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### Data Article

# Gene expression profiling of *brakeless* mutant *Drosophila* embryos



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#### ABSTRACT

The transcriptional co-regulator Brakeless performs many important functions during *Drosophila* development, but few target genes have been identified. Here we use Affymetrix microarrays to identify Brakeless-regulated genes in 2–4 h old *Drosophila* embryos. Robust multi-array analysis (RMA) and statistical tests revealed 240 genes that changed their expression more than 1.5 fold. We find that up- and down-regulated genes fall into distinct gene ontology categories. In our associated study [2] we demonstrate that both up- and down-regulated genes can be direct Brakeless targets. Our results indicate that the co-repressor and co-activator activities of Brakeless may result in distinct biological responses. The microarray data complies with MIAME guidelines and is deposited in GEO under accession number GSE60048.

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#### Specifications Table [please fill in right-hand column of the table below]

Subject area

Biology Developmental Biology, Molecular Biology

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	http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi? token=olqdikeqjnutraf&acc=GSE60048
Data accessibility	The data is deposited in GEO under accession number GSE60048.
Data source location	
Experimental features	RNA was extracted from 2–4 h old embryos, converted to cDNA and hybri- dized to Affymetrix arrays
factors	Comparison of control embryos and embryos derived from brakeless germ- line clones
Data format	Analyzed with RMA and statistical tests
How data was acquired	Affymetrix Drosophila gene chip (version 2) array
Type of data	Table and figure
More specific sub- ject area	

#### Value of the data

- This data significantly extends the number of Brakeless-target genes
- The data shows that more genes are down-regulated than up-regulated in *brakeless* mutant embryos
- Down-regulated and up-regulated Brakeless target genes fall into distinct gene ontology clusters



**Fig. 1.** Brakeless-repressed and Brakeless-activated genes fall into distinct GO-categories. Grouping of genes into enrichment clusters revealed by DAVID functional annotation analysis. (A) Genes down-regulated in *bks* mutant embryos from the expression array and (B) up-regulated genes. The enrichment score depicts the geometric mean in log<sub>2</sub>-scale of the member's *P*-values within a cluster.

#### 1. Data

To identify Brakeless target genes in the early embryo, RNA was isolated from embryos derived from *brakeless* (*bks*<sup>278</sup>) germline clones, which lack the maternal contribution of Brakeless. This was compared to RNA from germline clone embryos generated with the unmutagenized FRT chromosome on which the *bks*<sup>278</sup> allele was induced [8]. The RNA was converted to cDNA and hybridized to an Affymetrix array. Misregulated genes that change their expression more than 1.5 fold were identified (supplementary material Table 1). We compared our gene list to an RNA-seq dataset that distinguishes maternal from zygotic transcripts in *Drosophila* embryos using polymorphisms [7]. The Brakeless-regulated genes were categorized as being maternally, zygotically, or maternally and zygotically (matzyg) derived (supplementary material Table 1). They were also subjected to functional annotation analysis using DAVID [3], which groups genes into clusters based on co-association with gene ontology (GO) terms (supplementary material Table 1). As shown in Fig. 1, up-regulated and down-regulated gene fall into distinct GO clusters.

#### 2. Experimental design, materials and methods

#### 2.1. Germline clones

The FLP–FRT dominant female sterile technique previously described in Ref. [1] was used to generate *brakeless* germline clones. The *bks*<sup>278</sup> allele was used, which has a 345 bp deletion that causes a frame shift at aa 741 resulting in addition of 79 novel amino acids [5]. It was outcrossed with a *w*<sup>1118</sup> strain to remove potential second-site mutations. *FRT*<sup>2R</sup>G13 *bks*<sup>278</sup>/CyO females were crossed with males of the genotype *hs*-*FLP*/Y; *FRT*<sup>2R</sup>G13 *ovo*<sup>D1</sup>/CyO, derived from *FRT*<sup>2R</sup>G13 *ovo*<sup>D1</sup>/T(1;2)OR64/CyO (Bloomington stock #4344). Off-spring larvae were heat-shocked for 3 h at 37 °C on days 3, 4 and 5 after egg-laying to induce expression of the FLP recombinase. *Cy* + females were crossed to *FRT*<sup>2R</sup>G13 *bks*<sup>278</sup>/CyO males and embryos were collected, dechorionated using bleach, and directly frozen in Trizol at  $-80^{\circ}$ C for RNA isolation. Corresponding crosses, embryo collection and RNA isolation was performed with an unmutagenized *FR1*<sup>2R</sup>G13 chromosome.

#### 2.2. Microarray analysis

Staged 2–4 h old embryos were collected and immediately frozen at  $-80^{\circ}$ C prior to RNA extraction. Total RNA was isolated using TRIzol (Invitrogen) and purified using an RNeasy kit (Qiagen) according to the manufacturer's protocols. Forty ul of embryos were used for each of three biological replicates of embryos derived from  $FRT^{2R}G13 \ bks^{278}$  or  $FRT^{2R}G13 \ c \ px \ sp$  control germline clones. cDNA probes were hybridized to an Affymetrix *Drosophila* gene chip (version 2). The intensity values were normalized and summarized with the robust multi-array analysis (RMA) method [6], using R (www. R-project.org) and the Bioconductor package [4].

#### 2.3. Statistical analysis

After RMA normalization, 640 probes passed a > 1.5 fold difference in median expression levels between the conditions investigated. They were subjected to a two tailed unpaired Student's *t*-test with a 95% confidence level, followed by Bonferroni's correction for multiple tests, resulting in a *P*-value cut-off at 0.000078125. After removal of probe-sets targeting duplicates and pseudogenes, 240 genes remained.

#### 2.4. GO analysis

The lists of genes with significantly changed expression levels containing 174 down- and 66 upregulated identifiers were used as input lists for the DAVID Functional Annotation Clustering tool [3]. The tool provides analysis of annotation content and gene ontology term enrichments, to highlight the most relevant GO terms associated with a gene list. The enrichment score is a geometric mean of the member's *P*-values in a -log scale within an annotation cluster.

#### Acknowledgments

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#### Appendix A. Supplementary material

Supplementary material associated with this article can be found in the online version at doi:10. 1016/j.dib.2015.08.033.

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