# A current update on the phytopharmacological aspects of *Houttuynia cordata* Thunb

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purposes traditionally.<sup>[8]</sup>

TAXONOMICAL CLASSIFICATION

Houttuynia Thunb.; Species: H. cordata.<sup>[9]</sup>

**BOTANICAL DESCRIPTION** 

# ABSTRACT

The present review is an attempt to put an insight into a medicinal plant *Houttuynia cordata* Thunb, which is indigenous to North-East India and China. It is an aromatic medicinal herb belonging to family *Saururaceae* and is restricted to specialized moist habitats. The review provides detailed information regarding the morphology, distribution, phytochemistry, ethnopharmacological uses and also describes various pharmacological activities reported on the plant *H. cordata*. The review describes therapeutic efficacy of the whole plant and its extracts, fractions and isolated compounds in different diseased condition. Among the important pharmacological activities reported includes, anti-mutagenic, anti-cancer, adjuvanticity, anti-obesity, hepatoprotective, anti-viral, anti-bacterial, anti-inflammatory, free radical scavenging, anti-microbial, anti-allergic, anti-leukemic, chronic sinusitis and nasal polyps activities. Thus, the present review will act as a source of referential information to researchers to perform clinical studies on isolated compounds that may serve the society and will help in improving human health care system.

Key words: Anti-viral, aristolactams, Houttuynia cordata, Saururaceae

# **INTRODUCTION**

In most of the developing countries, 70-95% of the population rely on traditional medicines for primary health-care and out of these 85% of people use plants or their extracts as the active substance.<sup>[1]</sup> The search for new biologically active compounds from plants usually depends on the specific ethnic and folk information obtained from local practitioners and is still regarded as an important source for drug discovery. In India, approximately 2000 drugs are of plant origin.<sup>[2]</sup> In view of the widespread interest on using medicinal plants, the present review on *Houttuynia cordata* Thunb. provides up-to-date information with reference to botanical, commercial, ethnopharmacological, phytochemical and pharmacological studies that appears in the literature. *H. cordata* Thunb. belongs to the family *Saururaceae* and is commonly known as Chinese lizard tail. It is a perennial herb

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with stoloniferous rhizome having two distinct chemotypes.<sup>[3,4]</sup> The Chinese chemotype of the species is found in wild and semi-wild conditions in the North-East of India from April

to September.<sup>[5-7]</sup> H. cordata is available in India, especially in

Brahmaputra valley of Assam and is utilized by various tribes of Assam in the form of vegetable as well as in various medicinal

Kingdom: Plantae; Phylum: Magnoliophyta; Class: Magnoliopsida;

Sub-class: Magnoliidae; Order: Piperales; Family: Saururaceae; Genus:

The plant H. cordata is an aromatic medicinal herb with creeping

root stock. It grows about 20-50 cm in height with leaves

measuring 4-8 cm in length, 3-6 cm in width and are broad,

ovate-cordate. Stipular sheath are 1-2.5 cm, 1/4-1/2 as long as petiole, usually ciliate, base enlarged and slightly clasping; petiole

usually purplish abaxial, base cordate, apex shortly acuminate; veins 5-7, basal or innermost pair arising ca. 5 mm above the base, if 7-veined, then outermost pair very slender or inconspicuous; reticulate veins  $\pm$  conspicuous. Inflorescences 1.5-2.5 cm long,

3-6 mm wide; peduncles 1.5-3 cm, subglabrous; involucral bracts oblong or obovate, 10-15 mm long, 5-7 mm wide, apex rounded. Flowers are naked with dense spikes, subtended by four white and petalloid bracts, involucres, non-petal, yellow inflorescences ca. 1-3 cm long, three stamens, flowering in June-July.<sup>[10]</sup> Rhizomes are creeping, thin while basal part of stems creeping rooted in whorls at nodes, apical part erect, glabrous or pubescent on nodes, sometimes purplish red.<sup>[11]</sup> The chromosomal number of *H. cordata* was reported as 2n = 96.<sup>[12]</sup>

# ETHNOMEDICAL USES

In the North-East region of India, whole plant of H. cordata is eaten raw as a medicinal salad for lowering the blood sager level and is commonly known by the name Jamyrdoh.<sup>[13]</sup> Moreover, leaf juice is taken for the treatment of cholera, dysentery, curing of blood deficiency and purification of blood.<sup>[14]</sup> Young shoots and leaves are eaten raw or cooked as a pot-herb. A decoction of this plant is used internally for the treatment of many ailments including cancer, coughs, dysentery, enteritis and fever. Externally, it is used for the treatment of snake bites and skin disorders. The leaves and stems are harvested during the growing season and are used as fresh decoctions. The leaf juice is also used as antidote and astringent.<sup>[15]</sup> The root, young shoots, leaves and sometimes the whole plant is traditionally used to cure various human ailments throughout South-East Asia. In Indo-China region, the entire plant is considered for its cooling, resolvent and emmenagogue properties. The leaves are recommended for the treatment of measles, dysentery and gonorrhea. The plant is also used in the treatment of eve troubles, skin diseases, hemorrhoids, relieving fever, resolving toxin, reducing swelling, draining pus, promoting urination and in certain diseases of women.[16]

## PHARMACOLOGY

*H. cordata* possess a number of medicinally important activities such as anti-leukemic,<sup>[17]</sup> anti-cancer,<sup>[18]</sup> adjuvanticity,<sup>[19]</sup> anti-oxidant<sup>[20]</sup> and inhibitory effects on anaphylactic reaction and mast cell activation.<sup>[21]</sup> Moreover, *H. cordata* has also been utilized for the treatment of herpes simplex virus type 1 (HSV-1), influenza virus, human immunodeficiency virus type 1,<sup>[22]</sup> and chronic sinusitis and nasal polyps.<sup>[21]</sup>

### Anaphylactic inhibitory activity

Oral administration of *H. cordata* water extract inhibited compound 48/80-induced systemic anaphylaxis in mice. Water extract of *H. cordata* at 100 mg; p.o. also inhibited the local allergic reaction, passive cutaneous anaphylaxis (PCA), activated by anti-dinitrophenyl (DNP) immunoglobulin E (IgE) antibody in rats. It also reduced the compound 48/80-induced mast cell degranulation and colchicine-induced deformation of rat peritoneal mast cells (RPMC). Moreover, *H. cordata* water extracts dose-dependently inhibited histamine release and calcium uptake of RPMC induced by compound 48/80 or anti-DNP IgE. The same extract also increased the level of intracellular cyclic adenosine monophosphate (cAMP) and inhibited significantly the compound 48/80-induced cAMP reduction in RPMC. These results suggest that water extracts of *H. cordata* may be beneficial in the treatment of mast cell-mediated anaphylactic responses.<sup>[21]</sup>

### Anti-mutagenic activity

Aqueous extracts of *H. cordata* has been reported to possess anti-mutagenic effect on benzo (a) pyrene, aflatoxin B1 and oxidized frying oil (OFO), which demonstrated a dose-dependent response using the Ames test in Sprague-Dawley rats, which were fed with a diet of 0, 2, or 5% *H. cordata* and 15% fresh oil or OFO for 28 days. After administration of OFO, it was observed that there was a significant decrease in polyphenol content in plasma which increased in the faces showing an apparent decrease in absorption of polyphenol. On treatment with *H. cordata*, the polyphenol content in plasma improved, which may be due to the presence of higher polyphenol concentration in the aqueous extracts of *H. cordata*.<sup>[23]</sup>

### Anti-inflammatory activity

Essential oil from *H. cordata* was reported to exhibit anti-inflammatory activity by a mechanism of action similar to that of non-steroidal anti-inflammatory drugs (NSAIDs). They inhibited the release of lipopolysaccharide (LPS)-induced prostaglandin  $E_2$  (PGE<sub>2</sub>) from mouse peritoneal macrophages (IC<sub>50</sub> value: 44.8 µg/mL). Moreover, the inhibitory activity of *H. cordata* essential oil elicited a dose-dependent inhibition of cyclooxygenase-2 (COX-2) enzyme activity (IC<sub>50</sub> value: 30.9 µg/mL). *H. cordata* essential oil was also found to elicit reduction in LPS-induced COX-2 messenger ribonucleic acid (mRNA) and protein expression, but did not affect COX-1 expression. NSAID and specific COX-2 inhibitor NS398 functioned similarly in LPS-induced mouse peritoneal macrophages.<sup>[24]</sup>

Water extracts of *H. cordata* exhibits anti-inflammatory activity on lipoteichoic acid (LTA)-induced inflammation in dermal fibroblasts by blocking the tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) pathway. *H. cordata* (20 µg/mL) suppressed the level of LTA-induced TNF- $\alpha$ , mRNA and LTA-induced COX-2 protein by up to 40% and 52% respectively. Moreover, TNF- $\alpha$ -induced COX-2 expression was also down-regulated by *H. cordata* treatment up to 65%.<sup>[25]</sup>

Ethanolic extract of whole plant of *H. cordata* (10  $\mu$ g/mL for 24 h) showed marked effect in treating mast cell-induced inflammatory diseases by a significant decrease in chemotactic index (63%) of human mast cells (HMC-1) in response to stem cell factor by inhibiting the nuclear factor-kappa B (NF- $\kappa$ B) activation.<sup>[26]</sup>

Anti-inflammatory activity of *H. cordata* injection (HCI), which constituted a mixture of essential oil from *H. cordata*, aqueous solution of sodium chloride solution and tween-80 was reported in carrageenan induced inflammation in the rat pleurisy model and by xylene in the mice ear edema model. The fluid volume,

protein concentration, C-reactive protein and cell infiltration were attenuated by HCI at all doses and touched bottom at a dose of 0.54 mL/100 g. This drug was also effective in inhibiting xylene induced ear edema and the percentage of inhibition came to 50% at a test dose of 80  $\mu$ L/20 g.<sup>[27]</sup> Another anti-inflammatory study on supercritical extract of *H. cordata* (HSE) in a carrageenan-air pouch model showed that HSE (200 mg/kg; p.o.) exerts anti-inflammatory effects by inhibiting both TNF- $\alpha$ -nitric oxide (NO) and COX-2-PGE, pathways.<sup>[28]</sup>

### **Anti-viral activity**

The optimal dosage of HCI showed direct inhibitory activity on cell infection by pseudorabies herpesvirus using Vero cells (a monkey kidney cell line) and swine testis cells as a model. While at high dosage *H. cordata* alone caused cell apoptosis.<sup>[29]</sup>

Aqueous extract of *H. cordata* showed immunomodulatory and anti-severe acute respiratory syndrome (SARS) activities. *H. cordata* also stimulated the proliferation of mouse splenic lymphocytes significantly and dose-dependently. By flow cytometry, it was revealed that *H. cordata* increased the proportion of CD4<sup>+</sup> and CD8<sup>+</sup> T cells. Moreover, it caused a significant increase in the secretion of interleukin (IL)-2 and (IL)-10 by mouse splenic lymphocytes. In the anti-viral aspect, *H. cordata* exhibited significant inhibitory effects on SARS coronavirus (SARS-CoV) 3C-like protease (3CL<sup>pro</sup>) and RNA-dependent RNA polymerase.<sup>[30]</sup>

A 4-5 dioxoaporphin namely norcepharadione B isolated from the n-hexane fraction of *H. cordata* was found to possess good inhibitory activity against the replication of HSV-1.<sup>[31]</sup>

In particular, quercetin 7-rhamnoside (Q7R), a flavonoid present in *H. cordata*, has been reported to elicit anti-viral activities against porcine epidemic diarrhea virus (PEDV), which is the predominant cause of severe enteropathogenic diarrhea. Q7R actively inhibited PEDV replication with a 50% inhibitory concentration (IC<sub>50</sub>: 0.014  $\mu$ g/mL). The 50% cytotoxicity concentration (CC<sub>50</sub>) of Q7R was over 100  $\mu$ g/mL and the derived therapeutic index was 7142. Therefore, Q7R could be considered to be a lead compound for development of anti-PEDV drugs, which may be used to arrest the early stage of PEDV replication.<sup>[32]</sup>

Quercetin 3-rhamnoside (Q3R) from *H. cordata* possessed strong inhibitory effects on influenza A/WS/33 virus by reducing the formation of a visible cytopathic effect. Moreover, Q3R also inhibited virus replication in the initial stage of virus infection by indirect interaction with virus particles.<sup>[33]</sup>

One of the finding claimed that aqueous extract of *H. cordata* possess anti-viral activity against dengue virus serotype 2 (DEN-2), strain 16681. *H cordata* (10-100 mg/mL) was found to exhibit significant reduction in intracellular DEN-2 RNA production correlating with the decrease in dengue protein expression after pre- and post-incubation with HepG2 cells. Moreover,

in the direct blocking mode, the extract bound with DEN-2 strongly inhibited the intracellular viral RNA replication with an effective dose (EC<sub>50</sub>) of 0.8 mg/mL. Concentrations as low as 10-40 mg/mL of *H. cordata* extract also exhibited protective effect on virion release from infected monkey kidney cell line (LLC-MK2) cells.<sup>[34]</sup> Hot water extracts of *H. cordata* blocked HSV-2 infection through inhibition of NF- $\kappa$ B activation by the presence of major water extractable flavonoids quercetin or isoquercitrin at 10  $\mu$ M.<sup>[35]</sup>

### Anti-obesity effect

Administration of an aqueous extract of leaves of *H. cordata* (1 g/kg; p.o.) significantly inhibited the corn oil-induced increase in plasma triglyceride levels in mice. It also inhibited the oleic acid and glycerol induced increase in the levels of plasma non-esterified fatty acids and glycerol, respectively. Moreover, an anti-obesity effect of *H. cordata* leaf extract has also been reported in mice with high-fat-diet-induced obesity.<sup>[36]</sup>

### **Anti-bacterial activities**

Water extract of H. cordata showed the anti-bacterial effects against murine salmonellosis. The anti-bacterial activity of H. cordata water extract was also examined in a Salmonella enterica serovar (Salmonella typhimurium) and was found to increase in a dose-dependent manner at concentrations from 25 to 100 mg/mL during 8-h incubation. Water extract of H. cordata also affected RAW 264.7 cells including morphologic changes and bacterial uptake, but there was no significant difference in bacterial replication in RAW 264.7 cells. With H. cordata water extract alone, NO production by RAW 264.7 cells did not increase, but when RAW 264.7 cells were infected by S. typhimurium, with or without extract, NO production with extract was 2-fold higher than that without extract. Treatment with aqueous extract of H. cordata did not affect inducible nitrous oxide synthase (iNOS) mRNA expression by RAW 264.7 cells, but when RAW 264.7 cells with extract were infected by S. typhimurium, iNOS mRNA expression was increased during 8-h incubation. Furthermore, water extract of H. cordata showed virulence reduction effects in S. typhimurium-infected BALB/c mice. After a lethal dose of S. typhimurium, the mortality rate in the extract untreated group was 100% at 7th day, but at the doses 25, 50 and 100  $\mu$ g/mL of extract groups were survived until 11, 17 and 23 days respectively. These data suggest that H. cordata water extract is stable and beneficial in the treatment of bacterial infection including intracellular replicating pathogens and may solve anti-microbial misuse and overuse.[37] Houttuynin (decanoyl acetaldehyde), a  $\beta$ -dicarbonyl compound, is reported as a major anti-bacterial constituent in the volatile oil of H. cordata.[38]

# Effects on xenobiotic-metabolizing enzyme system of rodents

One of the finding on *H. cordata* claimed that the OFO feeding produced a significant increase in phase I and II enzyme systems, including the content of CYP450 and microsomal protein. The oil was also reported to play a significant role in activities of nicotinamide adenine dinucleotide phosphate reductase, ethoxyresorufin O-deethylase (EROD), pentoxyresorufin O-dealkylase, aniline hydroxylase (ANH), aminopyrine demethylase (AMD) and quinone reductase (QR) (Phase-II enzyme) in Sprague-Dawley rats. In addition, the activities of EROD, ANH and AMD decreased and QR increased after feeding with *H. cordata* in OFO-fed group. Feeding with *H. cordata* diet also resulted in better regulation of the xenobiotic-metabolizing enzyme system.<sup>[39]</sup>

### **Anti-cancer activity**

It is reported that *H. cordata* induced apoptotic cell death in human primary colorectal cancer cells through a mitochondria-dependent signaling pathway. *H. cordata* at 250 µg/mL showed chromatin condensation in the treated cells. Moreover, *H. cordata* increased reactive oxygen species production and decreased the mitochondrial membrane potential ( $\Delta \Psi$  (m)) in examined cells. Mitochondria-dependent apoptotic signaling pathway was shown to be involved as determined by the increase in the levels of cytochrome c, Apaf-1 and caspase-3 and -9. The decrease in the level of  $\Delta \Psi$  (m) was associated with an increase in the BAX/BCL-2 ratio which led to activation of caspase-3 and -9.<sup>[40]</sup>

Six bioactive alkaloids, aristolactam B, piperolactam A, aristolactam A, norcepharadione B, cepharadione B and splendidine were isolated by bioactivity-guided fractionation of a methanolic extract of the aerial part of *H. cordata*. All the isolates exhibited moderate cytotoxicity against the five human cancer cell lines (A-549, SK-OV-3, SKMEL-2, XF-498 and HCT-15) examined *in vitro*. Among them, splendidine was found to exhibit significant activity against each cell line and aristolactam B exhibited selective activity against XF-498 (central nerve system cell) (ED<sub>50</sub>, 0.84 µg/mL).<sup>[18]</sup>

### **Anti-allergic activity**

Aqueous extracts of *H. cordata* (10 or 100 mg/kg; p.o.) show a significant effect on mast cell-mediated anaphylactic reaction, which is involved in many allergic diseases such as asthma and allergic rhinitis. *H. cordata* aqueous extract inhibited the compound 48/80-induced systemic anaphylaxis in mice. It also inhibited the local allergic reaction, PCA, activated by anti-DNP IgE antibody in rats. Compound 48/80-induced mast cell degranulation and colchicine-induced deformations of RPMC were also reduced by the *H. cordata* extract. Moreover, the extract dose-dependently inhibited histamine release and calcium uptake of RPMC induced by compound 48/80 or anti-DNP IgE. Aqueous extract of *H. cordata* increased the level of intracellular cAMP and inhibited significantly the compound 48/80-induced cAMP reduction in RPMC.<sup>[21]</sup>

Ethanolic extract of *H. cordata* showed beneficial therapeutic effects on the T helper 2-mediated or allergic skin disorders. Ethanolic extract inhibited the expression of IL-4 and (IL)-5 in response to phorbol 12-myristate 13-acetate (PMA) and calcium ionophore (CaI) in Jurkat T cells and the HMC-1 line, HMC-1. IL-4-5 and TNF- $\alpha$  (TNF- $\gamma$ )-induced thymus activation

regulated chemokine (TARC) production was also blocked by ethanolic extract of *H. cordata* in skin fibroblast CCD-986 sk cells. Stimulants included in PMA, phytohemagglutinin and CaI, increased the mRNA level of CC chemokine receptor 4 (CCR4), a receptor of TARC, in Jurkat T cells and the ethanolic extract weakly blocked the increased mRNA level. However, the stimulants and *H. cordata* ethanolic extract had no effect on the CCR4 protein level. The ethanolic extract also inhibited the TARC-induced migration, as well as basal migration of Jurkat T cells.<sup>[41]</sup> It has been also reported that water extract of *H. cordata* suppressed anaphylactic reaction and IgE-mediated allergic response through inhibition of cytokines and multiple events of FcERI-dependent signaling cascades in mast cells.<sup>[42]</sup>

### **Anti-diabetic activity**

*H. cordata* water extract was also reported to exhibit significant decrease in the urinary protein, urinary albumin, monocyte chemo-attractant protein expression level and renal connective tissue growth factor (CTGF). It also showed a significant improvement in insulin resistance after giving 8 weeks of treatment to streptozotocine (STZ) induced type II diabetic mellitus rats.<sup>[43,44]</sup>

A recent study has shown that the volatile oil from *H. cordata* restored the alterations in blood glucose, insulin, adiponectin and CTGF levels in diabetic rats, induced by the combination of a high-carbohydrate and high-fat diet and STZ injection, which may be attributed to the reduced insulin resistance, adiponectin and CTGF levels.<sup>[45]</sup>

### **Anti-oxidant activity**

Methanolic extract of *H. cordata* in an *in vitro* model has shown to possess free radical scavenging activity using 2, 2-diphenyl-1-picrylhydrazyl (DPPH), ferric reducing anti-oxidant power and trolox equivalent anti-oxidant capacity assays.<sup>[46]</sup> Anti-oxidant activity of *H. cordata* is mainly attributed to the presence of chlorogenic acids and its derivatives, catechin and procyanidin B, which were also characterized using on-line liquid chromatography-electrospray ionization mass spectrometer coupled with DPPH assay.<sup>[47]</sup>

One of the finding showed anti-oxidant activity of fermentation product of five indigenous plants *Phyllanthus emblica* Linn, *Morinda citrifolia* Linn, *H. cordata* Thunb. *Terminalia chebula* Retz and *Kaempferia parviflora* Wall. On oxidative stress in Wistar rats with STZ-induced type II diabetes. Fermentation product at the dose level 2 and 6 mL/kg body weight/day for 6 weeks significantly (P < 0.05) decreased the diabetes-associated oxidative stress to a large extent through the inhibition of lipid peroxidation.<sup>[48]</sup>

### **Dietary effects**

A recent investigation has reported that methanolic extract of *H. cordata* powder (1 g/kg p.o.) increased growth performance, dry matter, nitrogen digestibility, white blood cell concentration, meat longissimus muscle area and thiobarbituric acid reactive substances value in finishing pigs.<sup>[49]</sup>

# **PHYTOCHEMISTRY**

Phytochemical investigations on plant *H. cordata* up to 2012 have reported number of phytoconstituents present in the plant. Various types of chemical constituents such as aristolactams, 5,4-dioxoaporphines, oxoaporphines, amides, indoles, ionones, flavonoids, benzenoids, steroids and different volatile oils have been isolated from *H. cordata*. Houttuynoside  $A^{[31]}$  and houttuynamide  $A^{[31]}$  have also been isolated from this plant. Among the isolated compounds, some have been evaluated for their anti-oxidant and anti-tyrosinase activity. Cepharadione B showed strong inhibitory activity against tyrosinase with an  $IC_{50}$  value of 170 mM. Quercitrin, quercetin-3-*O*- $\beta$ -D-galactop yranoside showed excellent DPPH radical-scavenging property with  $IC_{50}$  values of 31 and 63 mM, respectively.<sup>[31]</sup> Table 1 and Figure 1 demonstrate detailed information regarding the phytoconstituents with their specific classes isolated from *H. cordata*.

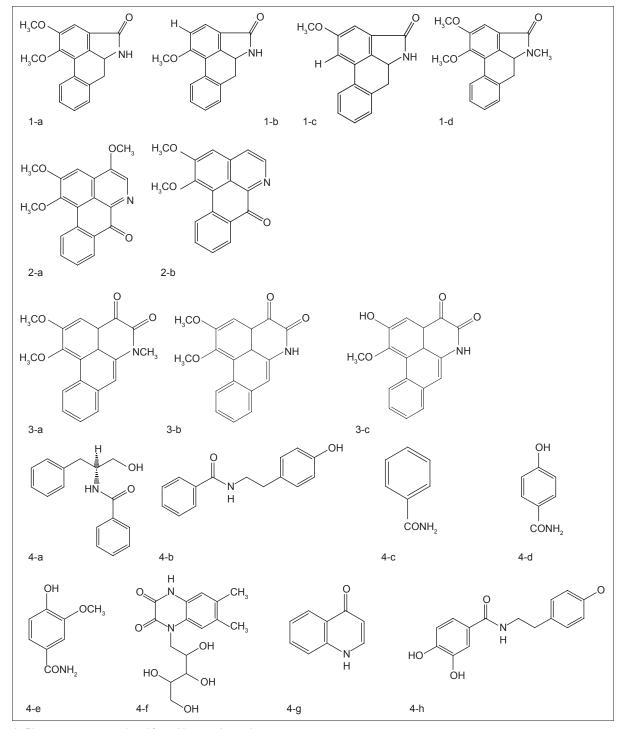


Figure 1: Phytoconstituents isolated from Houttuynia cordata

# CONCLUSION

According to Florae Reipublicae Popularis Sinicae and Flora

Sichuanica, *H. cordata* Thunb. (Yuxingcao in Chinese) is the only species in the genus *Houttuynia*.<sup>[87,88]</sup> Its medicinal importance is well-described in Chinese system of medicine in fever, to

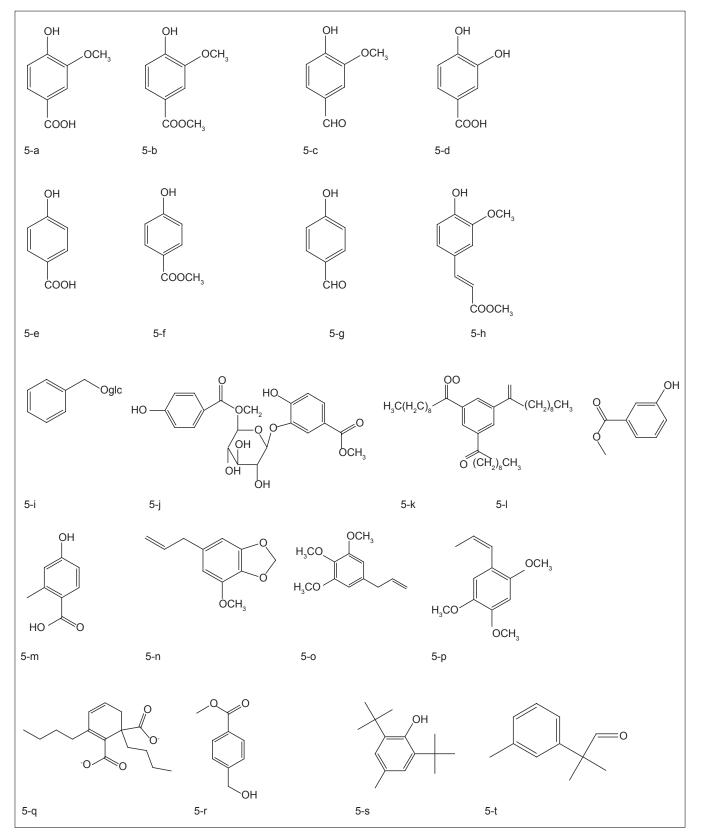


Figure 1: Phytoconstituents isolated from Houttuynia cordata

ease malnutrition, clearing of body toxins, anti-bacterial and in treatment of lung carbuncles. In addition, the plant is widely used as vegetable in North-Eastern parts of India and China and has been identified as one of the most potential medical and edible plant genetic resources by the Chinese State Health Department. Thus, the information provided in the present review may act as

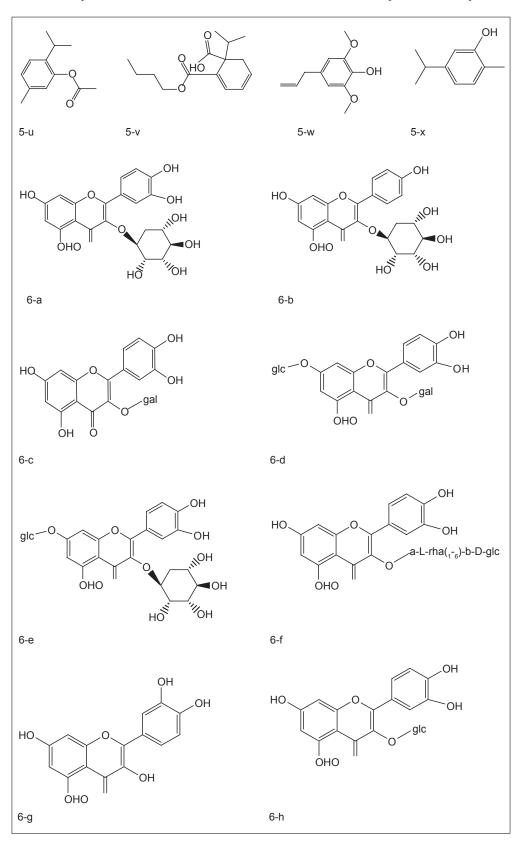


Figure 1: Phytoconstituents isolated from Houttuynia cordata

a contributing factor to the fact that at least 25% of all modern medicines are derived, either directly or indirectly, from medicinal

plants, primarily through the application of modern technology to traditional knowledge.<sup>[1]</sup>

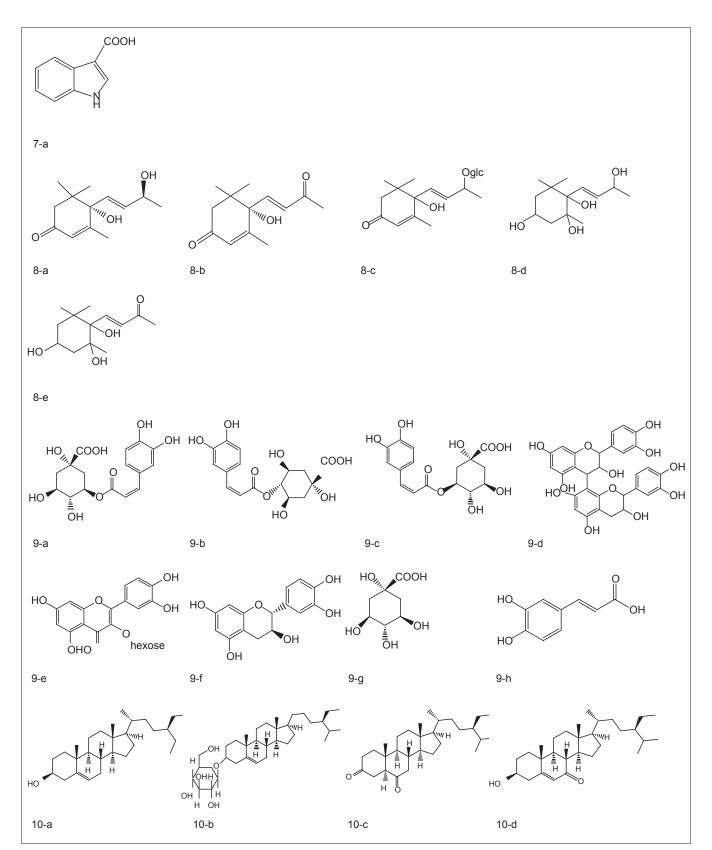


Figure 1: Phytoconstituents isolated from Houttuynia cordata

*H. cordata* offers an overall greater therapeutic value. The plethora of activities reported for the extracts, fractions and compounds isolated from *H. cordata* provide promising evidence

for future research, which could achieve an important place in the world of modern drugs. Isolation on a large scale, chemical transformations and synthesis of the active compounds will

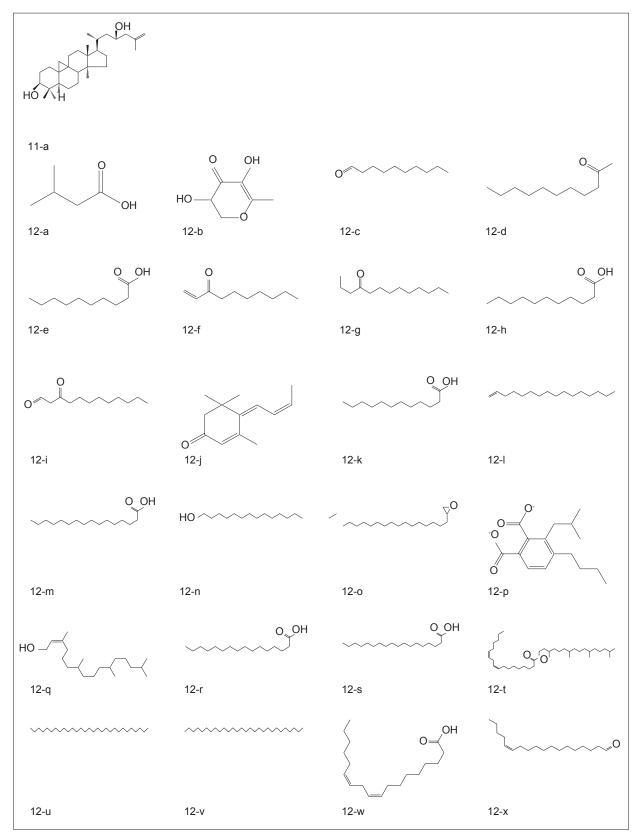
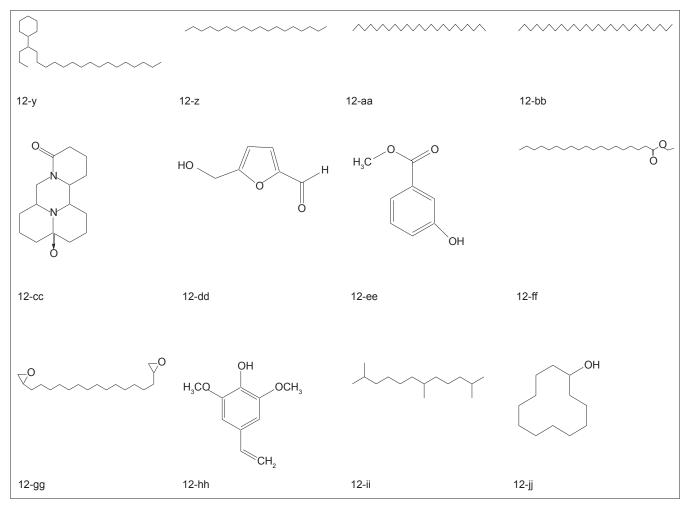


Figure 1: Phytoconstituents isolated from Houttuynia cordata

definitely enhance their pharmacological value. The provided information will also help in developing pharmacophores of many isolates that have not yet been identified and will also help in performing clinical trials using various active compounds against a variety of diseased conditions. Thus, the present review will elaborate the significance of *H. cordata* in human health-care system and will promote natural product research to its optimum height.



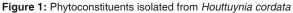


Table 1: List of phytoconstituents isolated from
H. cordata till year 2012

Si No.	Phytoconstituents	Part of plant used for isolation	Reference
1	Aristolactams		
	a. aristolactam B II (Cepharanone B)	Dried aerial parts	a. <sup>[18,50]</sup>
	b. Aristolactam A II	Dried aerial parts	b. <sup>[18,50]</sup>
	c. Piperolactam A	Dried aerial parts	C. <sup>[18,50]</sup>
	d. Caldensin	Whole plant	d. <sup>[31,51]</sup>
2	Oxoaporphines		
	a. Splendidine	Dried aerial parts	a. <sup>[22]</sup>
	b. Lysicamine/oxonuciferine	Whole plant	b. <sup>[52]</sup>

t of plant used for isolation Dioxoaporphines Cepharadione B Norcepharadione B	Part of plant used for isolation Dried aerial parts	a. [18,50]
Cepharadione B	2.1.00 0.01101	
	2.1.00 0.01101	
Norcepharadione B		I. [19.52]
		b. <sup>[18,53]</sup>
Noraritolodione		C. <sup>[22,53]</sup>
ides		
<i>I</i> -(1-hydroxymethyl-2- nylethyl) benzamide	Whole plant	a. <sup>[54]</sup>
<i>N</i> -(4-hydroxyphenylethyl)		b. <sup>[55]</sup>
Phenyl carboxamide		C. <sup>[56]</sup>
4-Hydroxybenzamide		d. <sup>[57]</sup>
4-Hydroxy-3-		e. <sup>[58]</sup>
	enzamide Phenyl carboxamide 4-Hydroxybenzamide	enzamide Phenyl carboxamide 4-Hydroxybenzamide

# Table 1: Contd.

No.	Part of plant used for isolation	Part of plant used for isolation	Reference
	methoxybenzamide		
	f. 6,7-Dimethyl-1-ribitol-1-yl-1, 4-dihydroquinoxaline-2,3-dione		f. <sup>[59]</sup>
	g. (1 <i>H</i> )-quinolinone		g. <sup>[60]</sup>
	h. Houttuynamide A		h. <sup>[31]</sup>
5	Benzenoids		a. <sup>[31,61]</sup>
	a. Vanillic acid b. Methyl vanillate	Whole plant	a. <sup>[31,62]</sup>
	c. Vanillin		D. <sup>[31,62]</sup>
	d. Protocatechuic acid		d. <sup>[31,63]</sup>
	e. 4-Hydroxybenzoic acid		e. <sup>[64]</sup>
	f. Methylparaben		f. <sup>[65]</sup>
	g. <i>p</i> -Hydroxybenzaldehyde		g. <sup>[67]</sup>
	h. Cis- and trans-methyl ferulate		h. <sup>[31,68]</sup>
	i. Benzyl-β-D-glucopyranoside		i. <sup>[31,69]</sup>
	j. Houttuynoside A		j. <sup>[31]</sup>
	k. 1,3,5,-Tridecanoylbenzene I. Methyl-3-hydroxybenzoate		k. <sup>[66]</sup>
	m. Methylaparaben		m. <sup>[66]</sup>
	n. Myristicin		n. <sup>[71]</sup>
	o. Elemicine		O. <sup>[71]</sup>
	p. α-Asarone		p. <sup>[71]</sup>
	q. 1,3-Dibutylphthalate		q. <sup>[70,72]</sup>
	r. Methyl-4-		r. <sup>[72,73]</sup>
	hydroxymethylbenzoate		
	s. 2,6-Bis-(1,1-dimethyl)-		S. <sup>[72]</sup>
	4-methylphenol		. [70 70]
	t. m-Tolyl-dimethylacetaldehyde		t. <sup>[72,73]</sup> u. <sup>[72]</sup>
	u. Thymylacetate v. Butyl-2-isopropyl phthalate		u. <sup>[70,73]</sup>
	w. 2,6-Dimethoxy-4-(2-propenyl)		W. <sup>[70]</sup>
	phenol		
	x. 2-Methyl-5-(1-methylethyl)		X. <sup>[72]</sup>
	phenol		
6	Flavonoids		
	a. Quercitrin	Dried aerial	a. <sup>[18,74]</sup>
		parts	. (76 70)
	b. Afzelin		b. <sup>[75,76]</sup>
	c. Hyperin		c. <sup>[76]</sup> d. <sup>[75,76]</sup>
	d. Quercetin-3-O-β-D-galacto pyranosyl-7-O-β-D-glactopyra		d. [ <sup>13,10]</sup>
	noside		(70)
	e. Quercetin-3-O- $\alpha$ -L-rhamno pyranosyl-7-O- $\beta$ -D-glucopyran		e. <sup>[76]</sup>
	oside		£ [77 74]
	f. Rutin		f. <sup>[77,74]</sup> g. <sup>[78,31]</sup>
	g. Quercetin		g. [79] h. <sup>[79]</sup>
7	h. Isoquercitrin Indoles		11. 19
1	a. Indole-3-carboxylic acid	Whole plant	a. <sup>[31,80]</sup>
8	lonones	Trifolo plant	u.
-*	a. Vomifoliol	Whole plant	a. <sup>[66]</sup>
	b. Dehydrovomifoliol	·	b. <sup>[31,81]</sup>
	c. Reseoside		C. <sup>[31,82]</sup>
	d. 6-(9-Hydroxy-but-7-ethyl)-1,		d. <sup>[31,83]</sup>
	1,5-trimethylcyclohexane-		
	3,5,6-triol		104.042
			- (31.84)
	e. 7-(3,5,6-Trihydroxy-2,6,		e. <sup>[31,84]</sup>
	e. 7-(3,5,6-Trihydroxy-2,6, 6-trimethylcyclohexyl)-		e. <sup>[01,04]</sup>
۵	e. 7-(3,5,6-Trihydroxy-2,6, 6-trimethylcyclohexyl)- byt-3-en-2-one		e. [01,04]
9	e. 7-(3,5,6-Trihydroxy-2,6, 6-trimethylcyclohexyl)-	Dried aerial	[47]

# Table 1: Contd...

Tak	Table 1: Contd				
Si No.	Part of plant used for isolation	Part of plant used for isolation	Reference		
	part b. Crypto-chlorogenic acid c. Neo-chlorogenica acid d. Procynanidin B e. Catechin f. Quinic acid g. Caffeic acid				
10	Steroids a. β-Sitosterol b. β-Sitosteryl glucoside c. 5-α-Stigmastane-3,6-dione d. 3-Hydroxy-β-sitost-5-en-7-one	Rhizome	a. <sup>[61,77]</sup> b. <sup>[85]</sup> c. <sup>[66]</sup> d. <sup>[66]</sup>		
11	Triterpenoids a) Cycloart-25-ene-3b, 24-diol	Whole plant	a. <sup>[31,86]</sup>		
12	Volatile oil a. 3-Methyl-butanoic acid b. 2,3-Dihydro-3,5-dihydroxy-6 methyl- 4 <i>H</i> -pyran-4-one c. Decanal d. 2-Undecanone e. <i>n</i> -Decanoic acid f. 1-Decen-3-one g. 4-Tridecanone h. Undecanoic acid	Whole plant	[70]		
	<ul> <li>i. Decanoyl acetaldehyde</li> <li>j. Megastigmatrienone</li> <li>k. Dodecanoic acid</li> <li>l. 1-Hexadecene</li> <li>m. <i>n</i>-Hexadecanoic acid</li> <li>n. Pentadecanol</li> <li>o. Hexadecyl-oxirane</li> <li>p. Butyl-2-methylpropylphthalate</li> </ul>				
	<ul> <li>q. 3,7,11,15-Tetramethyl-</li> <li>2-hexadecen-1-ol</li> <li>r. Palmitic acid</li> <li>s. Octadecanoic acid</li> <li>t. Phytol, (<i>Z</i>, <i>Z</i>) 9,12-</li> <li>octadecadienoic acid</li> <li>u. Nonacosane</li> <li>v. Octacosane</li> </ul>				
	<ul> <li>w. Linoleic acid</li> <li>x. (Z)-13-octadecenal</li> <li>y. (1-Propylheptadecyl)</li> <li>cyclohexane</li> <li>z. Octadecane</li> <li>aa. Tricosane</li> </ul>				
	bb. Hexacosane cc. <i>N</i> -oxide matrine <i>dd</i> . 5-(Hydroxymethyl)-2- furancarboxalder ee. 3-Hydroxy-benzoate methyl				
	ff. Nonadecanoate ethyl gg. Tettadecyl oxirane hh. 2,6-Dimethhoxy-4- (2-propenyl)-phenol ii. 2,6,11-Timethyl-dodecane jj. Cyxlododecanol				

## REFERENCES

- World Health Organization. The World Medicines Situation: WHO/EMP/MIE/2011.2.3. Geneva: World Health Organization; 2011. Available from: http://digicollection.org/hss/documents/ s18063en/s18063en.pdf.
- Bhattacharyya N, Sarma S. Assessment of availability, ecological feature, and habitat preference of the medicinal herb *Houttuynia cordata* Thunb. in the Brahmaputra Valley of Assam, India. Environ Monit Assess 2010;160:277-87.
- Tutupalli LV, Chaubal MG. Saururaceae V. Composition of essential oil from foliage of *Houttuynia cordata* and chemo systematics of *Saururaceae*. Lloydia 1975;38:92-6.
- Brown D. Encyclopedia of Herbs and Their Uses. London: Dorling Kindersley; 1995.
- 5. Kanjilal PC, Dev RN. Flora of Assam. Vol. III. New Delhi: Omsons Publishers; 1937. p. 113.
- 6. Dev DB. The Flora of Tripura. Vol. II. New Delhi: Today and Tomorrow's Publisher; 1983. p. 139-40.
- Mukherjee A, Roy SD. An account of *Piperaceae* and *Saururaceae* in the hills of Darjeeling district, West Bengal (India). J Econ Tax Bot 1987;9:367-72.
- Chakraborti S, Sinha S, Sinha RK. High-frequency induction of multiple shoots and clonal propagation from rhizomatous nodal segments of *Houttuynia Cordata* Thunb. An ethnomedicinal herb of India. *In Vitro* Cell Dev Biol Plant 2006;42:394-8.
- Watson L, Dallwitz MJ. The families of flowering plants: description, illustrations, identification and information retrieval, 1992. Available from: http://biodiversity.uno.edu/delta/, 14<sup>th</sup> December 2000.
- Bora C. Ethnobotany of lower Subansiri District (Nishi Tribe) of Arunachal Pradesh, Ph.D. Thesis. Gauhati University, Assam, India, 2001.
- Meng SW, Chen ZD, Li DZ, Liang H ×. Phylogeny of Saururaceae based on mitochondrial matR gene sequence data. J Plant Res 2002;115:71-6.
- 12. Okada H. Karyomorphology and relationships in some genera of *Saururaceae* and *Piperaceae*. Bot Mag 1986;99:289-99.
- Friht.Org.in. Medicinal Plants Conservation and Sustainable Utilisation-Meghalaya, India. Annexure-C. 72-5, 2003. Available from: http://friht.org.in/html/reports/meghalayaslpc.pdf.
- Hynniewta SR, Kumar Y. Herbal remedies among the Khasi traditional healers and village folks in Meghalaya. Indian J Tradit Knowl 2008;7:581-6.
- Tapan S. Determination of nutritive value, mineral contents and anti-oxidant activity of some wild edible plants from Meghalaya state, India. Asian J Appl Sci 2011;4:238-46.
- Lu HM, Liang YZ, Yi LZ, Wu XJ. Anti-inflammatory effect of Houttuynia cordata injection. J Ethnopharmacol 2006;104:245-9.
- Chang JS, Chiang LC, Chen CC, Liu LT, Wang KC, Lin CC. Antileukemic activity of *Bidens pilosa* L. var. minor (Blume) Sherff and *Houttuynia cordata* Thunb. Am J Chin Med 2001;29:303-12.
- Kim SK, Ryu SY, No J, Choi SU, Kim YS. Cytotoxic alkaloids from *Houttuynia cordata*. Arch Pharm Res 2001;24:518-21.
- Wang D, Yu Q, Eikstadt P, Hammond D, Feng Y, Chen N. Studies on adjuvanticity of sodium houttuyfonate and its mechanism. Int Immunopharmacol 2002;2:1411-8.
- Cho EJ, Yokozawa T, Rhyu DY, Kim SC, Shibahara N, Park JC. Study on the inhibitory effects of Korean medicinal plants and their main compounds on the 1,1-diphenyl-2-picrylhydrazyl radical. Phytomedicine 2003;10:544-51.
- 21. Li GZ, Chai OH, Lee MS, Han EH, Kim HT, Song CH. Inhibitory effects of *Houttuynia cordata* water extracts on

anaphylactic reaction and mast cell activation. Biol Pharm Bull 2005;28:1864-8.

- 22. Hayashi K, Kamiya M, Hayashi T. Virucidal effects of the steam distillate from *Houttuynia cordata* and its components on HSV-1, influenza virus, and HIV. Planta Med 1995;61:237-41.
- Chen YY, Liu JF, Chen CM, Chao PY, Chang TJ. A study of the antioxidative and antimutagenic effects of *Houttuynia cordata* Thunb. using an oxidized frying oil-fed model. J Nutr Sci Vitaminol (Tokyo) 2003;49:327-33.
- 24. Li W, Zhou P, Zhang Y, He L. *Houttuynia cordata*, a novel and selective COX-2 inhibitor with anti-inflammatory activity. J Ethnopharmacol 2011;133:922-7.
- Jee YC, Jung AL, Jee BL, Sook JY, Seung CL. Anti-inflammatory activity of *Houttuynia cordata* against lipoteichoic acid-induced inflammation in human dermal fibroblasts. Chonnam Med J 2010;46:140-7.
- Kim IS, Kim JH, Kim JS, Yun CY, Kim DH, Lee JS. The inhibitory effect of *Houttuynia cordata* extract on stem cell factor-induced HMC-1 cell migration. J Ethnopharmacol 2007;112:90-5.
- Lu HM, Liang YZ, Wu XJ, Qiu P. Tentative fingerprint-efficacy study of *Houttuynia cordata* injection in quality control of traditional Chinese medicine. Chem Pharm Bull (Tokyo) 2006;54:725-30.
- Shin S, Joo SS, Jeon JH, Park D, Jang MJ, Kim TO, et al. Anti-inflammatory effects of a *Houttuynia cordata* supercritical extract. J Vet Sci 2010;11:273-5.
- Ren X, Sui X, Yin J. The effect of *Houttuynia cordata* injection on pseudorabies herpesvirus (PrV) infection *in vitro*. Pharm Biol 2011;49:161-6.
- Lau KM, Lee KM, Koon CM, Cheung CS, Lau CP, Ho HM, et al. Immunomodulatory and anti-SARS activities of *Houttuynia* cordata. J Ethnopharmacol 2008;118:79-85.
- Chou SC, Su CR, Ku YC, Wu TS. The constituents and their bioactivities of *Houttuynia cordata*. Chem Pharm Bull (Tokyo) 2009;57:1227-30.
- Choi HJ, Kim JH, Lee CH, Ahn YJ, Song JH, Baek SH, et al. Antiviral activity of quercetin 7-rhamnoside against porcine epidemic diarrhea virus. Antiviral Res 2009;81:77-81.
- Choi HJ, Song JH, Park KS, Kwon DH. Inhibitory effects of quercetin 3-rhamnoside on influenza A virus replication. Eur J Pharm Sci 2009;37:329-33.
- Leardkamolkarn V, Sirigulpanit W, Phurimsak C, Kumkate S, Himakoun L, Sripanidkulchai B. The inhibitory actions of *Houttuynia cordata* aqueous extract on dengue virus and dengue-infected cell. J Food Biochem 2012;36:86-92.
- Chen X, Wang Z, Yang Z, Wang J, Xu Y, Tan RX, *et al. Houttuynia cordata* blocks HSV infection through inhibition of NF-κB activation. Antiviral Res 2011;92:341-5.
- Miyata M, Koyama T, Yazawa K. Water extract of *Houttuynia* cordata Thunb. leaves exerts anti-obesity effects by inhibiting fatty acid and glycerol absorption. J Nutr Sci Vitaminol (Tokyo) 2010;56:150-6.
- Kim GS, Kim DH, Lim JJ, Lee JJ, Han DY, Lee WM, *et al.* Biological and antibacterial activities of the natural herb *Houttuynia cordata* water extract against the intracellular bacterial pathogen *Salmonella* within the RAW 264.7 macrophage. Biol Pharm Bull 2008;31:2012-7.
- Duan X, Zhong D, Chen X. Derivatization of beta-dicarbonyl compound with 2, 4-dinitrophenylhydrazine to enhance mass spectrometric detection: Application in quantitative analysis of houttuynin in human plasma. J Mass Spectrom 2008;43:814-24.
- Chen YY, Chen CM, Chao PY, Chang TJ, Liu JF. Effects of frying oil and *Houttuynia cordata* thumb. on xenobiotic-metabolizing enzyme system of rodents. World J Gastroenterol 2005;11:389-92.

- Lai KC, Chiu YJ, Tang YJ, Lin KL, Chiang JH, Jiang YL, et al. Houttuynia cordata Thunb. extract inhibits cell growth and induces apoptosis in human primary colorectal cancer cells. Anticancer Res 2010;30:3549-56.
- 41. Lee JS, Kim IS, Kim JH, Kim JS, Kim DH, Yun CY. Suppressive effects of *Houttuynia cordata* Thunb. (*Saururaceae*) extract on Th2 immune response. J Ethnopharmacol 2008;117:34-40.
- 42. Han EH, Park JH, Kim JY, Jeong HG. *Houttuynia cordata* water extract suppresses anaphylactic reaction and IgE-mediated allergic response by inhibiting multiple steps of FcepsilonRI signaling in mast cells. Food Chem Toxicol 2009;47:1659-66.
- Wang HY, Lu M, Xiu YF. *Houttuynia cordata* modulates connective tissue growth factor and insulin resistance in rats with diabetes mellitus. Chin J New Drug 2009;16:1540-1544.
- 44. Liu Y, Wang H. Mechanism of herba *Houttuyniae* on relieving renal impairment in streptozotocin-induced diabetic rats. Tradit Chin Drug Res Clin Pharmacol 2010;02:107-110.
- 45. Wang HY, Bao JL. Effect of *Houttuynia cordata* aetherolea on adiponectin and connective tissue growth factor in a rat model of diabetes mellitus. J Tradit Chin Med 2012;32:58-62.
- Li HB, Wonga CC, Chenga KW, Chena F. Antioxidant properties in vitro and total phenolic contents in methanol extracts from medicinal plants. LWT 2008;41:385-90.
- Nuengchamnong N, Krittasilp K, Ingkaninan K. Rapid screening and identification of antioxidants in aqueous extracts of *Houttuynia cordata* using LC-ESI-MS coupled with DPPH assay. Food Chem 2009;117:750-6.
- Chaiyasut C, Kusirisin W, Lailerd N, Lerttrakarnnon P, Suttajit M, Srichairatanakool S. Effects of phenolic compounds of fermented Thai indigenous plants on oxidative stress in streptozotocin-induced diabetic rats. Evid Based Complement Alternat Med 2011;2011:749307.
- 49. Yan L, Meng QW, Kim IH. The effects of dietary *Houttuynia cordata* and *Taraxacum officinale* extract powder on growth performance, nutrient digestibility, blood characteristics and meat quality in finishing pigs. Livest Sci 2011;141:188-93.
- Prôbstle A, Bauer R. Aristolactams and a 4,5-dioxoaporphine derivative from *Houttuynia cordata*. Planta Med 1992;58:568-9.
- Achari B, Bandyopadhyay S, Chakravarty AK, Pakrashi SC. Carbon-13 NMR spectra of some phenanthrene derivatives from *Aristolochia indica* and their analogues. Org Magn Reson 1984;22:741-6.
- 52. Chen CY, Chang FR, Wu YC. The constituents from the stems of *Annona cherimola*. J Chin Chem Soc 1997;44:313-9.
- 53. Jong TT, Jean MY. Alkaloids from *Houttuynia cordata*. J Chin Chem Soc 1993;40:301-3.
- 54. Bate RB, Janda KD. A convenient synthesis of  $\alpha$ -acyl amino alcohols from azlactones. Synthesis 1984;4:310-1.
- Ghosh P, Ghosh MK, Thakur S, Dan J, Akihisa T, Tamura T, et al. Dihydroxy acidissiminol and acidissiminol epoxide, two tyramine derivatives from *Limonia acidissima*. Phytochemistry 1994;37:757-60.
- Lampert H, Mikenda W, Karpfen A, Kahlig H. NMR shieldings in benzoyl and 2-hydroxy-benzoyl compounds. Experimental versus GIAO calculated data. J Phys Chem 1997;A101:9610-7.
- Pouchert CJ, Behnke J. *The Aldrich Library of* <sup>13</sup>C and <sup>1</sup>H-FTNMR Spectra, Aldrich Chemical Company, America. Vol. II: 1394C (1993).
- Kergomard A, Renard MF. Action of two strains of *Streptomyces* on aromatic substrates. Agric Biol Chem 1986;50:2913-4.
- Miles HT, Smyrniotis PZ, Stadtman ER. Bacterial degradation products of riboflavin. III. Isolation, structure determination and biological transformations of 1-ribityl-2,3-diketo-1,2,3,4-tetrahyd ro-6,7-dimethyl-quinoxaline. J Am Chem Soc 1959;81:1946-9.

- Zalibera L, Milata V, Llavsky D. <sup>1</sup>H and <sup>13</sup>C NMR spectra of 3-substituted 4-quinolones. Magn Reson Chem 1998;36:681-4.
- 61. Lee CK, Lu CK, Kuo YH, Chen JZ, Sun GZ. The new prenylated flavones from the roots of *Ficus beecheyana*. J Chin Chem Soc 2004;51:437-42.
- Wilson SC, Howard PW, Forrow SM, Hartley JA, Adams LJ, Jenkins TC, *et al.* Design, synthesis, and evaluation of a novel sequence-selective epoxide-containing DNA cross-linking agent based on the pyrrolo[2, 1-c] [1,4]benzodiazepine system. J Med Chem 1999;42:4028-41.
- Zhang HL, Nagatsu A, Okuyama H, Mizukami H, Sakakibara J. Sesquiterpene glycosides from cotton cake oil. Phytochemistry 1998;48:665-8.
- Wen LL, Chang FR, Hsieh TJ, Wu YC. The constituents of Euchresta formosana. J Chin Chem Soc 2004;49:421-6.
- Carter MJ, Fleming I, Percival A. The Diels-Alder route to allylsilanes from 1-trimethylsilyl butadienes. J Chem Soc Perkin 1 1981;1:2415-34.
- Jong TT, Jean MY. Constituents of *Houttuynia cordata* and the crystal structure of vomifoliol. J Chin Chem Soc 1993;40:399-402.
- Schmitt B, Schneider B. Dihydrocinnamic acids are involved in the biosynthesis of phenylphenalenones in *Anigozanthos* preissii. Phytochemistry 1999;52:45-53.
- Babu KS, Raju BC, Srinivas PV, Rao AS, Kumar SP, Rao JM. A simple, effective and highly selective cleavage of 3-methylbut-2-enyl (prenyl) ethers using *p*-toluenesulfonic acid. Chem Lett 2003;32:704-5.
- Withopf B, Richling E, Roscher R, Schwab W, Schreier P. Sensitive and selective screening for 6'-O-malonylated glucoconjugates in plants. J Agric Food Chem 1997;45:907-11.
- Qi M, Ge X, Liang M, Fu R. Flash gas chromatography for analysis of volatile compounds from *Houttuynia cordata* Thunb. Anal Chim Acta 2004;527:69-72.
- Zeng Z, Zhi JG, Zeng HP, Lai WL. Application of organic mass spectrometry in studies on *Houttuynia cordata*, a traditional Chinese medicine. Fenxi Huaxue 2003;20:399-404.
- Zeng HY, Jiang LJ, Zhang YC. Chemical constituents of volatile oil from *Houttuynia cordata* Thunb. Zhiwu Ziyuan Yu Huanjing Xuebao 2003;12:50-2.
- Liang M, Qi M, Zhang C, Zhou S, Fu R, Huang J. Gas chromatography-mass spectrometry analysis of volatile compounds from *Houttuynia cordata* Thunb. after extraction by solid-phase microextraction, flash evaporation and steam distillation. Anal Chim Acta 2005;531:97-104.
- Xu X, Ye H, Wang W, Yu L, Chen G. Determination of flavonoids in *Houttuynia cordata* Thunb. and *Saururus chinensis* (Lour.) Bail. by capillary electrophoresis with electrochemical detection. Talanta 2006;68:759-64.
- 75. Wu TS, Chan YY. Constituents of leaves of *Uncaria hirsuta* Haviland. J Chin Chem Soc 1994;41:209-12.
- Meng J, Leung KS, Dong XP, Zhou YS, Jiang ZH, Zhao ZZ. Simultaneous quantification of eight bioactive components of *Houttuynia cordata* and related *Saururaceae* medicinal plants by on-line high performance liquid chromatography-diode array detector-electrospray mass spectrometry. Fitoterapia 2009;80:468-74.
- Takagi S, Yamaki M, Masuda K, Kunota M. On the constituents of the terrestrial part of *Houttuynia cordata* Thunb. Shoyakugaku Zasshi 1978;32:123-5.
- Shimura M, Zhou Y, Asada Y, Yoshikawa T, Hatake K, Takaku F, et al. Inhibition of Vpr-induced cell cycle abnormality by quercetin: A novel strategy for searching compounds targeting Vpr. Biochem Biophys Res Commun 1999;261:308-16.
- 79. Meng J, Leung KS, Jiang Z, Dong X, Zhao Z, Xu LJ. Establishment

of HPLC-DAD-MS fingerprint of fresh *Houttuynia cordata*. Chem Pharm Bull (Tokyo) 2005;53:1604-9.

- Chiji H, Arakawa Y, Ueda S, Kuroda M, Izawa M. 5,2-Dihydroxy-6,7-methylenedioxyisoflavone from seed balls of sugar beet. Phytochemistry 1986;25:281-2.
- Netting AG, Millborrow BV, Duffield AM. Determination of abscisic acid in *Eucalyptus haemastoma* leaves using gas chromatography/mass spectrometry and deuterated internal standards. Phytochemistry 1982;21:385-9.
- Chen KS, Chang FR, Chia YC, Wu TS, Wu YC. Chemical constituents of *Neolitsea parvigemma* and *Neolitsea knoishii*. J Chin Chem Soc 1998;45:103-10.
- Kijima H, Otsuka H, Ide T, Ogimi C, Hirata E, Takushi A, *et al.* Glycosides of megastigmane and of the simple alcohols from *Alangium premnifolium*. Phytochemistry 1996;42:723-7.
- Broom SJ, Ede RM, Wilkins AL. Synthesis of (+/-)-E-4-(1,2,4-trihydroxy -2,6,6-trimethylcyclohexyl)-but-3-en-2-one: A novel degraded carotenoid isolated from New Zealand thyme

(Thymus vulgaris) honey. Tetrahedron Lett 1992;33:3197-200.

- Nozakim H, Suzuki H, Hirayana T, Kasai R, Wu RY, Lee KH. Antitumour triterpenes of *Maytenus diversifolia*. Phytochemistry 1986;25:479.
- Kuo YH, Li YC. Constituents of the bark of *Ficus microcarpa* Lf. J Chin Chem Soc 1997;44:321-5.
- Yungchien T. Florae Reipublicae Popularis Sinicae. 20<sup>th</sup> ed. Beijing: Science Press; 1982. p. 8.
- Wenpei F. Flora Sichuanica. 1<sup>st</sup> ed. Chengdu: Sichuan People Press; 1981. p. 126-7.

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