Brief Communication

Assessing floral trait variation in *Platanthera dilatata* (Orchidaceae) across an elevational gradient

Mortiz Plendl¹ · Genevieve E. van der Voort^{1,2} · Jasmine K. Janes^{1,2,3}

Received: 16 April 2024 / Accepted: 11 December 2024 Published online: 23 December 2024 © The Author(s) 2024 OPEN

Abstract

Flower morphology often changes over altitude, although the patterns themselves can be variable, with flowers being either smaller or larger. Floral trait variation is often considered in the context of pollinator-mediated selection. However, other explanations, including underlying genetics and plasticity, resource availability and floral enemies have been proposed. Here, we assess 10 floral traits in *Platanthera dilatata* var. *dilatata* across an elevational gradient on Vancouver Island, British Columbia, Canada, to determine if floral traits vary with altitude. We find that floral traits are larger at the lowest elevation site. However, much of the floral trait variation appears to be driven by temperature, which is not necessarily correlated with the altitudinal gradient. Given the intrinsic link between climate and resource availability, we suggest that resource availability confers a local selection pressure on floral trait size that may be balanced at larger spatial scales by antagonistic pressure from shared pollinators. Direct investigations of the environmental and genetic factors driving floral trait variation are recommended.

Keywords Orchid · Climate · Resource availability · Morphometrics · Pollinators

1 Introduction

Floral diversity is often thought to be driven by pollinators [1], leading to many theories and studies in relation to the most effective pollinator principle [2] and its impacts on floral evolution. Numerous examples exist of pollinator-mediated speciation occurring among sympatric populations as a result of different pollinators selecting for diverging floral traits (e.g., [3, 4]) including floral size (e.g., [5–7]). However, several other hypotheses have been put forward to help explain interspecific floral size variation. For example, the 'escape hypothesis' suggests that many floral parts serve a protective function from herbivores, pollen and nectar thieves, and fungal infection in addition to their role as pollinator attractants [8]. The 'resource cost hypothesis' acknowledges heterogeneous resource availability and posits that flower size will increase to attract pollinators only if resources are sufficient [8]. The role of genetics and plasticity on development have also been suggested, as increased cell division and cell elongation have been observed contributing to floral size changes in some species [9]. One plant family that seems particularly amenable to floral variation studies is the Orchidaceae [1, 10, 11].

[☐] Jasmine K. Janes, Jasmine.Janes@unbc.ca | ¹Biology Department, Vancouver Island University, 900 Fifth St, Nanaimo, BC V9R 5S5, Canada. ²Ecosystem Science and Management, University of Northern British Columbia, Prince George, BC V2N 4Z9, Canada. ³IUCN, Species Survival Commission, Orchid Specialist Group, Gland, Switzerland.





Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s44372-024-00071-6.

Platanthera dilatata (Pursh) Lindl. ex Beck (Orchidaceae; white bog orchid) is native to North America [12]. The plants range in height (11–130 cm), have many white flowers in a single inflorescence, and the number of leaves at the base varies (typically 2–5) [13]. Three variants of P. dilatata are formally recognized and differentiated based on floral spur length— P. dilatata var. leucostachys (Lindl.) Luer has a spur longer than the labellum, P. dilatata var. albiflora (Cham.) Ledeberg. has a spur shorter than the labellum, and *P. dilatata* var. dilatata (Pursh) Lindl. ex Beck spurs are similar in length to the labellum [12] (Table 1). It has been suggested that variation in the length of the nectar-producing floral spurs drives, and responds to, pollinator efficacy as the length of the pollinator's proboscis/tongue must be compatible with the length of the spur [13]. Thus, variation in floral traits, such as spur length, could prevent gene flow among populations of P. dilatata by excluding certain pollinators, ultimately leading to reproductive isolation. Considerable variation has been observed among populations of *P. dilatata*, especially in spur and labellum lengths (see descriptions from [12, 14–17]) (Table 1).

The floral morphology of *P. dilatata* sensu lato has been assessed in previous studies in the continental USA [14], but not in Canada, which represents a substantial proportion of its range. Western Canada, in particular, is believed to exhibit great variation in floral form [17]. Assessments of population-level variation of the spur, and comparisons among other floral features of *P. dilatata* var. *dilatata* are sparse [15]. In addition, information on pollinators and important abiotic factors throughout the range of *P. dilatata* is limited. However, recent observations of *P. dilatata* show that a number of insect families are likely pollinators [18], that different habitats (e.g., bog versus fen) often result in different mycorrhizal associations [15], and that populations exhibiting morphological variation are often genetically differentiated but also contain hybrids [15]. These observations suggest a number of antagonistic processes may be at work on P. dilatata. For example, pollinators may provide a directional selection pressure while abiotic factors may confer balancing or disruptive selection pressure.

After observing considerable variation in *P. dilatata* plant height between low and high elevation sites (pers. obs.), we used floral morphometrics to investigate variation among populations of P. dilatata var. dilatata. Floral morphometrics has been successfully used to identify new plant species [19], and to better understand the evolutionary relationships among sympatric and closely related European Platanthera species [20]. We were particularly interested in understanding how variable floral traits are along the aforementioned elevational gradient, and if floral trait variation is a response to biotic or abiotic factors. This work provides an important stepping stone in teasing apart the factors contributing to the evolutionary mosaic of *P. dilatata* populations.

2 Methods

All plants were identified as Platanthera dilatata var. dilatata based on the spur being roughly equal in length to the labellum [17]. From here on we refer simply to P. dilatata. Table 2 provides full population details including the habitat type. Three flowers were sampled from five plants per population (N = 75) across an elevational gradient (> 1,000 m). Samples collected within the Strathcona Provincial Park boundary were collected in 2020 under a Letter of Authorization (98700-20/Strathcona). Fully expanded flowers were collected from the lower, middle, and upper portion of the inflorescence to capture any developmental variation [13]. Within a population, individual plants were at least 1 m apart to avoid sampling clones. Each flower was preserved in 70% ethanol until measurements could be made.

Table 1	Literature-derived
labellur	n and spur lengths for
variants	s of Platanthera dilatata

Species	Spur length (mm)	Labellum length (mm)	Literature source
P. dilatata var. albiflora	Up to 10.0	Longer than spur	Leur [14]
	2.0-7.0	6.0–10.0	Sheviak [17]
	2.5–3.7	3.9–5.0	Wallace [16]
P. dilatata var. dilatata	5.0-10.0	5.0–10.0	Leur [14]
	4.0-12.0	5.0–10.0	Sheviak [17]
	3.5–5.8	3.3–6.1	Wallace [16]
P. dilatata—no variant specified	6.5–12.0	5.25–9.5	Wallace and Bowles [15]
	2.0–12.9	3.3–8.17	Adhikari and Wallace [12]
P. dilatata var. leucostachys	10.0-20.0	5.0–13.3	Leur [14]
	8.0-20.0	4.0-11.0	Sheviak [17]
	7.0–10.6	4.8–7.1	Wallace [16]



		•	•			
Population	Altitude (m)	Latitude	Longitude	Habitat	Total plants	Total flowers
Ranger cabin (Rang)	1,229	49.70504	– 125.33553	Alpine meadow	5	15
Paradise meadows (Para)	1,071	49.73801	- 125.31448	Alpine meadow	5	15
Strathcona 800 (S800)	800	49.74280	- 125.21096	Roadside seepage	5	15
Strathcona 400 (S400)	400	49.74271	- 125.24601	Roadside seepage	5	15
Quadra Island (Quad)	88	50.17060	- 125.22848	Estuary	5	15
				Total	25	75

 Table 2
 Site information for Platanthera dilatata samples used in this study.

Three flowers were sampled from five plants per population (N = 75)

Flowers were dissected under 16X magnification (Wild M3B, Heerburgg, Switzerland), and 10 floral traits were measured with digital calipers (Neoteck 150 mm Digital Vernier Caliper, China; resolution 0.01 mm, accuracy to 0.03 mm) to the nearest 0.01 mm. Measured traits are listed in Table 3. These 10 traits were selected for two reasons: (1) preliminary assessments in which a subsample of flowers were measured twice showed that measurement replicates deviated from one another by < 5%; and (2) similar floral traits are commonly used in orchid morphometrics (e.g., [21, 22]). We did not include measurements of the gymnostemium, even though the distance between the anthers is often considered an important mechanism for reproductive isolation [23], because the percent deviation for these measurements was > 5% in preliminary assessments.

Climate variables for each population were obtained from ClimateBC v7.42 [24]. This application downscales gridded monthly climate data from PRISM [25] in order to provide scale-free seasonal and annual climate variables. Climate variables included measurements related to temperature, precipitation, radiation, degree days, evaporation, humidity, and moisture index. Supplementary Table 1 contains a full list of the seasonal and annual climate variables.

2.1 Data analysis

Analyses were performed in R 4.2.2 (R Core Team, 2021) and RStudio 2022.12.0.353 (Posit Team, 2022). Summary statistics (e.g., mean, standard deviation, standard error) were calculated for each floral trait. A mean value for the three flowers per plant was calculated to avoid non-independence in some analyses. The normality of morphometric measurements for each floral trait were assessed using a Shapiro–Wilk test [26]. The rstatix package [27] was used to perform one-way ANOVAs for parametric variables and Kruskal–Wallis [28] tests for nonparametric variables, with *post-hoc* Tukey [29] and Dunn [30] tests with Bonferroni correction [31], respectively. ANOVA results were visualized using ggplot2 [32]. Additionally, we performed a linear mixed model using all flower replicates per plant in the Ime4 package [33]. Here, population was the fixed effect and plant was the random effect for each floral trait (response variable).

To visualize relationships among the morphological data, and reduce dimensionality, a principal component analysis (PCA) was performed using the packages vegan [34] and ellipse [35] in R. The number of principal components to be retained was assessed by screeplot and the influence of each trait on the PCA was determined by the PCA loadings. A

Trait	Description	Range (mm)	Mean (±SE)
Dorsal sepal length (dor.s.l)	Longest part of the dorsal sepal from perianth attachment to distal end	3.76–7.46	5.40 (±0.10)
Dorsal sepal width (dor.s.w)	Widest part of the dorsal sepal	2.24-3.98	3.08 (±0.04)
Lateral sepal length (lat.s.l)	Longest part of a lateral sepal from perianth attachment to distal end	4.29-8.18	6.55 (±0.11)
Lateral sepal width (lat.s.w)	Widest part of a lateral sepal	1.75–3.45	2.59 (±0.04)
Lateral petal length (lat.p.l)	Longest part of a lateral petal from perianth attachment to distal end	3.63–7.58	5.66 (±0.10)
Lateral petal width (lat.p.w)	Widest part of a lateral petal	1.74-4.08	2.64 (±0.06)
Labellum length (lab.l)	Longest part of the labellum from perianth attachment to distal end	2.37–9.97	7.10 (±0.18)
Labellum width (lab.w)	Widest part of the dilated portion of the labellum	1.95–6.82	2.98 (±0.10)
Spur length (spu.l)	Total length of the spur from perianth attachment to distal end	5.04-10.72	7.40 (±0.14)
Spur width (spu.w)	Width of the spur near the point of perianth attachment	0.47–1.46	0.93 (±0.02)

Table 3 Descriptions of floral traits measured, the range of values, and the mean measurement per trait (mm)



canonical correspondence analysis (CCA) was performed to assess the relationships among floral traits within populations and climate variables [36].

3 Results

Considerable variation in each floral trait was observed (Table 3). The majority of floral traits increased in size with decreasing altitude (Fig. 1). Dorsal sepal, lateral sepal, and spur width values were greater at the lowest altitude (Quadra Island) but these width values were also high at one of the mid-elevation sites (Strathcona 800) (Fig. 1). The greatest labellum lengths were observed at the second lowest altitude site, Strathcona 400 (Fig. 1). Using the mean from three flowers to avoid non-independence, the Shapiro–Wilk test of normality indicated that all floral traits except spur width were parametric. One-way ANOVA results of parametric floral traits showed a significant difference among populations for all floral traits (P < 0.05) except labellum width (Table 3; Fig. 1). Tukey test comparisons showed significant interactions for the following floral traits: dorsal sepal length (Rang-S400, P < 0.01; Rang-Quad, P < 0.01; Para-S400, P < 0.05; Para-Quad, P < 0.05; Fig. 1A), lateral petal length (Rang-S400, P < 0.05; Rang-Quad, P < 0.05; Para-Quad, P < 0.05; Fig. 1C), lateral petal width (Rang-Quad, P < 0.01; Para-Quad, P < 0.05; Fig. 1D), lateral sepal length (Rang-S400, P < 0.01; Rang-Quad, P < 0.01; Para-S400, P < 0.01; Para-Quad, P < 0.01; Fig. 1E), lateral sepal width (Para-Quad, P < 0.05; Fig. 1F), spur length (Rang-Quad, P < 0.05; Fig. 1G), and labellum length (Rang-S400, P < 0.05; Fig. 1I). Dorsal sepal width showed a significant difference among populations (F = 3.364, 207 P = 0.02) but the post hoc Tukey test comparisons showed no significant interactions (Fig. 1B). Spur width, which was nonparametric, was significantly different among populations according to the Kruskal–Wallis test (df=4; X^2 =13.2; P=0.01; Table 4). A significant interaction was shown by the post hoc Dunn test (Para-Quad, P < 0.05; Fig. 1H). Results from the linear mixed effect models supported those obtained from the one-way ANOVAs (results not shown).

The first two PCA axes explained 77% of the variation among populations (Fig. 2A). The PCA loadings showed that lateral petal length and labellum width had the most influence (maximum and minimum, respectively) on PC1, while labellum width and labellum length had the most influence (maximum and minimum, respectively) on PC2 (Fig. 2B). All floral width traits had a positive influence on PC2 while all length traits had a negative influence (Fig. 2B). In ordination space, PC1 separated populations by elevation rather than habitat type, with higher elevation sites (e.g., Paradise Meadows and Ranger Cabin) being more similar to each other than to lower elevation sites (Fig. 2A). The lowest elevation population, Quadra Island (Quad), is influenced strongly by floral trait widths, whereas length of floral trait appears to influence the Strathcona 400 (S400) population (Fig. 2A).

The CCA biplot for annual climate variables shows that the temperature difference between the mean warmest and coolest months (TD—or continentality effect) has a strong influence on the lowest elevation site and width of floral traits (Fig. 3A, B). Mean annual temperature (MAT), mean warmest and mean coldest month temperatures (MWMT and MCMT, respectively) had a strong negative influence on the high elevation sites (Ranger Cabin and Paradise Meadows) (Fig. 3A). Maximum seasonal temperature variables (Tmax wt = winter mean max. temp.; Tmax at = autumn mean max. temp.; Tmax_sp = spring mean max. temp.; Tmax_sm = summer mean max. temp.) exhibit a similar influence on populations and floral trait widths (Fig. 3C, D). One mid-elevation site (Strathcona 800) experienced temperatures that were comparable to the lowest elevation site (Quadra Island) (Fig. 3).

4 Discussion

We assessed floral variation among plants of P. dilatata var. dilatata across an elevational gradient. Generally, floral traits were larger at low elevation sites, although some mid-elevation sites from roadside habitats displayed equally wide (i.e., spur, lateral sepal, dorsal sepal) or long (i.e., lateral sepal, labellum, dorsal sepal) floral traits (Fig. 1). The PCA separated populations according to elevation rather than habitat type, based on the fact that two well-separated mid-elevation populations were from the same habitat type (Fig. 2A; Table 2). The PCA, which explained 77% of the variation, showed that lateral petal length and labellum width were the most influential variables contributing to axes 1 and 2, respectively (Fig. 2). The CCA indicated that annual and seasonal climate variables associated with temperature explained much of the variation in floral traits, particularly the width of floral traits (Fig. 3).



Fig. 1 Box plots displaying ANOVA results with Tukey multiple comparison post hoc test significance (**A–G**, **I**) and Kruskal–Wallis results with Dunn multiple comparison post hoc test significance (**H**). Significance indicated by *>0.05; **>0.01. The tenth floral trait, labellum width, is not displayed as ANOVA results were non-significant (refer to Table 4 for significance values)



We observed considerable variation among floral traits (Fig. 1; Table 3). Our spur length range was within those previously recorded by [12, 14, 15, 17] (Table 1) but longer than the spur lengths recorded by [16]. The labellum lengths observed here were largely within the range of other recorded labellum lengths (Table 1) but we did observe some that were quite small by comparison (2.37 mm; Table 3). While floral traits appear quite variable across the range of *P. dilatata*, they do not appear to have changed over time (e.g., measures from 1975 are still comparable to measurements today),



Discover Plants (202

(2024) 1:63

https://doi.org/10.1007/s44372-024-00071-6

Table 4Results from one-wayANOVA and Kruskal–Wallistests

Floral trait	Test	F _{4, 20}	X ² (df)	P-value
Dorsal sepal length	One-way ANOVA	7.66		0.00***
Dorsal sepal width	One-way ANOVA	3.36		0.02*
Lateral sepal length	One-way ANOVA	9.05		0.00***
Lateral sepal width	One-way ANOVA	4.60		0.00**
Lateral petal length	One-way ANOVA	5.46		0.00**
Lateral petal width	One-way ANOVA	0.74		0.00**
Labellum length	One-way ANOVA	4.32		0.01*
Labellum width	One-way ANOVA	0.74		0.58
Spur length	One-way ANOVA	3.61		0.02*
Spur width	Kruskal–Wallis		13.2(4)	0.01*

Significance levels: * 0.05; ** 0.01; *** 0.001





O Discover



Fig. 3 Canonical correspondence analysis relationships among sites and annual climatic variables (A), morphological flower traits and annual climatic variables (B), sites and seasonal climatic variables (C), and morphological flower traits and seasonal climatic variables (D)

suggesting that different selection pressures operate at local scales (i.e., evolutionary mosaic) but an overall balancing selection pattern is maintained at larger spatial scales.

Altitudinal variation in floral traits has been observed in a number of different plant species [37-39], including orchids [40, 41]. Several explanations have been put forward to explain the changes, including: pollinator shifts [37], genetic variation [42], resource availability [43], and climate [37]. With respect to pollinators, recent studies of P. dilatata are revealing that the species likely has a number of pollinators in the Hymenoptera and Lepidoptera [15, 18]. In the context of this study area, previous work suggests that Bombus flavifrons, B. melanopygus, B. mixtus, B. sitkensis, and B. vancouverensis of the Hymenoptera, and Argynnis hydaspe (syn. Speyeria hydaspe), Rhuemaptera sp., and a Pterophorid of the Lepidoptera are pollinators [18], at least at the higher elevation sites. Records from iNaturalist indicate that all of these insect species have been observed across the elevational gradient at some point in time but general information on A. hydaspe indicates that this species prefers higher altitude habitats [44]. The Bombus spp. are cited as having tongues in the 4.6–5.5 mm range [45, 46]), while Argynnis spp. may have proboscis lengths of 12–14.6 mm [47]. We were unable to find specific length observations for Rheumaptera or Pterophoridae except general descriptions of 'well-developed' [48]. Given the observed tongue lengths for many Bombus spp. relative to the spur lengths observed for the lower altitude sites, this genus may not be a particularly effective pollinator at these sites. Typically, the length of the tongue or proboscis must be compatible in order for the pollinator to obtain its nectar reward [13, 49, 50]. However, nectar production in P. chlorantha can reach a point where the spur overflows [51]. Thus, ample nectar production, coupled with wider and longer spurs at lower elevations, may eliminate incompatibility among pollinators and spur lengths that might otherwise prevent gene



flow between populations at different altitudes. Nectar production in *P. dilatata* appears unstudied so further work on correlations between spur length/width on nectar production and reproductive success would be valuable.

Research on *P. dilatata* populations in Alaska identified two broad categories of floral trait variation—large versus small flowers [15]. These morphotypes were found to be genetically distinct, although admixed individuals were identified in each population [15]. The prevalence of admixture suggests that pollinators and their efficacy were not negatively impacted by changes in floral size. While a genetic basis to floral size variation in *P. dilatata* has been identified, it is unclear how much of the phenotypic variation observed is due to plasticity from GxE interactions. The influence of herbivores and nectivores may also have a direct link as genetically distinct populations and morphotypes may exhibit differential palatability and susceptibility to floral enemies [8]. In this context, selection pressure from floral enemies (i.e., disruptive or directional—maintaining different genotypes) may provide an antagonistic pressure to pollinators (i.e., stabilizing—mixing genotypes). References to nectivores and herbivores that target flowers in the Orchidaceae are not common (e.g., [52]); most studies report damage to leaves [13, 53]. Direct investigations of floral enemies and common garden experiments could provide considerable insight into the ecological and genetic processes at work.

Another explanation for floral trait variation over elevation is the availability of resources [8] although, it is often difficult to disentangle the link between resource availability (i.e., water, nutrients) and climate [54]. Studies show that populations at higher altitudes typically experience shorter growing seasons [55] and greater seasonal fluctuation in resources [56], resulting in smaller floral traits and plant sizes [43]. We observe an interaction among annual and seasonal temperature variables with floral trait sizes, particularly the width of floral traits (Fig. 3). One of the mid-elevation sites (Strathcona 800), experienced annual and seasonal temperatures that were comparable to the lowest elevation site (Quadra Island). This may be due to slight differences in aspect and/or distance to the road, which may reduce albedo and contribute to slightly warmer temperatures. Warmer temperatures have been shown to have mixed effects on flowering traits across a broad range of species, with some species producing fewer [57] or more [58], or larger [59] or smaller [60] flowers in response. Further, temperature is known to impact numerous plant physiological responses [61] and the availability of resources both directly (e.g., evaporation of water) and indirectly (e.g., cycling of nutrients by microbial communities) [54]. Interactions among climate and resources could lead to changes in gene expression and phytohormone production [61] resulting in changes to cell division and anisotropy (e.g., differential growth) in floral meristems. Anisotropy is known to be the main driver of spur length diversity in several species [9, 62]. It would be valuable to design studies that systematically assess the effects of environmental or climatic variables correlated with altitude on the development of orchid flowers.

5 Conclusion

Our study reveals that floral traits of *P. dilatata* var. *dilatata* typically increase in size at lower elevations. Evaluations of climate variables along this elevational gradient suggest that temperature has the greatest influence, although temperature is not necessarily correlated with altitude in this study. Thus, the 'resource cost hypothesis' appears to be the best explanation for floral trait variation in *P. dilatata* given the strong interactions among climate and resources. This study provides an important basis for future work investigating the ecological and genetic mechanisms behind floral trait variation contributes to or diminishes local adaptation, and what consequences this may have on pollinator-mediated selection moving forward.

Acknowledgements The authors would like to acknowledge the assistance of MacIntyre Barrera in the field, and the assistance of Andy Smith and Erica McLaren for their support in obtaining the relevant Provincial Park permissions.

Author contributions MP performed data collection, measurements, analysis, and writing. GEV assisted with measurements and writing. JKJ collected samples, funded and supervised the project, and assisted with writing.

Funding This research was supported by Canada Summer Jobs (2021) and a Natural Sciences and Engineering Research Council of Canada Discovery Grant (RGPIN-2020-04475) to JKJ. GEV was funded by a Natural Sciences and Engineering Research Council of Canada Undergraduate Student Research Award (2022) through Vancouver Island University.

Data availability All data is available on request.

Code availability No custom code was used.



Declarations

Ethics and consent to participate Plant samples collected from within the Strathcona Provincial Park boundary were collected in 2020 under a Letter of Authorization (98700-20/Strathcona). Samples collected outside of the Strathcona Provincial Park boundary were on Crown Land and did not require a permit.

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- 1. Trunschke J, Sletvold N, Ågren J. The independent and combined effects of floral traits distinguishing two pollination ecotypes of a moth-pollinated orchid. Ecol Evol. 2019;9(3):1191–201. https://doi.org/10.1002/ece3.4808.
- 2. Stebbins GL. Adaptive radiation of reproductive characteristics in angiosperms, l: pollination mechanisms. Annu Rev Ecol Syst. 1970;1(1):307–26. https://doi.org/10.1146/annurev.es.01.110170.001515.
- 3. Cardona J, Lara C, Ornelas JF. Pollinator divergence and pollination isolation between hybrids with different floral color and morphology in two sympatric Penstemon species. Sci Rep. 2020;10(1):8126. https://doi.org/10.1038/s41598-020-64964-8.
- 4. Cuevas E, Espino J, Marques I. Reproductive isolation between Salvia elegans and S. fulgens, two hummingbird-pollinated sympatric sages. Plant Biol. 2018;20(6):1075–82. https://doi.org/10.1111/plb.12874.
- 5. Boberg E, Alexandersson R, Jonsson M, Maad J, Ågren J, Nilsson LA. Pollinator shifts and the evolution of spur length in the mothpollinated orchid Platanthera bifolia. Ann Bot-Iondon. 2014;113(2):267–75. https://doi.org/10.1093/aob/mct217.
- 6. Pleasants JM, Moe S. Floral display size and pollination of the western prairie fringed orchid, Platanthera praeclara (Orchidaceae). Lindleyana. 1983;8:32–8.
- 7. Robertson JL, Wyatt R. Evidence for pollination ecotypes in the yellow-fringed orchid, Platanthera ciliaris. Evolution. 1990;44(1):121–33. https://doi.org/10.1111/j.1558-5646.1990.tb04283.x.
- 8. Galen C. Why do flowers vary? Bioscience. 1999;49(8):631-40. https://doi.org/10.2307/1313439.
- 9. Puzey JR, Gerbode SJ, Hodges SA, Kramer EM, Mahadevan L. Evolution of spur-length diversity in Aquilegia petals is achieved solely through cell-shape anisotropy. Proc R Soc B: Biol Sci. 2012;279(1733):1640–5. https://doi.org/10.1098/rspb.2011.1873.
- 10. Gögler J, et al. Increased divergence in floral morphology strongly reduces gene flow in sympatric sexually deceptive orchids with the same pollinator. Evol Ecol. 2015;29(5):703–17. https://doi.org/10.1007/s10682-015-9779-2.
- 11. Xu S, Schlüter PM, Schiestl FP. Pollinator-driven speciation in sexually deceptive orchids. Int J Ecol. 2012;2012:1–9. https://doi.org/ 10.1155/2012/285081.
- 12. Adhikari B, Wallace LE. Does the Platanthera dilatata (Orchidaceae) complex contain cryptic species or continuously variable populations? Plant Syst Evol. 2014;300(6):1465–76. https://doi.org/10.1007/s00606-013-0974-8.
- 13. Boland J. The floral biology of Platanthera dilatata (Pursh) Lindl. (Orchidaceae). 1993.
- 14. Luer CA. The native orchids of the United States and Canada, excluding Florida. New York: The New York Botanical Garden; 1975.
- 15. Wallace LE, Bowles ML. Floral and genetic divergence across environmental gradients is moderated by inter-population gene flow in Platanthera dilatata (Orchidaceae). Front Ecol Evol. 2023;11:1085938. https://doi.org/10.3389/fevo.2023.1085938.
- 16. Wallace LE. Molecular evidence for allopolyploid speciation and recurrent origins in Platanthera huronensis (Orchidaceae). Int J Plant Sci. 2003;164(6):907–16. https://doi.org/10.1086/378658.
- 17. Sheviak CJ. Platanthera, Flora of North America North of Mexico, vol. 26. Oxford: Oxford University Press; 2002. p. 551–71.
- 18. van der Voort GE, Gilmore SR, Gorrell JC, Janes JK. Continuous video capture, and pollinia tracking, in Platanthera (Orchidaceae) reveal new insect visitors and potential pollinators. PeerJ. 2022;10: e13191. https://doi.org/10.7717/peerj.13191.
- 19. Pryor L, Williams E, Gunn B. A morphometric analysis of Eucalyptus urophylla and related taxa with descriptions of two new species. Aust Syst Bot. 1995;8(1):57–70. https://doi.org/10.1071/sb9950057.
- Bateman RM, James KE, Rudall PJ. Contrast in levels of morphological versus molecular divergence between closely related Eurasian species of Platanthera (Orchidaceae) suggests recent evolution with a strong allometric component. New J Bot. 2012;2(2):110–48. https://doi.org/10.1179/2042349712y.0000000013.
- 21. Bateman RM, Rudall PJ, Moura M. Systematic revision of Platanthera in the Azorean archipelago: not one but three species, including arguably Europe's rarest orchid. PeerJ. 2013;1: e218. https://doi.org/10.7717/peerj.218.
- 22. Esposito F, Vereecken NJ, Gammella M, Rinaldi R, Laurent P, Tyteca D. Characterization of sympatric Platanthera bifolia and Platanthera chlorantha (Orchidaceae) populations with intermediate plants. PeerJ. 2018;6: e4256. https://doi.org/10.7717/peerj.4256.
- 23. Schiestl FP, Schlüter PM. Floral Isolation, specialized pollination, and pollinator behavior in orchids. Entomology. 2009;54(1):425–46. https://doi.org/10.1146/annurev.ento.54.110807.090603.
- 24. Spittlehouse D. ClimateBC: your access to interpolated climate data for BC. Stream Watershed Manag Bull. 2006;99:16–21.



- 25. Daly C, et al. Physiographically sensitive mapping of climatological temperature and precipitation across the conterminous United States. Int J Clim. 2008;28(15):2031–64. https://doi.org/10.1002/joc.1688.
- 26. Shapiro SS, Wilk MB. An analysis of variance test for normality (complete samples). Biometrika. 1965;52(3–4):591–611. https://doi. org/10.1093/biomet/52.3-4.591.
- 27. A. Kassambara, *rstatix: pipe-friendly framework for basic statistical tests*. 2023. https://CRAN.R-project.org/package=rstatix. Accessed 18 January 2024.
- 28. Kruskal WH, Wallis WA. Use of Ranks in one-criterion variance analysis. J Am Stat Assoc. 1952;47:583–621.
- 29. Keselman HJ, Rogan JC. The Tukey multiple comparison test: 1953–1976. Psychol Bull. 1977;84(5):1050–6. https://doi.org/10.1037/0033-2909.84.5.1050.
- 30. Dunn OJ. Multiple comparisons using rank sums. Technometrics. 1964;6:241–52. https://doi.org/10.1080/00401706.1964.10490181.
- 31. Holm S. A simple sequentially rejective multiple test procedure. Scand J Stat. 1979;6:65–70.
- 32. Wickam H. ggplot2: elegant graphics for data analysis. New York: Springer-Verlag; 2016.
- 33. Bates D, Mächler M, Bolker B, Walker S. Fitting linear mixed-effects models using Ime4. J Stat Softw. 2015. https://doi.org/10.18637/jss.v067.i01.
- 34. Oksanen J, et al. Vegan: Community Ecology Package. R package version 2.0-10. 2013. Accessed 18 January 2024.
- 35. Murdoch D, Chow ED. Ellipse: Functions for drawing ellipses and ellipse-like confidence regions. 2022. https://CRAN.R-project.org/packa ge=ellipse. Accessed 18 January 2024.
- 36. ter Braak CJF. Canonical correspondence analysis: a new eigenvector technique for multivariate direct gradient analysis. Ecology. 1986;67(5):1167–79. https://doi.org/10.2307/1938672.
- 37. Egawa S, Hattori M, Itino T. Elevational floral size variation in *Prunella vulgaris*. Am J Plant Sci. 2015;06(13):2085–91. https://doi.org/ 10.4236/ajps.2015.613209.
- Pi H, Quan Q, Wu B, Lv X, Shen L, Huang S. Altitude-related shift of relative abundance from insect to sunbird pollination in Elaeagnus umbellata (Elaeagnaceae). J Syst Evol. 2021;59(6):1266–75. https://doi.org/10.1111/jse.12685.
- 39. Zhao Z-G, Wang Y-K. Selection by pollinators on floral traits in generalized Trollius ranunculoides (Ranunculaceae) along altitudinal gradients. PLoS ONE. 2015;10(2): e0118299. https://doi.org/10.1371/journal.pone.0118299.
- 40. Balfour DA, Linder HP. Morphological variation in populations of Disa uniflora (Diseae: Orchidaceae) in the southwestern Cape, South Africa". Can J Bot. 1990;68:2361–70. https://doi.org/10.1139/b90-301.
- 41. Neto LM, den Berg CV, Forzza RC. Linear and geometric morphometrics as tools to resolve species circumscription in the Pseudolaelia vellozicola complex (Orchidaceae, Laeliinae). Plant Ecol Evol. 2019;152(1):53–67. https://doi.org/10.5091/plecevo.2019.1531.
- 42. Hoffmann AA, Griffin PC, Macraild RD. Morphological variation and floral abnormalities in a trigger plant across a narrow altitudinal gradient. Austral Ecol. 2009;34(7):780–92. https://doi.org/10.1111/j.1442-9993.2009.01984.x.
- 43. Kiełtyk P. Patterns of floral allocation along an elevation gradient: variation in Senecio subalpinus growing in the Tatra Mountains. Alp Bot. 2021;131(1):117–24. https://doi.org/10.1007/s00035-021-00247-w.
- 44. Pyle RM. Butterflies of Cascadia. Washington: Seattle Audubon Society.
- 45. Inouye DW. The effect of proboscis and corolla tube lengths on patterns and rates of flower visitation by bumblebees. Oecologia. 1980;45(2):197–201. https://doi.org/10.1007/bf00346460.
- 46. Patt JM, Merchant MW, Williams DRE, Meeuse BJD. Pollination biology of Platanthera stricta (Orchidaceae) in Olympic National Park, Washington. Am J Bot. 1989;76(8):1097–106. https://doi.org/10.2307/2444821.
- 47. Herrera CM. Pollinator abundance, morphology, and flower visitation rate: analysis of the 'quantity' component in a plant-pollinator system. Oecologia. 1989;80(2):241–8. https://doi.org/10.1007/bf00380158.
- 48. Fernald CH. The Pterophoridae of North America. Boston: Wright and Potter Printing; 1898.
- 49. Borkowsky CL. Enhancing pollination of the endangered western prairie fringed orchid (Platanthera praeclara) by sphinx moths (Lepidoptera: Sphingidae) in tall grass prairie in southeastern Manitoba and an examination of orchid nectar production. Winnipeg: University of Manitoba; 2006.
- 50. van der Pijl L, Dodson CH. Orchid flowers: their pollination and evolution. Oxford: University of Miami Press; 1966.
- 51. Stpiczynska M. Floral longevity and nectar secretion of Platanthera chlorantha (Custer) Rchb. (Orchidaceae). Ann Bot. 2003;92(2):191– 7. https://doi.org/10.1093/aob/mcg124.
- 52. Nunes CEP, Maruyama PK, Azevedo-Silva M, Sazima M. Parasitoids turn herbivores into mutualists in a nursery system involving active pollination. Curr Biol. 2018;28(6):980-986.e3. https://doi.org/10.1016/j.cub.2018.02.013.
- 53. Light MHS, MacConaill M. In plain sight. Discovering insect herbivores of orchids. Eur J Environ Sci. 2014;11(2):13–9. https://doi.org/ 10.14712/23361964.2015.54.
- 54. Chapin FS, Matson PA, Vitousek PM. Principles of terrestrial ecosystem ecology. New York: Springer, New York; 2011. p. 259–96. https://doi.org/10.1007/978-1-4419-9504-9_9.
- 55. Inouye DW, Barr B, Armitage KB, Inouye BD. Climate change is affecting altitudinal migrants and hibernating species. Proc Natl Acad Sci. 2000;97(4):1630–3. https://doi.org/10.1073/pnas.97.4.1630.
- 56. Lee MA, Burger G, Green ER, Kooij PW. Relationships between resource availability and elevation vary between metrics creating gradients of nutritional complexity. Oecologia. 2021;195(1):213–23. https://doi.org/10.1007/s00442-020-04824-4.
- 57. Saavedra F, Inouye DW, Price MV, Harte J. Changes in flowering and abundance of Delphinium nuttallianum (Ranunculaceae) in response to a subalpine climate warming experiment. Global Change Biology. 2003;9:885–894. https://doi.org/10.1046/j.1365-2486. 2003.00635.x.
- Arft AM, Walker MD, Gurevitch J, Alatalo JM, Bret-Harte MS. Responses of tundra plants to experimental warming: meta-analysis of the international tundra experiment. Ecol Monogr. 1999;69:491–511. https://doi.org/10.1890/0012-9615(1999)069[0491:rotpte]2.0. co;2.
- 59. Hoover SER, Ladley JJ, Shchepetkina AA, Tisch M, Gieseg SP, Tylianakis JM. Warming, CO₂, and nitrogen deposition interactively affect a plant-pollinator mutualism. Ecol Lett. 2012;15(3):227–34. https://doi.org/10.1111/j.1461-0248.2011.01729.x.



- 60. Murcia C. Effect of floral morphology and temperature on pollen receipt and removal in ipomoea trichocarpa. Ecology. 1990;71(3):1098–109. https://doi.org/10.2307/1937378.
- 61. Khodorova NV, Boitel-Conti M. The role of temperature in the growth and flowering of geophytes. Plants. 2013;2(4):699–711. https://doi. org/10.3390/plants2040699.
- 62. Mack J-LK, Davis AR. The relationship between cell division and elongation during development of the nectar-yielding petal spur in Centranthus ruber (Valerianaceae). Ann Bot. 2015;115(4):641–9. https://doi.org/10.1093/aob/mcu261.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

