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Pest categorisation of potato leafroll virus (non-EU isolates)

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Abstract

Following a request from the EU Commission, the Panel on Plant Health has addressed the pest categorisation of non-EU isolates of potato leafroll virus (PLRV). The information currently available on geographical distribution, biology, epidemiology, potential entry pathways, potential additional impact and availability of control measures of non-EU isolates of PLRV has been evaluated with regard to the criteria to qualify as a potential Union quarantine pest. Because non-EU isolates of PLRV are absent from the EU, they do not meet one of the requirements to be regulated as a regulated non-quarantine pest (RNQP) (presence in the EU); as a consequence, the Panel decided not to evaluate the other RNQP criteria for these isolates. This categorisation was performed considering two groups of PLRV isolates: those associated with the tomato yellow top disease (PLRV-TYTV), not reported from the EU, and all other isolates (hereafter referred to as PLRV), with a worldwide distribution. Isolates of PLRV-TYTV could potentially have an additional impact over the current situation in the EU and therefore meet all the criteria to qualify as a potential Union quarantine pest. All other non-EU PLRV isolates, should they be introduced, are not expected to have additional impact and therefore do not meet this criterion to qualify as a potential Union quarantine pest.

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Keywords: European Union, pest risk, plant pest, PLRV, tomato yellow top disease, TYTV, quarantine

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Figure 1: © CABI



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1. Introduction

1.1. Background and Terms of Reference as provided by the requestor

1.1.1. Background

Council Directive 2000/29/EC¹ on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community establishes the present European Union plant health regime. The Directive lays down the phytosanitary provisions and the control checks to be carried out at the place of origin on plants and plant products destined for the Union or to be moved within the Union. In the Directive's 2000/29/EC annexes, the list of harmful organisms (pests) whose introduction into or spread within the Union is prohibited, is detailed together with specific requirements for import or internal movement.

Following the evaluation of the plant health regime, the new basic plant health law, Regulation (EU) 2016/2031² on protective measures against pests of plants, was adopted on 26 October 2016 and will apply from 14 December 2019 onwards, repealing Directive 2000/29/EC. In line with the principles of the above mentioned legislation and the follow-up work of the secondary legislation for the listing of EU regulated pests, EFSA is requested to provide pest categorisations of the harmful organisms included in the annexes of Directive 2000/29/EC, in the cases where recent pest risk assessment/pest categorisation is not available.

1.1.2. Terms of reference

EFSA is requested, pursuant to Article 22(5.b) and Article 29(1) of Regulation (EC) No 178/2002³, to provide scientific opinion in the field of plant health.

EFSA is requested to prepare and deliver a pest categorisation (step 1 analysis) for each of the regulated pests included in the appendices of the annex to this mandate. The methodology and template of pest categorisation have already been developed in past mandates for the organisms listed in Annex II Part A Section II of Directive 2000/29/EC. The same methodology and outcome is expected for this work as well.

The list of the harmful organisms included in the annex to this mandate comprises 133 harmful organisms or groups. A pest categorisation is expected for these 133 pests or groups and the delivery of the work would be stepwise at regular intervals through the year as detailed below. First priority covers the harmful organisms included in Appendix 1, comprising pests from Annex II Part A Section I and Annex II Part B of Directive 2000/29/EC. The delivery of all pest categorisations for the pests included in Appendix 1 is June 2018. The second priority is the pests included in Appendix 2, comprising the group of *Cicadellidae* (non-EU) known to be vector of Pierce's disease (caused by *Xylella fastidiosa*), the group of *Tephritidae* (non-EU), the group of potato viruses and virus-like organisms, the group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L., and the group of *Margarodes* (non-EU species). The delivery of all pest categorisations for the pests included in Appendix 2 is end 2019. The pests included in Appendix 3 cover pests of Annex I part A section I and all pest categorisations should be delivered by end 2020.

For the above mentioned groups, each covering a large number of pests, the pest categorisation will be performed for the group and not the individual harmful organisms listed under "such as" notation in the Annexes of the Directive 2000/29/EC. The criteria to be taken particularly under consideration for these cases, is the analysis of host pest combination, investigation of pathways, the damages occurring and the relevant impact.

Finally, as indicated in the text above, all references to 'non-European' should be avoided and replaced by 'non-EU' and refer to all territories with exception of the Union territories as defined in Article 1 point 3 of Regulation (EU) 2016/2031.

¹ Council Directive 2000/29/EC of 8 May 2000 on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community. OJ L 169/1, 10.7.2000, p. 1–112.

² Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants. OJ L 317, 23.11.2016, p. 4–104.

³ Regulation (EC) No 178/2002 of the European Parliament and of the Council of 28 January 2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. OJ L 31/1, 1.2.2002, p. 1–24.

1.1.2.1. Terms of Reference: Appendix 1

List of harmful organisms for which pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

Annex IIAI

(a) Insects, mites and nematodes, at all stages of their development

<i>Aleurocanthus</i> spp.	<i>Numonia pyrivorella</i> (Matsumura)
<i>Anthonomus bisignifer</i> (Schenkling)	<i>Oligonychus perditus</i> Pritchard and Baker
<i>Anthonomus signatus</i> (Say)	<i>Pissodes</i> spp. (non-EU)
<i>Aschistonyx eppoi</i> Inouye	<i>Scirtothrips aurantii</i> Faure
<i>Carposina niponensis</i> Walsingham	<i>Scirtothrips citri</i> (Moultex)
<i>Enarmonia packardi</i> (Zeller)	<i>Scolytidae</i> spp. (non-EU)
<i>Enarmonia prunivora</i> Walsh	<i>Scrobipalopsis solanivora</i> Povolny
<i>Grapholita inopinata</i> Heinrich	<i>Tachypterellus quadrigibbus</i> Say
<i>Hishomonus phycitidis</i>	<i>Toxoptera citricida</i> Kirk.
<i>Leucaspis japonica</i> Ckll.	<i>Unaspis citri</i> Comstock
<i>Listronotus bonariensis</i> (Kuschel)	

(b) Bacteria

Citrus variegated chlorosis	<i>Xanthomonas campestris</i> pv. <i>oryzae</i> (Ishiyama)
<i>Erwinia stewartii</i> (Smith) Dye	Dye and pv. <i>oryzicola</i> (Fang, et al.) Dye

(c) Fungi

<i>Alternaria alternata</i> (Fr.) Keissler (non-EU pathogenic isolates)	<i>Elsinoe</i> spp. Bitanc. and Jenk. Mendes
<i>Anisogramma anomala</i> (Peck) E. Müller	<i>Fusarium oxysporum</i> f. sp. <i>albedinis</i> (Kilian and Maire) Gordon
<i>Apiosporina morbosa</i> (Schwein.) v. Arx	<i>Guignardia piricola</i> (Nosa) Yamamoto
<i>Ceratocystis virescens</i> (Davidson) Moreau	<i>Puccinia pittieriana</i> Hennings
<i>Cercoseptoria pini-densiflorae</i> (Hori and Nambu) Deighton	<i>Stegophora ulmea</i> (Schweinitz: Fries) Sydow & Sydow
<i>Cercospora angolensis</i> Carv. and Mendes	<i>Venturia nashicola</i> Tanaka and Yamamoto

(d) Virus and virus-like organisms

Beet curly top virus (non-EU isolates)	Little cherry pathogen (non- EU isolates)
Black raspberry latent virus	Naturally spreading psorosis
Blight and blight-like	Palm lethal yellowing mycoplasma
Cadang-Cadang viroid	Satsuma dwarf virus
Citrus tristeza virus (non-EU isolates)	Tatter leaf virus
Leprosis	Witches' broom (MLO)

Annex IIB

(a) Insect mites and nematodes, at all stages of their development

<i>Anthonomus grandis</i> (Boh.)	<i>Ips cembrae</i> Heer
<i>Cephalcia lariciphila</i> (Klug)	<i>Ips duplicatus</i> Sahlberg
<i>Dendroctonus micans</i> Kugelán	<i>Ips sexdentatus</i> Börner
<i>Gilpinia hercyniae</i> (Hartig)	<i>Ips typographus</i> Heer
<i>Gonipterus scutellatus</i> Gyll.	<i>Sternochetus mangiferae</i> Fabricius
<i>Ips amitinus</i> Eichhof	

(b) Bacteria

Curtobacterium flaccumfaciens pv. *flaccumfaciens*
(Hedges) Collins and Jones

(c) Fungi

Glomerella gossypii Edgerton

Hypoxyton mammatum (Wahl.) J. Miller

Gremmeniella abietina (Lag.) Morelet

1.1.2.2. Terms of Reference: Appendix 2

List of harmful organisms for which pest categorisation is requested per group. The list below follows the categorisation included in the annexes of Directive 2000/29/EC.

Annex IAI**(a) Insects, mites and nematodes, at all stages of their development**

Group of Cicadellidae (non-EU) known to be vector of Pierce's disease (caused by *Xylella fastidiosa*), such as:

- | | |
|--|---|
| 1) <i>Carneocephala fulgida</i> Nottingham | 3) <i>Graphocephala atropunctata</i> (Signoret) |
| 2) <i>Draeculacephala minerva</i> Ball | |

Group of Tephritidae (non-EU) such as:

- | | |
|--|---|
| 1) <i>Anastrepha fraterculus</i> (Wiedemann) | 12) <i>Pardalaspis cyanescens</i> Bezzi |
| 2) <i>Anastrepha ludens</i> (Loew) | 13) <i>Pardalaspis quinaria</i> Bezzi |
| 3) <i>Anastrepha obliqua</i> Macquart | 14) <i>Pterandrus rosa</i> (Karsch) |
| 4) <i>Anastrepha suspensa</i> (Loew) | 15) <i>Rhacochlaena japonica</i> Ito |
| 5) <i>Dacus ciliatus</i> Loew | 16) <i>Rhagoletis completa</i> Cresson |
| 6) <i>Dacus curcurbitae</i> Coquillett | 17) <i>Rhagoletis fausta</i> (Osten-Sacken) |
| 7) <i>Dacus dorsalis</i> Hendel | 18) <i>Rhagoletis indifferens</i> Curran |
| 8) <i>Dacus tryoni</i> (Froggatt) | 19) <i>Rhagoletis mendax</i> Curran |
| 9) <i>Dacus tsuneonis</i> Miyake | 20) <i>Rhagoletis pomonella</i> Walsh |
| 10) <i>Dacus zonatus</i> Saund. | 21) <i>Rhagoletis suavis</i> (Loew) |
| 11) <i>Epochra canadensis</i> (Loew) | |

(c) Viruses and virus-like organisms

Group of potato viruses and virus-like organisms such as:

- | | |
|----------------------------------|--|
| 1) Andean potato latent virus | 5) Potato virus T |
| 2) Andean potato mottle virus | 6) non-EU isolates of potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc) and Potato leafroll virus |
| 3) Arracacha virus B, oca strain | |
| 4) Potato black ringspot virus | |

Group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L., such as:

- | | |
|--------------------------------------|--|
| 1) Blueberry leaf mottle virus | 8) Peach yellows mycoplasma |
| 2) Cherry rasp leaf virus (American) | 9) Plum line pattern virus (American) |
| 3) Peach mosaic virus (American) | 10) Raspberry leaf curl virus (American) |
| 4) Peach phony rickettsia | 11) Strawberry witches' broom mycoplasma |
| 5) Peach rosette mosaic virus | 12) Non-EU viruses and virus-like organisms of <i>Cydonia</i> Mill., <i>Fragaria</i> L., <i>Malus</i> Mill., <i>Prunus</i> L., <i>Pyrus</i> L., <i>Ribes</i> L., <i>Rubus</i> L. and <i>Vitis</i> L. |
| 6) Peach rosette mycoplasma | |
| 7) Peach X-disease mycoplasma | |

Annex IIAI**(a) Insects, mites and nematodes, at all stages of their development**

Group of *Margarodes* (non-EU species) such as:

- | | |
|--|--|
| 1) <i>Margarodes vitis</i> (Phillipi) | 3) <i>Margarodes prieskaensis</i> Jakubski |
| 2) <i>Margarodes vredendalensis</i> de Klerk | |

1.1.2.3. Terms of Reference: Appendix 3

List of harmful organisms for which pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

Annex IAI**(a) Insects, mites and nematodes, at all stages of their development**

<i>Acleris</i> spp. (non-EU)	<i>Longidorus diadecturus</i> Eveleigh and Allen
<i>Amauromyza maculosa</i> (Malloch)	<i>Monochamus</i> spp. (non-EU)
<i>Anomala orientalis</i> Waterhouse	<i>Myndus crudus</i> Van Duzee
<i>Arrhenodes minutus</i> Drury	<i>Nacobbus aberrans</i> (Thorne) Thorne and Allen
<i>Choristoneura</i> spp. (non-EU)	<i>Naupactus leucoloma</i> Boheman
<i>Conotrachelus nenuphar</i> (Herbst)	<i>Premnotrypes</i> spp. (non-EU)
<i>Dendrolimus sibiricus</i> Tschetverikov	<i>Pseudopityophthorus minutissimus</i> (Zimmermann)
<i>Diabrotica barberi</i> Smith and Lawrence	<i>Pseudopityophthorus pruinosus</i> (Eichhoff)
<i>Diabrotica undecimpunctata howardi</i> Barber	<i>Scaphoideus luteolus</i> (Van Duzee)
<i>Diabrotica undecimpunctata undecimpunctata</i> Mannerheim	<i>Spodoptera eridania</i> (Cramer)
<i>Diabrotica virgifera zea</i> Krysan & Smith	<i>Spodoptera frugiperda</i> (Smith)
<i>Diaphorina citri</i> Kuway	<i>Spodoptera litura</i> (Fabricus)
<i>Heliothis zea</i> (Boddie)	<i>Thrips palmi</i> Karny
<i>Hirschmanniella</i> spp., other than <i>Hirschmanniella gracilis</i> (de Man) Luc and Goodey	<i>Xiphinema americanum</i> Cobb sensu lato (non-EU populations)
<i>Liriomyza sativae</i> Blanchard	<i>Xiphinema californicum</i> Lamberti and Bleve-Zacheo

(b) Fung

<i>Ceratocystis fagacearum</i> (Bretz) Hunt	<i>Mycosphaerella larici-leptolepis</i> Ito et al.
<i>Chrysomyxa arctostaphyli</i> Diете	<i>Mycosphaerella populorum</i> G. E. Thompson
<i>Cronartium</i> spp. (non-EU)	<i>Phoma andina</i> Turkensteen
<i>Endocronartium</i> spp. (non-EU)	<i>Phyllosticta solitaria</i> Ell. and Ev.
<i>Guignardia laricina</i> (Saw.) Yamamoto and Ito	<i>Septoria lycopersici</i> Speg. var. <i>malagutii</i> Ciccarone and Boerema
<i>Gymnosporangium</i> spp. (non-EU)	<i>Thecaphora solani</i> Barrus
<i>Inonotus weirii</i> (Murril) Kotlaba and Pouzar	<i>Trechispora brinkmannii</i> (Bresad.) Rogers
<i>Melampsora farlowii</i> (Arthur) Davis	

(c) Viruses and virus-like organisms

Tobacco ringspot virus	Pepper mild tigré virus
Tomato ringspot virus	Squash leaf curl virus
Bean golden mosaic virus	Euphorbia mosaic virus
Cowpea mild mottle virus	Florida tomato virus
Lettuce infectious yellows virus	

(d) Parasitic plants

Arceuthobium spp. (non-EU)

Annex I A I I**(a) Insects, mites and nematodes, at all stages of their development**

Meloidogyne fallax Karssen

Rhizoecus hibisci Kawai and Takagi

Popillia japonica Newman

(b) Bacteria

Clavibacter michiganensis (Smith) Davis et al. ssp. *Ralstonia solanacearum* (Smith) Yabuuchi et al. *sepedonicus* (Spieckermann and Kotthoff) Davis et al.

(c) Fungi

Melampsora medusae Thümen

Synchytrium endobioticum (Schilbersky) Percival

Annex I B**(a) Insects, mites and nematodes, at all stages of their development**

Leptinotarsa decemlineata Say

Liriomyza bryoniae (Kaltenbach)

(b) Viruses and virus-like organisms

Beet necrotic yellow vein virus

1.2. Interpretation of the Terms of Reference

EFSA is asked to develop pest categorisations for non-EU isolates of seven potato viruses, i.e. potato leafroll virus and potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc), which are defined by their geographical origin outside the EU. As such, isolates of these viruses occurring outside the EU territory are considered as non-EU isolates. Accordingly, a virus isolate infecting a plant originating in a non-EU country is considered to be a non-EU isolate. All seven viruses are important pathogens of potato and, therefore, there is no uncertainty about the fact that non-EU isolates have an impact on potato crops in absolute terms. However, EU isolates of these viruses already have an impact in the EU; consequently, the Panel decided to evaluate whether the non-EU isolates would have an additional impact compared to the current situation, upon introduction and spread in the EU. This interpretation was agreed with the European Commission.

This scientific opinion presents the pest categorisation of non-EU isolates of potato leafroll virus (PLRV). Non-EU isolates of PLRV are listed in the Appendices of the Terms of Reference (ToR) to be subject to pest categorisation to determine whether they fulfil the criteria of a quarantine pest for the area of the EU excluding Ceuta, Melilla and the outermost regions of Member States (MSs) referred to in Article 355 (1) of the Treaty on the Functioning of the European Union (TFEU), other than Madeira and the Azores.

Because non-EU isolates of PLRV are absent from the EU, they do not meet one of the requirements to be regulated as a regulated non-quarantine pest (RNQP) (presence in the EU); as a consequence, the Panel decided not to evaluate the other RNQP criteria for these isolates.

Despite the fact that *Solanum phureja* is considered by some authorities as an invalid taxon that should be renamed *Solanum tuberosum* Phureja Group,⁴ the Panel considered the uncertainty on this aspect high enough that it decided, in line with the EPPD Global Database, to separately address *S. phureja* as a distinct entity regulated within the 'potato and other tuber forming *Solanum* species' in Directive 2000/29/EC.

The new Plant Health Regulation (EU) 2016/2031⁵, on the protective measures against pests of plants, will be applying from December 2019. The regulatory status sections (Section 3.3.) of the

⁴ See https://ec.europa.eu/food/sites/food/files/plant/docs/sc_spmah_20160205_sum.pdf

⁵ Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants, amending Regulations (EU) 228/2013, (EU) 652/2014 and (EU) 1143/2014 of the European Parliament and of the Council and repealing Council Directives 69/464/EEC, 74/647/EEC, 93/85/EEC, 98/57/EC, 2000/29/EC, 2006/91/EC and 2007/33/EC. OJ L 317, 23.11.2016, pp. 4–104.

present opinion are still based on Council Directive 2000/29/EC, as the document was adopted in November 2019.

2. Data and methodologies

2.1. Data

2.1.1. Literature search

A literature search on PLRV was conducted in the ISI Web of Science bibliographic database. The scientific name of the pest was used as search term. Relevant papers were reviewed with a focus on potential differences between isolates and strains. Further references and information were obtained from experts, as well as from citations in the reviewed papers and grey literature. The search was continued until no further information could be found or until the collected information was considered sufficient to perform the pest categorisation; consequently, the presented data is not necessarily exhaustive.

2.1.2. Database search

Information on hosts, vectors and distribution at species level, was retrieved from CABI Crop Protection Compendium (CABI cpc), the EPPO Global Database, and relevant publications. Additional data on isolates distribution was obtained from the literature.

Data about the import of commodity types that could potentially provide a pathway for the pest to enter the EU and about the area of hosts grown in the EU were obtained from EUROSTAT (Statistical Office of the European Communities).

The Europhyt database was consulted to identify interceptions of non-EU isolates of PLRV. Europhyt is a web-based network run by the Directorate General for Health and Food Safety (DG SANTÉ) of the European Commission and is a subproject of PHYSAN (Phyto-Sanitary Controls) specifically concerned with plant health information. The Europhyt database manages notifications of interceptions of plants or plant products that do not comply with EU legislation, as well as notifications of plant pests detected in the territory of the MSs and the phytosanitary measures taken to eradicate or avoid their spread.

2.2. Methodologies

The Panel performed the pest categorisation for non-EU isolates of PLRV, following the guiding principles and steps presented in the EFSA guidance on quantitative pest risk assessment (EFSA PLH Panel, 2018) and in the International Standard for Phytosanitary Measures No 11 (FAO, 2013) and No 21 (FAO, 2004).

General information on PLRV will be provided at species level. Further information will be added at the level of strains and/or non-EU isolates when available and applicable.

This work was initiated following an evaluation of the EU plant health regime. Therefore, to facilitate the decision-making process, in the conclusions of the pest categorisation, the Panel addresses explicitly each criterion for a Union quarantine pest in accordance with Regulation (EU) 2016/2031 on protective measures against pests of plants, and includes additional information required in accordance with the specific terms of reference received by the European Commission. As explained in the interpretation of the Terms of Reference, the criterion on impact focuses on additional impact of non-EU isolates of PLRV. For each conclusion, the Panel provides a short description of its associated uncertainty.

Table 1 presents the Regulation (EU) 2016/2031 pest categorisation criteria on which the Panel bases its conclusions. All relevant criteria have to be met for the pest to potentially qualify as a quarantine pest. If one of the criteria is not met, the pest will not qualify.

It should be noted that the Panel's conclusions are formulated respecting its remit and particularly with regard to the principle of separation between risk assessment and risk management (EFSA founding regulation (EU) No 178/2002); therefore, the Panel will present a summary of the reported impacts. Impacts are expressed in terms of yield and quality losses and not in monetary terms, whereas addressing social impacts is outside the remit of the Panel.

Table 1: Pest categorisation criteria under evaluation, as defined in Regulation (EU) 2016/2031 on protective measures against pests of plants (the number of the relevant sections of the pest categorisation is shown in brackets in the first column)

Criterion of pest categorisation	Criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Criterion in Regulation (EU) 2016/2031 regarding protected zone quarantine pest (articles 32–35)	Criterion in Regulation (EU) 2016/2031 regarding Union regulated non-quarantine pest
Identity of the pest (Section 3.1)	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?
Absence/ presence of the pest in the EU territory (Section 3.2)	Is the pest present in the EU territory? If present, is the pest widely distributed within the EU? Describe the pest distribution briefly!	Is the pest present in the EU territory? If not, it cannot be a protected zone quarantine organism	Is the pest present in the EU territory? If not, it cannot be a RNQP. (A regulated non-quarantine pest must be present in the risk assessment area)
Regulatory status (Section 3.3)	If the pest is present in the EU but not widely distributed in the risk assessment area, it should be under official control or expected to be under official control in the near future	The protected zone system aligns with the pest free area system under the International Plant Protection Convention (IPPC) The pest satisfies the IPPC definition of a quarantine pest that is not present in the risk assessment area (i.e. protected zone)	Is the pest regulated as a quarantine pest? If currently regulated as a quarantine pest, are there grounds to consider its status could be revoked?
Pest potential for entry, establishment and spread in the EU territory (Section 3.4)	Is the pest able to enter into, become established in, and spread within, the EU territory? If yes, briefly list the pathways!	Is the pest able to enter into, become established in, and spread within, the protected zone areas? Is entry by natural spread from EU areas where the pest is present possible?	Is spread mainly via specific plants for planting, rather than via natural spread or via movement of plant products or other objects? Clearly state if plants for planting is the main pathway!
Potential for consequences in the EU territory (Section 3.5)	Would the pests' introduction have an economic or environmental impact on the EU territory?	Would the pests' introduction have an economic or environmental impact on the protected zone areas?	Does the presence of the pest on plants for planting have an economic impact as regards the intended use of those plants for planting?
Available measures (Section 3.6)	Are there measures available to prevent the entry into, establishment within or spread of the pest within the EU such that the risk becomes mitigated?	Are there measures available to prevent the entry into, establishment within or spread of the pest within the protected zone areas such that the risk becomes mitigated? Is it possible to eradicate the pest in a restricted area within 24 months (or a period longer than 24 months where the biology of the organism so justifies) after the presence of the pest was confirmed in the protected zone?	Are there measures available to prevent pest presence on plants for planting such that the risk becomes mitigated?

Criterion of pest categorisation	Criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Criterion in Regulation (EU) 2016/2031 regarding protected zone quarantine pest (articles 32–35)	Criterion in Regulation (EU) 2016/2031 regarding Union regulated non-quarantine pest
Conclusion of pest categorisation (Section 4)	A statement as to whether (1) all criteria assessed by EFSA above for consideration as a potential quarantine pest were met and (2) if not, which one(s) were not met	A statement as to whether (1) all criteria assessed by EFSA above for consideration as potential protected zone quarantine pest were met, and (2) if not, which one(s) were not met	A statement as to whether (1) all criteria assessed by EFSA above for consideration as a potential RNQP were met, and (2) if not, which one(s) were not met

The Panel will not indicate in its conclusions of the pest categorisation whether to continue the risk assessment process, but following the agreed two-step approach, will continue only if requested by the risk managers. However, during the categorisation process, experts may identify key elements and knowledge gaps that could contribute significant uncertainty to a future assessment of risk. It would be useful to identify and highlight such gaps so that potential future requests can specifically target the major elements of uncertainty, perhaps suggesting specific scenarios to examine.

2.3. Nomenclature

Virus nomenclature is reported using the latest release of the official classification by the International Committee on Taxonomy of Viruses (ICTV, Release 2018b.v1, <https://talk.ictvonline.org/taxonomy/>). Virus names are not italicised throughout this opinion, corresponding to ICTV instructions.

3. Pest categorisation

3.1. Identity and biology of the pest

3.1.1. Identity and taxonomy

Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?

Yes. PVY is a well-known virus and the definition of 'non-EU isolates', as used in the present opinion, has been clarified (See Section 1.2).

Potato leafroll virus is a well-characterised virus in the genus *Polerovirus*, family Luteoviridae (Domier, 2011). It has a single-stranded positive-sense RNA genome, which contains six open reading frames (ORFs) numbered from 0 to 5 (Haupt et al., 2005). Complete and/or partial genomic sequences are available for a number of isolates.

3.1.2. Biology of the pest

PLRV, including PLRV-TYTV, is neither known to be transmitted by pollen or true seeds, nor mechanically (Hassan et al., 1985; Mink, 1993; Mayo et al., 2000; Sastry, 2013). It is transmitted by vegetative propagation (via tubers) (Loebenstein and Gaba, 2012) and most of the isolates, including PLRV-TYTV, are reported to be transmitted by aphids (Hemiptera: Aphididae) such as *Aphis fabae* (Scopoli), *Aphis gossypii* (Glover), *Macrosiphum euphorbiae* (Thomas) and *Myzus persicae* (Sulzer) in a persistent and circulative manner (Hassan and Thomas, 1984; Khaled et al., 2018; CABI cpc, 2019). Aphid transmission failed for some isolates or is at least reported as inefficient (Tamada et al., 1984; Rouze-Jouan et al., 2001).

3.1.3. Intraspecific diversity

Viruses generally exist as quasispecies, which means that they accumulate as a cluster of closely related sequence variants in a single host (Andino and Domingo, 2015). This is likely due to competition among the genomic variants that are generated as a consequence of the error-prone viral replication (higher in RNA than in DNA viruses) and the ensuing selection of the most fit variants in a

given environment (Domingo et al., 2012). This genetic variability may have consequences on the virus' biological properties (e.g. host range, transmissibility, and pathogenicity) as well as on the reliability of detection methods, especially when they target variable genomic regions.

This pest categorisation focuses on taxonomic levels below the species level, i.e. on isolates and strains, which are defined as follows:

- **Isolate:** virus population as present in a plant;
- **Strain:** group of isolates sharing biological, molecular, and/or serological properties (Garcia-Arenal et al., 2001).

ICTV does not address taxonomic levels below species level and, therefore, the names of strains are based on reports in literature. In the past, the term 'strain' has also often been used as a synonym for 'isolate'. As a consequence of this inconsistent use of terminology, the literature is often unclear.

The overall sequence variation among PLRV isolates from around the world is low and no strains have been distinguished (Keese et al., 1990; Faccioli et al., 1995; Guyader and Ducray, 2002; Mukherjee et al., 2003; Hühnlein et al., 2016). ORF0 is reported as the least conserved coding region among polioviruses (Guyader and Ducray, 2002), and would therefore be most suitable to examine genetic diversity below the species level. ORF0 encodes a host range determinant that functions as a suppressor of RNA silencing (Domier, 2011), thus being linked to symptom expression (van der Wilk et al., 1997), and virus accumulation (Sadowy et al., 2001), which provides a potential link to impact, should there be genetic diversity. Based on phylogenetic analysis of ORF0 at nucleotide level, PLRV isolates could be divided in three groups (Group 1, 2, and 3) (Guyader and Ducray, 2002; Khouadja et al., 2005, 2014; Plchova et al., 2009). Based on the amino acid sequence of the protein encoded by ORF0, two groups were distinguished, corresponding to group 1 and group 2+3 of the previously mentioned division (Hühnlein et al., 2016). However, this grouping is based on a limited number of isolates and is not reported to be related to biological properties or geographical distribution; therefore, non-EU PLRV isolates will be categorised as a single ensemble without considering these phylogenetic groups.

There are two other groups of isolates reported, i.e. the South African (Roos, 2013) and PLRV-TYTV (Thomas, 1984; van den Heuvel et al., 1990). In an MSc thesis, Roos and Bellstedt (2013) report the existence of PLRV isolates in South-Africa that are different in genomic sequence and pathogenicity compared to other PLRV isolates. However, no data are reported to support the enhanced severity of these isolates. In addition, these isolates are not further mentioned in the literature and will therefore not be further considered in this pest categorisation.

PLRV-TYTV isolates were originally reported in *Solanum lycopersicum* as tomato yellow top virus (TYTV) (Thomas, 1984). A TYTV isolate from Brazil was later shown to belong to the species *Potato leafroll virus* (van den Heuvel et al., 1990). Isolates of TYTV were reported to produce more severe symptoms on tomato than other PLRV isolates (Hassan and Thomas, 1984), but to produce only mild symptoms in potato (Thomas, 1993). Therefore, PLRV-TYTV will be categorised separately (see table 2) while, as indicated above, all other isolates of PLRV will be categorised as a single group hereafter named 'PLRV'.

Table 2: Categorised virus and isolate in the present opinion

Group of isolates	Acronym	Other information	Key references
Potato leafroll virus isolates	PLRV	All PLRV isolates apart from PLRV-TYTV	Guyader and Ducray (2002), Hühnlein et al. (2016)
Potato leafroll virus-tomato yellow top isolates	PLRV-TYTV		Thomas (1984), van den Heuvel et al. (1990)

3.1.4. Detection and identification of the pest

Are detection and identification methods available for the pest?

Yes. Methods are available for detection and identification of PLRV at species level and therefore for the identification of non-EU isolates. Apart from a bioassay, there is currently no detection and identification method for PLRV-TYTV. Based on the single sequence available, a molecular method could be developed to identify PLRV-TYTV, with uncertainty on its specificity (inclusivity and exclusivity).

As mentioned in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), virus detection and identification is complicated by several recurrent uncertainties. ICTV lists species demarcation criteria, but it is not always clear whether these are met in diagnostic tests. Furthermore, in the absence or near absence of information on genetic variability, it is not possible to guarantee that a given test will detect all variants of a species. On the contrary, generic tests may detect closely related viruses in addition to the target species. This implies that the reliability of a test depends on its validation for the intended use. For initial screening, it is important to prevent false negative results, which means that the following performance characteristics are most relevant: analytical sensitivity, inclusivity of analytical specificity (coverage of the intraspecies variability) and selectivity (matrix effects). For identification, it is important to prevent false positives and, therefore, the possible occurrence of cross reactions should be determined, i.e. the exclusivity of the analytical specificity (the resolution should be sufficient to discriminate between related species).

PLRV is a well-known virus for which detection methods are available. Enzyme-linked immunosorbent assay (ELISA) and/or real-time polymerase chain reaction (RT-PCR) are available for the detection and identification of PLRV at species level (Agindotan et al., 2007; Chomič et al., 2010).

Inoculation on tomato and the evaluation of severity of symptoms could form the basis of a PLRV-TYTV-specific bioassay but its application would be cumbersome and impractical (inoculation by grafting or aphids and need of control isolates).

Apart from this bioassay, no specific method is currently available for the identification of PLRV-TYTV. However, based on the single sequence available (NCBI GenBank accession AF453397) (Guyader and Ducray, 2002), a method could be developed to identify PLRV-TYTV, with uncertainty on its specificity (inclusivity and exclusivity).

3.2. Pest distribution

3.2.1. Pest distribution outside the EU

PLRV occurs worldwide wherever potato is grown (Jeffries, 1998; Loebenstein and Gaba, 2012; CABI cpc, 2019).

PLRV-TYTV is reported in Australia, Brazil, New Zealand and the USA (Thomas, 1984).

3.2.2. Pest distribution in the EU

Is the pest present in the EU territory? If present, is the pest widely distributed within the EU?

Yes. PLRV is present in the EU.

No. PLRV-TYTV is not reported in the EU.

As indicated in the previous section, PLRV is reported worldwide, including many EU MSs (Cyprus, France, Germany, Greece, Poland, the Netherlands, Spain and the United Kingdom) (van der Wilk et al., 1989; Palucha et al., 1994; Loebenstein and Gaba, 2012; Hühnlein et al., 2016).

PLRV-TYTV is not reported in the EU. The absence of PLRV-TYTV is associated with uncertainties due to the absence of specific surveys. In addition, isolates might have been ('mis-') identified as PLRV when detected in other hosts than tomato.

3.3. Regulatory status

3.3.1. Council Directive 2000/29/EC

Non-EU isolates of PLRV are specifically listed in Council Directive 2000/29/EC and are regulated in Annex IAI (See Table 3).

Table 3: Non-EU isolates of PLRV in Council Directive 2000/29/EC

Annex I, Part A	Harmful organisms whose introduction into, and spread within, all member states shall be banned
Section I	Harmful organisms not known to occur in any part of the community and relevant for the entire community
(d)	Viruses and virus-like organisms
2.	Potato viruses and virus-like organisms such as: (g) non-European isolates of potato viruses A, M, S, V, X and Y (including Y ^o , Y ⁿ and Y ^c) and Potato leafroll virus

3.3.2. Legislation addressing potato

Table 4 reports on the articles in Council Directive 2000/29/EC which address potato or tuber-forming species of *Solanum* L. PLRV may also infect other hosts; references to the corresponding legislation are reported in Table 5 (see section 3.4.1).

Table 4: Overview of the regulation in Annexes III, IV and V of Council Directive 2000/29/EC that applies to potato or tuber-forming *Solanum* species

Annex III, Part A	Plants, plant products and other objects the introduction of which shall be prohibited in all Member States	
	Description	Country of origin
10.	Tubers of <i>Solanum tuberosum</i> L., seed potatoes	Third countries other than Switzerland
11.	Plants of stolon- or tuber-forming species of <i>Solanum</i> L. or their hybrids, intended for planting, other than those tubers of <i>Solanum tuberosum</i> L. as specified under Annex III A (10)	Third countries
12.	Tubers of species of <i>Solanum</i> L., and their hybrids, other than those specified in points 10 and 11	Without prejudice to the special requirements applicable to the potato tubers listed in Annex IV, Part A Section I, third countries other than Algeria, Egypt, Israel, Libya, Morocco, Syria, Switzerland, Tunisia and Turkey, and other than European third countries which are either recognised as being free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., in accordance with the procedure referred to in Article 18 (2), or in which provisions recognised as equivalent to the Community provisions on combating <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. in accordance with the procedure referred to in Article 18(2), have been complied with
Annex IV, Part A	Special requirements which shall be laid down by all member states for the introduction and movement of plants, plant products and other objects into and within all Member States	
Section I	Plants, plant products and other objects originating outside the Community	
	Plants, plant products and other objects	Special requirements
25.1	Tubers of <i>Solanum tuberosum</i> L., originating in countries where <i>Synchytrium endobioticum</i> (Schilbersky) Percival is known to occur	Without prejudice to the prohibitions applicable to the tubers listed in Annex III(A) (10), (11) and (12), official statement that: (a) the tubers originate in areas known to be free from <i>Synchytrium endobioticum</i> (Schilbersky) Percival (all races other than Race 1, the common European race), and no symptoms of <i>Synchytrium endobioticum</i> (Schilbersky) Percival have been observed either at the place of production or in its immediate vicinity since the beginning of an adequate period; or (b) provisions recognised as equivalent to the Community provisions on combating <i>Synchytrium endobioticum</i> (Schilbersky) Percival in accordance with the procedure referred to in Article 18(2) have been complied with, in the country of origin

25.2.	Tubers of <i>Solanum tuberosum</i> L.	<p>Without prejudice to the provisions listed in Annex (A) (10), (11) and (12) and Annex IV(A)(I) (25.1), official statement that:</p> <p>(a) the tubers originate in countries known to be free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al.;</p> <p>or</p> <p>(b) provisions recognised as equivalent to the Community provisions on combating <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. in accordance with the procedure referred to in Article 18 (2), have been complied with, in the country of origin</p>
25.3.	Tubers of <i>Solanum tuberosum</i> L., other than early potatoes, originating in countries where Potato spindle tuber viroid is known to occur	<p>Without prejudice to the provisions applicable to the tubers listed in Annex III(A) (10), (11) and (12) and Annex IV(A)(I) (25.1) and (25.2), suppression of the faculty of germination</p>
25.4.	Tubers of <i>Solanum tuberosum</i> L., intended for planting	<p>Without prejudice to the provisions applicable to the tubers listed in Annex III(A)(10), (11) and (12) and Annex IV(A)(I) (25.1), (25.2) and (25.3), official statement that the tubers originate from a field known to be free from <i>Globodera rostochiensis</i> (Wollenweber) Behrens and <i>Globodera pallida</i> (Stone) Behrens and</p> <p>(aa) either, the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known not to occur;</p> <p>or</p> <p>(bb) in areas where <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known to occur, the tubers originate from a place of production found free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al., or considered to be free thereof, as a consequence of the implementation of an appropriate procedure aiming at eradicating <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. which shall be determined in accordance with the procedure referred to in Article 18(2)</p> <p>and</p> <p>(cc) either the tubers originate in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known not to occur; or</p> <p>(dd) in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known to occur,</p> <p>— either the tubers originate from a place of production which has been found free from <i>Meloidogyne chitwoodi</i> Golden et al. (all populations), and <i>Meloidogyne fallax</i> Karssen based on an annual survey of host crops by visual inspection of host plants at appropriate times and by visual inspection both externally and by cutting of tubers after harvest from potato crops grown at the place of production, or</p> <p>— the tubers after harvest have been randomly sampled and, either checked for the presence of symptoms after an appropriate method to induce symptoms, or laboratory tested, as well as inspected visually both externally and by cutting the tubers, at appropriate times and in all cases at the time of closing of the packages or containers before marketing according to the provisions on closing in Council Directive 66/403/EEC of 14 June 1996 on the</p>

		marketing of seed potatoes (1) and no symptoms of <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen have been found
25.4.1.	Tubers of <i>Solanum tuberosum</i> L., other than those intended for planting	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (12) and Annex IV(A)(I) (25.1), (25.2) and (25.3), official statement that the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is not known to occur
25.4.2.	Tubers of <i>Solanum tuberosum</i> L.	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (10), (11) and (12) and Annex IV(A) (I) (25.1), (25.2), (25.3), (25.4) and (25.4.1), official statement that: (a) the tubers originate in a country where <i>Scrobipalopsis solanivora</i> Povolny is not known to occur; or (b) the tubers originate in an area free from <i>Scrobipalopsis solanivora</i> Povolny, established by the national plant protection organisation in accordance with relevant International Standards for Phytosanitary Measures
25.5.	Plants of Solanaceae, intended for planting, other than seeds, originating in countries where Potato stolbur mycoplasma is known to occur	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (10), (11), (12) and (13), and Annex IV(A)(I) (25.1), (25.2), (25.3) and (25.4), official statement that no symptoms of Potato stolbur mycoplasma have been observed on the plants at the place of production since the beginning of the last complete cycle of vegetation
Section II	Plants, plant products and other objects originating in the Community	
	Plants, plant products and other objects	Special requirements
18.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting	Official statement that: (a) the Union provisions to combat <i>Synchytrium endobioticum</i> (Schilbersky) Percival have been complied with; and (b) either the tubers originate in an area known to be free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. or the Union provisions to combat <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. have been complied with; and (d) (aa) either, the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known not to occur; or (bb) in areas where <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known to occur, the tubers originate from a place of production found free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al., or considered to be free thereof, as a consequence of the implementation of an appropriate procedure aiming at eradicating <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al.; and (e) either, the tubers originate in areas in which <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known not to occur, or in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known to occur: — either, the tubers originate from a place of production which has been found free from <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen based on an annual survey of host crops by visual inspection of host plants at appropriate times and by visual inspection both externally and by cutting of tubers after harvest from potato crops grown at the place of production, or

		<p>— the tubers after harvest have been randomly sampled and, either checked for the presence of symptoms after an appropriate method to induce symptoms or laboratory tested, as well as inspected visually both externally and by cutting the tubers, at appropriate times and in all cases at the time of closing of the packages or containers before marketing according to the provisions on closing in Council Directive 66/403/EEC, and no symptoms of <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen have been found</p>
18.1.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting, other than those to be planted in accordance with Article 4.4(b) of Council Directive 2007/33/EC	Without prejudice to the requirements applicable to the tubers of <i>Solanum tuberosum</i> L., intended for planting in Annex IV, Part A, Section II (18.1), official statement that the Union provisions to combat <i>Globodera pallida</i> (Stone) Behrens and <i>Globodera rostochiensis</i> (Wollenweber) Behrens are complied with
18.2	Tubers of <i>Solanum tuberosum</i> L., intended for planting, other than tubers of those varieties officially accepted in one or more Member States pursuant to Council Directive 70/457/EEC of 29 September 1970 on the common catalogue of varieties of agricultural plant species (1)	<p>Without prejudice to the special requirements applicable to the tubers listed in Annex IV(A)(II) (18.1), official statement that the tubers:</p> <ul style="list-style-type: none"> — belong to advanced selections such a statement being indicated in an appropriate way on the document accompanying the relevant tubers, — have been produced within the Community, <p>and</p> <ul style="list-style-type: none"> — have been derived in direct line from material which has been maintained under appropriate conditions and has been subjected within the Community to official quarantine testing in accordance with appropriate methods and has been found, in these tests, free from harmful organisms
18.3	Plants of stolon or tuber-forming species of <i>Solanum</i> L., or their hybrids, intended for planting, other than those tubers of <i>Solanum tuberosum</i> L. specified in Annex IV(A)(II) (18.1) or (18.2), and other than culture maintenance material being stored in gene banks or genetic stock collections	<p>(a) The plants shall have been held under quarantine conditions and shall have been found free of any harmful organisms in quarantine testing;</p> <p>(b) the quarantine testing referred to in (a) shall:</p> <ul style="list-style-type: none"> (aa) be supervised by the official plant protection organisation of the Member State concerned and executed by scientifically trained staff of that organisation or of any officially approved body; (bb) be executed at a site provided with appropriate facilities sufficient to contain harmful organisms and maintain the material including indicator plants in such a way as to eliminate any risk of spreading harmful organisms; (cc) be executed on each unit of the material; <ul style="list-style-type: none"> — by visual examination at regular intervals during the full length of at least one vegetative cycle, having regard to the type of material and its stage of development during the testing programme, for symptoms caused by any harmful organisms, — by testing, in accordance with appropriate methods to be submitted to the Committee referred to in Article 18: <ul style="list-style-type: none"> — in the case of all potato material at least for: <ul style="list-style-type: none"> — Andean potato latent virus, — Arracacha virus B. oca strain,

		<ul style="list-style-type: none"> — Potato black ringspot virus, — Potato spindle tuber viroid, — Potato virus T, — Andean potato mottle virus, — common potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc) and Potato leaf roll virus, — <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., — <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al., — in the case of true seed potato of least for the viruses and viroid listed above; <p>(dd) by appropriate testing on any other symptom observed in the visual examination in order to identify the harmful organisms having caused such symptoms;</p> <p>(c) any material, which has not been found free, under the testing specified under (b) from harmful organisms as specified under (b) shall be immediately destroyed or subjected to procedures which eliminate the harmful organism(s);</p> <p>(d) each organisation or research body holding this material shall inform their official Member State plant protection service of the material held</p>
18.3.1.	Seeds of <i>Solanum tuberosum</i> L., other than those specified in point 18.4	<p>Official statement that:</p> <p>The seeds derive from plants complying, as applicable, with the requirements set out in points 18.1., 18.1.1, 18.2 and 18.3;</p> <p>and</p> <p>(a) the seeds originate in areas known to be free from <i>Synchytrium endobioticum</i> (Schilbersky) Percival, <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. and Potato spindle tuber viroid;</p> <p>or</p> <p>(b) the seeds comply with all of the following requirements:</p> <ul style="list-style-type: none"> (i) they have been produced in a site where, since the beginning of the last cycle of vegetation, no symptoms of disease caused by the harmful organisms referred to in point (a) have been observed; (ii) they have been produced at a site where all of the following actions have been taken: <ul style="list-style-type: none"> 3 separation of the site from other solanaceous plants and other host plants of Potato spindle tuber viroid; 4 prevention of contact with staff and items, such as tools, machinery, vehicles, vessels and packaging material, from other sites producing solanaceous plants and other host plants of Potato spindle tuber viroid, or appropriate hygiene measures concerning staff or items from other sites producing solanaceous plants and other host plants of Potato spindle tuber viroid to prevent infection; 5 only water free from all harmful organisms referred to in this point is used

18.4	Plants of stolon, or tuber-forming species of <i>Solanum</i> L., or their hybrids, intended for planting, being stored in gene banks or genetic stock collections	Each organisation or research body holding such material shall inform their official Member State plant protection service of the material held
18.5.	Tubers of <i>Solanum tuberosum</i> L., other than those mentioned in Annex IV(A)(II) (18.1), (18.1.1), (18.2), (18.3) or (18.4)	There shall be evidence by a registration number put on the packaging, or in the case of loose-loaded potatoes transported in bulk, on the vehicle transporting the potatoes, that the potatoes have been grown by an officially registered producer, or originate from officially registered collective storage or dispatching centres located in the area of production, indicating that the tubers are free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. and that (a) the Union provisions to combat <i>Synchytrium endobioticum</i> (Schilbersky) Percival, and (b) where appropriate, the Union provisions to combat <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., and (c) the Union provisions to combat <i>Globodera pallida</i> (Stone) Behrens and <i>Globodera rostochiensis</i> (Wollenweber) Behrens are complied with

Annex IV, Part B			
Special requirements which shall be laid down by all member states for the introduction and movement of plants, plant products and other objects into and within certain protected zones			
	Plants, plant products and other objects	Special requirements	Protected zone(s)
20.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting	Without prejudice to the provisions applicable to the plants listed in Annex III(A) (10), (11), Annex IV(A)(I) (25.1), (25.2), (25.3), (25.4), (25.5), (25.6), Annex IV(A)(II) (18.1), (18.2), (18.3), (18.4), (18.6), official statement that the tubers: (a) were grown in an area where Beet necrotic yellow vein virus (BNYVV) is known not to occur; or (b) were grown on land, or in growing media consisting of soil that is known to be free from BNYVV, or officially tested by appropriate methods and found free from BNYVV; or (c) have been washed free from soil	F (Britanny), FI, IRL, P (Azores), UK (Northern Ireland)
20.2.	Tubers of <i>Solanum tuberosum</i> L., other than those mentioned in Annex IV(B) (20.1)	(a) The consignment or lot shall not contain more than 1% by weight of soil, or (b) the tubers are intended for processing at premises with officially approved waste disposal facilities which ensures that there is no risk of spreading BNYVV	F (Britanny), FI, IRL, P (Azores), UK (Northern Ireland)

Annex V	Plants, plant products and other objects which must be subject to a plant health inspection (at the place of production if originating in the Community, before being moved within the Community—in the country of origin or the consignor country, if originating outside the Community) before being permitted to enter the Community
Part A	Plants, plant products and other objects originating in the Community
Section I	Plants, plant products and other objects which are potential carriers of harmful organisms of relevance for the entire Community and which must be accompanied by a plant passport
1.3.	Plants of stolon- or tuber-forming species of <i>Solanum</i> L. or their hybrids, intended for planting
	Plants, plant products and other objects which are potential carriers of harmful organisms of relevance for certain protected zones and which must be accompanied by a plant passport valid for the appropriate zone when introduced into or moved within that zone
Section II	Without prejudice to the plants, plant products and other objects listed in Part I
1.5.	Tubers of <i>Solanum tuberosum</i> L., intended for planting
Part B	Plants, plant products and other objects originating in territories, other than those territories referred to in Part A
Section I	Plants, plant products and other objects which are potential carriers of harmful organisms of relevance for the entire Community
4.	Tubers of <i>Solanum tuberosum</i> L.

3.3.3. Legislation addressing the organisms that vector PLRV (Directive/2000/29/EC)

Most PLRV isolates (see Section 3.1.2) are reported to be transmitted by aphid vectors which are not subject to specific regulation.

3.4. Entry, establishment and spread in the EU

3.4.1. Host range

Table 5 provides information on reports of natural hosts of PLRV and PLRV-TYTV, including the associated regulation.

Due to the absence of specific surveys for PLRV-TYTV, any PLRV-TYTV isolate in a host other than tomato would likely have been identified as a PLRV isolate. Therefore, it cannot be excluded that PLRV-TYTV's natural host range is similar to that of PLRV.

Table 5: Natural hosts of PLRV. Data regarding natural hosts was retrieved from the CABI cpc and literature up to September 18, 2019

Virus	Hosts	Rationale and/or uncertainty	Regulation
PLRV	<i>Capsicum annuum</i> (Knierim et al., 2013), <i>Corchorus olitorius</i> (Biswas et al., 2014), <i>Datura stramonium</i> (de Souza-Dias et al., 1999), <i>Nicandra physalodes</i> (Thomas, 1993), <i>Physalis sp.</i> (Thomas, 1993), <i>Solanum betaceum</i> (Álvarez et al., 2011; Jaramillo et al., 2011), <i>S. lycopersicum</i> (Thomas, 1993), <i>S. tuberosum</i> (Guyader and Ducray, 2002), <i>S. phureja</i> (Franco-Lara and Barker, 1999), <i>S. sarrachoides</i> (Thomas and Hassan, 2002), <i>Ullucus tuberosus</i> (Lizarraga, 1996), <i>Zinnia elegans</i> (Thomas and Hassan, 2002)	Experimental host reported in several botanical families (Thomas and Hassan, 2002) <i>Ullucus tuberosus</i> as natural host is associated with uncertainties; reported once based on only one identification technique (ELISA) (Lizarraga, 1996); Fox et al. (2019) reported a new polerovirus in <i>U. tuberosus</i> which showed cross-reactions with PLRV antibodies in ELISA	<i>Capsicum sp.</i> : IVAI 16.6, 25.7, 36.3, IVAII 18.6.1, 18.7; VBI 1.3. <i>Solanum sp.</i> : IIIA 10,11,12; IVAI 25.1, 25.2, 25.3, 25.4, 25.4.1, 25.4.2, 25.5, 25.6, 25.7, 25.7.1, 25.7.2, 28.1, 36.2, 45.3, 48; IVAII 18.1, 18.1.1, 18.2, 18.3, 18.3.1, 18.4, 18.5, 18.6, 18.6.1, 18.7, 26.1, 27; IVBI 20.1, 20.2; VAI 1.3, 2.4; VAII 1.5; VBI 1, 3, 4 <i>Solanaceae</i> : IIIA 13
PLRV-TYTV	<i>Solanum lycopersicum</i> (Thomas, 1984)	Limited information. Several experimental hosts, including <i>S. tuberosum</i> , are reported (Hassan and Thomas, 1984; Hassan et al., 1985)	

ELISA: enzyme-linked immunosorbent assay; PLRV: potato leafroll virus; TYTV: tomato yellow top disease.

3.4.2. Entry

Is the pest able to enter into the EU territory? If yes, identify and list the pathways.

Yes. Non-EU isolates of PLRV may enter the EU territory via plants for planting, i.e. seed potatoes (tubers) and/or microplants. Additional pathways include ware potatoes (i.e. tubers intended for consumption or processing), plants for planting and fruits of other hosts, and viruliferous aphid vectors.

The following pathways can be considered for entry of non-EU isolates of PLRV into the EU: potato plants for planting (seed potatoes, microplants), ware potatoes (i.e. tubers intended for consumption or processing), plants for planting and fruits of other natural hosts, and viruliferous aphid vectors (see Table 6 for the major pathways).

PLRV is transmitted by vegetative propagation and therefore seed potatoes and more generally, potato plants for planting, are considered the most important pathway for entry. The potential pathways for entry of non-EU isolates via seed potatoes of *S. tuberosum* and plants for planting of

other tuber-forming *Solanum* species and their hybrids is addressed by the current EU legislation (Table 5; (EU) 2000/29 Annex IIIA, 10 and 11), which sets that import is not allowed from third countries except Switzerland. However, import of seed potatoes from Canada into Greece, Spain, Italy, Cyprus, Malta and Portugal is allowed by a derogation (2011/778/EU, 2014/368/EU, document C (2014) 3878). PLRV is reported in Canada (Keese et al., 1990) and Switzerland (Steinger et al., 2014) and, by definition, the PLRV isolates present in these countries are considered to be non-EU isolates. Therefore, the pathway of plants for planting of potato is considered partially regulated for PLRV. PLRV-TYTV is not known to naturally infect potato but since potato has been shown to be an experimental host (Hassan and Thomas, 1984; Hassan et al., 1985), natural infections in potato cannot be ruled out. Would PLRV-TYTV isolates be able to naturally infect potato, the potential pathway of plants for planting of potato should be considered closed by legislation given their geographical distribution.

Entry of ware potatoes is addressed by the current EU legislation (Table 4, Annex IIIA, 12). Import of ware potatoes is prohibited from third countries other than Algeria, Egypt, Israel, Libya, Morocco, Syria, Switzerland, Tunisia and Turkey, and from European non-EU countries which do not meet a series of requirements addressing several other pathogens (see Table 4). PLRV is or should be considered present in these specified countries given its worldwide distribution. By definition, the PLRV isolates present in these countries are considered non-EU isolates. They can in principle enter the EU via the ware potato pathway as there are no specific measures in place that mitigate the risk of entry. As reported in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), the majority of the imported ware potatoes comes from Egypt and Israel (47 and 47.2% respectively). Note that as long as ware potatoes are used for the intended use (consumption or processing) the ability of the non-EU isolates of PLRV to establish is very low. In addition, there are specific measures in place (Annex IV 25.3) for countries where potato spindle tuber viroid is known to occur (according to EPPO: Egypt, Israel and Turkey) aimed at mitigating the risk of establishment by suppression of the faculty of germination of ware potatoes, other than early potatoes, from these countries. When considering PLRV and PLRV-TYTV separately, only PLRV is known to be present in the countries for which derogations apply. Therefore, the ware potato pathway is considered partially regulated for non-EU isolates of PLRV but closed by legislation for PLRV-TYTV, irrespective whether potato is a natural host for PLRV-TYTV.

PLRV has a relatively wide natural and experimental host range (see Section 3.4.1), of which some are not regulated (*Corchorus olitorius* and *Ullucus tuberosus*). For PLRV-TYTV only *S. lycopersicum* is reported as a natural host, which is regulated, but it cannot be excluded that the natural host range of PLRV-TYTV is comparable to that of PLRV (see previous section). As a consequence, the pathway of plants for planting of other hosts is considered partially regulated for PLRV. For PLRV-TYTV, this pathway is closed for solanaceous plants for planting and possibly open for the other potential non-solanaceous hosts.

Viruliferous aphid vectors are a possible pathway of entry for non-EU isolates of PLRV (Section 3.1.3). Since the relevant aphid species, such as *M. persicae* (Sulzer), are not subject to specific regulation, this pathway is open for non-EU isolates of PLRV and PLRV-TYTV. PLRV is transmitted by aphids in a persistent way, which implies that viruliferous aphids are able to transmit the virus for the rest of their life; it is worth noticing that aphids may engage in passive long distance migrations using air currents.

Import of fruits of hosts other than potato (*Capsicum annuum* and *Solanum lycopersicum*) is possible from countries, where both PLRV and PLRV-TYTV isolates have been reported and could therefore provide an additional pathway for entry of non-EU isolates of PLRV and PRLV-TYTV. However, this potential pathway is considered as minor because PRLV and PRLV-TYTV are not known to be seed transmitted and because aphid transmission from an imported fruit to a cultivated plant is considered unlikely given the relatively unlikely set of events involved (aphids feeding on imported fruits then moving to susceptible plants). This pathway is not included in Table 6.

Table 6: Identified major pathways for potential entry of non-EU isolates of PLRV, PLRV-TYTV and the extent to which these pathways are addressed by current legislation

Group of isolates	Potato plants for planting ⁽¹⁾	Ware potatoes ⁽¹⁾	Plants for planting of other hosts ^{(1),(2)}	Viruliferous vectors ⁽¹⁾	Uncertainties
PLRV	Pathway partially regulated: plants for planting of potato can be imported from Canada and Switzerland	Pathway partially regulated: ware potatoes can be imported from a range of non-EU European countries where PLRV is reported	Pathway partially regulated: regulated and unregulated hosts	Pathway open: among other aphid species, the main vector <i>Myzus persicae</i> is not regulated and widespread	Geographic distribution Existence of other natural hosts
PLRV-TYTV	Not a pathway: potato is not reported as a natural host. Should potato be a host, the pathway would be closed by legislation given the geographical distribution of these isolates	Not a pathway: potato is not reported as a natural host. Should potato be a host, the pathway would be closed by legislation given the geographical distribution of these isolates	Pathway closed for solanaceous plants for planting: import is banned from countries where PLRV-TYTV is reported Pathway possibly open for non-solanaceous hosts: the existence of additional natural hosts not subject to import restrictions cannot be excluded	Pathway open: among other aphid species, the main vector <i>Myzus persicae</i> is not regulated and widespread	Geographic distribution Potato as natural host Existence of other natural hosts

(1): '**Pathway open**': no regulation or ban that prevents this pathway, '**Pathway closed**' (as opposed to 'pathway open'): ban that prevents entry. '**Pathway possibly open**': no direct evidence of the existence of the pathway (not closed by current legislation), but existence cannot be excluded based on comparisons with the biology of closely related viruses (in the same genus or family). '**Pathway regulated**': regulations exist that limit the probability of entry along the pathway, but there is not a complete ban on imports. '**Pathway partially regulated**': pathway consists of several sub-pathways, some are open, while others are closed (e.g. regulation for some hosts, but not for others; a ban exists for some non-EU MSs but not for all). '**Not a pathway**': no evidence supporting the existence of the pathway.

(2): Plants for planting, including seeds and pollen, of other hosts which are listed in Table 5.

The Europhyt database does not report any interception of PLRV by EU MSs between 1995 and August 8, 2019.

3.4.3. Establishment

Is the pest able to become established in the EU territory?

Yes. Non-EU isolates of PLRV, including PLRV-TYTV, are likely to become established in the EU territory, as EU isolates and the main hosts are already present in the EU.

3.4.3.1. EU distribution of main host plants

Potato is widely grown in the EU, as reported in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020).

3.4.3.2. Climatic conditions affecting establishment

Except for those conditions affecting survival of the host plants, no eco-climatic constraints exist for the PLRV isolates categorised here. Therefore, it is expected that these isolates are able to establish wherever their hosts may live. Potato is widely cultivated in the EU and therefore the Panel considers that climatic conditions will not impair the ability of the viruses addressed here to establish in the EU. However, it must be taken into consideration that virus impact, accumulation and distribution within natural hosts are dependent on environmental conditions. The same applies to expression of symptoms, vector populations and virus transmission being affected by climatic conditions.

3.4.4. Spread

Is the pest able to spread within the EU territory following establishment?

Yes. Non-EU isolates of PLRV, including PLRV-TYTV, can spread via plants for planting, and in addition, most of them can be spread by aphid vectors.

Most non-EU isolates of PLRV, including PLRV-TYTV, can be transmitted by aphids (see Section 3.1.3), including *Myzus persicae* (Sulzer), which is widespread in and outside the EU (see Figure 1).

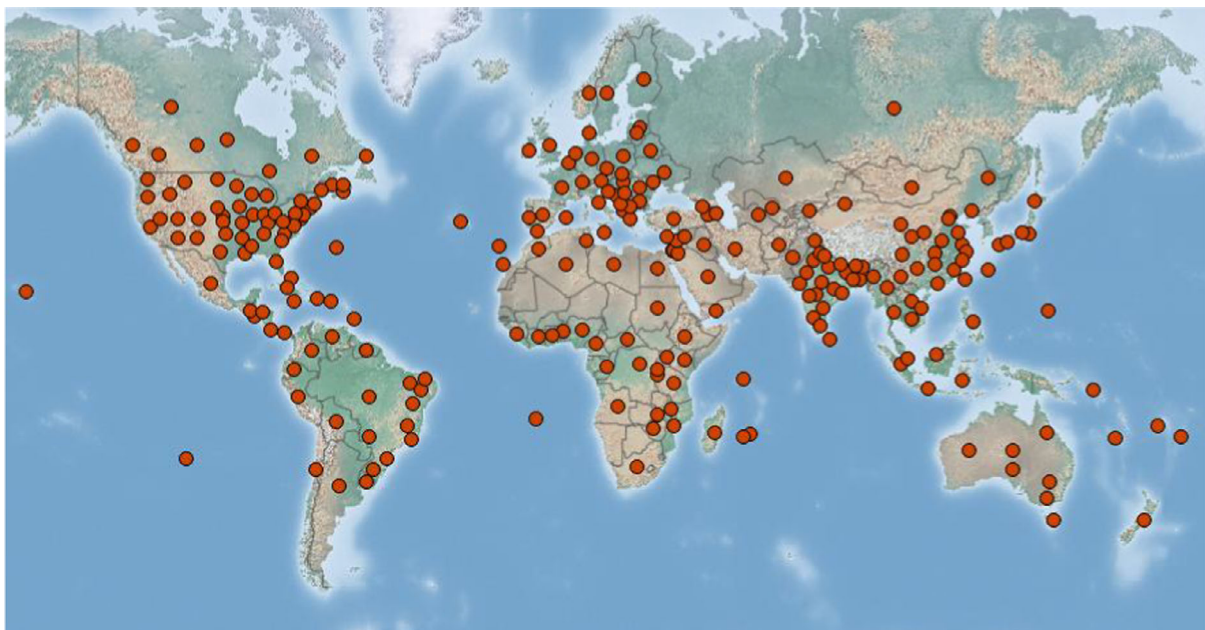


Figure 1: Global distribution map of *Myzus persicae* (Sulzer). Extracted from CABI cpc on 8 August 2019

3.5. Impacts

Would the pests' introduction have an economic or environmental impact on the EU territory?

Yes. Isolates of PLRV-TYTV can be expected to have an additional impact on the EU territory, although the magnitude of the impact is uncertain.

No. Non-EU isolates of PLRV are not known to differ from PLRV isolates already present in the EU and no additional impact is therefore expected on the EU territory.

As mentioned in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), symptoms caused by viruses are influenced by different factors, such as the isolate of the virus, the host and variety, and environmental conditions. A causal relation between a virus and reported symptoms is not always clear, for example in the case of mixed infections. Mixed infections are especially common in vegetative-propagated crops such as potato and the presence of additional viruses might increase or attenuate the observed symptoms. Therefore, reports on the symptomatology of individual viruses might not be conclusive, leading to uncertainties on the causal relation between a virus and the symptoms reported.

PLRV is reported to cause yield reduction of potato by 30–50% in secondary infections (Loebenstein and Gaba, 2012). However, symptoms and impact are dependent on potato cultivar and environmental conditions, for example the percentage of infected tubers is reported to vary at different altitudes (Bertschinger et al., 2017). PLRV isolates occur worldwide and there is no evidence for differences in molecular or biological properties between EU and non-EU PLRV isolates. Therefore, non-EU PLRV isolates, if introduced, are not expected to have an additional impact over the present situation.

PLRV-TYTV causes the tomato yellow top disease in *S. lycopersicum*, including symptoms such as growth reduction, leaf chlorosis and necrosis of flower buds (Thomas, 1984). PLRV-TYTV is reported to cause no, or only weak, symptoms on potato (Hassan and Thomas, 1984; Hassan et al., 1985). Tomato yellow top disease is not known to occur in the EU, with uncertainty. Therefore, PLRV-TYTV is expected to have an additional impact in tomato over the current situation, should it be introduced in the EU. However, the magnitude of the impact is unclear.

3.6. Availability and limits of mitigation measures

Are there measures available to prevent the entry into, establishment within or spread of the pest within the EU such that the risk becomes mitigated?

Yes. See Section 3.3 for measures already implemented in the current legislation. Additional measures could be implemented to further regulate the identified pathways or to limit entry, establishment or spread of non-EU isolates of PLRV.

3.6.1. Identification of additional measures

Phytosanitary measures are currently applied to potato and other hosts (see Sections 3.3 and 3.4.1). Potential additional measures to mitigate the risk of entry of the isolates categorised in this opinion may include:

- Repel import derogations for potato plants for planting;
- Extension of phytosanitary measures to specifically include hosts other than potato;
- Banning import of non-potato hosts plants for planting from countries where PLRV is present;
- Extension of certification schemes or testing requirements to non-solanaceous natural hosts;
- Extension of plant passport requirements to specifically include hosts other than stolon- and tuber-forming *Solanum* species.

In addition, non-EU isolates of PLRV may enter into the EU through viruliferous aphids. Measures against aphids may include chemical treatment of consignments identified as potential entry pathways.

3.6.1.1. Additional control measures

Table 7 reports on the potential additional control measures to reduce the likelihood of entry, establishment and/or spread of the categorised non-EU isolates of PLRV. The additional control

measures are selected from a longer list reported in EFSA PLH Panel (2018). Control measures are measures that have a direct effect on pest abundance.

Table 7: Selected additional control measures to consider to reduce the likelihood of pest entry, establishment and/or spread of non-EU isolates of PLRV

Information sheet (with hyperlink to information sheet if available)	Control measure summary	Risk component	Rationale
Growing plants in isolation	Description of possible exclusion conditions that could be implemented to isolate the crop from pests and if applicable relevant vectors. E.g. a dedicated structure such as glass or plastic greenhouses	Spread	Growing plants in insect proof greenhouses may prevent infestation by viruliferous aphid vectors. This measure would not be applicable for potato, with the exception of early stages of seed potato production Production of seed potatoes in areas with low aphid pressure (e.g. high altitude) would minimise the risk of infestation
Chemical treatments on consignments or during processing	Use of chemical compounds that may be applied to plants or to plant products after harvest, during process or packaging operations and storage The treatments addressed in this information sheet are: a) fumigation; b) spraying/dipping pesticides; c) surface disinfectants; d) process additives; e) protective compounds	Entry	a), b) and c) could remove viruliferous aphid vectors PLRV is transmitted by aphids in a persistent and circulative manner, which implies that viruliferous aphids will keep the ability to transmit the virus throughout their life. Therefore, the additional effect on preventing entry might be substantial
Roguing and pruning	Roguing is defined as the removal of infested plants and/or uninfested host plants in a delimited area, whereas pruning is defined as the removal of infested plant parts only, without affecting the viability of the plant	Establishment and spread	Roguing of infested plants is efficient, in particular to prevent spread of PLRV via aphid vectors. Pruning is not effective to remove a virus from infected plants
Crop rotation, associations and density, weed/volunteer control	Crop rotation, associations and density, weed/volunteer control are used to prevent problems related to pests and are usually applied in various combinations to make the habitat less favourable for pests The measures deal with (1) allocation of crops to field (over time and space) (multi-crop, diversity cropping) and (2) to control weeds and volunteers as hosts of pests/vectors	Spread and impact	Viruses are maintained by vegetative propagation and, therefore, control of volunteers is important. Control of weed hosts may be of relevance
Timing of planting and harvesting	The objective is to produce phenological asynchrony in pest/crop interactions by acting on or benefiting from specific cropping factors such as: cultivars, climatic conditions, timing of the sowing or planting, and level of maturity/age of the plant seasonal timing of planting and harvesting	Spread and impact	Relevant to prevent transmission by aphid vectors

Information sheet (with hyperlink to information sheet if available)	Control measure summary	Risk component	Rationale
Chemical treatments on crops including reproductive material	Chemical treatments on crops may prevent infestations by vectors and seed transmission	Spread and impact	Desiccation/removal of the foliage reduces the risk of transmission via aphid vectors and may prevent transport to the tubers of infected plants
Post-entry quarantine and other restrictions of movement in the importing country	This information sheet covers post-entry quarantine of relevant commodities; temporal, spatial and end-use restrictions in the importing country for import of relevant commodities; Prohibition of import of relevant commodities into the domestic country Relevant commodities are plants, plant parts and other materials that may carry pests, either as infection, infestation or contamination	Entry and spread	Identifying virus-infected plants and banning their movement limit the risks of entry and spread in the EU

3.6.1.2. Additional supporting measures

Table 8 reports on the possible additional supporting measures which are selected from the list reported in EFSA PLH Panel (2018). Supporting measures are organisational measures or procedures supporting the choice of appropriate risk reduction options that do not directly affect pest abundance.

Table 8: Selected supporting measures in relation to currently unregulated hosts and pathways. Supporting measures are organisational measures or procedures supporting the choice of appropriate risk reduction options that do not directly affect pest abundance

Information sheet title (with hyperlink to information sheet if available)	Supporting measure summary	Risk component	Comments
Inspection and trapping	Inspection is defined as the official visual examination of plants, plant products or other regulated articles to determine if pests are present or to determine compliance with phytosanitary regulations (ISPM 5) The effectiveness of sampling and subsequent inspection to detect pests may be enhanced by including trapping and luring techniques	Entry and spread	Visual inspection may detect potentially infected material Only applicable when visible symptoms on leaves and/or propagating tissues occur, which is dependent on the isolate, host/cultivar, and environmental conditions
Laboratory testing	Examination, other than visual, to determine if pests are present using official diagnostic protocols. Diagnostic protocols describe the minimum requirements for reliable diagnosis of regulated pests	Entry and spread	Laboratory testing may detect/ identify non-EU isolates of PLRV on sampled material

Information sheet title (with hyperlink to information sheet if available)	Supporting measure summary	Risk component	Comments
Certified and approved premises	Mandatory/voluntary certification/ approval of premises is a process including a set of procedures and of actions implemented by producers, conditioners and traders contributing to ensure the phytosanitary compliance of consignments. It can be a part of a larger system maintained by a National Plant Protection Organization in order to guarantee the fulfilment of plant health requirements of plants and plant products intended for trade. Key property of certified or approved premises is the traceability of activities and tasks (and their components) inherent the pursued phytosanitary objective. Traceability aims to provide access to all trustful pieces of information that may help to prove the compliance of consignments with phytosanitary requirements of importing countries	Entry and spread	Certified and approved premises may guarantee the absence of the harmful viruses imported for research and/or breeding purposes
Delimitation of Buffer zones	ISPM 5 defines a buffer zone as 'an area surrounding or adjacent to an area officially delimited for phytosanitary purposes in order to minimize the probability of spread of the target pest into or out of the delimited area, and subject to phytosanitary or other control measures, if appropriate' (ISPM 5). The objectives for delimiting a buffer zone can be to prevent spread from the outbreak area and to maintain a pest free production place, site or area	Spread	Buffer zones may contribute to reduce the spread of non-EU isolates of PLRV after entry in the EU
Sampling	According to ISPM 31, it is usually not feasible to inspect entire consignments, so phytosanitary inspection is performed mainly on samples obtained from a consignment. It is noted that the sampling concepts presented in this standard may also apply to other phytosanitary procedures, notably selection of units for testing	Spread	
Phytosanitary certificate and plant passport	An official paper document or its official electronic equivalent, consistent with the model certificates of the IPPC, attesting that a consignment meets phytosanitary import requirements (ISPM 5) a) export certificate (import) b) plant passport (EU internal trade)	Entry and spread	

Information sheet title (with hyperlink to information sheet if available)	Supporting measure summary	Risk component	Comments
Certification of reproductive material (voluntary/official)	Certification of reproductive material when not already implemented would contribute to reduce the risk associated with spread	Spread	
Surveillance	Official surveillance may contribute to early detection of non-EU isolates of PLRV, favouring immediate adoption of control measures if they come to establish	Spread	

3.6.1.3. Biological or technical factors limiting the effectiveness of measures to prevent the entry, establishment and spread of the pest

- Symptomless infections for some of the non-EU isolates of PLRV in some hosts;
- Uneven virus distribution or low concentrations limiting the reliability of the detection;
- Absence of a validated diagnostic protocol allowing the specific identification of PLRV-TYTV.

3.7. Uncertainty

The Panel identified the following knowledge gaps and uncertainties:

Identity and biology

- Limited data on biological diversity within the PLRV species;
- Uncertainty on the existence of other non-EU isolates of PLRV that have not yet been identified and might have additional impact on the EU territory;
- Lack of information on the molecular diversity of PLRV-TYTV (only one sequence available).

Pest distribution

- Uncertainty on the geographical distribution and prevalence of the categorised groups of isolates of PLRV because of the absence of systematic surveys and, additionally, because PLRV-TYTV isolates might have been ('mis-') identified as PLRV when detected in other hosts than tomato.

Regulatory status

- The concept of 'non-EU isolates' leaves some room for interpretation, which may create confusion or difficulties when enforcing the legislation (see Section 1.2).

Entry, establishment and spread in the EU (host range, entry, establishment, spread)

- Uncertainty on the host range of the categorised groups of isolates of PLRV, particularly in the case of PLRV-TYTV.

Impact

- Uncertainty on the magnitude of the impact of non-EU isolates and whether this impact would exceed that of the isolates already present in the EU.

4. Conclusions

The information currently available on geographical distribution, biology, epidemiology, potential additional impact over the present situation, and potential entry pathways of non-EU isolates of PLRV has been evaluated with regard to the criteria to qualify as a potential Union quarantine pest. The conclusions of the Panel are summarised in Table 9.

Non-EU isolates of PLRV other than PLRV-TYTV do not meet one of the criteria evaluated by EFSA to be regarded as a potential Union quarantine pest, since they are not expected to have an additional impact in the EU.

Isolates of PLRV-TYTV meet all the criteria evaluated by EFSA to qualify as a potential Union quarantine pest.

The Panel wishes to stress that these conclusions are associated with uncertainties because of limited information on distribution, biology and impact of PLRV isolates at strain level. In particular, the magnitude of the potential additional impact over the present situation is generally unknown. Furthermore, other potentially harmful non-EU isolates of PLRV might exist that have not been discovered yet.

Table 9: The Panel's conclusions on the pest categorisation criteria defined in Regulation (EU) 2016/2031 on protective measures against pests of plants (the number of the relevant sections of the pest categorisation is shown in brackets in the first column) for non-EU isolates of PLRV

Criterion of pest categorisation	Panel's conclusions against criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Key uncertainties
Identity of the pest (Section 3.1)	The identity of PLRV is well established Methods are available for detection and identification of PLRV at species level. Apart from a bioassay, there is currently no identification method available for PLRV-TYTV	Uncharacterised PLRV isolates may exist
Absence/presence of the pest in the EU territory (Section 3.2)	PLRV is present in the EU PLRV-TYTV is not reported in the EU	Unreported presence of PLRV-TYTV isolates in the EU
Regulatory status (Section 3.3)	Non-EU isolates of PLRV are currently regulated in Annex IAI	Interpretation of the concept of 'non-EU isolate'
Pest potential for entry, establishment and spread in the EU territory (Section 3.4)	<p>Non-EU isolates of PLRV are able to enter into the EU</p> <p>The pathway of plants for planting of potato is partially regulated for non-EU isolates of PLRV (plants for planting of potato can be imported from countries in which these isolates are present). Potato plants for planting are not a pathway for PLRV-TYTV, since potato has not been reported to be a natural host for this group of isolates. Should potato be a natural host for PLRV-TYTV, the pathway would be closed by legislation</p> <p>For ware potatoes, the pathway of entry is partially regulated for non-EU isolates of PLRV (ware potatoes can be imported from countries in which these isolates are present). Import of ware potatoes is not a pathway of entry for PLRV-TYTV, since potato has not been reported to be a natural host for this group of isolates. Should potato be a natural host for PLRV-TYTV, the pathway would be closed by legislation</p> <p>The pathway of plants for planting of other hosts is partially regulated for non-EU isolates of PLRV; For isolates of PLRV-TYTV, it is closed for Solanaceous plants for planting, and possibly open for other unregulated non-Solanaceous hosts plants, the existence of which cannot be excluded</p>	<ul style="list-style-type: none"> – Geographical distribution – Existence of other natural hosts, in particular for PLRV-TYTV – Existence and relevance of trade of plants for planting of non-Solanum hosts

Criterion of pest categorisation	Panel's conclusions against criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Key uncertainties
	<p>The pathways of viruliferous aphids and of fruits of host species are open for non-EU isolates of PLRV and of PLRV-TYTV</p> <p>If non-EU isolates of PLRV were to enter the EU territory, they could become established and spread</p>	
Potential for consequences in the EU territory (Section 3.5)	<p>Isolates of PLRV-TYTV can be expected to have an additional impact on the EU territory</p> <p>Non-EU isolates of PLRV are not known to differ from PLRV isolates already present in the EU and no additional impact is therefore expected on the EU territory</p>	Uncertainty on the magnitude of impact of non-EU isolates of PLRV
Available measures (Section 3.6)	Phytosanitary measures are available to reduce the likelihood of entry and spread of non-EU isolates of PLRV, including PLRV-TYTV, in the EU	No uncertainty
Conclusion on pest categorisation (Section 4)	<p>Non-EU isolates of PLRV other than PLRV-TYTV do not meet one of the criteria evaluated by EFSA to be regarded as a potential Union quarantine pest, since they are not expected to have an additional impact in the EU</p> <p>Isolates of PLRV-TYTV meet all the criteria evaluated by EFSA to qualify as a potential Union quarantine pest: they are expected to have an additional impact over the current situation in the EU</p>	
Aspects of assessment to focus on/scenarios to address in future if appropriate	<p>The main knowledge gaps or uncertainties identified concern:</p> <ul style="list-style-type: none"> Limited data on biological diversity within the PLRV species Lack of information on the molecular diversity of PLRV-TYTV (only one sequence available) Uncertainty on the geographical distribution and prevalence of the categorised groups of isolates of PLRV Uncertainty on the host range of the categorised groups of isolates of PLRV, particularly of PLRV-TYTV. Uncertainty on the magnitude of the impact of non-EU isolates of PLRV and whether this impact would exceed that of the isolates already present in the EU <p>Given the limited information available on possible biological differences between PLRV-TYTV and other PLRV isolates, the development of a full PRA is unlikely to allow to resolve the uncertainties attached to the present categorisation until more data become available</p>	

References

- Agindotan BO, Shiel PJ and Berger PH, 2007. Simultaneous detection of potato viruses, PLRV, PVA, PVX and PVY from dormant potato tubers by TaqMan real-time RT-PCR. *Journal of Virological Methods*, 142, 1–9.
- Álvarez JA, Cotes JM and Marin M, 2011. Detection of viruses associated to planting material of tamarillo in Colombia. *Biotechnología en el Sector Agropecuario y Agroindustrial*, 9, 43–50.
- Andino R and Domingo E, 2015. Viral quasispecies. *Virology*, 479–480, 46–51.
- Bertschinger L, Bühler L, Dupuis B, Duffy B, Gessler C, Forbes GA, Keller ER, Scheidegger UC and Struik PC, 2017. Incomplete infection of secondarily infected potato plants – an environment dependent underestimated mechanism in plant virology. *Frontiers in Plant Science*, 8, 74.
- Biswas C, Dey P, Mitra S, Bera A, Satpathy S and Karmakar PG, 2014. First Report of Potato leaf roll virus (PLRV) Naturally Occurring on Jute (*Corchorus olitorius*) in India. *Plant Disease*, 98, 1592.
- CABI cpc, 2019. Datasheet of potato leafroll virus (PLRV). Available online: <https://www-cabi-org/cpc/datasheet/42783> [Accessed 17 September 2019].

- Chomić A, Pearson MN, Clover GRG, Farreyrol K, Saul D, Hampton JG and Armstrong KF, 2010. A generic RT-PCR assay for the detection of Luteoviridae. *Plant Pathology*, 59, 429–442.
- Domier LL, 2011. Luteoviridae. In: King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (eds.). *Virus Taxonomy*, 9th Report of the International Committee on Taxonomy of Viruses, pp. 1045–1053.
- Domingo E, Sheldon J and Perales C, 2012. Viral quasispecies evolution. *Microbiology and Molecular Biology Reviews*, 76, 159–216.
- EFSA PLH Panel (EFSA Panel on Plant Health), 2018. Guidance on quantitative pest risk assessment. *EFSA Journal* 2018;16(8):5350, 86 pp. <https://doi.org/10.2903/j.efsa.2018.5350>.
- EFSA PLH Panel (EFSA Panel on Plant Health), 2020. Pest categorisation of non-EU viruses and viroids of potato. *EFSA Journal* 2020;18(1):5853, 134 pp. <https://doi.org/10.2903/j.efsa.2020.5853>
- Faccioli G, Rosner A and Forni M, 1995. Use of the polymerase chain reaction to clone the potato leafroll virus coat protein gene directly from the total RNA of infected plants. *Potato Research*, 38, 211–218.
- FAO (Food and Agriculture Organization of the United Nations), 1995. ISPM (International standards for phytosanitary measures) No 4. Requirements for the establishment of pest free areas. Available online: <https://www.ippc.int/en/publications/614/>
- FAO (Food and Agriculture Organization of the United Nations), 2004. ISPM (International Standards for Phytosanitary Measures) 21—Pest risk analysis of regulated non-quarantine pests. FAO, Rome, 30 pp. Available online: https://www.ippc.int/sites/default/files/documents/1323945746_ISPM_21_2004_En_2011-11-29_Refor.pdf
- FAO (Food and Agriculture Organization of the United Nations), 2013. ISPM (International Standards for Phytosanitary Measures) 11—Pest risk analysis for quarantine pests. FAO, Rome, 36 pp. Available online: https://www.ippc.int/sites/default/files/documents/20140512/ispm_11_2013_en_2014-04-30_201405121523-494.65%20KB.pdf
- FAO (Food and Agriculture Organization of the United Nations), 2017. ISPM (International standards for phytosanitary measures) No 5. Glossary of phytosanitary terms. Available online: <https://www.ippc.int/en/publications/622/>
- Fox A, Fowkes AR, Skelton A, Harju V, Buxton-Kirk A, Kelly M, Forde SMD, Pufal H, Conyers C, Ward R, Weekes R, Boonham N and Adams IP, 2019. Using high-throughput sequencing in support of a plant health outbreak reveals novel viruses in *Ullucus tuberosus* (Basellaceae). *Plant Pathology*, 68, 576–587.
- Franco-Lara L and Barker H, 1999. Characterisation of resistance to potato leafroll virus accumulation in *Solanum phureja*. *Euphytica*, 108, 137–144.
- Garcia-Arenal F, Fraile A and Malpica JM, 2001. Variability and genetic structure of plant virus populations. *Annual Review of Phytopathology*, 39, 157–186.
- Guyader S and Ducray DG, 2002. Sequence analysis of Potato leafroll virus isolates reveals genetic stability, major evolutionary events and differential selection pressure between overlapping reading frame products. *Journal of General Virology*, 83, 1799–1807.
- Hassan S and Thomas PE, 1984. Etiological distinctions between tomato yellow top virus and potato leafroll and beet western yellows viruses. *Plant Disease*, 68, 684–685.
- Hassan S, Thomas PE and Mink GI, 1985. Tomato Yellow Top Virus: host Range, Symptomatology, Transmission, and Variability. *Phytopathology*, 75, 287–291.
- Haupt S, Stroganova T, Ryabov E, Kim SH, Fraser G, Duncan G, Mayo MA, Barker H and Taliensky M, 2005. Nucleolar localization of potato leafroll virus capsid proteins. *Journal of General Virology*, 86, 2891–2896.
- van den Heuvel JEJM, De Blank CM, Golbach R and Peters D, 1990. A characterization of epitopes on potato leafroll virus coat protein. *Archives of Virology*, 115, 185–197.
- Hühnlein A, Schubert J, Zahn V and Thieme T, 2016. Examination of an isolate of Potato leaf roll virus that does not induce visible symptoms in the greenhouse. *European Journal of Plant Pathology*, 145, 829–845.
- Jaramillo M, Gutierrez PA, Lagos LE, Cotes JM and Marin M, 2011. Detection of a complex of viruses in tamarillo (*Solanum betaceum*) orchards in the Andean region of Colombia. *Tropical Plant Pathology*, 36, 150–159.
- Jeffries CJ, 1998. FAO-IPGRI Technical guidelines for the safe movement of germplasm no 19 potato_IPGRI.
- Keese P, Martin RR, Kawchuk LM, Waterhouse PM and Gerlach WL, 1990. Nucleotide sequences of an Australian and a Canadian isolate of potato leafroll luteovirus and their relationships with two European isolates. *Journal of General Virology*, 71(Pt 3), 719–724.
- Khaled W, Fekih IB, Nahdi S, Souissi R and Boukhris-Bouhachem S, 2018. Transmission efficiency of potato leafroll virus by four potato colonizing Aphid species in Tunisian Potato Fields. *Potato Research*, 61, 89–96.
- Khouadja FD, Rouze-Jouan J, Guyader S, Marrakchi M and Fakhfakh H, 2005. Biological and molecular characterization of Tunisian isolates of Potato leafroll virus. *Journal of Plant Pathology*, 87, 91–99.
- Khouadja FD, Rouzé-Jouan J, Guyader S and Fakhfakh H, 2014. Possible correlations between the characteristics of Potato leafroll virus isolates occurring in different geographical regions in Tunisia. *Phytoparasitica*, 42, 259–267.
- Knierim D, Tsai WS and Kenyon L, 2013. Analysis of sequences from field samples reveals the presence of the recently described pepper vein yellows virus (genus *Polerovirus*) in six additional countries. *Archives of Virology*, 158, 1337–1341.
- Lizarraga C, 1996. First Report of Potato Leafroll Virus in Ulluco (*Ullucus tuberosus* Caldas). *Plant Disease*, 80, 344.
- Loebenstein G and Gaba V, 2012. Viruses of Potato. *Advances Virus Research*, 84, 209–246.

- Mayo M, Ryabov E, Fraser G and Taliansky M, 2000. Mechanical transmission of Potato leafroll virus. *Journal of General Virology*, 81, 2791–2795.
- Mink GI, 1993. Pollen and seed-transmitted viruses and viroids. *Annual Review of Phytopathology*, 31, 375–402.
- Mukherjee K, Verma Y, Chakrabarti SK, Singh MN and Khurana SMP, 2003. Cloning and Sequencing of Coat Protein Gene of an Indian Potato Leaf Roll Virus (PLRV) Isolate and its Similarity with other Members of Luteoviridae. *Virus Genes*, 26, 247–253.
- Palucha A, Sadowy E, Kujawa A, Juszczuk M, Zagorski W and Hulanicka D, 1994. Nucleotide sequence of RNA of a Polish isolate of potato leafroll luteovirus. *Acta Biochimica Polonica*, 41, 405–414.
- Plchova H, Cerovska N, Moravec T and Dedic P, 2009. Short communication: molecular analysis of potato leafroll virus isolates from the Czech Republic. *Virus Genes*, 39, 153–155.
- Roos WG, 2013. An investigation of prevalence and the detection and race identification of South African potato viruses. Thesis presented in fulfillment of the requirements for the degree of Master of Science (Biochemistry) at the University of Stellenbosch.
- Rouze-Jouan J, Terradot L, Pasquer F, Tanguy S and Giblot Ducrey-Bourdin DD, 2001. The passage of Potato leafroll virus through *Myzus persicae* gut membrane regulates transmission efficiency. *Journal of General Virology*, 82, 17–23.
- Sadowy E, Maasen A, Juszczuk M, David C, Zagorski-Ostojka W, Gronenborn B and Hulanicka MD, 2001. The ORF0 product of Potato leafroll virus is indispensable for virus accumulation. *Journal of General Virology*, 82, 1529–1532.
- Sastry KS, 2013. Introduction. In: *Seed-borne plant virus diseases*. pp. 1–53.
- de Souza-Dias JAC, Russo P, Miller L and Slack SA, 1999. Comparison of nucleotide sequences from three potato leafroll virus (PLRV) isolates collected in Brazil. *American Journal of Potato Research*, 76, 17–24.
- Steinger T, Gilliland H and Hebeisen T, 2014. Epidemiological analysis of risk factors for the spread of potato viruses in Switzerland. *Annals of Applied Biology*, 164, 200–207.
- Tamada T, Harrison BD and IM R, 1984. Variation among British isolates of potato leafroll virus. *Annals of Applied Biology*, 104, 107–116.
- Thomas JE, 1984. Characterisation of an Australian isolate of tomato yellow top virus. *Annals of Applied Biology*, 104, 79–86.
- Thomas JE, 1993. Alternative hosts and the epidemiology of potato leafroll virus in Queensland. *Australian Journal of Agricultural Research*, 44, 1905–1916.
- Thomas PE and Hassan S, 2002. First Report of Twenty-two New Hosts of Potato leafroll virus. *Plant Disease*, 86, 561.
- van der Wilk F, Huisman MJ, Cornelissen BJC, Huttinga H and Goldbach R, 1989. Nucleotide sequence and organization of potato leafroll virus genomic RNA. *FEBS Letters*, 245, 51–56.
- van der Wilk F, Houterman P, Molthoff J, Hans F, Dekker B, van den Heuvel J, Huttinga H and Goldbach R, 1997. Expression of the potato leafroll virus ORF0 induces viral-disease-like symptoms in transgenic potato plants. *Molecular Plant-Microbe Interactions*, 10, 153–159.

Abbreviations

CABI cpc	CABI Crop Protection Compendium
DG SANTÉ	Directorate General for Health and Food Safety
EPPO	European and Mediterranean Plant Protection Organization
FAO	Food and Agriculture Organization
ICTV	International Committee on Taxonomy of Viruses
IPPC	International Plant Protection Convention
ISPM	International Standards for Phytosanitary Measures
MS	Member State
ORF	open reading frame
PLH	EFSA Panel on Plant Health
PLRV	potato leafroll virus
PZ	Protected Zone
RT-PCR	real-time polymerase chain reaction
TFEU	Treaty on the Functioning of the European Union
ToR	Terms of Reference
TYTV	tomato yellow top disease

Glossary

Containment (of a pest)	Application of phytosanitary measures in and around an infested area to prevent spread of a pest (FAO, 1995, 2017)
Control (of a pest)	Suppression, containment or eradication of a pest population (FAO, 1995, 2017)

Entry (of a pest)	Movement of a pest into an area where it is not yet present, or present but not widely distributed and being officially controlled (FAO, 2017)
Eradication (of a pest)	Application of phytosanitary measures to eliminate a pest from an area (FAO, 2017)
Establishment (of a pest)	Perpetuation, for the foreseeable future, of a pest within an area after entry (FAO, 2017)
Impact (of a pest)	The impact of the pest on the crop output and quality and on the environment in the occupied spatial units
Introduction (of a pest)	The entry of a pest resulting in its establishment (FAO, 2017)
Isolate	Virus population as present in a plant
Measures	Control (of a pest) is defined in ISPM 5 (FAO 2017) as 'Suppression, containment or eradication of a pest population' (FAO, 1995). Control measures are measures that have a direct effect on pest abundance. Supporting measures are organisational measures or procedures supporting the choice of appropriate Risk Reduction Options that do not directly affect pest abundance.
Pathway	Any means that allows the entry or spread of a pest (FAO, 2017)
Phytosanitary measures	Any legislation, regulation or official procedure having the purpose to prevent the introduction or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests (FAO, 2017)
Protected zones (PZ)	A Protected zone is an area recognised at EU level to be free from a harmful organism, which is established in one or more other parts of the Union
Quarantine pest	A pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled (FAO, 2017)
Regulated non-quarantine pest	A non-quarantine pest whose presence in plants for planting affects the intended use of those plants with an economically unacceptable impact and which is therefore regulated within the territory of the importing contracting party (FAO, 2017)
Risk reduction option (RRO)	A measure acting on pest introduction and/or pest spread and/or the magnitude of the biological impact of the pest should the pest be present. A RRO may become a phytosanitary measure, action or procedure according to the decision of the risk manager
Spread (of a pest)	Expansion of the geographical distribution of a pest within an area (FAO, 2017)
Strain	Group of isolates sharing biological, molecular and/or serological properties