

Recruitment of the γ -Tubulin Ring Complex to *Drosophila* Salt-stripped Centrosome Scaffolds

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Abstract. Extracting isolated *Drosophila* centrosomes with 2 M KI generates salt-resistant scaffolds that lack the centrosomal proteins CP190, CP60, centrosomin, and γ -tubulin. To clarify the role of these proteins in microtubule nucleation by centrosomes and to identify additional centrosome components required for nucleation, we have developed an in vitro complementation assay for centrosome function. Centrosome aster formation is reconstituted when these inactive, salt-stripped centrosome scaffolds are supplemented with a soluble fraction of a *Drosophila* embryo extract. The CP60 and CP190 can be removed from this extract without effect, whereas removing the γ -tubulin destroys the complementing activity. Consistent with

these results, we find no evidence that these three proteins form a complex together. Instead, γ -tubulin is found in two distinct protein complexes of 240,000 and \sim 3,000,000 D. The larger complex, which is analogous to the *Xenopus* γ -tubulin ring complex (γ TuRC) (Zheng, Y., M.L. Wong, B. Alberts, and T. Mitchison. 1995. *Nature*. 378:578–583), is necessary but not sufficient for complementation. An additional factor found in the extract is required. These results provide the first evidence that the γ TuRC is required for microtubule nucleation at the centrosome.

Key words: centrosome • γ -tubulin • microtubule nucleation • microtubule-organizing center • cytoskeleton

IN animal cells, centrosome-nucleated microtubule arrays are essential for a wide variety of cellular processes including cell division and chromosome segregation, directed cell movement and interphase cytoplasmic organization (for reviews see Mazia, 1987; Vorobjev and Nadezhdina, 1987; Kellogg et al., 1994). EM studies have shown that centrosomes consist of a pair of centriolar cylinders surrounded by an electron-dense cloud of pericentriolar material (PCM),¹ and that the PCM originates the many microtubules that are nucleated by the centrosome (Rieder and Borisy, 1982; Vorobjev and Chentsov, 1982; Keryer et al., 1984).

The molecular characterization of the centrosome and its ability to nucleate microtubules is still in its early stages. The centrosome may contain as many as 100 different proteins (for review see Kalt and Schliwa, 1993; Kellogg et al., 1994), but it is not known how many of these are actual com-

ponents of the PCM, with direct or indirect roles in microtubule nucleation. The identification of centrosomal components is further confounded by the fact that the centrosome, as the focus of the cell's microtubule array, is also a hub for intracellular trafficking. This makes it difficult to distinguish actual components of the PCM from molecules recruited by the microtubule array. To simplify this problem, we define the "core" centrosome as the structure that remains when all of its microtubules have been depolymerized.

The discovery of one such core centrosomal protein, γ -tubulin, has led to a breakthrough in our understanding of microtubule nucleation by centrosomes. γ -Tubulin is a highly conserved member of the tubulin family shown to be involved in microtubule nucleation (Oakley et al., 1990; Stearns et al., 1991; Zheng et al., 1991; Joshi et al., 1992; Felix et al., 1994; Stearns and Kirschner, 1994). Recently, a γ -tubulin-containing ring complex (γ TuRC), capable of nucleating microtubules in vitro, was purified from *Xenopus* eggs (Zheng et al., 1995). EM tomography on centrosomes isolated from *Drosophila* revealed the presence of rings containing γ -tubulin in the PCM in both the presence and absence of nucleated microtubules. The γ -tubulin rings are found at the microtubule minus ends in centrosome-nucleated microtubule asters (Moritz et al., 1995b).

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1. *Abbreviations used in this paper:* CNN, centrosomin; PCM, pericentriolar material; TDB, tubulin dilution buffer; γ TuRC, γ -tubulin ring complex.

Ring structures are also visible in the PCM of centrosomes of the surf clam, *Spisula* (Vogel et al., 1997). These results suggest that the γ TuRC is a highly conserved structure responsible for the microtubule-nucleating capacity of the PCM (Moritz et al., 1995b; Zheng et al., 1995).

Although these studies indicate that the γ TuRC is likely to be essential for microtubule nucleation by centrosomes, many important questions remain. These include: What is the structural organization of the PCM and how is it assembled? How is the γ TuRC anchored within the PCM? Is the attachment of the γ TuRC to the centrosome matrix important for its activity? Do other centrosomal proteins contribute to microtubule nucleation?

Other core centrosomal proteins that may have direct or indirect roles in microtubule nucleation include pericentrin, CP190, CP60, and centrosomin (CNN). Pericentrin is a human autoimmune antigen that has also been identified in mouse and *Xenopus*. It is thought to be a structural component of the PCM that may play an essential role in its organization (Doxsey et al., 1994). CP60 and CP190 are two core centrosomal proteins of unknown function identified in *Drosophila*. CP190 is a novel, zinc-finger-containing protein identified by microtubule affinity chromatography (Kellogg et al., 1989; Whitfield et al., 1995). Native CP190 localizes primarily to nuclei during interphase, becoming prominent at centrosomes upon nuclear envelope breakdown at the onset of mitosis (Frasch et al., 1986; Whitfield et al., 1988; Oegema et al., 1997). CP60, a novel protein identified by immunoaffinity chromatography by virtue of its ability to interact with CP190 (Kellogg and Alberts, 1992), also localizes to both nuclei and centrosomes in a cell cycle-dependent manner, but with slightly different timing (Kellogg et al., 1995; Oegema et al., 1997). Previous work from our laboratory suggested that CP190, CP60, and γ -tubulin are components of a soluble protein complex present in embryo extracts (Raff et al., 1993), but further studies show that this is not the case (see below). In *Drosophila*, the centrosome core also contains at least one developmentally important component, called CNN, which is the target of a homeotic gene and is essential for proper centrosome function (Li and Kaufman, 1996).

To examine the role of the γ TuRC in microtubule nucleation at centrosomes and to test the potential contributions to nucleation by other known centrosomal components, we sought to develop an in vitro complementation assay for aster formation using isolated *Drosophila* centrosomes (Moritz et al., 1995a; Moritz and Alberts, 1998). Previous work has shown that the microtubule-nucleating activity of mammalian centrosomes can be destroyed by salt or urea treatments, and that the activity can be restored by injecting the treated centrosomes into *Xenopus* eggs, or by mixing them with egg extract. This suggests that factors in the egg cytoplasm can associate with the damaged centrosomes, restoring their ability to nucleate microtubules (Klotz et al., 1990; Buendia et al., 1992). With this information in mind, we developed an assay in which microtubule nucleation by *Drosophila* centrosomes is reconstituted from two components, inactive salt-stripped centrosome scaffolds and the high speed supernatant of a *Drosophila* embryo extract.

In this paper, we characterize both the salt-stripped scaffolds and the soluble components in the extract that are

necessary for nucleation. In particular, we test for a role in nucleation for CP190, CP60 and the *Drosophila* γ TuRC. Our assay also allows us to begin to address what components, if any, are required for attachment of the γ TuRC to the salt-stripped scaffolds.

Materials and Methods

Buffers

BRB80: 80 mM K-Pipes, pH 6.8, 1 mM MgCl₂, 1 mM Na₃EGTA (prepare as a 5 \times stock, dilute to 1 \times for use). Hepes buffer: 50 mM K-Hepes, pH 7.6, 1 mM MgCl₂, 1 mM Na₃EGTA. Hepes block: Hepes buffer + 100 mM KCl, 10 mg/ml BSA (fraction V; Sigma Chemical Co., St. Louis, MO), and 1 mM β -mercaptoethanol. Embryo extract buffer for making complementing extract: Hepes buffer + 100 mM KCl, 10% glycerol, 1:100 protease inhibitor stock, and 1 mM PMSF. Protease inhibitor stock: 10 mM benzamide-HCl, 0.1 mg/ml phenanthroline, 1 mg/ml aprotinin, 1 mg/ml leupeptin, 1 mg/ml pepstatin A in ethanol. GTP stock: 0.5 M GTP (Sigma Chemical Co.) in 1 \times BRB80. Tubulin dilution buffer (TDB): 1 \times BRB80, 10% glycerol, 1 mM GTP. TDB wash: TDB + 10 mg/ml BSA (fraction V; Sigma Chemical Co.). Extract buffer for characterizations of CP60, CP190, γ -tubulin protein complexes: 50 mM K-Hepes, pH 7.6, 75 mM KCl, 1 mM Na₃EGTA, 1 mM EDTA, 0.05% NP-40, 1:50 protease inhibitor stock, 2 mM PMSF. Gradient buffer: Hepes buffer + 1 mM β -mercaptoethanol, 1:200 protease inhibitor stock. Column buffer: Hepes buffer + 2% wt/vol glycerol, 1 mM β -mercaptoethanol, 1:200 protease inhibitor stock. PBS: 5.4 mM Na₂HPO₄, 1.8 mM KH₂PO₄, 137 mM NaCl, 2.7 mM KCl, adjusted to pH 7.2. PBST: PBS + 0.1% Tween-20. Sample buffer: 63 mM Tris-HCl, pH 6.8, 3% SDS, 5% β -mercaptoethanol, 10% glycerol. TBS: 10 mM Tris-Cl, pH 8, 150 mM NaCl, 0.05% Tween-20.

Centrosome Isolation

Drosophila centrosomes were isolated on sucrose gradients from 0–3.5-h-old embryos and tested for activity as previously described (Moritz et al., 1995a; Moritz and Alberts, 1998).

Tubulin

Tubulin was purified from bovine brain (Mitchison and Kirschner, 1984) and labeled with *N*-hydroxy-succinimidyl-rhodamine (Hyman et al., 1991) as described previously.

Acid-washed, Poly-lysine-coated Coverslips

Acid-washed, 12-mm round glass coverslips were prepared in large batches by incubating the coverslips in a large glass beaker with 1 N HCl at 65°C for 4 h to overnight with occasional swirling. The coverslips were rinsed extensively in ddH₂O, until the pH was neutral, and then incubated in 0.1% wt/vol poly-L-lysine for 20 min. The coverslips were dried by laying them out on a large piece of filter paper, or in a drying oven.

On-Glass Complementation Assay

To destroy the microtubule-nucleating activity of centrosomes, an equal volume of 4 M KI in 1 \times BRB80 was mixed with the centrosomes, and the mixture was incubated on ice for 10 min. 20 μ l of this mixture was then applied to a 12-mm round acid-washed, poly-lysine-coated glass coverslip, which was placed on a piece of Parafilm inside a humidified Petri dish kept in a 30°C water bath. The centrosomes were allowed to bind to the coverslip for 5 min, and then washed briefly by pipetting on and aspirating off three times with 60 μ l Hepes block. (For controls in which the centrosomes were omitted, the coverslips were washed in the same way with Hepes block, and then the sample was applied and treated in the same way as coverslips with centrosomes.) The final wash was allowed to incubate on the coverslip for 5 min. Depending on the experiment, the dish containing the coverslips was either kept at 30°C, or transferred to a 0°C ice bath. The final wash was then replaced with 10–60 μ l of the sample to be tested. The centrosomes were incubated with the sample for 10 min (at 0° or 30°C), and then the sample was washed away briefly three times with 60 μ l TDB wash. A 25- μ l mixture of unlabeled and rhodamine-labeled tubulin (usually in a 7:1 ratio) diluted to 2 mg/ml in TDB was then incubated

on the coverslips for 10 min at 30°C. Any resulting microtubules or asters were fixed by a 3-min incubation with 60 μ l 1% glutaraldehyde in BRB80 (EM grade; Ted Pella, Inc., Redding, CA), followed by a 3-min incubation with -20°C methanol. The coverslips were then inverted and mounted on slides on drops of mounting medium (80% glycerol in PBS + 1 mg/ml *para*-phenylenediamine). The slides were viewed on a Nikon Microphot-FXA, 100 \times objective (1.4 NA), and either photographed using Kodak Ektachrome 400 Elite or Ektachrome P1600 film, or on a Nikon Optiphot-2, 60 \times or 100 \times objective (1.4 NA) using a cooled CCD camera (Princeton Scientific Instruments, Inc., Monmouth Junction, NJ). Micrographs were processed using Photoshop (Adobe Systems, Inc., Mountain View, CA). For quantitation, asters were counted in 50 randomly selected, 100 \times microscope fields.

For immunofluorescence, unlabeled tubulin was used during the microtubule regrowth step, and the samples on coverslips were rehydrated after methanol fixation by washing in TBS. Residual glutaraldehyde from the fixation step was reduced by incubation with 0.1% sodium borohydride in TBS for 7 min. The samples were washed and blocked in TBS + 3% BSA for 5 min, and incubated simultaneously for 1 h with rabbit anti- γ -tubulin and DM1 α (mouse anti- α -tubulin, T2096; Sigma Chemical Co.), each diluted 1:1,000. After washing, the coverslips were incubated for 1 h with a mixture of fluorescein-labeled goat anti-rabbit (1:500) and Texas red-labeled donkey anti-mouse (1:50), washed, and then mounted for viewing under the fluorescence microscope. Images were obtained on a Nikon Optiphot-2 (60 \times objective, 1.4 NA) using a cooled CCD camera (Princeton Scientific Instruments, Inc.). WinView software (Princeton Scientific Instruments, Inc.) was used to quantitate fluorescence intensity (see Fig. 3*h*).

Drosophila Embryo Extracts

Drosophila embryos between 0- and 2-h old (for preparation of complementing extract), or 0- and 4.5-h old (for characterization of protein complexes) were harvested, dechorionated, and then washed as described previously (Moritz and Alberts, 1998). The embryos were dried by blotting with paper towels, weighed, and then resuspended in 1 vol of extract buffer. The embryos were immediately homogenized by five passes of a motor-driven Teflon pestle in a glass Dounce homogenizer. The extract could be frozen in liquid nitrogen at this point and stored at -80°C. To prepare high speed supernatant for complementation tests and their associated immunodepletions, the crude extract was centrifuged for 20 min at ~228,000 *g* (TL100; Beckman Instruments, Inc., Fullerton, CA), the supernatant was transferred to a new tube, and then centrifuged again in the same way. For complementation tests, nocodazole (100 μ M final concentration) was added to the extract before centrifugation. To prepare the supernatant for characterizations of protein complexes, the crude extract was centrifuged for 10 min at 30,000 rpm in a Beckman TLA 100.3 rotor, transferred to a new tube and centrifuged again at 100,000 rpm for 8 minutes in the same rotor.

Antibodies

The rabbit antibodies to CP60 and to amino acids 385-508 of CP190 have been previously described (Kellogg et al., 1995; Oegema et al., 1995). The rabbit antibody to amino acids 705-789 of CP190 was prepared according to Oegema et al. (1995). One of the rabbit anti- γ -tubulin antibodies used was raised against the full-length maternal form of *Drosophila* γ -tubulin (these sequence data are available from GenBank/EMBL/DBJ under accession number P42271) expressed in baculovirus. The second antibody recognizing γ -tubulin was raised against the COOH-terminal peptide QIDYPQWSPAWEASKAG of the maternal form of *Drosophila* γ -tubulin. The production and purification of these antibodies will be described elsewhere.

Immunoprecipitations

To prepare the antibodies used for immunoprecipitation, 20-30 μ g of antibody was coupled to 50 μ l of packed Affiprep protein A beads (Bio-Rad Laboratories, Hercules, CA). The beads were first mixed by gentle rotation with antibody in PBST for 0.5-1 h at room temperature, and then washed three times with PBST, followed by three washes and resuspension in 0.2 M sodium borate, pH 9.0. To covalently attach the antibodies to the beads, dimethyl pimelimidate was added to 20 mM and the beads were incubated while rotating the tube gently for 0.5-1 h at room temperature. To inactivate residual cross-linker, the beads were washed into 0.2 M eth-

anolamine, pH 8.0, and rotated at room temperature for 2 h to overnight before use. The beads were then pre-eluted three times with 100 mM glycine, pH 2.3, before washing into extract buffer. To begin the immunoprecipitation, 50 μ l of packed beads were rotated with 300 μ l of concentrated embryo extract for 1 h at 4°C. The beads were pelleted and the supernatants sampled. The beads were washed four times with column buffer (or with extract buffer) plus 75 mM KCl, 0.05% NP-40 or 0.05% Triton X-100, and 1:200 protease inhibitor stock, and then once with the same buffer without detergent. In some cases (for immunoprecipitations to characterize protein complexes), proteins were eluted three times sequentially with 150 μ l of 100 mM glycine, pH 2.3. The elutions were pooled and neutralized by addition of 200 μ l 0.5 M K-Hepes, pH 7.6. For gel analysis, 20 μ g of porcine insulin was sometimes added as carrier and the samples were precipitated with TCA.

In experiments to test the ability of immunodepleted extracts to complement salt-stripped centrosomes, the beads were pelleted, washed as above, and then boiled in sample buffer for SDS-PAGE and Western analysis. Samples of the supernatants were kept for this purpose as well, and the remainder of the supernatant was used in the on-glass complementation assay.

Sucrose Gradient Sedimentation and Gel-filtration Chromatography

Sucrose gradients were poured as step gradients (five 950- μ l steps) that were allowed to diffuse into continuous gradients overnight at 4°C before use. The gradients were formed from 5-20% or 5-40% sucrose (Ultrapure; ICN Biomedicals, Costa Mesa, CA) in gradient buffer plus 75 mM, 100 mM, or 500 mM KCl, as indicated for each experiment. A 50-75- μ l aliquot of sample was loaded onto each gradient, and the gradients were centrifuged at 4°C at 50,000 rpm in a Beckman SW55 rotor for 4 to 8 h, as indicated. The gradients were fractionated from the top by hand into 16 300- μ l fractions. Protein standards (0.5 mg/ml each) were loaded in an equivalent volume and were run in parallel over identical gradients for each experiment.

Gel-filtration chromatography was carried out on a Superose-6 column by FPLC (Pharmacia Biotech Sevrage, Uppsala, Sweden) in column buffer plus 75 mM, 100 mM, or 500 mM KCl, as indicated. The column was calibrated with standards of known Stokes radii as indicated in the legend to Fig. 5. The size and shape (Stokes radii) of protein complexes were estimated according to (Siegel and Monty, 1966). The axial ratios of the equivalent prolate ellipsoids of revolution, $\{a/b\}_p$, were estimated according to (Laue et al., 1992), using the method of Kuntz (1971) to estimate the degree of hydration from amino acid sequence.

Sucrose Gradient Quantitation

Standards were precipitated by the addition of TCA to 10%, resuspended in sample buffer, separated by 8.5 or 11% PAGE, and then stained with Coomassie blue. Gels were scanned into the computer using a UMAX scanner (Fremont, CA), and the program NIH Image was used to quantitate band intensities. The peak fraction was assigned for each standard using Kaleidagraph (Synergy Software, Reading, PA). Standard curves of peak fraction versus sedimentation coefficient were then used to convert fraction number to S value (essentially $S_{20,w}$) for each sucrose gradient to allow direct comparison of protein complexes sedimented in 75 mM and 500 mM KCl. This use of standards to correct to $S_{20,w}$ from different buffers is valid as long as the partial specific volumes are the same for the standard proteins and the protein complexes being studied (Martin and Ames, 1961).

Quantitative Immunoblotting

For immunoblots, samples were precipitated by the addition of TCA to 10% and resuspended in Sample buffer before separation by SDS-PAGE on 10 or 11% gels. Proteins were then transferred to nitrocellulose (pore size 0.1 μ m) in the presence of 25% methanol, 0.15 M glycine, 0.02% SDS. The blots were incubated for 20 min in block (TBS + 0.1% Tween-20, 3% nonfat dry milk, 10% glycerol). A chemiluminescent substrate system (SuperSignal CL-HRP; Pierce Chemical Co., Rockford, IL) was used to detect the HRP-conjugated secondary antibodies. The developed film was scanned into the computer using a UMAX scanner and NIH Image was used to quantitate band intensities. Serial dilutions of CP190, CP60, and γ -tubulin were blotted simultaneously with all experimental fractions, allowing us to determine the relative concentrations of CP190, CP60, and γ -tubulin in each fraction, as shown in Fig. 6.

Results

In Vitro Complementation of Salt-stripped Centrosomes

To study the role of the γ TuRC in microtubule nucleation at centrosomes and to test for potential contributions to nucleation by other centrosomal components, we developed an *in vitro* assay in which the nucleating activity of salt-stripped *Drosophila* centrosomes is restored by incubation with embryo extract (Fig. 1). Initially, several different salts, including NaCl, KCl, and KI, as well as urea at various concentrations were tested for their ability to inactivate *Drosophila* centrosomes. Whereas all of the salts and urea were destructive to some extent (data not shown), we found that treatment with 2 M KI consistently destroyed the microtubule-nucleating activity of the centrosomes.

Using this complementation assay, we determined that a 228,000 *g* supernatant of an extract made from 0–2-h-old embryos was able to complement the KI-stripped centrosomes (Fig. 2). When extract was incubated on the coverslip in the absence of centrosomes, many microtubules, but virtually no asters, formed (Fig. 2, *b* and *c*). When KI-stripped centrosomes were incubated with buffer instead of extract, very few microtubules and no asters formed (Fig. 2 *d*). When KI-stripped centrosomes were incubated

with the complementing extract before the tubulin incubation, asters that look very similar to those that formed on buffer-treated centrosomes were produced (compare Fig. 2, *a* and *e*). Complementation occurred when the KI-treated centrosomes were incubated with extract containing nocodazole at either 0° or 30°C, indicating that aster regrowth was not merely due to the elongation of microtubules initiated during incubation with the extract (Fig 2, *e* and *f*).

To be confident that centrosomes were indeed present even when few or no asters could be found, the number of centrosomes on the coverslips was independently verified by immunofluorescent staining of α -tubulin in the centrioles (and γ -tubulin where possible). The number of centrosomes (whether intact or salt-stripped) bound to individual coverslips was found to be quite consistent (Fig. 2 *g*).

An initial characterization of the complementing extract showed that its activity could be destroyed by heating to 60°C or boiling, and that ATP was not required for complementation. In addition, when the extract was prepared under conditions that promote microtubule polymerization and the microtubules were removed by centrifugation, the extract did not complement. This suggests that at least one component required for the complementation binds to microtubules (data not shown).

The Centrosomal Proteins CP60, CP190, CNN, and γ -Tubulin Are Removed from Centrosomes by 2 M KI

Since it was possible to complement the salt-stripped centrosomes with soluble factors present in embryo extract, yet these same factors were not capable of inducing aster formation in the absence of the KI-treated centrosomes (Fig. 2), it appeared that a remnant structure must persist after treatment with 2 M KI. This remnant might be a scaffolding to which the soluble factors necessary for restoring microtubule nucleation attach. Therefore, we characterized what remains at the centrosomes after salt treatment, and how the known centrosomal proteins behave under such conditions. We began by determining whether the salt treatment removed the known centrosomal proteins CP60, CP190, CNN, and γ -tubulin (Kellogg et al., 1989; Oakley and Oakley, 1989; Kellogg and Alberts, 1992; Whitfield et al., 1995; Li and Kaufman, 1996). Centrosomes were incubated with 2 M KI, as described above for the complementation assay, and then pelleted. The centrosomal protein profile becomes simpler after KI treatment (Fig. 3 *a*). Immunoblotting of the pellets and supernatants of buffer- and KI-treated centrosomes shows that virtually all detectable CP60, CP190, CNN, and γ -tubulin are removed by the salt (Fig. 3 *b*). Electron microscopic examination of the KI-stripped centrosomes shows that the centrioles are destroyed to varying degrees by the salt, but that there is little obvious structural abnormality in the PCM (data not shown).

γ -Tubulin, but Not CP60 or CP190, Are Required for Restoring Microtubule-nucleating Activity to KI-treated Centrosomes

The fact that treatment of centrosomes with KI both led to a loss of microtubule-nucleating activity and extracted CP60, CP190, CNN, and γ -tubulin, suggested that one or more of these proteins might be needed for this activity. Therefore, we tested the effect of immunodepleting CP60,

Complementation Assay

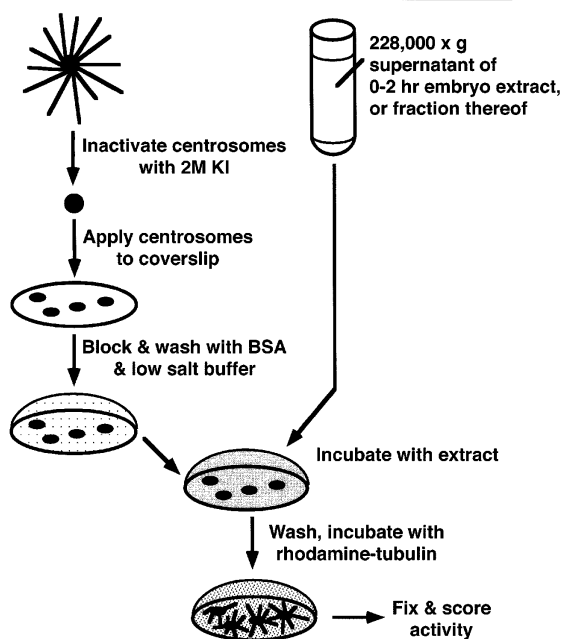


Figure 1. Complementation assay. Centrosomes isolated from *Drosophila* embryos are inactivated by incubation with 2 M KI. The KI-treated centrosomes are allowed to bind to a glass coverslip. The coverslip is washed and blocked with a low-salt, BSA-containing buffer, and then incubated with the extract or fraction to be tested. The extract/fraction is washed away and the coverslip is incubated with rhodamine-labeled tubulin. Any resulting asters are fixed sequentially with glutaraldehyde and methanol. The number of asters per 50 microscope fields (100 \times objective) is determined by counting samples while viewing through a fluorescence microscope. See Materials and Methods for details.

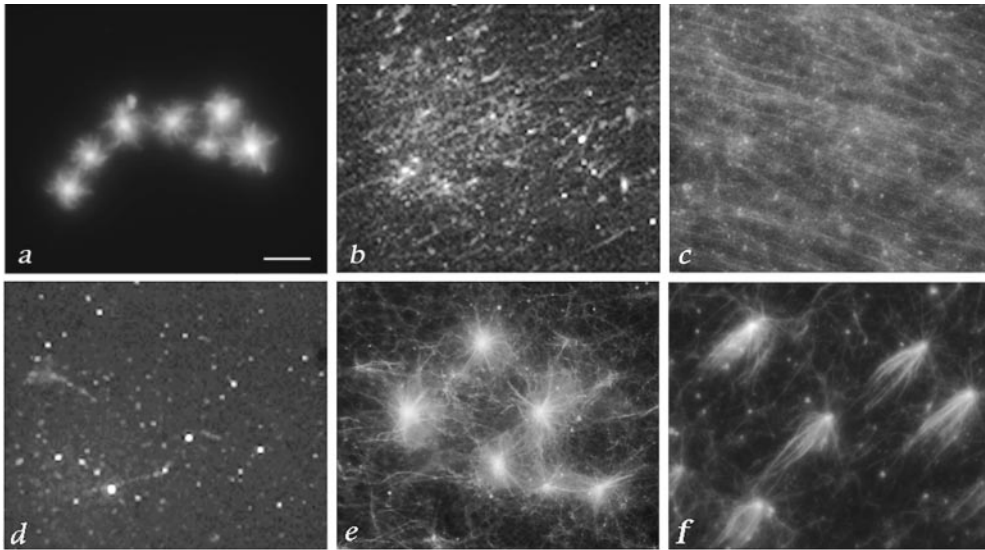
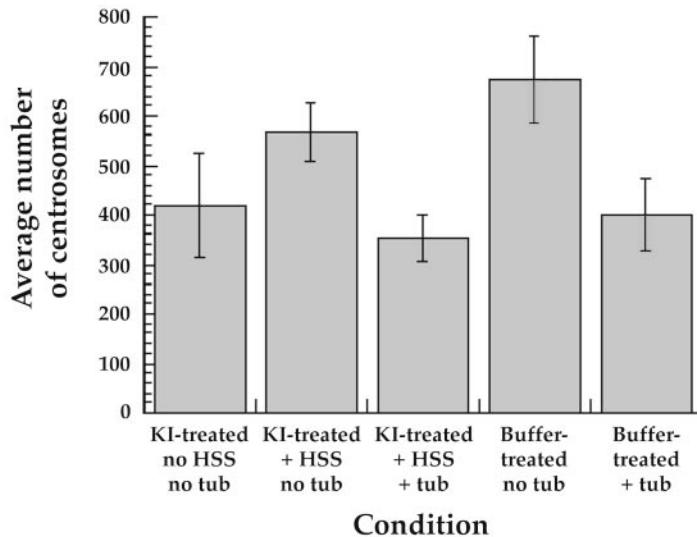


Figure 2. Examples of complementation of KI-treated centrosomes. The complementation assay was carried out as outlined in Fig. 1 and Materials and Methods. (a) Microtubule asters regrew on buffer-treated centrosomes that were incubated with rhodamine-labeled tubulin at 30°C. (b and c) Microtubules, but no asters, formed from a 228,000 g supernatant from a 0–2 h embryo extract was incubated at 30°C (b) or 0°C (c) on coverslips in the absence of centrosomes, followed by a 30°C incubation with rhodamine-tubulin. (d) When KI-treated centrosomes were incubated with buffer instead of extract, followed by rhodamine-tubulin, few microtubules and no asters formed. (e and f) Asters formed when KI-treated centrosomes were first incubated with extract at 30°C (e) or 0°C (f) and then with rhodamine-tubulin at 30°C. (g) KI- and buffer-treated centrosomes bind consistently to coverslips. The number of centrosomes bound to coverslips under the typical experimental conditions used throughout this study was determined by counting structures that were stained with antibodies against α - and/or γ -tubulin. The average number counted in 50 and 100 \times microscope fields is shown. Three to five separate experiments were counted for each condition. Bar, 10 μ m.

g



CP190, and γ -tubulin on the ability of the extract to complement salt-stripped centrosomes (we were not able to deplete CNN to a sufficient extent for this assay). Each protein was quantitatively depleted (Fig. 3 i), and the resulting extracts were tested in the *in vitro* assay (Fig. 1). Only the depletion of γ -tubulin had an effect on the ability of the extract to complement KI-inactivated centrosomes, and this activity was consistently destroyed by the removal of γ -tubulin (Table I; see Fig. 8 c).

We used immunofluorescence to further examine the effect of the KI treatment and extract complementation on the presence of γ -tubulin at centrosomes. In these experiments, staining centrosomes with antibodies against γ -tubulin and α -tubulin (which recognize PCM and centrioles, respectively) confirmed that γ -tubulin is removed from centrosomes by treatment with KI (Fig. 3, c and d; c' and d'). We also found that γ -tubulin reassociates with centrosomes during incubation with extract (Fig. 3, e and f; e'

and f'). To rule out the possibility that spurious asters form in the absence of centrosomes resulting from clustering of γ -tubulin into foci, we also stained coverslips that were incubated with extract followed by tubulin in the absence of centrosomes, and found no foci of γ -tubulin staining (Fig. 3, g and g'). These data were quantitated by measuring the fluorescence intensity of α - and γ -tubulin staining at intact (buffer-treated) and KI-treated centrosomes (Fig. 3 h).

Cumulatively, these results suggest that γ -tubulin is required for aster formation, whereas CP60 and CP190 are not. We had initially expected that CP60 and CP190 might also be at least indirectly required since our laboratory had previously found evidence for the existence of a protein complex containing these three proteins (Raff et al., 1993). These results led us to re-evaluate the interactions of CP60, CP190, and γ -tubulin. In addition, we were interested in characterizing further the γ -tubulin component required for complementation.

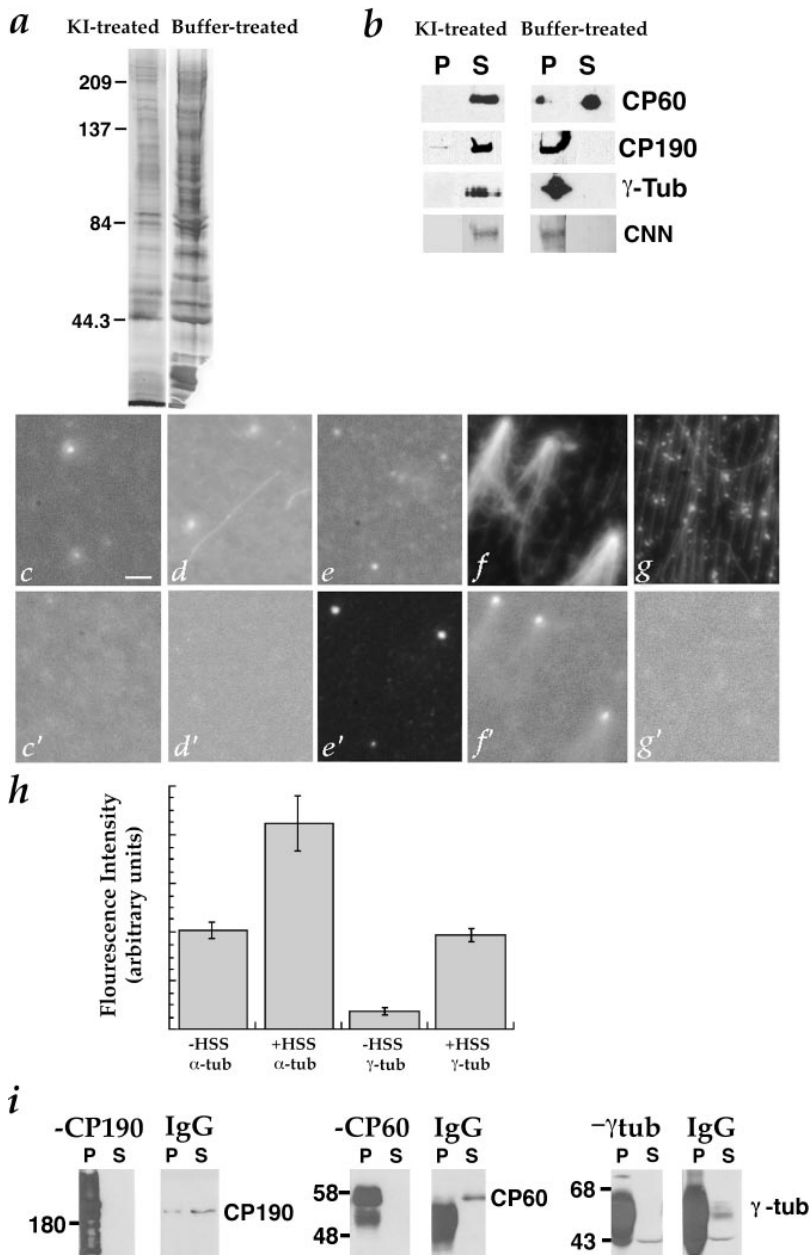


Figure 3. Effects of 2 M KI treatment on centrosomes. (a) The profile of centrosomal proteins is simplified by treatment with 2 M KI. Isolated centrosomes ($\sim 3 \times 10^7$) were mixed with an equal volume of $1 \times$ BRB80 + 4 M KI (left) or $1 \times$ BRB80 (right), incubated on ice for 10 min, and then pelleted by centrifugation at 30,000 g for 1 h. The pellets were washed three times with $1 \times$ BRB80 and resuspended in sample buffer, boiled, and then separated by SDS-PAGE on a 10% gel. The gel was silver stained. (b) Removal of CP60, CP190, CNN, and γ -tubulin from centrosomes by 2 M KI. $\sim 5 \times 10^6$ centrosomes were mixed with an equal volume of either $1 \times$ BRB80 + 4 M KI (left-hand panels) or $1 \times$ BRB80 (right-hand panels), and incubated on ice for 10 min. Centrosomes were pelleted by centrifugation at 30,000 g for 15 min, washed with $1 \times$ BRB80, and then resuspended in sample buffer for SDS-PAGE. Proteins released from centrosomes into the supernatants by the KI or buffer treatments were precipitated with 10% TCA and resuspended in sample buffer for SDS-PAGE. The presence of the centrosomal proteins CP60, CP190, CNN, and γ -tubulin in the pellets (P) and supernatants (S) was determined by immunoblotting. Top, CP60 was completely solubilized by KI, and partly solubilized by buffer. Middle, Most CP190 was solubilized by KI, but not by buffer. Bottom, γ -tubulin and CNN were completely solubilized by KI, but not by buffer. (c–g) Immunofluorescence (Texas red, donkey anti-mouse) localization of α -tubulin (DM1 α , to label centrioles and microtubules) on KI-treated centrosomes before and after complementation and with and without regrown microtubules, under the conditions indicated below. (c–g) Immunofluorescence (fluorescein, goat anti-rabbit) localization of γ -tubulin (rabbit anti- γ -tubulin) in the same fields shown in (c–g). The samples were stained after the following complementation assays were performed: (c, c') KI-treated centrosomes incubated with buffer alone (no extract, no tubulin). Note that α - (c) but not γ -tubulin (c') is present, indicating that KI has removed γ -tubulin from the centrosomes. (d, d') KI-treated centrosomes incubated with buffer followed by tubulin. (e, e') KI-treated centrosomes incubated with complementing extract followed by buffer (no additional tubulin). Note that γ -tubulin has re-associated with the centrosomes. (f, f') KI-treated centrosomes incubated with complementing extract followed by tubulin. (g, g') Coverslip without centrosomes, incubated with complementing extract followed by tubulin. Note the lack of γ -tubulin-staining foci. (h) 2 M KI removes some of the α -tubulin and most of the γ -tubulin from centrosomes, but these proteins reassociate with centrosomes during incubation with complementing extract (HSS). Fluorescence intensity was quantitated on KI-treated centrosomes costained with antibodies against α - and γ -tubulin. 50 centrosomes were measured for each condition. (i) Immunoblots showing quantitative immunodepletion of CP190, CP60, or γ -tubulin from the 228,000 g supernatant of a 0–2 h embryo extract that complements KI-treated centrosomes (see Materials and Methods for details). Left, CP190 was completely immunodepleted from the extract by affinity-purified anti-CP190 antibody, but not by non-immune IgG. Multiple forms of CP190 are visible in the –CP190 pellet. Middle, CP60 was immunodepleted by CP60 antibody, but not by non-immune IgG. (Bands in pellet lanes are antibody and/or CP60, which are of similar size). Right, γ -tubulin was immunodepleted by γ -tubulin antibody, but not by non-immune IgG. (γ -Tubulin comigrates with antibody, so bands in pellet lanes are antibody and/or γ -tubulin. In addition, the γ -tubulin antibody used for immunoblotting cross-reacts with a second ~ 43 -kD band). Bar, 10 μ m.

centrosomes incubated with complementing extract followed by buffer (no additional tubulin). Note that γ -tubulin has re-associated with the centrosomes. (f, f') KI-treated centrosomes incubated with complementing extract followed by tubulin. (g, g') Coverslip without centrosomes, incubated with complementing extract followed by tubulin. Note the lack of γ -tubulin-staining foci. (h) 2 M KI removes some of the α -tubulin and most of the γ -tubulin from centrosomes, but these proteins reassociate with centrosomes during incubation with complementing extract (HSS). Fluorescence intensity was quantitated on KI-treated centrosomes costained with antibodies against α - and γ -tubulin. 50 centrosomes were measured for each condition. (i) Immunoblots showing quantitative immunodepletion of CP190, CP60, or γ -tubulin from the 228,000 g supernatant of a 0–2 h embryo extract that complements KI-treated centrosomes (see Materials and Methods for details). Left, CP190 was completely immunodepleted from the extract by affinity-purified anti-CP190 antibody, but not by non-immune IgG. Multiple forms of CP190 are visible in the –CP190 pellet. Middle, CP60 was immunodepleted by CP60 antibody, but not by non-immune IgG. (Bands in pellet lanes are antibody and/or CP60, which are of similar size). Right, γ -tubulin was immunodepleted by γ -tubulin antibody, but not by non-immune IgG. (γ -Tubulin comigrates with antibody, so bands in pellet lanes are antibody and/or γ -tubulin. In addition, the γ -tubulin antibody used for immunoblotting cross-reacts with a second ~ 43 -kD band). Bar, 10 μ m.

Drosophila γ -Tubulin Is in a Protein Complex That Is Similar to the *Xenopus* γ TuRC

We used immunoprecipitations, gel filtration, and sucrose gradient sedimentation to investigate centrosomal protein complexes containing γ -tubulin, CP60, and/or CP190, which

might restore microtubule-nucleating activity to salt-stripped centrosomes. CP60, CP190, or γ -tubulin were each immunoprecipitated from concentrated embryo extracts and analyzed by SDS-PAGE (Fig. 4 a) and immunoblotting (Fig. 4, b and c). Antibodies recognizing γ -tubulin (Zheng, Y., unpublished observations) immunoprecipi-

Table I. Ability of Immunodepleted Extracts to Restore Microtubule-nucleating Activity to KI-treated Centrosomes

KI-treated centrosomes incubated with:	No. of asters per 50 fields*
Buffer	0
Extract + random IgG	94
Extract + protein A beads	93
Extract - γ -tubulin	5
Extract - CP60	88
Extract - CP190	100

The microtubule-nucleating activity of centrosomes was destroyed by incubation with 2 M KI. Inactivated centrosomes were incubated with the indicated buffer or extract followed by rhodamine-labeled tubulin. Samples were then fixed and examined under a fluorescence microscope and the No. of asters per 50 microscope fields (100 \times objective) was counted. (As described in Fig. 1.)

*Averages of 3 experiments.

tated γ -tubulin and a group of associated proteins that are components of the *Drosophila* γ TuRC. The protein profile of this complex is very similar to the that of the *Xenopus* γ TuRC (Fig. 4 a, right two lanes; Oegema, K., and Y. Zheng, manuscript in preparation) (Zheng et al., 1995). CP190 antibodies brought down CP190 and a large fraction of CP60 (Fig. 4 a, middle two lanes). Antibodies to CP60 immunoprecipitated CP60 and a small fraction of the CP190 found in extracts (Fig. 4 a, second lane from left).

Immunoblots of the supernatants and pellets from the immunoprecipitations confirmed the identity of the Coomassie-stained bands. Analysis of the immunoprecipitation supernatants showed that each protein was depleted by its corresponding antibody (Fig. 4 b). In addition, CP60 is largely depleted in the supernatants of extracts treated with antibodies against CP190 (Fig. 4 b).

Immunoprecipitation pellets from concentrated extract (+) or from buffer controls (-) are shown in Fig. 4 c. Neither CP190 nor CP60 was detected in the pellets from immunoprecipitations performed with the antibodies to γ -tubulin, nor was any γ -tubulin present in the pellets of immunoprecipitations performed with antibodies against CP60 or CP190. The majority of the CP60 in extracts coimmunoprecipitated with CP190, however, and only a small fraction of the CP190 coimmunoprecipitated with CP60. These results suggest that, although CP60 and CP190 associate in these extracts, neither CP190 nor CP60 are in a cytoplasmic complex with γ -tubulin.

In Embryo Extracts, γ -Tubulin Is Found in Two Distinct Complexes, Neither of Which Contain CP190 or CP60

To characterize potential protein complexes involving γ -tubulin, CP60, and CP190 further, we analyzed the behavior of these proteins by gel-filtration chromatography and by sucrose gradient sedimentation. The same concentrated embryo extract was simultaneously fractionated by both techniques in identical buffers containing either 75 or 500 mM KCl (Fig. 5). To facilitate the comparison between complexes containing γ -tubulin and those containing CP190 and CP60, we used a quantitative blotting technique that allowed us to determine the relative concentrations of CP190, CP60 and γ -tubulin in each fraction (Fig. 6). Sucrose gradients run in different salt concentrations cannot be directly compared because of differences in buffer den-

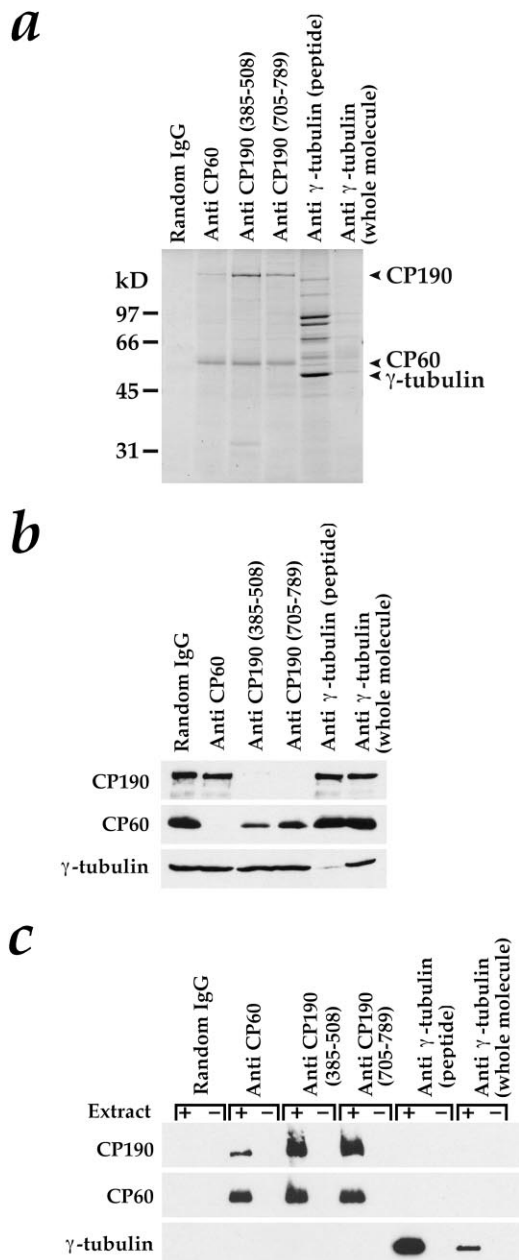


Figure 4. Tests for complex formation by immunoprecipitation. Since *Drosophila* γ -tubulin is the same size as IgG heavy chain, the immunoprecipitations in a-c were carried out with special care to avoid any IgG contamination in the pellets (see Materials and Methods). (a) Immunoprecipitation pellets after separation by SDS-PAGE on an 11% gel and staining with Coomassie blue. Anti-CP60 immunoprecipitated CP60 and a small fraction of the CP190. Both anti-CP190 antibodies immunoprecipitated CP190 and a large percentage of the CP60. Antibodies to γ -tubulin immunoprecipitated γ -tubulin and a group of γ -tubulin-associated proteins that are components of the *Drosophila* γ TuRC. The anti- γ -tubulin COOH-terminal peptide antibody was much more effective in immunoprecipitations than the antibody made to the whole γ -tubulin molecule. (b and c) Western blots to detect CP190, CP60, and γ -tubulin in immunoprecipitation supernatants (b) and pellets (c). In c, beads were incubated in the presence (+) or absence (-) of extract to control for antibody contamination in the pellets.

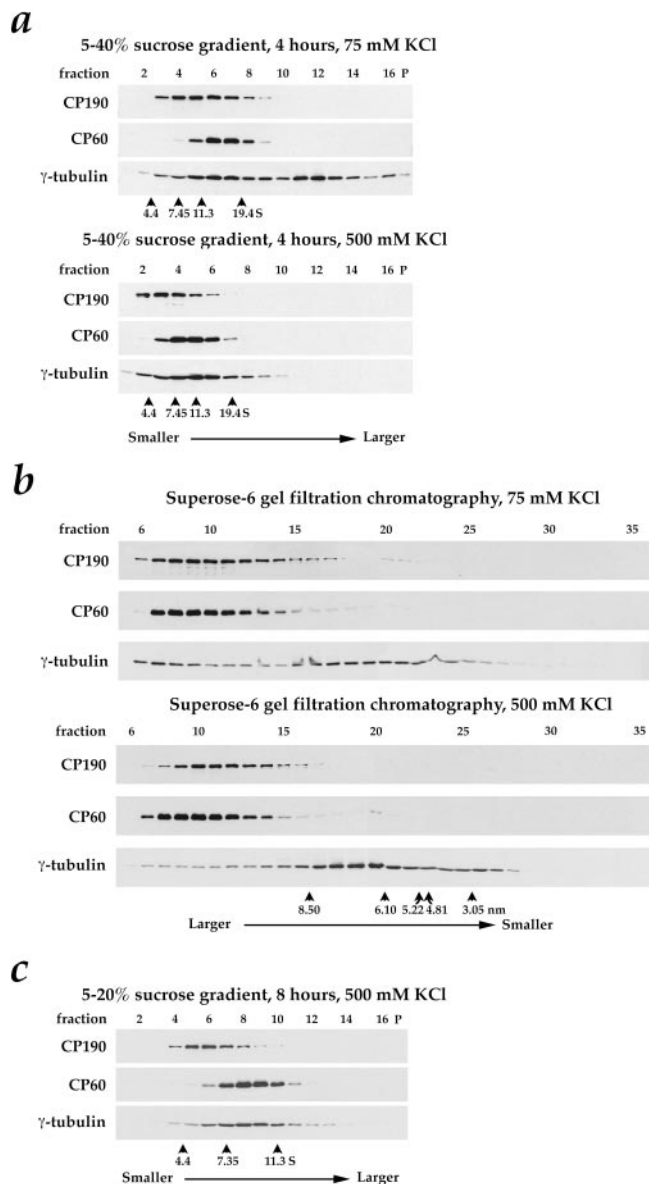


Figure 5. Behavior of CP60, CP190, and γ -tubulin during sucrose gradient sedimentation and Superose-6 gel filtration of concentrated *Drosophila* embryo extracts. Each fraction was immunoblotted for CP190, CP60, and γ -tubulin after separation by SDS-PAGE on an 11% gel. A single extract was first buffer exchanged using spin columns into column buffer containing either 75 mM or 500 mM KCl. An aliquot was then sedimented through 5–40% sucrose gradients for 4 h (*a*) or fractionated by Superose-6 gel filtration chromatography (*b*) in buffers containing 75 mM or 500 mM KCl. In *c*, a separate extract was sedimented on a 5–20% sucrose gradient for 8 h in 500 mM KCl to increase the separation between the smaller complexes present in the high salt. Sucrose gradient fractions were collected from the top of the gradient; gradient pellets are also shown (*P*). Standards were run in parallel with the extract over identical sucrose gradients. The location of the peak for each standard is indicated with an arrowhead above its S value (see Materials and Methods for determination of peak fractions). The sucrose gradient standards used were BSA (4.4 S), rabbit muscle aldolase (7.35 S), bovine liver catalase (11.3 S), and porcine thyroglobulin (19.4 S). The Superose-6 column was calibrated with bovine thyroglobulin (Stokes radius = 8.5 nm), horse spleen ferritin (6.1 nm), bovine liver catalase (5.22 nm), rabbit muscle aldolase (4.81 nm), and hen egg ovalbumin (30.5 nm). The location of the peak for each standard is indicated with an arrowhead above its Stokes radius in *b*.

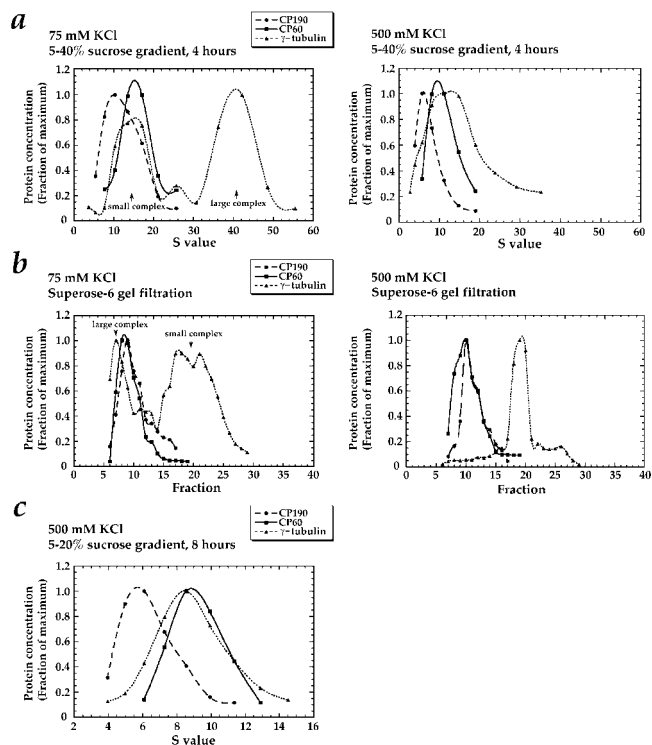
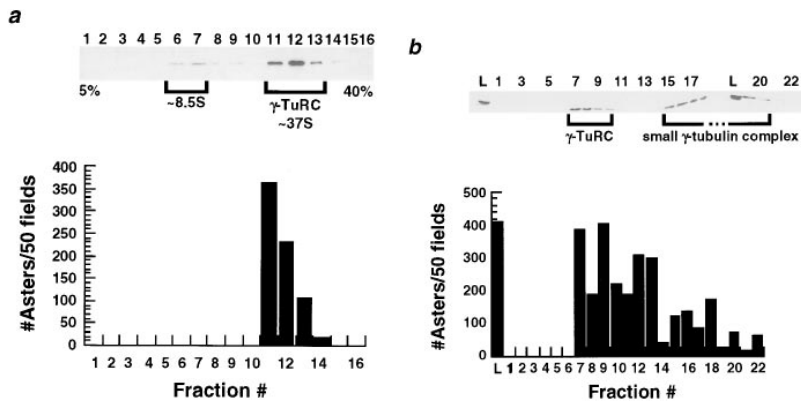


Figure 6. A graphical representation of the sucrose gradient (*a*) and gel filtration data (*b* and *c*) in Fig. 5. The γ -tubulin is present in two distinct complexes; CP190 and CP60 are not components of either complex. For each fraction, standard curves were used to determine the corresponding S value and the relative concentrations of CP190, CP60, and γ -tubulin (see Materials and Methods).

sity and viscosity that affect sedimentation rates. Therefore, standard curves of peak fraction versus S value, generated by loading proteins of known S value on identical sucrose gradients run in parallel with each experimental gradient, were used to convert fraction number to S value for each gradient.

In buffer containing 75 mM KCl, most of the γ -tubulin is found in two complexes that can be separated by both sucrose gradient sedimentation and by gel filtration (Figs. 5, *a* and *b*; and 6, *a* and *b*, left panels). The large γ -tubulin complex can be converted to the small γ -tubulin complex by raising the KCl concentration to 500 mM (compare γ -tubulin migration in Fig. 5 *a*, top and bottom; *b*, top and bottom; Fig. 6, *a* and *b*, compare left and right). In addition, the gel-filtration peak corresponding to the small γ -tubulin complex, which appears heterogeneous in 75 mM KCl, becomes much more homogeneous in 500 mM KCl. These results suggest that the small γ -tubulin complex is a subunit of the large γ -tubulin complex.

The sedimentation coefficients of the large and small γ -tubulin complexes are 36.9 S and 8.5 S, respectively. (The sedimentation coefficient of the large γ -tubulin complex was determined on a separate sucrose gradient using the 30 S ribosome particle as a standard, in addition to the standards mentioned in the legend to Fig. 5 [data not shown]). The Stokes radii of the small and large γ -tubulin complexes, estimated from our gel-filtration results, are 6.9 nm and ~20 nm, respectively (the large γ -tubulin complex fractionates close to the void volume of the Superose-6



The γ TuRC partially purified by FPLC on a Superose-6 gel-filtration column run in Column buffer + 100 mM KCl complements KI-treated centrosomes. *Top*, immunoblot showing that two γ -tubulin-containing complexes can be separated by gel filtration. The γ TuRC elutes just after the void volume. 50 μ l of the extract that complements KI-treated centrosomes was loaded on a 24 ml Superose-6 column. 0.5-ml fractions were collected. The column load (L) and fractions 1–22 are shown. *Bottom*, column fractions were tested for their ability to complement KI-treated centrosomes (Fig. 1). Fractions containing the γ TuRC were able to complement.

Figure 7. (a) The γ TuRC partially purified on a 5–40% sucrose gradient complements KI-treated centrosomes. The small γ -tubulin complex does not. 100 μ l of the extract that complements KI-treated centrosomes was loaded on a 5-ml 5–40% gradient made in column buffer + 100 mM KCl. See Materials and Methods for details on running and fractionating gradients. *Top*, immunoblot showing that two γ -tubulin-containing complexes can be separated on a 5–40% sucrose gradient. *Bottom*, sucrose gradient fractions were tested (Fig. 1) for their ability to complement KI-treated centrosomes and the number of asters that formed in 50 microscope fields was counted. Fractions 11–14, which contain the γ TuRC were able to complement KI-treated centrosomes. (b)

column, preventing a more precise determination). From these sedimentation coefficients and Stokes radii, the masses of the small and large γ -tubulin complexes were estimated to be 240,000 and \sim 3,000,000 D, respectively (Siegel and Monty, 1966).

On sucrose gradients, CP60 comigrates with the small γ -tubulin complex in both high and low salt (compare CP60 with γ -tubulin in Figs. 5 a, and 6 a, left panel). However, under the same buffer conditions, CP60 can be separated easily from the small γ -tubulin complex by gel filtration (compare CP60 and γ -tubulin peaks in Figs. 5 b and 6 b). By gel filtration in 75 mM KCl (Figs. 5 b, and 6 b, left panel), CP60 elutes with the large γ -tubulin complex, but under identical conditions, CP60 and the γ -tubulin large complex are easily separated on sucrose gradients (Figs. 5 a, and 6 a, left panel). Similarly, CP190 can be separated from the small γ -tubulin complex by gel filtration (Figs. 5 b, and 6 b), and from the large γ -tubulin complex on sucrose gradients (Figs. 5 a, and 6 a, left panel). These experiments indicate that neither CP60 nor CP190 are among the components of either the large or small γ -tubulin complexes. Interestingly, even in high salt conditions where CP190 and CP60 do not associate (data not shown), both proteins are found in large complexes. The properties of

these centrosomal protein complexes are summarized in Table II.

The γ -Tubulin Ring Complex Is Necessary, but Not Sufficient for Complementation of Salt-stripped Centrosomes

Since our immunodepletion studies demonstrated a requirement for soluble γ -tubulin in the complementation assay, we used the assay to determine if either the small or large γ -tubulin-containing complexes could complement the centrosome scaffolds. For this test we fractionated embryo extract on sucrose gradients or by gel filtration and then assayed the fractions for complementing activity. The sucrose gradient fractions containing the large complex complemented the salt-stripped centrosomes, whereas fractions containing the small complex did not (Fig. 7 a). Gel filtration fractions containing the large complex also gave good complementation; in addition, we observed some less robust complementation by the gel filtration fractions containing the small complex (Fig. 7 b).

These results suggested that the γ -tubulin large complex is not only essential, but possibly sufficient for complementation. To determine if this was the case, we used an antibody that recognizes the COOH-terminal 17 amino acids of the *Drosophila* maternal form of γ -tubulin to purify the large complex (we will refer to this subsequently as the *Drosophila* γ TuRC) in a manner similar to that described for the purification of the *Xenopus* γ TuRC (Zheng, Y., manuscript in preparation; Zheng et al., 1995), see Fig. 4 a for an immunoprecipitation showing the protein composition of this complex). The purified *Drosophila* γ TuRC has a ring structure very similar to that of the *Xenopus* complex (Zheng, Y., manuscript in preparation). To our surprise, we found that the immunoaffinity-purified γ TuRC was not able to complement.

We reasoned that an additional factor in the extract that was removed upon immunoaffinity purification of the γ TuRC was required to allow the γ TuRC to complement. To test this idea, equal volumes of pure γ TuRC and extract that had been immunodepleted using an anti- γ -tubu-

Table II. Properties of Centrosomal Protein Complexes in Extracts

	$S_{20,w}$	R_s	Mol wt (estimated)	{a/b}
		\AA		
γ -Tubulin large complex (75 mM KCl)	36.9 S	\sim 200	\sim 3,000,000	
γ -Tubulin small complex (500 mM KCl)	8.5 S	68.7	240,000	
CP190 (500 mM KCl)	5.7 S	146	343,000	19
CP60 (500 mM KCl)	8.9 S	149	546,000	23*

*Suggests that the CP60 oligomer is highly asymmetric.

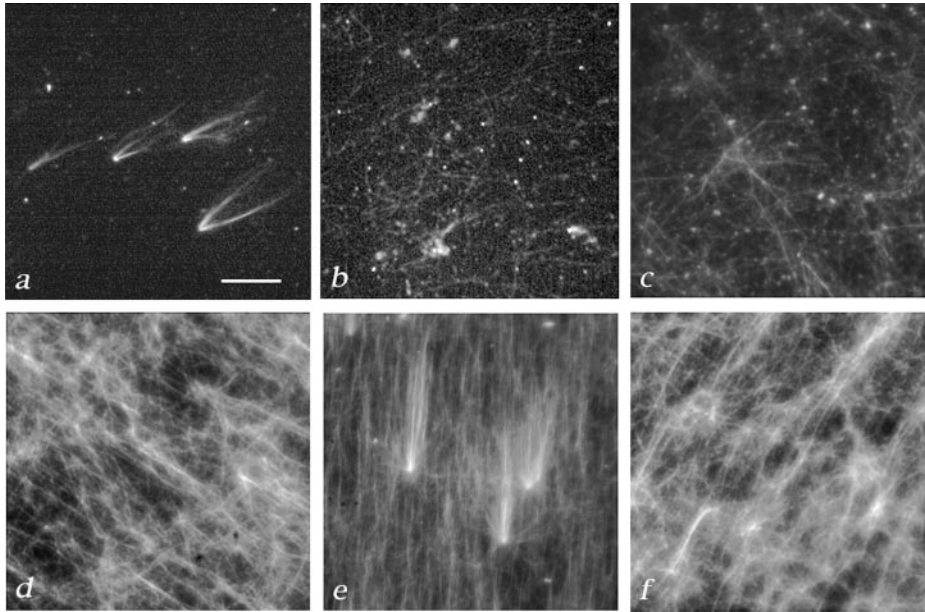


Figure 8. The γ TuRC is necessary, but not sufficient for complementation of salt-stripped centrosomes. Complementation tests were carried out as described in Fig. 1 and Materials and Methods. (a) Asters formed when KI-treated centrosomes were incubated with the partially purified large γ -tubulin complex from a Superose-6 column followed by rhodamine-tubulin. (b) Some free microtubules, but no asters formed when the same fraction was incubated without KI-treated centrosomes. (c) KI-treated centrosomes were not complemented by γ -tubulin-depleted extract. (4 asters were counted in 50 $100\times$ microscope fields). (d) KI-treated centrosomes were not complemented by immunoaffinity-purified γ TuRC, although microtubules could form (6 asters/50 microscope fields). (e) Asters formed after incubation of

KI-treated centrosomes with a 1:1 mixture of immunoaffinity-purified γ TuRC and γ -tubulin-depleted extract (151 asters/50 fields). (Other ratios of extract to γ TuRC were tested; although some activity is still detectable at a 20:1 ratio of extract to γ TuRC (data not shown). (f) Microtubules, but no asters formed when a 1:1 mixture of immunoaffinity-purified γ TuRC and γ -tubulin-depleted extract were incubated in the absence of centrosomes. Bar, 10 μ m.

lin antibody were mixed, and then tested for complementation of salt-stripped centrosomes in the on-glass assay (Fig. 8). Asters formed when KI-treated centrosomes were incubated with the partially purified large γ -tubulin complex from a Superose-6 column (Fig. 8 a). Some free microtubules, but no asters formed when the centrosomes were omitted (Fig. 8 b). If KI-stripped centrosomes were incubated with a mixture of pure γ TuRC and γ -tubulin-depleted extract, asters formed (Fig. 8 e). However, centrosomes incubated with pure γ TuRC or γ -tubulin-depleted extract alone were not able to produce asters (Fig. 8, c and d). In the absence of centrosomes, the mixture of pure γ TuRC and γ -tubulin-depleted extract formed microtubules, but no asters (Fig. 8 f).

We concluded that the immunisolated γ TuRC requires an additional factor found in extracts to complement salt-stripped centrosomes. To determine whether the order of addition of these two components is important, we incubated salt-stripped centrosomes sequentially with γ -tubulin-depleted extract followed by a washing step, and then by pure γ TuRC, or vice versa. The centrosomes only regained their ability to nucleate microtubule asters when they were first incubated with the γ -tubulin-depleted extract, followed by pure γ TuRC, or when they were incubated with a mixture of the two. The complementation worked best when the two components were added simultaneously (data not shown). Initial characterization by sucrose gradient sedimentation and gel filtration of this additional required factor indicates that it has an estimated molecular weight of 220,000 D (data not shown).

Discussion

To begin to understand microtubule nucleation in the context of the centrosome, we developed an in vitro comple-

mentation assay in which centrosome aster formation is reconstituted from salt-stripped centrosome scaffolds and a soluble fraction provided by a *Drosophila* embryo extract. This assay opens several new avenues for studying the structure and composition of the centrosome matrix. Extraction of centrosomes with the strongly chaotropic salt, KI, removes all of the known *Drosophila* components of core centrosomes, reducing the centrosomes to a simpler structure that appears to contain a scaffolding on which microtubule-nucleating sites may reassemble. The proteins left in this salt-resistant structure are unknown, but it should be possible to identify them by peptide sequencing.

Electron microscopy of the salt-stripped centrosome scaffolds did not reveal any striking modifications of the PCM, suggesting that CP60, CP190, CNN, and γ -tubulin are distributed throughout the PCM in intact centrosomes rather than being confined to particular regions. This is consistent with our previous EM observation that γ -tubulin is found at all levels of the PCM (Moritz et al., 1995b).

We have also characterized the soluble components contributed by the embryo extract that are required for centrosome aster reconstitution. By immunodepleting the *Drosophila* centrosomal proteins CP60, CP190, and γ -tubulin from the complementing extract, we found that γ -tubulin is absolutely required for aster formation in our assay. This requirement is similar to the requirement for γ -tubulin in the aster assembly assay in *Xenopus* extracts (Felix et al., 1994; Stearns and Kirschner, 1994). Previous work from our laboratory suggested that CP60, CP190, and γ -tubulin are in a protein complex together (Raff et al., 1993), yet we found that CP60 and CP190 were not necessary for complementation.

We therefore characterized the γ -tubulin-containing complexes found in *Drosophila* embryo extracts to deter-

mine if we could identify a specific complex required for complementation and to re-evaluate the association of γ -tubulin with CP190 and CP60. Protein complexes were immunoprecipitated or fractionated by sucrose-gradient sedimentation and gel-filtration chromatography at high and low salt concentrations. We found that in low salt, most of the γ -tubulin is found in two distinct complexes of 240,000 and \sim 3,000,000 D, which can be separated by either fractionation technique. The larger γ -tubulin complex can be converted into the smaller complex by 500 mM KCl, suggesting that the small γ -tubulin complex is a subunit of the larger one. The large γ -tubulin complex seen in these extracts has been purified to near homogeneity and is the *Drosophila* analogue of the γ -tubulin ring complex (γ TuRC) isolated from *Xenopus* egg extracts (Zheng, Y., and K. Oegema, manuscript in preparation; Zheng et al., 1995).

These experiments also demonstrated that although CP60, CP190, and γ -tubulin are all found in large protein complexes neither CP60 nor CP190 are among the components of the γ -tubulin complexes. How can we reconcile these data with previous results favoring a cytoplasmic complex containing CP190, CP60, and γ -tubulin? In the previous study (Raff et al., 1993) the complexes were not examined by gel-filtration chromatography, experiments that turned out to be crucial in the current work to distinguish the CP60 complex from the γ -tubulin small complex. However, in the previous study, γ -tubulin was detected by Western blotting in the elutions from immunoaffinity columns constructed from antibodies to CP60 or CP190, although it was a very minor component of these elutions, since a Coomassie blue-staining γ -tubulin band was never detected (Raff et al., 1993). In contrast, in this study we were not able to immunoprecipitate γ -tubulin with antibodies to CP60 or CP190, nor could we immunoprecipitate CP60 or CP190 with antibodies to γ -tubulin. One possible explanation for these disparate results is that although γ -tubulin is not found in a soluble complex with CP60 or CP190 in extracts, these proteins may assemble with each other to form a higher order complex on immunoaffinity columns and in vivo at the centrosome under conditions where the concentrations of these centrosomal proteins are higher than they are in extracts.

To explore the role of the large and small γ -tubulin-containing complexes in aster formation, we tested embryo extracts fractionated on sucrose gradients or by gel filtration for complementing activity in our assay. Although a direct role for the small complex is unclear, both sucrose gradient and gel filtration fractions containing the large complex (the *Drosophila* γ TuRC) can complement salt-stripped centrosomes (Figs. 7 and 8). This result was surprising because treatment with KI extracts all of the known components of the pericentriolar material in *Drosophila* (CP190, CP60, CNN, and γ -tubulin) and appears to substantially simplify the protein composition of the isolated centrosomes. The fact that both sucrose gradient and gel filtration fractions containing the γ TuRC can complement suggests that the connection between the γ TuRC present in the extract and the salt-resistant scaffold is likely to be direct.

In contrast to the γ TuRC in the sucrose gradient and gel filtration fractions, purified *Drosophila* γ TuRC is unable

to complement, suggesting that a component or modification of the γ TuRC that is required for complementation is lost during immunoaffinity purification. The possibilities for what this component or modification is doing include: (a) providing a physical link between the centrosome and the γ TuRC, or (b) modifying the scaffolds or the γ TuRC in some way so that the γ TuRC can bind to, or become activated at the centrosome. The fact that the purified *Drosophila* γ TuRC can nucleate microtubules in solution (Zheng, Y., and C. Wiese, unpublished observation) and when bound to the coverslips in our assay (Fig. 8, *d* and *f*), suggests that this component or modification probably is not necessary for nucleating activity but may instead be required for attachment of the γ TuRC to the salt-stripped centrosome scaffolds. The order of addition experiment further supports the idea that γ -tubulin-depleted extract may supply an attachment factor required to link the purified γ TuRC to the salt-stripped scaffolds.

We have found that this putative attachment factor has an estimated molecular weight of \sim 220,000 D. Pericentrin is one possible candidate for the factor, since it is a large coiled-coil, core centrosome structural protein that may interact with γ -tubulin (Doxsey et al., 1994; Dichtenberg et al., 1998). However, testing this possibility awaits the development of pericentrin reagents that work well in *Drosophila*.

In summary, we have developed an assay in which microtubule nucleation by *Drosophila* centrosomes is reconstituted from inactive salt-stripped centrosome scaffolds and soluble components derived from a *Drosophila* embryo extract. We conclude that the γ TuRC is required for microtubule nucleation at centrosomes and that the connection between the γ TuRC and the simplified scaffolds is likely to be directly mediated by a factor found in extracts that is normally loosely associated with the γ TuRC. We believe that our approach of attempting to simplify centrosome structure and function by focusing on smaller centrosomal protein complexes in conjunction with an in vitro complementation assay will be valuable in understanding this important but complicated organelle.

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