary and extrapulmonary are showed in Table 1. Furthermore, 22 rifampicin resistance (7.7% of positive cases) and 13 rifampicin indeterminate (4.5% of positive cases) were also indicated by Xpert MTB/RIF.

Sensitivity, specificity, PPV, and NPV. The overall sensitivity, specificity, PPV and NPV of Xpert MTB/RIF assay for PTB were found to be 95.5%, 96.7%, 83.8%, and 99.1% respectively. For EPTB, the sensitivity, specificity, PPV, and NPV of Xpert MTB/RIF assay were found to be 76.5%, 95.9%, 62%, and 97.9%, respectively.

Among extrapulmonary specimens, osteoarticular and pericardial specimens have lower sensitivity rather than average EPTB sensitivity (p < 0.0001). A comparison of sensitivity between PTB and EPTB showed a significant difference with 95% and 76% respectively (p < 0.0001). With the same respect, NPV for both groups was very high, while comparison of PPV between PTB and EPTB revealed a significant difference (p < 0.0001) (Table 1). The forest plot (Fig. 1) shows the sensitivity, specificity, PPV, and NPV for each group of specimens. The inter-rater agreement test for PTB and EPTB showed Xpert MTB/RIF assay has substantial agreement (kappa 0.74) and moderate agreement (kappa 0.4) with standard conventional methods, respectively. The accuracy index for PTB and EPTB was 96% and 94%, respectively.

Based on the SROC curves (Fig. 2), the AUC for pulmonary and extrapulmonary specimens was 0.96 (95% CI: 0.94–0.97) and 0.86 (95% CI: 0.83–0.88), respectively (p < 0.0001).

Regression analysis. Based on our regression analysis, Xpert MTB/RIF results were used as regressand (dependent variables) and culture/direct smear results were used as regressors (independent variables). According to the Xpert MTB/RIF assay, results are semi-quantitative and are categorized as "Very low," "Low," "Medium," and "High" based on the levels of detection. Likewise, the results of conventional methods are also semi-quantitative, which could be categorized as 1+, 2+, and 3+ in direct smear assay, and 1+, 2+, 3+ and 4+ in culture assay. Thus, we were able to compute the relationships among these variables. Accordingly, as shown in Fig. 3, there were correlations between culture/direct smear results and Xpert MTB/RIF results. Therefore, we were able to assess the relationship between the results of Xpert MTB/RIF and culture/direct smear results.

Discussion

This is the first assessment of Xpert MTB/RIF performance conducted on a large sample size in Iran. In the current study, sensitivity for pulmonary specimens was higher than in extrapulmonary samples (95.5% vs. 76.5%).

Similarly, several studies evaluated the accuracy of Xpert MTB/RIF on pulmonary specimens; with sensitivity varies from 95% to 100% in smear-positive sputa^{29–34}.

Although, previous studies showed sufficient accuracy of Xpert MTB/RIF assay for diagnosis of PTB, an acceptable accuracy for EPTB has not been well established^{22,23,35-37}. In the current study, Xpert has high specificity but limited sensitivity for the detection of TB in extrapulmonary specimens. Although positive results may be useful in rapidly identifying the disease, negative results provide less certainty for ruling out EPTB. The sensitivity of Xpert MTB/RIF for TB detection in extrapulmonary samples varied widely across different sample types explain more (From 40% to 100%) (Table 1). As reported by other studies, it is possible that the decontamination step has determined the lowering of bacillary load and consequently the reduction of test sensitivity^{8,38}. In agreement with these findings, our study indicated that specimens, which need smashing and homogenization (i.e. osteoarticular and biopsy specimens), had lower sensitivity.

Xpert MTB/RIF assay is based on real-time bacon PCR and subsequently is sensitive to PCR inhibitors; therefore, it needs special protocols and/or treatment buffer for extrapulmonary specimens, which includes variety of specimens²³. The current Xpert MTB/RIF buffer has been developed for sputum. This may affect the outcome with false-negative results and decrease the sensitivity for specimens other than sputum.

We found that Xpert MTB/RIF results for pericardial and biopsy specimens are less sensitive with low PPV, which was likely due to the presence of blood and inhibitors for PCR reaction. Therefore, any treatment to remove blood before the reaction would increase the sensitivity of these specimens. Low PPV for osteoarticular specimens was also an important issue that should be addressed with further investigation.

Based on regression analysis, we were able to assess the relationship between the results of Xpert MTB/RIF and culture/direct smear results for PTB and EPTB specimens.

Limitation of the study. The current study is limited by a low sample size for blood and CSF specimens.

Conclusions

Xpert MTB/RIF could be used as the first-line diagnostic tool for PTB cases. However, the overall low sensitivity of Xpert MTB/RIF for extrapulmonary specimens precludes the use of this test to rule out EPTB with certainty.

Received: 1 November 2018; Accepted: 20 November 2019;

Published online: 06 December 2019

References

- 1. World Health Organization (WHO). Global tuberculosis report (2017).
- 2. Boehme, C. C. et al. Rapid molecular detection of tuberculosis and rifampin resistance. New England Journal of Medicine 363, 1005–1015 (2010).
- 3. Varahram, M., Nasiri, M. J., Farnia, P., Mozafari, M. & Velayati, A. A. A retrospective analysis of isoniazid-monoresistant tuberculosis: among Iranian pulmonary tuberculosis patients. *The open microbiology journal* 8, 1 (2014).
- Trébucq, A. et al. Xpert[®] MTB/RIF for national tuberculosis programmes in low-income countries: when, where and how? The International Journal of Tuberculosis and Lung Disease 15, 1567–1572 (2011).
- Dunn, J. J., Starke, J. R. & Revell, P. A. Laboratory diagnosis of Mycobacterium tuberculosis infection and disease in children. *Journal of clinical microbiology, JCM*. 03043-03015 (2016).
- 6. Organization, W. H. Early detection of tuberculosis: an overview of approaches, guidelines and tools. (2011).
- 7. Organization, W. H. In Xpert MTB/RIF implementation manual: technical and operational 'how-to'; practical considerations (2014).
- 8. Tortoli, E. et al. Clinical validation of Xpert MTB/RIF for the diagnosis of extrapulmonary tuberculosis. European Respiratory Journal, erj01763-02011 (2012).
- 9. Armand, S., Vanhuls, P., Delcroix, G., Courcol, R. & Lemaître, N. Comparison of the Xpert MTB/RIF test with an IS6110-TaqMan real-time PCR assay for direct detection of Mycobacterium tuberculosis in respiratory and nonrespiratory specimens. *Journal of clinical microbiology* (2011).
- Friedrich, S. O., von Groote-Bidlingmaier, F. & Diacon, A. H. Xpert MTB/RIF assay for diagnosis of pleural tuberculosis. *Journal of clinical microbiology* 49, 4341–4342 (2011).
- 11. Hillemann, D., Rüsch-Gerdes, S., Boehme, C. & Richter, E. Rapid molecular detection of extrapulmonary tuberculosis by automated GeneXpert[®] MTB/RIF system. *Journal of clinical microbiology* (2011).
- 12. Moure, R., Martín, R. & Alcaide, F. Effectiveness of an integrated real-time PCR method for detection of the Mycobacterium tuberculosis complex in smear-negative extrapulmonary samples in an area of low tuberculosis prevalence. *Journal of clinical microbiology* 50, 513–515 (2012).
- 13. Patel, V. B. et al. Diagnostic accuracy of quantitative PCR (Xpert MTB/RIF) for tuberculous meningitis in a high burden setting: a prospective study. *PLoS medicine* 10, e1001536 (2013).
- 14. Rufal, S. B. et al. Diagnostic usefulness of Xpert MTB/RIF assay for detection of tuberculous meningitis using cerebrospinal fluid. Journal of Infection 75, 125–131 (2017).
- 15. Metcalf, T. et al. Evaluation of the GeneXpert MTB/RIF in patients with presumptive tuberculous meningitis. PloS one 13, e0198695 (2018).

- 16. Boehme, C. C. et al. Feasibility, diagnostic accuracy, and effectiveness of decentralised use of the Xpert MTB/RIF test for diagnosis of tuberculosis and multidrug resistance: a multicentre implementation study. *The lancet* 377, 1495–1505 (2011).
- 17. Theron, G. et al. Evaluation of the Xpert MTB/RIF assay for the diagnosis of pulmonary tuberculosis in a high HIV prevalence setting. American journal of respiratory and critical care medicine 184, 132–140 (2011).
- 18. Vassall, A. et al. Rapid diagnosis of tuberculosis with the Xpert MTB/RIF assay in high burden countries: a cost-effectiveness analysis. PLoS medicine 8, e1001120 (2011).
- 19. Churchyard, G. J. et al. Xpert MTB/RIF versus sputum microscopy as the initial diagnostic test for tuberculosis: a cluster-randomised trial embedded in South African roll-out of Xpert MTB/RIF. The Lancet Global Health 3, e450–e457 (2015).
- 20. Creswell, J. et al. Results from early programmatic implementation of Xpert MTB/RIF testing in nine countries. BMC infectious diseases 14, 2 (2014).
- 21. Sachdeva, K. S. et al. Use of Xpert MTB/RIF in decentralized public health settings and its effect on pulmonary TB and DR-TB case finding in India. PloS one 10, e0126065 (2015).
- 22. Denkinger, C. M. et al. Xpert MTB/RIF assay for the diagnosis of extrapulmonary tuberculosis: a systematic review and meta-analysis. European Respiratory Journal, erj00078-02014 (2014).
- 23. Theron, G. et al. Determinants of PCR performance (Xpert MTB/RIF), including bacterial load and inhibition, for TB diagnosis using specimens from different body compartments. Scientific reports 4, 5658 (2014).
- 24. Pang, Y. et al. GeneXpert MTB/RIF assay in the diagnosis of urinary tuberculosis from urine specimens. Scientific reports 7, 6181 (2017).
- García-Basteiro, A. L. et al. The role of Xpert MTB/RIF in diagnosing pulmonary tuberculosis in post-mortem tissues. Scientific reports 6, 20703 (2016).
- 26. Pormohammad, A., Nasiri, M. J., McHugh, T. D., Riahi, S. M. & Bahr, N. C. A systematic review and meta-analysis of the diagnostic accuracy of nucleic acid amplification tests for tuberculous meningitis. *Journal of clinical microbiology* 57, e01113–01118 (2019).
- 27. Nasiri, M. J. et al. Drug resistance pattern of Mycobacterium tuberculosis isolates from patients of five provinces of Iran. Asian Pacific journal of tropical medicine 7, 193–196 (2014).
- 28. Marais, B. J. et al. Use of light-emitting diode fluorescence microscopy to detect acid-fast bacilli in sputum. Clinical Infectious Diseases 47, 203–207 (2008).
- 29. Rice, J. P., Seifert, M., Moser, K. S. & Rodwell, T. C. Performance of the Xpert MTB/RIF assay for the diagnosis of pulmonary tuberculosis and rifampin resistance in a low-incidence, high-resource setting. *PloS one* 12, e0186139 (2017).
- 30. Mei, Y. et al. Evaluation of Xpert MTB/RIF for the diagnosis of extrapulmonary tuberculosis in China. Biomedical and Environmental Sciences 29, 599–602 (2016).
- 31. Lombardi, G. et al. Diagnosis of smear-negative tuberculosis is greatly improved by Xpert MTB/RIF. PloS one 12, e0176186 (2017).
- 32. Helb, D. et al. Rapid detection of Mycobacterium tuberculosis and rifampin resistance by use of on-demand, near-patient technology. Journal of clinical microbiology 48, 229-237 (2010).
- 33. Marlowe, E. M. et al. Evaluation of the Cepheid Xpert MTB/RIF assay for direct detection of Mycobacterium tuberculosis complex in respiratory specimens. *Journal of clinical microbiology* 49, 1621–1623 (2011).
- 34. Kohli, M. et al. Xpert[®] MTB/RIF assay for extrapulmonary tuberculosis and rifampicin resistance. Cochrane Database of Systematic Reviews (2018).
- 35. Suzana, S. *et al.* Xpert MTB/Rif for the diagnosis of extrapulmonary tuberculosis—an experience from a tertiary care centre in South India. *Tropical Medicine & International Health* **21**, 385–392 (2016).
- 36. Maynard-Smith, L., Larke, N., Peters, J. A. & Lawn, S. D. Diagnostic accuracy of the Xpert MTB/RIF assay for extrapulmonary and pulmonary tuberculosis when testing non-respiratory samples: a systematic review. *BMC infectious diseases* 14, 709 (2014).
- 37. Schumacher, S. G. & Pai, M. Xpert *MTB/RIF for extra-pulmonary tuberculosis: time to look beyond accuracy. *The International Journal of Tuberculosis and Lung Disease* 19, 2–2 (2015).
- 38. Vadwai, V. et al. Xpert MTB/RIF, a new pillar in the diagnosis of extrapulmonary tuberculosis? *Journal of clinical microbiology, JCM*. 02319–02310 (2011).

Acknowledgements

This study was financially supported by Research Department of the School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran (Grant number:13097).

Author contributions

M.A.T., M.J.N., M.M., and G.H. conceived and designed the study. M.A.T., S.A. and M.Z. performed the experiments. M.A.T. and M.J.N. analyzed the data. M.A.T., M.J.N. and M.M. wrote the main manuscript text.

Competing interests

The authors declare no competing interests.

Additional information

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