

## **Supplementary Information**

### **Mechanically Tunable, Compostable, Healable and Scalable Engineered Living Materials**

Avinash Manjula-Basavanna<sup>1,2,3\*</sup>, Anna M. Duraj-Thatte<sup>2</sup>, Neel S. Joshi<sup>1\*</sup>

<sup>1</sup> Department of Chemistry and Chemical Biology, Northeastern University, Boston, Massachusetts, United States

<sup>2</sup> Department of Biological Systems Engineering, Virginia Polytechnic Institute and State University, Blacksburg, Virginia, United States

<sup>3</sup> Department of Bioengineering, Northeastern University, Boston, Massachusetts, United States

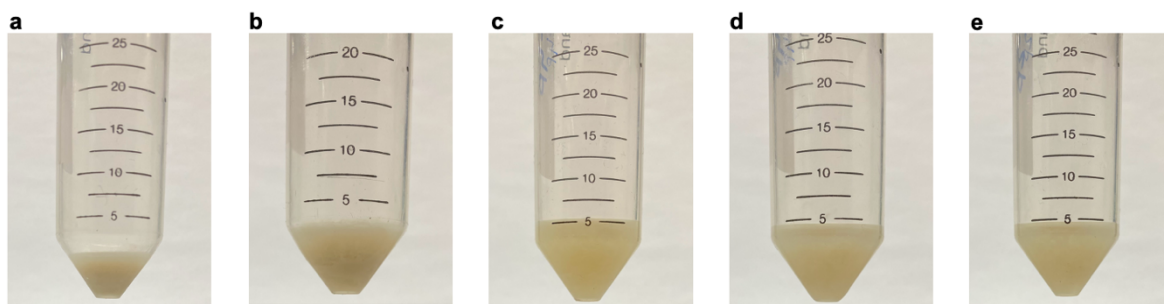
\*Corresponding author

mbavinash@northeastern.edu

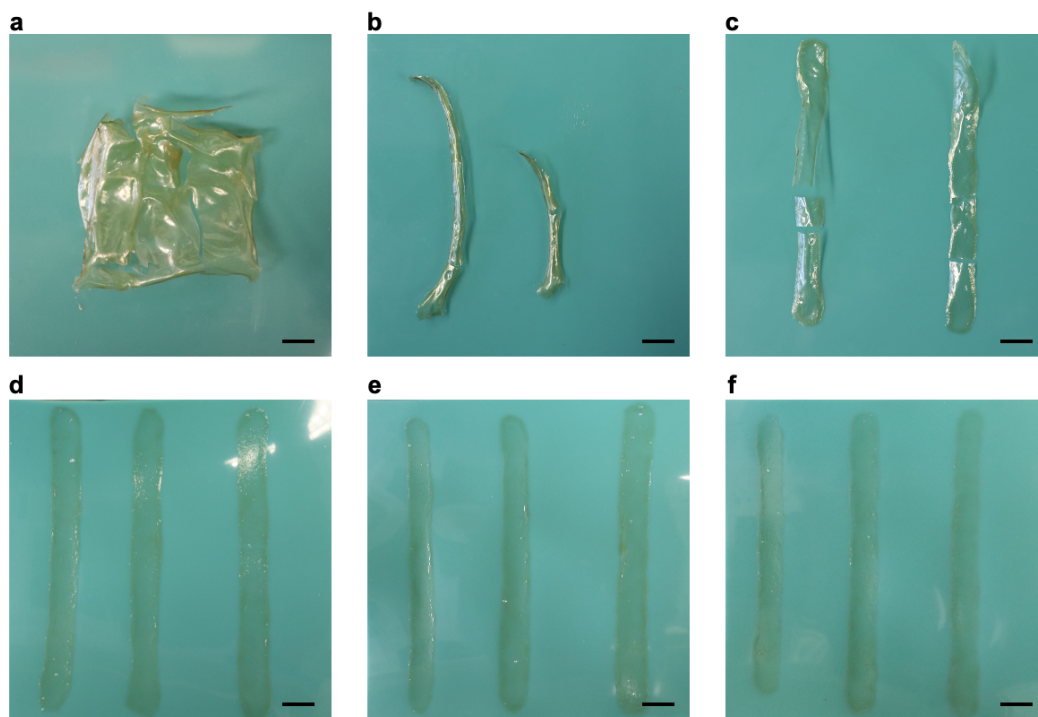
ne.joshi@northeastern.edu

**Supplementary Table 1. Genes and their DNA sequence.**

Gene	DNA Sequence
CsgA	GGTGTTCCTCAGTACGGCGGCGGCGGTAACACGGTGGTGGCGGTAATAA TAGCGGCCAAATTCTGAGCTGAACATTTACCAGTACGGTGGCGGTAACCTCTGC ACTTGCTCTGCAAATGATGCCCCGTAACCTGACTTGACTATTACCCAGCATGGC GGCGGTAATGGTGCAGATGTTGGTCAGGGCTCAGATGACAGCTCAATCGATCTG ACCCAACGTGGCTTCGGTAACAGCGCTACTCTTGATCAGTGGAACGGCAAAAAT TCTGAAATGACGGTTAAACAGTTCGGTGGTGGCAACGGTGCTGCAGTTGACCAG ACTGCATCTAACTCCTCCGTCAACGTGACTCAGGTTGGCTTTGTAACAACGCG ACCGCTCATCAGTAC
Linker	GGTGGATCTGGTAGCAGCGGCTCTGGTGGTTCTGGGGGCGGAAGTGGCTCCTC TGGGAGCGGGGGGTCGGGTGGTGGCTCGGGTTCATCTGGTAGTGGCGGTTCCG GGT
SpyTag	CGCGGCGTGCCGCATATTGTGATGGTGGATGCGTATAAACGCTATAAA
SpyCatcher	GTGACCACCCTGAGCGGCCTGAGCGGCGAACAGGGCCCCGAGCGGCGATATGA CCACCGAAGAAGATAGCGCGACCCATATTAATTTAGCAAACGCGATGAAGATG GCCGCGAACTGGCGGGCGCGACCATGGAAGTGGCGATAGCAGCGGCAAAAC CATTAGCACCTGGATTAGCGATGGCCATGTGAAAGATTTTTATCTGTATCCGGGC AAATATACCTTTGTGGAACCGCGGCGCCGGATGGCTATGAAGTGGCGACCCC GATTGAATTTACCGTGAACGAAGATGGCCAGGTGACCGTGGATGGCGAAGCGA CCGAAGGCGATGCGCATACC
Spacer	ATGAAAGTGCTGATTCTGGCGTGCCTGGTGGCGCTGGCGCTGGCGCGCGAAAC CATTGAAAGCCTGAGCAGCAGCGAAGAAAGCATTACCGAATATAAACAGAAAGT GGAAAAAGTGAAACATGAAGATCAGCAGCAGGGCGAAGATGAACATCAGGATAA AATTTATCCGAGCTTTCAGCCGCAGCCGCTGATTTATCCGTTTGTGGAACCGATT CCGTATGGCTTTCTGCCGCAGAACATTCTGCCGCTGGCGCAGCCGGCGGTGGT GCTGCCGGTGCCGCAGCCGGAATTATGGAAGTGCCGAAAGCGAAAGATAACCG TGTATACCAAAGGCCGCGTGATGCCGGTGCTGAAAAGCCCGACCATTCGTTTTT TTGATCCGCAGATTCCGAAACTGACCGATCTGGAAAACCTGCATCTGCCGCTGC CGCTGCTGCAGCCGCTGATGCAGCAGGTGCCGCAGCCGATTCCGCAGACCTG GCGCTGCCGCCGCAGCCGCTGTGGAGCGTGCCGCAGCCGAAAGTGCTGCCGA TTCCGCAGCAGGTGGTGGCGTATCCGCAGCGCGCGGTGCCGGTGACGGCGCT GCTGCTGAACCAGGAACTGCTGCTGAACCCGACCCATCAGATTTATCCGGTGAC CCAGCCGCTGGCGCCGGTGATACCCGATTAGCGTG

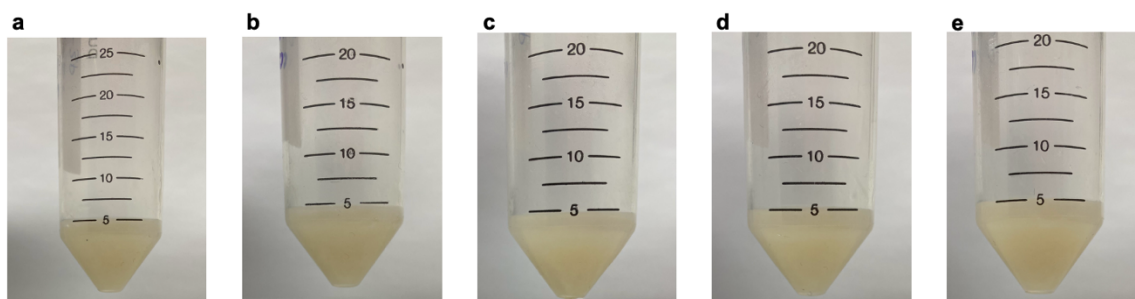


**Supplementary Figure 1. Gelator treated curli biomass.** Photographs of curli biomass obtained from **a** 1%, **b** 2%, **c** 3%, **d** 4% and **e** 5% gelator (sodium dodecyl sulfate).

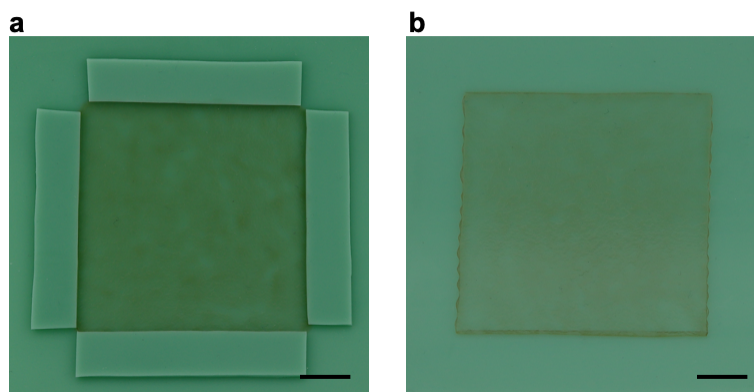


**Supplementary Figure 2. Gelator treated MECHS.** Photographs of MECHS obtained from **a** 0%, **b** 1%, **c** 2%, **d** 3%, **e** 4% and **f** 5% gelator (sodium dodecyl sulfate). Scale bar 1 cm.

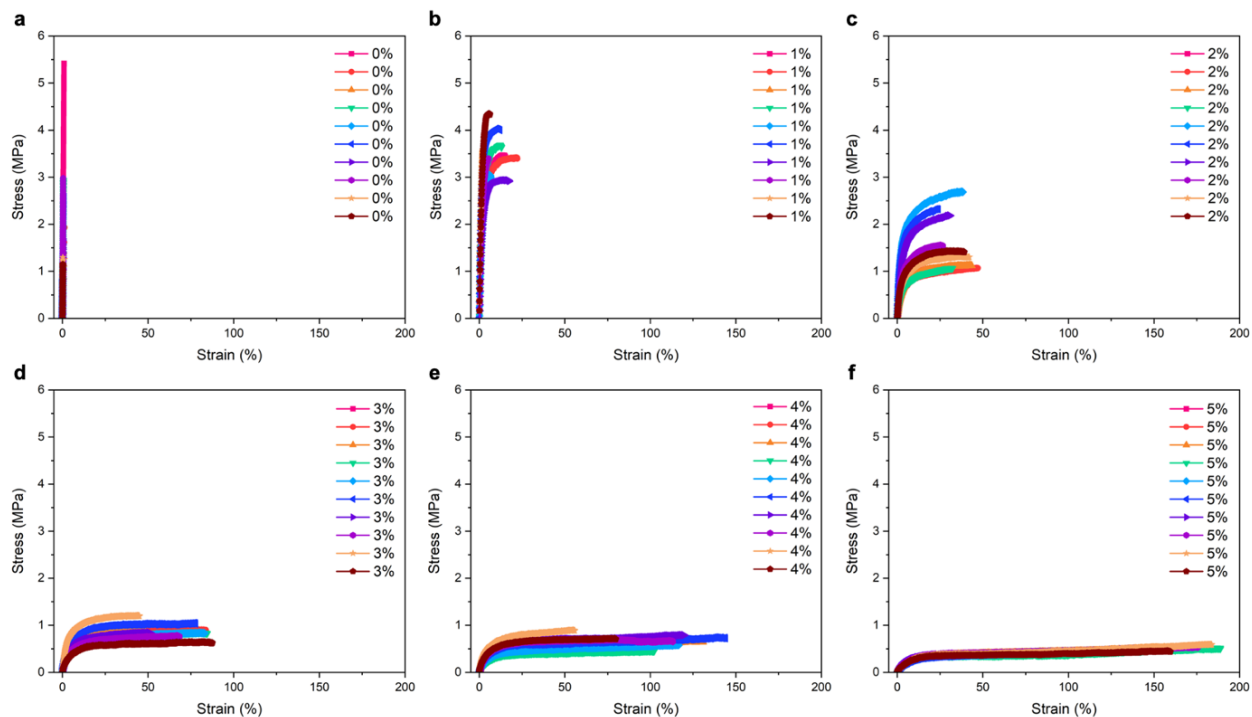




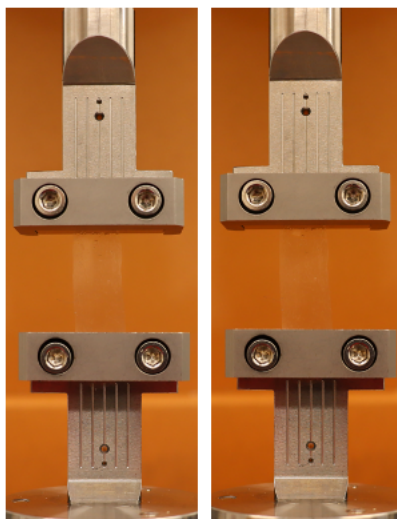
**Supplementary Figure 3. Plasticizer treated curli biomass.** Photographs of curli biomass obtained from **a** 1%, **b** 2%, **c** 3%, **d** 4% and **e** 5% plasticizer (glycerol).



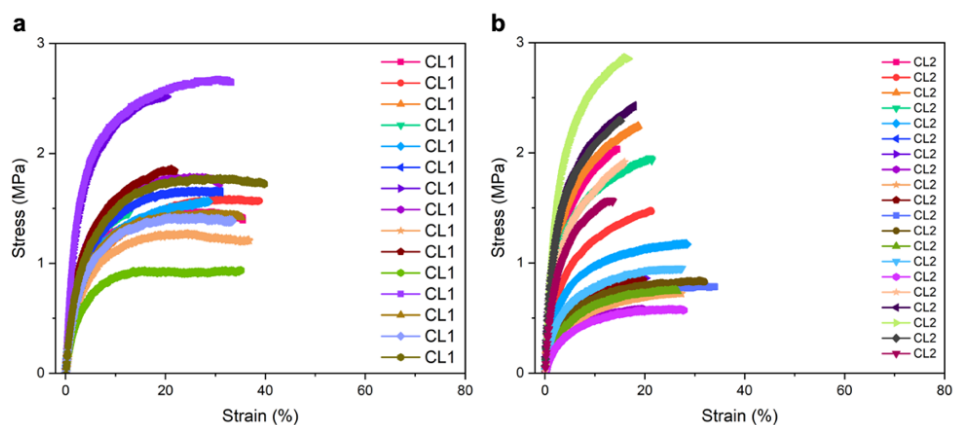
**Supplementary Figure 4. Plasticizer treated MECHS.** **a** and **b** Photographs of MECHS film obtained by treating with 3% plasticizer (glycerol) and casting on a silicone mold. Scale bar 1 cm.



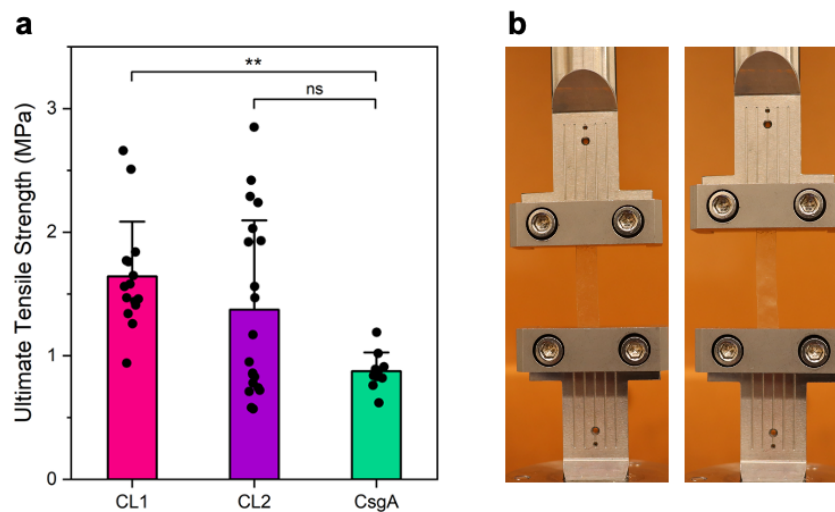
**Supplementary Figure 5. Tensile tests of MECHS.** Stress strain curves of MECHS films obtained from **a** 0%, **b** 1%, **c** 2%, **d** 3%, **e** 4% and **f** 5% plasticizer (glycerol).



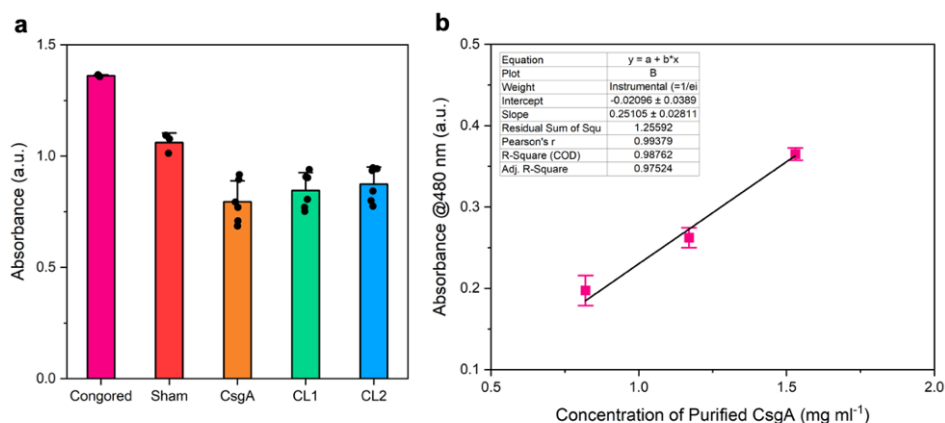
**Supplementary Figure 6. Tensile test of MECHS.** Representative photographs show the tensile test of MECHS film with the lateral dimension of 1 cm by 4 cm obtained from 3% of gelator. Left image: initial. Right image: before break



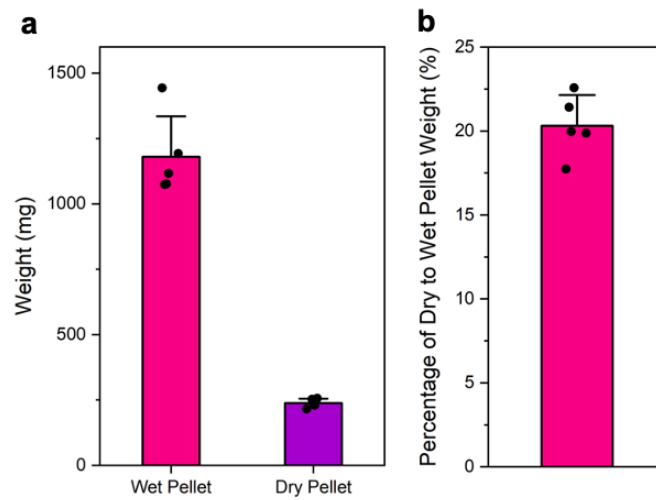
**Supplementary Figure 7. Tensile tests of MECHS.** Stress strain curves of MECHS obtained from **a** CL1 and **b** CL2.



**Supplementary Figure 8. Tensile tests of MECHS.** **a** Ultimate tensile strength of CsgA, CL1 and CL2 with 3% plasticizer. Data represented as mean  $\pm$  standard deviation. Biological replicates  $n = 10$  for CsgA,  $n = 15$  for CL1 and  $n = 20$  for CL2.  $**p = 0.0042$ ,  $^{ns}p = 0.0637$ . One-way ANOVA followed by Tukey's multiple comparisons test. **b** Representative photographs show the tensile test of CL2 film with the lateral dimension of 0.5 cm by 4 cm. Left image: initial. Right image: before break.

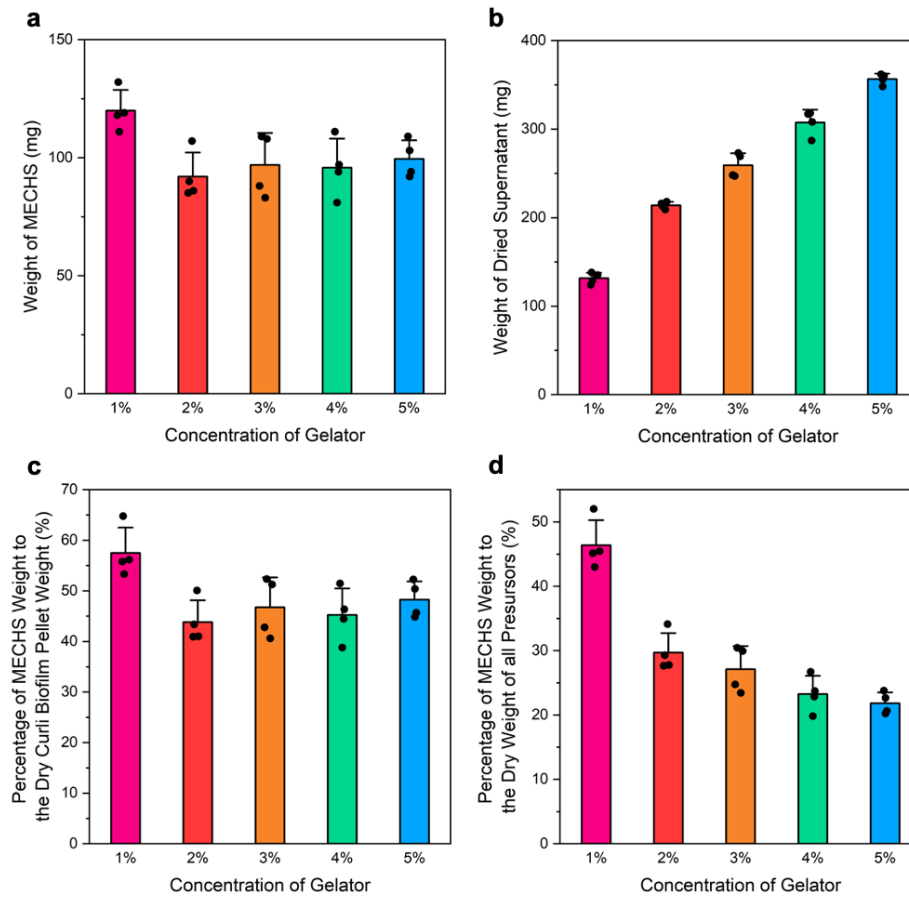


**Supplementary Figure 9. Congo Red assay.** **a** Congo Red absorbance at 480 nm for various samples. **b** Congo Red standard curve for purified CsgA (wet weight, CsgA-His). Biological replicates  $n = 3$  for Sham and  $n = 6$  for CsgA, CL1 and CL2. The net Congo Red absorbance of curli in CsgA, CL1 and CL2 were determined by subtracting the absorbance values of cell pellet having a sham plasmid (without curli operon), to account for the non-specific binding to other biomolecules. Data represented as mean  $\pm$  standard deviation.

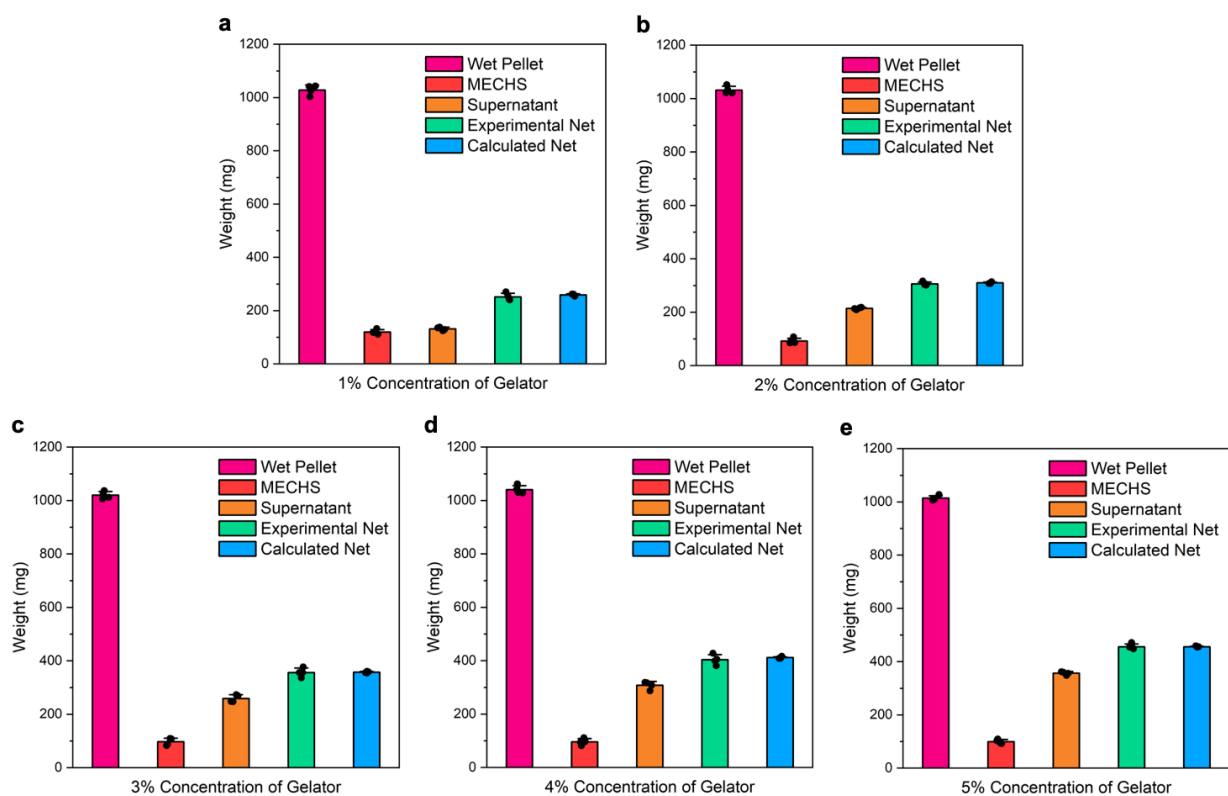


**Supplementary Figure 10. Weight analysis.** **a** Weight of wet and dry pellet of curli biofilm obtained from 500 ml cultures. **b** Percentage of dry to wet pellet weight of curli biofilm.  $n = 5$ . Data represented as mean  $\pm$  standard deviation.

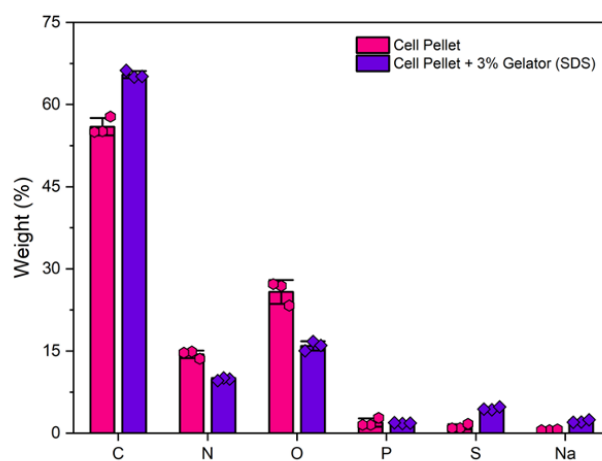




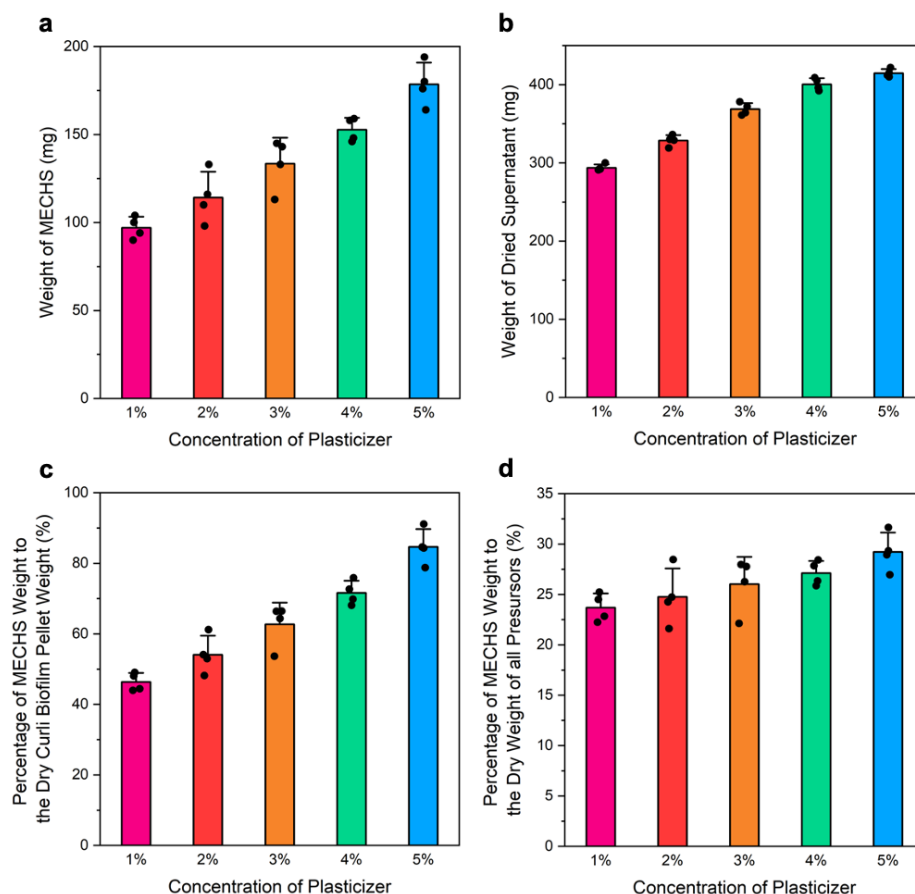
**Supplementary Figure 11. Weight analysis of gelator treated MECHS.** Weight of **a** MECHS films and **b** the dried supernatant obtained from 1 to 5% of gelator. Percentage of MECHS film weight to **c** dry curli biofilm pellet weight (20.3% of wet pellet) and **d** dry weight of all precursors for 1 to 5% of gelator. All precursors correspond to weights of cell pellet and that of 1-5% gelator.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.



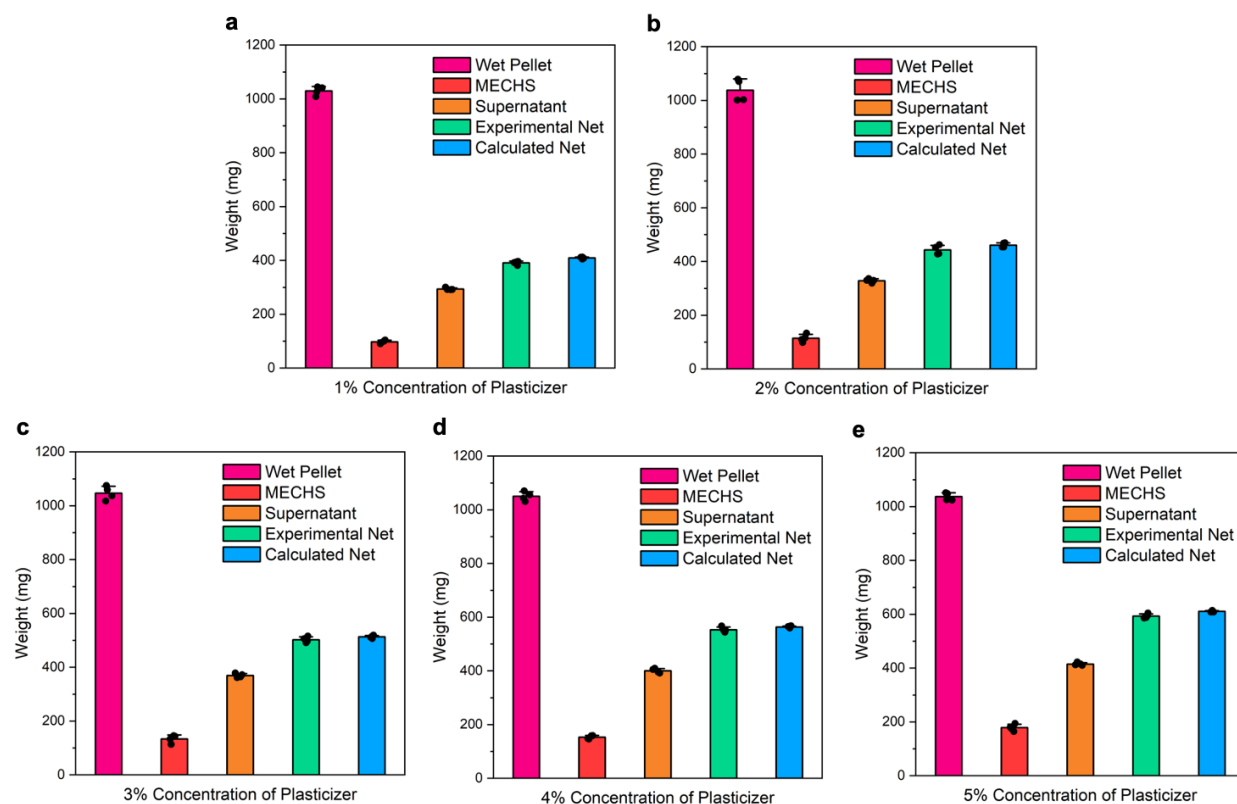
**Supplementary Figure 12. Weight analysis of gelator treated MECHS.** Weights of wet pellet, MECHS film, dried supernatant, experimental net (MECHS and dried Supernatant) and calculated theoretical net (20.3% of wet pellet) for **a** 1%, **b** 2%, **c** 3%, **d** 4% and **e** 5% of gelator.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.



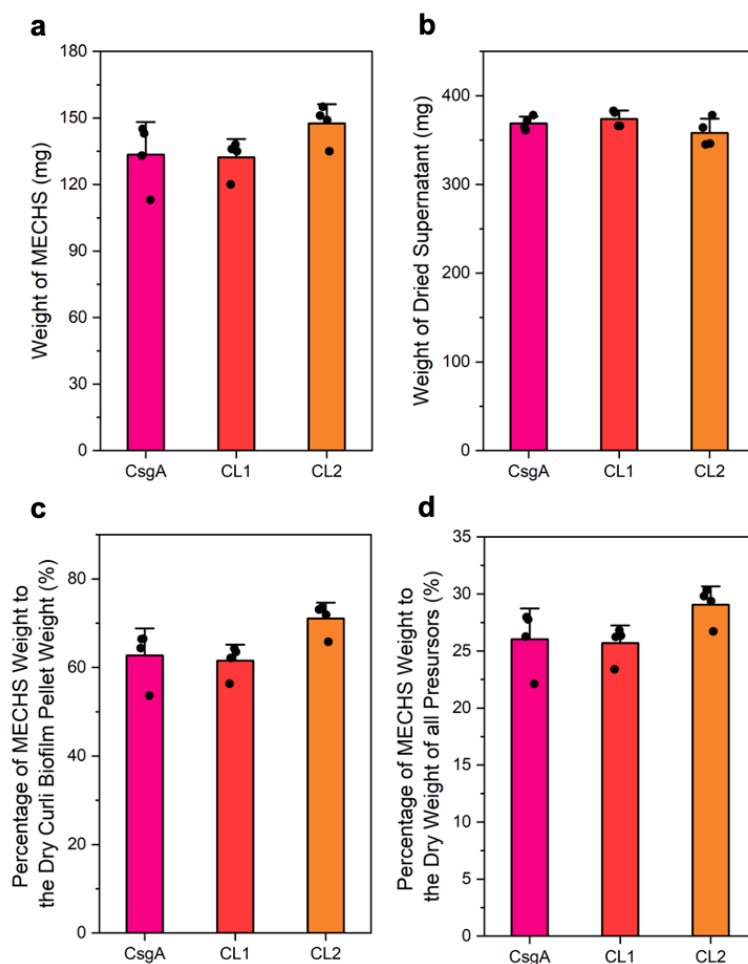
**Supplementary Figure 13. Compositional analysis of MECHS.** Energy Dispersive X-ray Analysis (EDAX) of *E. coli* curli biofilm cell pellets pretreated with or without 3% gelator (SDS) shows the weight percentage of various elements.  $n = 3$ . Data represented as mean  $\pm$  standard deviation.



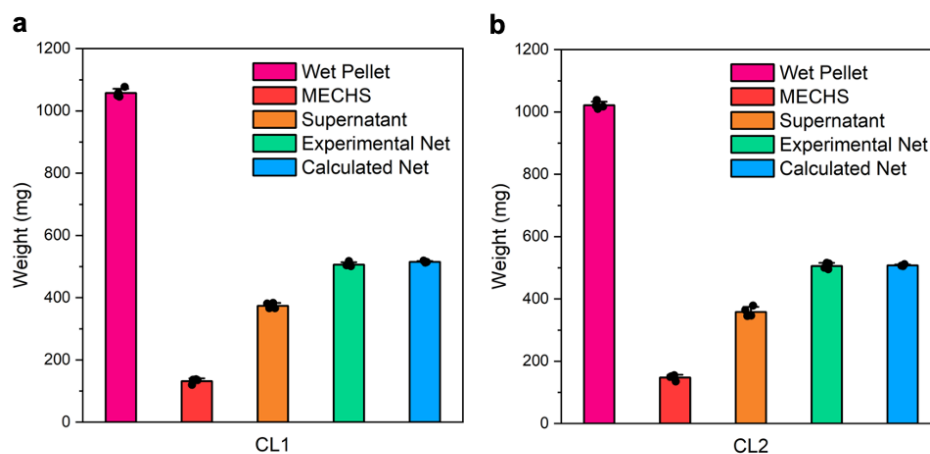
**Supplementary Figure 14. Weight analysis of plasticizer treated MECHS.** Weight of **a** MECHS films and **b** the dried supernatant obtained from 1 to 5% of plasticizer. Percentage of MECHS film weight to **c** dry curli biofilm pellet weight (20.3% of wet pellet) and **d** dry weight of all precursors for 1 to 5% of plasticizer. All precursors correspond to weights of cell pellet and that of 3% gelator and 1-5% plasticizer.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.



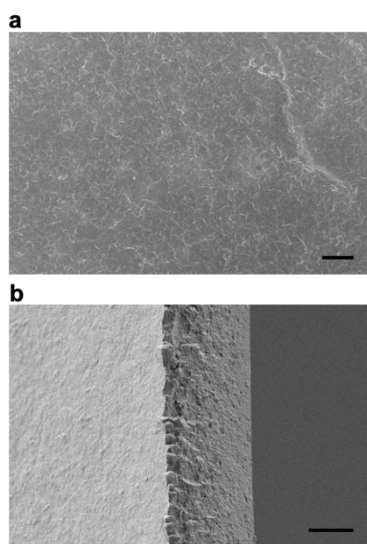
**Supplementary Figure 15. Weight analysis of plasticizer treated MECHS.** Weights of wet pellet, MECHS film, dried supernatant, experimental net (MECHS and dried Supernatant) and calculated theoretical net (20.3% of wet pellet) for **a** 1%, **b** 2%, **c** 3%, **d** 4% and **e** 5% of plasticizer.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.



**Supplementary Figure 16. Weight analysis of covalently crosslinked MECHS.** Weight of **a** MECHS films and **b** the dried supernatant of CsgA, CL1 and CL2. Percentage of MECHS weight to **c** dry curli biofilm pellet weight (20.3% of wet pellet) and **d** dry weight of all precursors for CsgA, CL1 and CL2. All precursors correspond to weights of cell pellet and that of 3% gelator and 3% plasticizer.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.

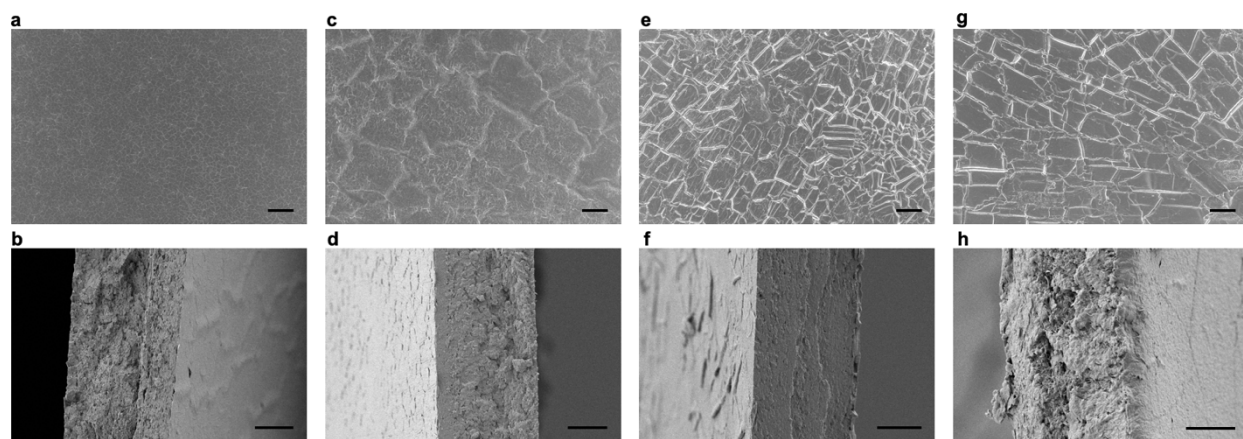


**Supplementary Figure 17. Weight analysis of covalently crosslinked MECHS.** Weights of wet pellet, MECHS, dried supernatant, experimental net (MECHS and dried Supernatant) and calculated theoretical net (20.3% of wet pellet) for **a** CL1 and **b** CL2.  $n = 4$ . Data represented as mean  $\pm$  standard deviation.



**Supplementary Figure 18. Morphological analysis of MECHS.** FESEM image of **a** top view and **b** side view of MECHS film obtained from 3% gelator. Scale bar **a** 20  $\mu\text{m}$  and **b** 10  $\mu\text{m}$ .

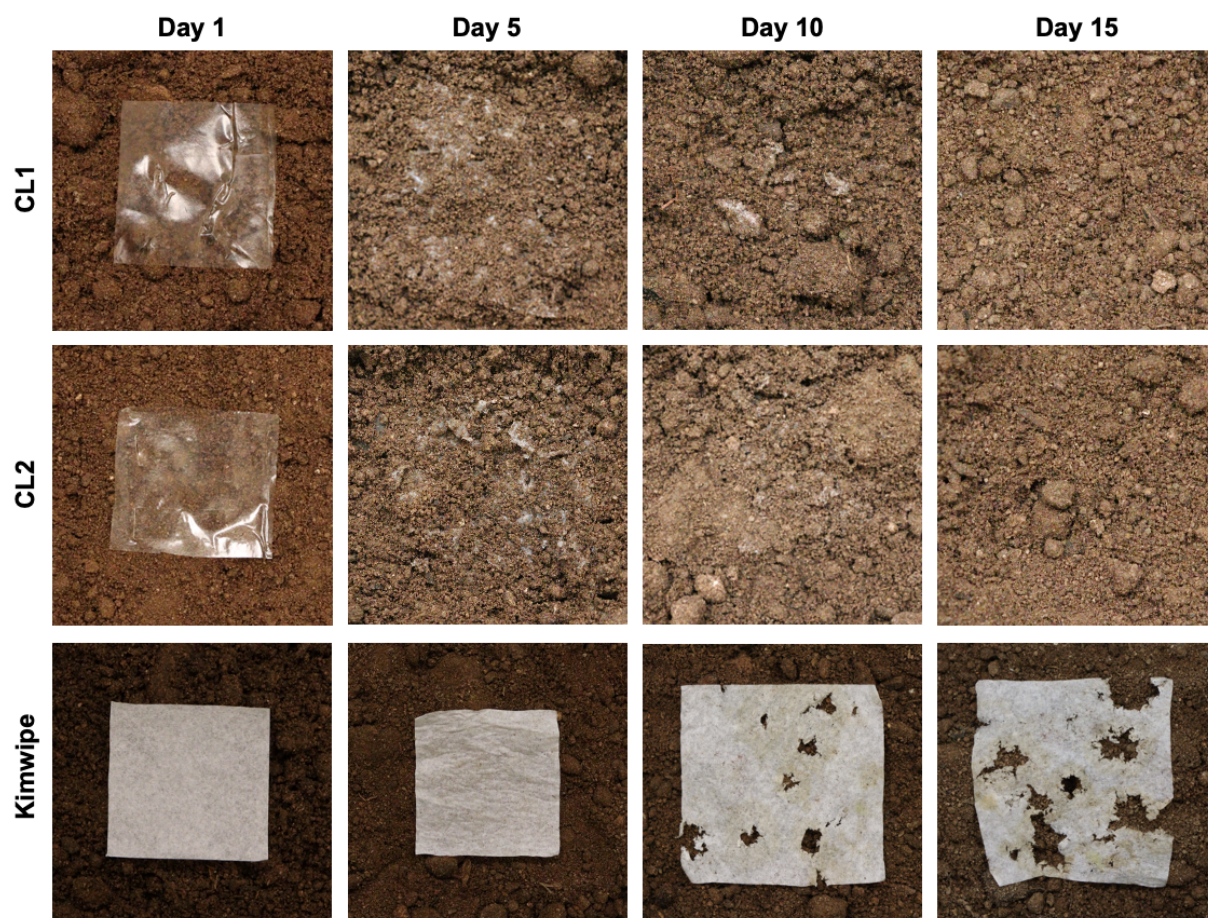




**Supplementary Figure 19. Morphological analysis of MECHS.** FESEM image of **a,c,e,g** top view and **b,d,f,h** side view of MECHS film obtained from **a,b** 1%, **c,d** 2%, **e,f** 4% and **g,h** 5% plasticizer. Scale bar Top Row: 20  $\mu\text{m}$  and Bottom Row: 10  $\mu\text{m}$ .

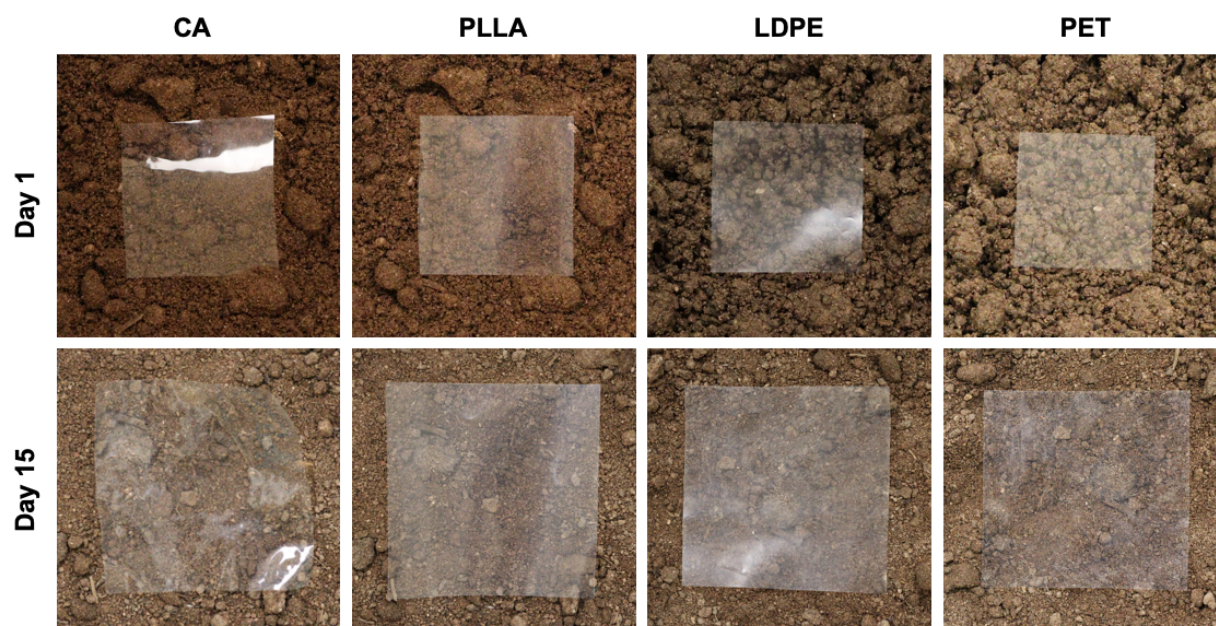


**Supplementary Figure 20. Greenhouse for biodegradation experiment.** Photograph of a mini greenhouse setup utilized for testing the compostability of MECHS.

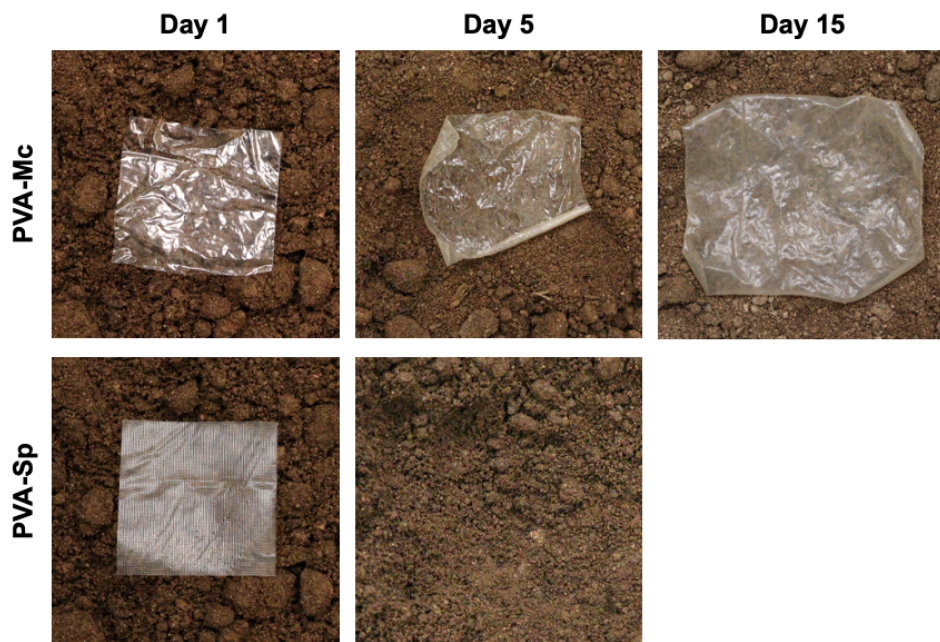


**Supplementary Figure 21. Compostability of MECHS and Kimwipe in a fresh fishnure.** Photographs show the biodegradation of CL1, CL2 and Kimwipe in a fresh fishnure. The lateral dimensions of the samples were 5 cm by 5 cm.

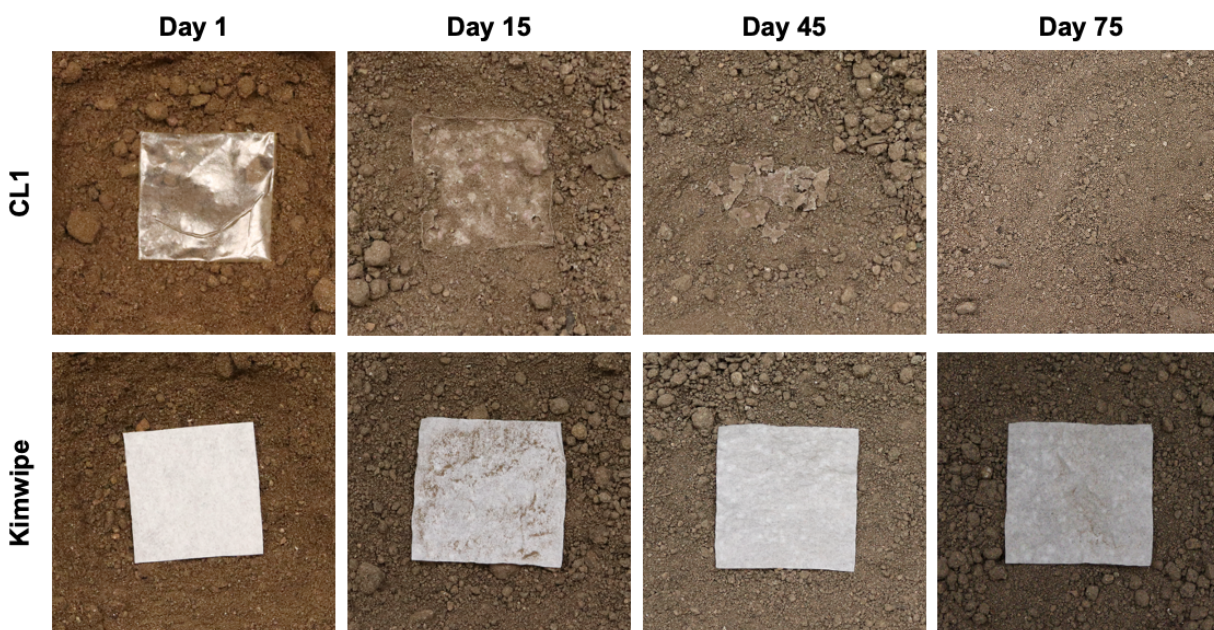




**Supplementary Figure 22. Compostability of Bioplastics and Plastics in a fresh fishnure.** Photographs show the biodegradation of cellulose acetate (CA), poly-L-lactic acid (PLLA), low density polyethylene (LDPE) and polyethylene terephthalate (PET) in a fresh fishnure. The lateral dimensions of the samples were 5 cm by 5 cm.

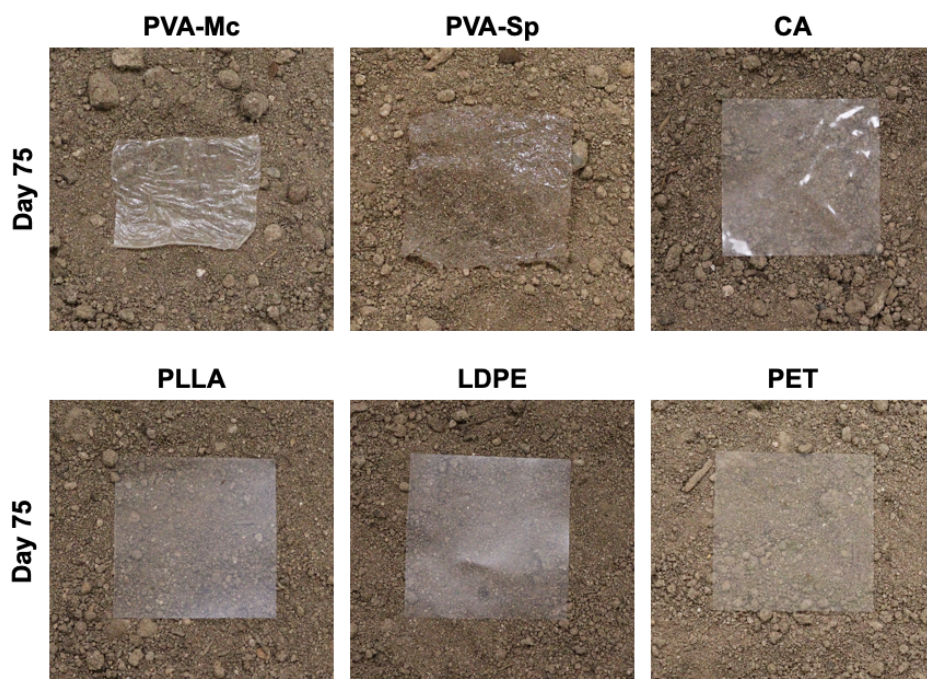


**Supplementary Figure 23. Compostability of Polyvinyl alcohol in a fresh fishnure.** Photographs show the films of Polyvinyl alcohol - Mckesson (PVA-Mc) and Polyvinyl alcohol - Superpunch (PVA-Sp) in a fresh fishnure. The lateral dimensions of the samples were 5 cm by 5 cm.

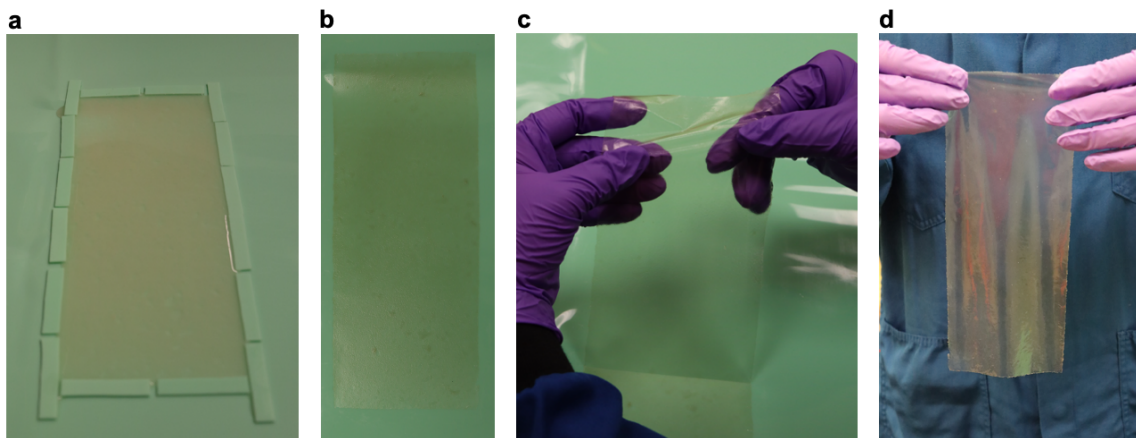


**Supplementary Figure 24. Compostability of MECHS and Kimwipe in a dry fishnure.** Photographs show the biodegradation of CL1 and Kimwipe in a dry fishnure. The lateral dimensions of the samples were 5 cm by 5 cm.



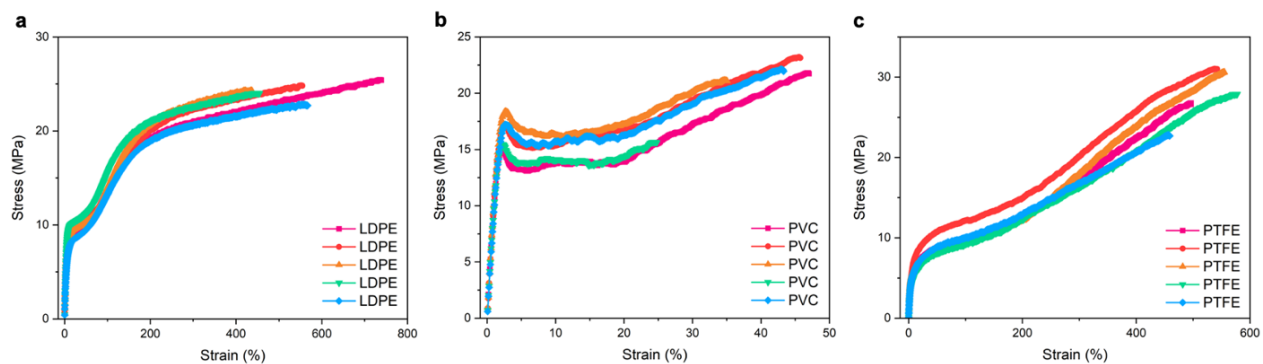


**Supplementary Figure 25. Compostability of Bioplastics and Plastics in a dry fishnure.** Photographs show the biodegradation of Polyvinyl alcohol - Mckesson (PVA-Mc) and Polyvinyl alcohol - Superpunch (PVA-Sp), cellulose acetate (CA), poly-L-lactic acid (PLLA), low density polyethylene (LDPE) and polyethylene terephthalate (PET) in a dry fishnure. The lateral dimensions of the samples were 5 cm by 5 cm.

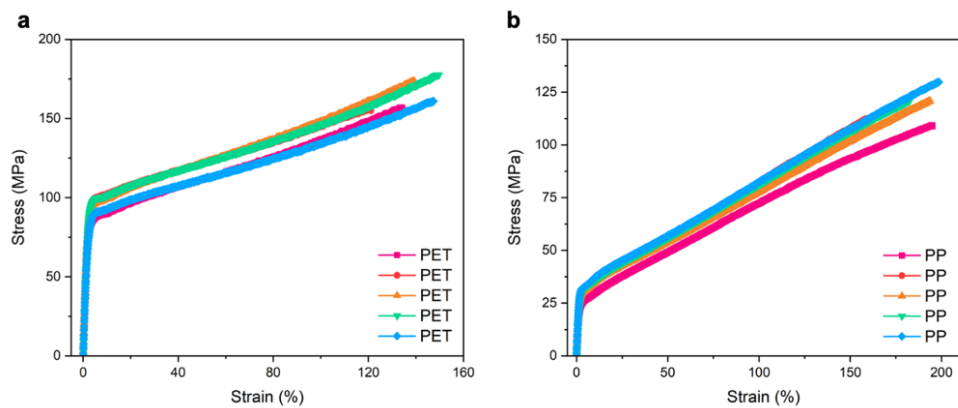


**Supplementary Figure 26. Biofabrication of large MECHS prototype.** Photographs show **a** casting of curli biomass on to a silicone mold, **b** ambient dried MECHS, **c** peeling of MECHS film from the silicone mold and **d** free-standing flexible MECHS film of lateral dimension 10 cm by 25 cm.

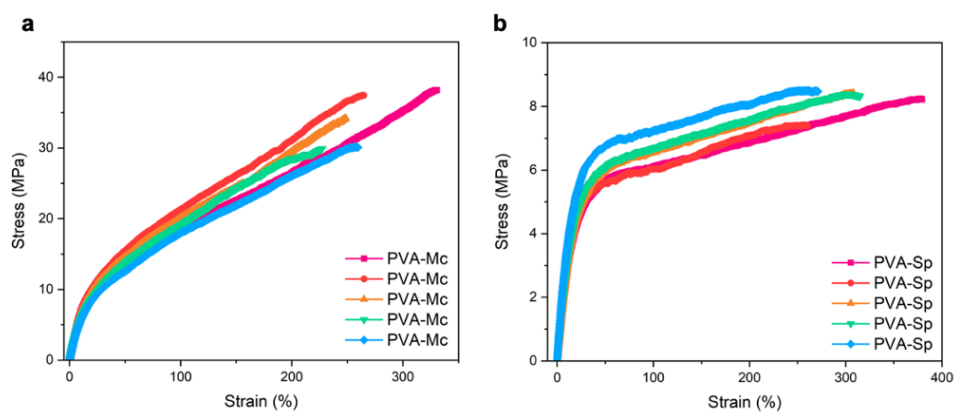




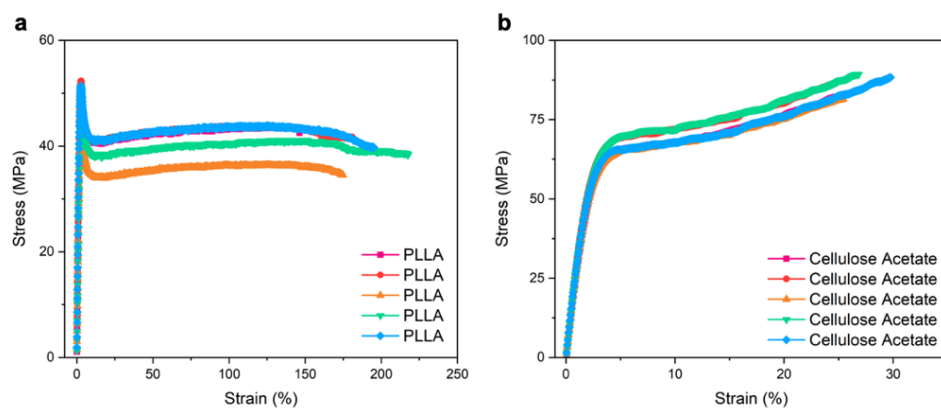
**Supplementary Figure 27. Tensile tests.** Stress strain curves of **a** Low density polyethylene (LDPE), **b** Polyvinyl chloride (PVC) and **c** Polytetrafluoroethylene (PTFE).



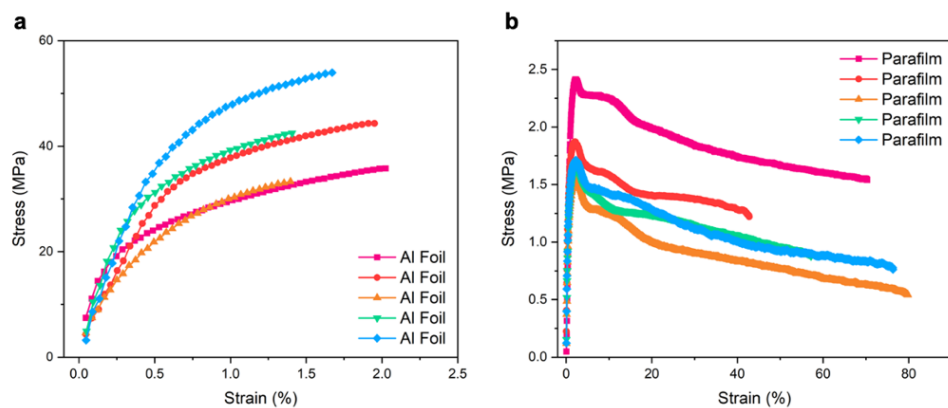
**Supplementary Figure 28. Tensile tests.** Stress strain curves of **a** Polyethylene terephthalate (PET) and **b** Polypropylene (PP).



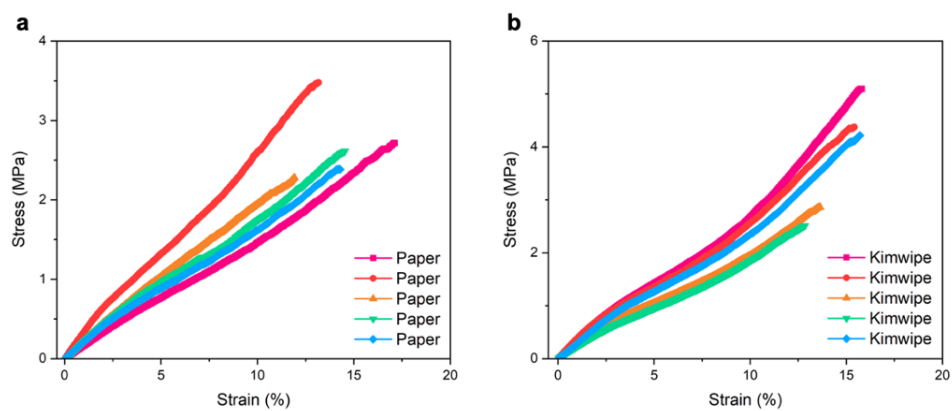
**Supplementary Figure 29. Tensile tests.** Stress strain curves of **a** PVA-McKesson and **b** PVA-Superpunch. PVA: Polyvinyl alcohol.



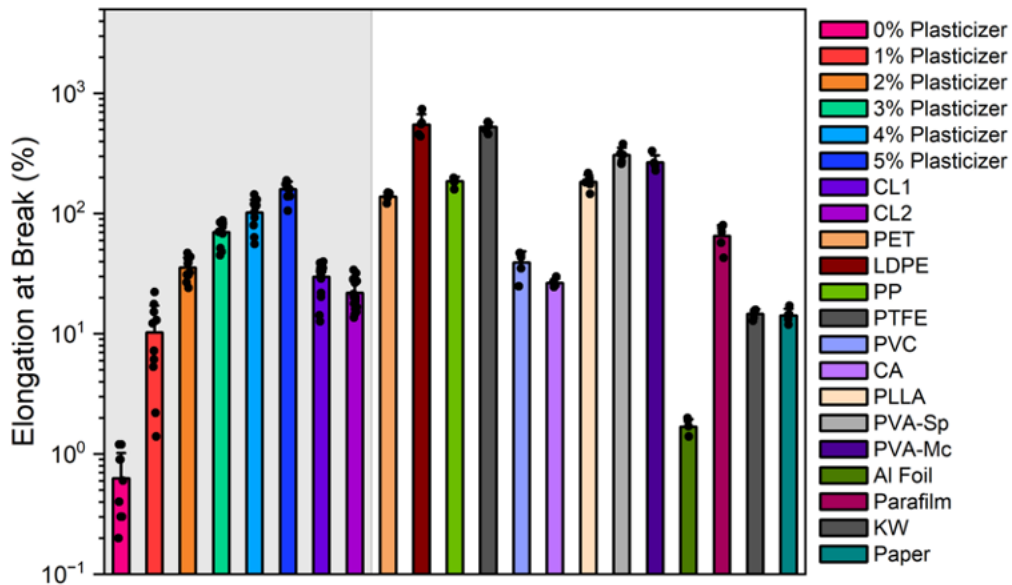
**Supplementary Figure 30. Tensile tests.** Stress strain curves of **a** Poly-L-lactic acid (PLLA) and **b** Cellulose Acetate.



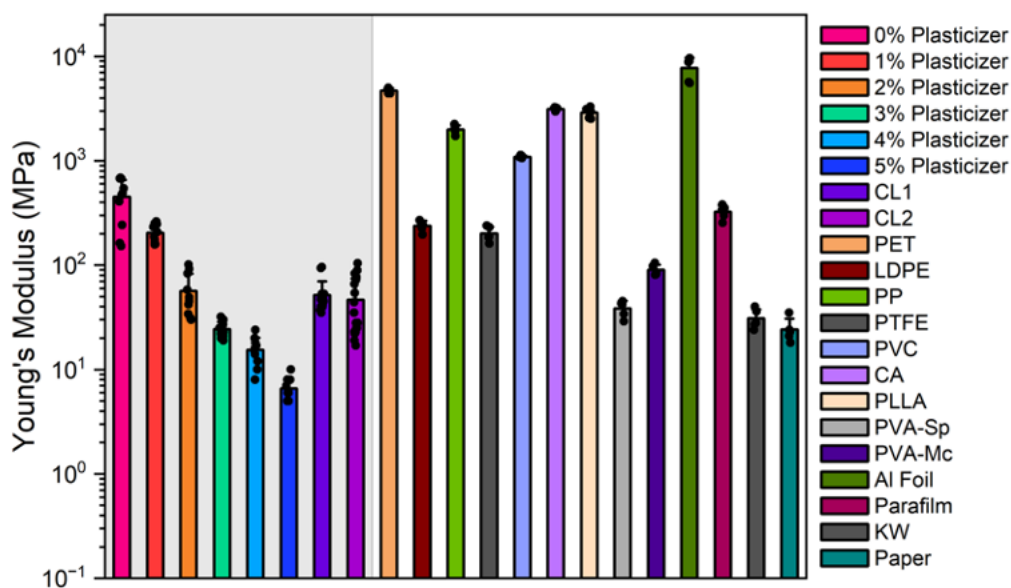
**Supplementary Figure 31. Tensile tests.** Stress strain curves of **a** Aluminum foil and **b** parafilm.



**Supplementary Figure 32. Tensile tests.** Stress strain curves of **a** Toilet paper and **b** Kimwipe.

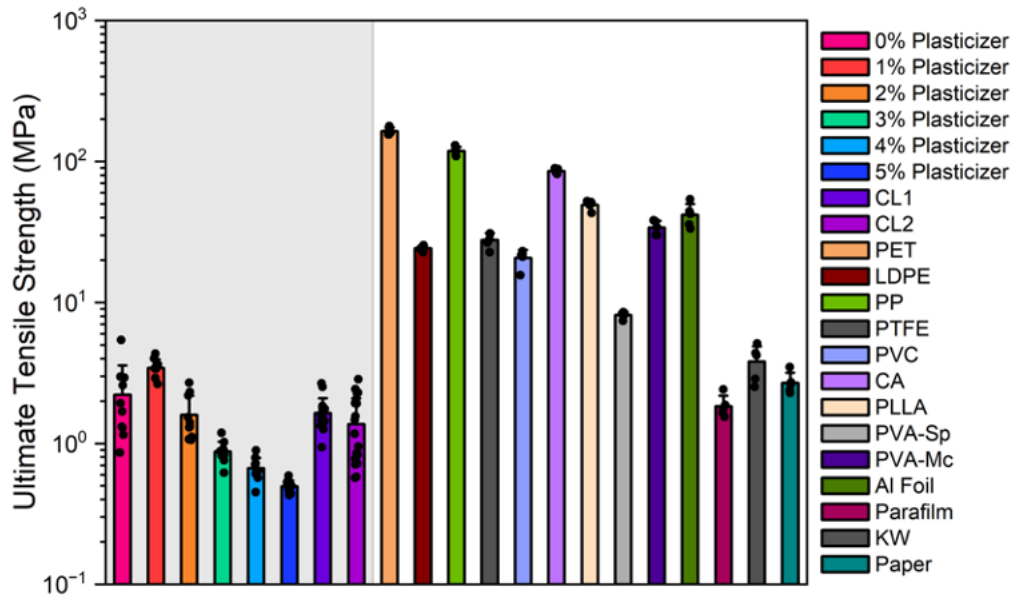


**Supplementary Figure 33. Tensile tests.** Plot shows the elongation at break for MECHS, various synthetic materials and biomaterials. Biological replicates  $n = 10$  for 0-5% plasticizer,  $n = 15$  for CL1,  $n = 20$  for CL2 and  $n = 5$  for all other samples. Low density polyethylene (LDPE), Polytetrafluoroethylene (PTFE), Poly-L-lactic acid (PLLA), Polyethylene terephthalate (PET), Cellulose acetate (CA), Polypropylene (PP), Polyvinyl chloride (PVC), Polyvinyl alcohol - Superpunch (PVA-Sp), Polyvinyl alcohol - Mckesson (PVA-Mc), Aluminum foil (Al Foil), Parafilm, Kimwipes (KW) and Toilet paper.



**Supplementary Figure 34. Tensile tests.** Plot shows the Young's modulus for MECHS, various synthetic materials and biomaterials. Biological replicates  $n = 10$  for 0-5% plasticizer,  $n = 15$  for CL1,  $n = 20$  for CL2 and  $n = 5$  for all other samples. Low density polyethylene (LDPE), Polytetrafluoroethylene (PTFE), Poly-L-lactic acid (PLLA), Polyethylene terephthalate (PET), Cellulose acetate (CA), Polypropylene (PP), Polyvinyl chloride (PVC), Polyvinyl alcohol - Superpunch (PVA-Sp), Polyvinyl alcohol - Mckesson (PVA-Mc), Aluminum foil (Al Foil), Parafilm, Kimwipes (KW) and Toilet paper.





**Supplementary Figure 35. Tensile tests.** Plot shows the ultimate tensile strength for MECHS, various synthetic materials and biomaterials. Biological replicates  $n = 10$  for 0-5% plasticizer,  $n = 15$  for CL1,  $n = 20$  for CL2 and  $n = 5$  for all other samples. Low density polyethylene (LDPE), Polytetrafluoroethylene (PTFE), Poly-L-lactic acid (PLLA), Polyethylene terephthalate (PET), Cellulose acetate (CA), Polypropylene (PP), Polyvinyl chloride (PVC), Polyvinyl alcohol - Superpunch (PVA-Sp), Polyvinyl alcohol - Mckesson (PVA-Mc), Aluminum foil (Al Foil), Parafilm, Kimwipes (KW) and Toilet paper.