



Article An Electrochemical Sensor Based on Amino Magnetic Nanoparticle-Decorated Graphene for Detection of Cannabidiol

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Abstract: For detection of cannabidiol (CBD)—an important ingredient in *Cannabis sativa* L.—amino magnetic nanoparticle-decorated graphene (Fe₃O₄-NH₂-GN) was prepared in the form of nanocomposites, and then modified on a glassy carbon electrode (GCE), resulting in a novel electrochemical sensor (Fe₃O₄-NH₂-GN/GCE). The applied Fe₃O₄-NH₂ nanoparticles and GN exhibited typical structures and intended surface groups through characterizations via transmission electron microscopy (TEM), scanning electron microscopy (SEM), X-ray powder diffraction (XRD), vibrating sample magnetometer (VSM), and Raman spectroscopy. The Fe₃O₄-NH₂-GN/GCE showed the maximum electrochemical signal for CBD during the comparison of fabricated components via the cyclic voltammetry method, and was systematically investigated in the composition and treatment of components, pH, scan rate, and quantitative analysis ability. Under optimal conditions, the Fe₃O₄-NH₂-GN/GCE exhibited a good detection limit (0.04 µmol L⁻¹) with a linear range of 0.1 µmol L⁻¹ to 100 µmol L⁻¹ ($r^2 = 0.984$). In the detection of CBD in the extract of *C. sativa* leaves, the results of the electrochemical method using the Fe₃O₄-NH₂-GN/GCE were in good agreement with those of the HPLC method. Based on these findings, the proposed sensor could be further developed for the portable and rapid detection of natural active compounds in the food, agricultural, and pharmaceutical fields.

Keywords: cannabidiol; Cannabis sativa L.; electrochemical sensor; graphene; magnetic nanoparticles

1. Introduction

Cannabis sativa L. (C. sativa) is an annual dioecious herb belonging to the Cannabinaceae family, which is cultivated worldwide, and was one of the original crops in China [1]. C. sativa can be simply divided into industrial hemp and marijuana-generally distinguished by the content of Δ^9 -Tetrahydrocannabinol (Δ^9 -THC) in the plant; it is considered to be industrial hemp when the content of Δ^9 -THC is lower than 0.3% (*w/w*), and otherwise is referred to as marijuana. Based on the existing legal requirements in China, the cultivation of marijuana is banned; all varieties of C. sativa planted in China belong to the industrial hemp category [2]. In recent years, the medicinal usage of cannabidiol (CBD) has received unprecedented attention in the pharmaceutical and cosmetics industries. Accordingly, as the natural extraction source of CBD, C. sativa has ushered in a new round of development [3]. As an isomeride of Δ^9 -THC, CBD is non-psychoactive and exhibits good pharmacological effects in treating chronic pain, anxiety, inflammation, depression, and many other symptoms [4,5]. Today, the quantitative analysis methods of cannabinoids---including CBD---are mainly chromatographic methods, such as high-performance liquid chromatography (HPLC), gas chromatography (GC), and mass spectrometry [6-8].

As a kind of ultrasensitive detection method, electrochemical sensors or biosensors are mainly reported for the detection of Δ^9 -THC, since Δ^9 -THC is a typical psychoactive drug and is strictly regulated [9]. However, electrochemical sensors developed for the detection of CBD are rare. Since the interest in (and market for) CBD and related products



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). are growing, the detection of CBD has also become more important [10]. A convenient and rapid strategy for the detection of CBD could meet many needs in various scenarios outside of the laboratory. Through customization in the size and composition of sensors, in combination with the design of a small and portable workstation, a preliminary and rapid detection of CBD in plants could be completed in the field, which could save a lot of work for farmers or researchers [11]. Therefore, the effort to develop electrochemical sensors for CBD using novel nanomaterials is worthwhile.

As a typical two-dimensional nanomaterial, graphene (GN) has displayed many properties, including large specific surface area, high chemical stability, and excellent electrochemical properties [12]. It is widely used to modify electrodes in order to achieve better results in applications of supercapacitors, potassium-ion batteries, and detectors for biomarkers, metabolites, viruses, etc. [13–16]. The beneficial effects of modification of electrochemical sensors have been proven, including route simplicity, high efficiency, good performance, and low cost [17]. Magnetic nanoparticles such as iron oxide also show characteristics such as low toxicity, ease of functionalization, high adsorption ability, and magnetic responsivity [18]. The introduction of magnetic nanoparticles in electrochemical sensors could facilitate of electron transfer and signal amplification. The combination of GN and magnetic nanoparticles, resulting in Fe_3O_4/GN nanocomposites, has been utilized in the construction of various electrochemical sensors [19]. The Fe_3O_4/GN nanocomposites have been applied with satisfactory performance in the electrochemical detection of arsenic ions, lobetyolin, dopamine, glucose, prostate-specific antigen, hepatitis C virus, etc. [20–25].

In this study, many materials were tested for the modification of electrodes in order to obtain higher signals in the electrochemical detection of CBD. Amino-group-modified Fe₃O₄ nanoparticles (Fe₃O₄-NH₂) were finally confirmed for the modification of a glassy carbon electrode (GCE) together with GN in the form of nanocomposites. After the characterizations of the materials, GN and Fe₃O₄-NH₂ were mixed as nanocomposites and modified on the GCE (Fe₃O₄-NH₂-GN/GCE) to develop a novel electrochemical sensor for the highly selective and sensitive detection of CBD (Figure 1). The composition and fabrication sequences of the modifiers were investigated and optimized. Under the optimal fabrication and analytical conditions, the proposed Fe₃O₄-NH₂-GN/GCE demonstrated enhanced electrochemical signals, good linearity, and satisfactory anti-interference ability for CBD. The CBD content of *C. sativa* leaf extract was detected using the proposed Fe₃O₄-NH₂-GN/GCE, and the results were compared with those of the conventional HPLC method. Hence, it can be expected that the Fe₃O₄-NH₂-GN/GCE has extensive potential applications in the detection of CBD and other natural ingredients.



Figure 1. Diagram of Fe₃O₄-NH₂-GN/GCE and electrochemical detection of CBD.

2. Materials and Methods

2.1. Reagents and Apparatus

Detailed information about the reagents and instrumentations can be found in the Supplementary Material.

2.2. Preparation of Fe₃O₄-NH₂ Nanoparticles

 Fe_3O_4 nanoparticles were prepared according to our previous report [26]. Typically, 1.35 g of ferric chloride, 3.60 g of sodium acetate, and 1.00 g of PEG 6000 were mixed in 50 mL of ethylene glycol. The mixture was stirred under ultrasonication for 30 min and poured into a Teflon-lined stainless steel autoclave (100 mL). The autoclave was put into a drying oven at 180 °C for 6 h. After reaction, the black products were poured out and washed with water and ethanol three times each.

The obtained Fe₃O₄ nanoparticles were then dispersed in 250 mL of ethanol and ultrasonicated for 30 min [27]. After that, the materials were poured into a round-bottomed flask, and 2 mL of 3-aminopropyltriethoxysilane was dripped slowly into the Fe₃O₄ nanoparticle dispersion under mechanical agitation. The reaction was performed at room temperature for 6 h. Finally, the Fe₃O₄-NH₂ nanoparticles were washed with ethanol three times and stored in ethanol at 4 °C.

2.3. Fabrication of the Fe₃O₄-NH₂-GN/GCE

Before modification, the GCE was polished using alumina powders (0.05 μ m) and cleaned via ultrasonication for 10 min. The surface of the GCE was dried with nitrogen gas and stored for further use. Next, 12.0 mg of Fe₃O₄-NH₂ nanoparticles and 12.0 mg of GN were mixed in 2.0 mL of water and ultrasonicated for 10 min to form a homogeneous solution, which was marked as Fe₃O₄-NH₂-GN nanocomposites. Then, 10 μ L of the Fe₃O₄-NH₂-GN suspension (6.0 mg/mL in water) was carefully dropped on the surface of GCE and air-dried to form an active layer on the surface of the electrode. The modified electrode was referred to as Fe₃O₄-NH₂-GN/GCE.

For comparison, 10 μ L of Fe₃O₄-NH₂ nanoparticles and GN (6.0 mg/mL in water) were fabricated on the GCE in the same procedures and conditions, which were designated as Fe₃O₄-NH₂/GCE and GN/GCE, respectively. For confirmation of the fabrication sequence, three kinds of sequences were compared. Fe₃O₄-NH₂ nanoparticles were firstly dropped on the surface of GCE; GN was then dropped on the surface when the nanomaterials were dried, the result of which was designated as GN/Fe₃O₄-NH₂/GCE. GN was firstly modified on the bare GCE, and then Fe₃O₄-NH₂ nanoparticles were modified, which was designated as Fe₃O₄-NH₂/GCE. These two electrodes were compared with Fe₃O₄-NH₂-GN/GCE for their electrochemical response under the same conditions.

2.4. Preparation of Real Sample

Dry *C. sativa* leaves were ground, passed through a 40-mesh sieve, and placed in an oven at 105 °C for 10 h. After these treatments, 0.5 g of *C. sativa* leaves was immersed in 50 mL of anhydrous methanol solution. The mixture was extracted for 20 min using an ultrasonic extractor at a power of 200 W (KQ5200DV, Kunshan Ultrasonic Instrument Co., Ltd., Kunshan, China). After extraction, the mixture was centrifuged at 4000 r/min for 5 min (TD5, Yingtai Instrument Co., Ltd., Changsha, China). Next, 2 mL of the upper transparent solution was diluted to 20 mL with phosphate buffer solution (PBs, 10 mmol L⁻¹, pH 5.0) and filtered with 0.45 μ m filter before analyses via HPLC and using the proposed sensor.

2.5. Determination of CBD by HPLC

For comparison of the detection results, the HPLC method was applied in the detection of samples as well. An isocratic elution program consisting of 0.1% acetic acid and 75% acetonitrile was applied for 30 min at 25 °C. The flow rate was set to 0.8 mL/min. The chromatogram was observed at 220 nm. The injection volume of the sample was 10 μ L. The

CBD content in the *C. sativa* leaf extract was calculated using the standard curve obtained by the measurement standards.

2.6. Electrochemical Measurements

The electrochemical measurements were performed using the three-electrode system in CBD solution, using PBs (10 mmol L⁻¹, pH 5.0, containing 10% methanol) as a solvent and supporting electrolyte. Cyclic voltammetry (CV) was used for the measurement, with a scan rate of 0.05 V s⁻¹ and a potential range from 0 V to 0.8 V. Electrochemical impedance spectroscopy (EIS) was applied to characterize the sensor conductivity in the solution containing 5.0 mmol L⁻¹ of K₃[Fe(CN)₆]/K₄[Fe(CN)₆] and 0.1 mol L⁻¹ of potassium chloride. The amplitude was 0.005 V with a frequency range of 0.1 to 10⁵ Hz. All experiments were carried out in three duplicates at 25 ± 2 °C.

3. Results and Discussion

3.1. Characterizations of Nanomaterials

3.1.1. TEM and SEM

The morphologies of Fe₃O₄ nanoparticles, GN, and Fe₃O₄-NH₂-GN were investigated via TEM (Figure 2). The round sphere of Fe₃O₄ nanoparticles, as well as the silk-like and wrinkled structures of GN, could be easily observed in the corresponding images (Figure 2a,b) [28]. The Fe₃O₄ nanoparticles showed sizes of about 430 nm, with good dispersion. After mixing, the Fe₃O₄-NH₂-GN nanocomposites retained the characteristics of Fe₃O₄ nanoparticles and GN. It can be seen in the TEM image that Fe₃O₄ nanoparticles were dispersed on the GN sheets (Figure 2c) [29].



Figure 2. TEM images of (a) Fe₃O₄, (b) GN, and (c) Fe₃O₄-NH₂-GN. (d) SEM image of Fe₃O₄-NH₂-GN.

An SEM image of the Fe_3O_4 - NH_2 -GN nanocomposites on the electrode surface was also provided in order to confirm the morphology and structure (Figure 2d), and showed the modified surface of the electrode. Though there was a kind of agglomeration in the nanocomposites, the existence of Fe_3O_4 nanoparticles (round spheres) on the GN could be confirmed. The irregular surface of the modified electrode might be one of the reasons for the improved electrochemical response.

3.1.2. XRD

The XRD patterns of Fe₃O₄ nanoparticles, GN, and Fe₃O₄-NH₂-GN were analyzed, and are shown in Figure 3a. The pattern of Fe₃O₄ nanoparticles exhibited typical peaks at 30.3°, 35.7°, 43.6°, 57.4°, and 62.9, which were attributed to the indices (220), (311), (400), (511), and (440) of the Fe₃O₄ crystal, respectively [30]. Meanwhile, in the pattern of Fe₃O₄-NH₂-GN, the related peaks became much weaker, which might be a result of the coating of GN and the modification of the amino groups [26]. Additionally, another peak at ~26° could be observed, belonging to the characteristic reflection of the existence of GN [31]. The XRD results confirmed the existence of Fe₃O₄ nanoparticles and GN in the Fe₃O₄-NH₂-GN nanocomposites.



Figure 3. (a) XRD patterns of GN, Fe₃O₄, and Fe₃O₄-NH₂-GN. (b) Raman spectra of GN and Fe₃O₄-NH₂-GN.

3.1.3. Raman

Figure 3b illustrates the Raman spectra of the GN and Fe_3O_4 -NH₂-GN nanocomposites. Both GN and Fe_3O_4 -NH₂-GN showed two peaks at around 1350 cm⁻¹ and 1570 cm⁻¹, which were designated as D band and G band; they represented the disordered sp3 carbon structure (D band) and the sp2 ordered crystalline structure (G band) of GN [32]. After the combination of the two nanomaterials, the intensities of the peaks reduced significantly, which might be a result of the introduction of Fe_3O_4 -NH₂ nanoparticles. However, the intensity ratio of the D to G peaks was maintained, showing that the structure of GN was not affected.

3.2. Electrochemical Characteristics

The electrochemical behavior of various modified electrodes in 100 μ mol L⁻¹ of CBD were compared via the CV method. As shown in Figure 4a, the electrochemical response of CBD on bare GCE was only 0.728 µA (black line). After the respective modifications with Fe₃O₄ nanoparticles and GN to the GCE, small oxidation peaks at around 0.5 V could be observed on the Fe₃O₄/GCE (blue line) and the GN/GCE (red line), which might be due to the electron transfer properties and the good conductivity of Fe₃O₄ nanomaterials and GN [33]. When Fe_3O_4 -GN suspensions were used to modify the GCE, resulting in the Fe₃O₄-GN/GCE, an apparent increase in peak current could be observed (5.659 μ A, green line). The advantages of GN and Fe₃O₄ nanoparticles were combined and enhanced. Moreover, when the Fe_3O_4 nanoparticles were functionalized by amino groups, the resulting modified electrode (Fe_3O_4 -NH₂-GN/GCE) showed the highest response among these electrodes (8.978 μ A, Pink line). In order to confirm the effect of amino groups on Fe₃O₄ nanoparticles, Fe₃O₄-nanoparticle- and Fe₃O₄-NH₂-nanoparticle-modified electrodes (Fe₃O₄/GCE and Fe₃O₄-NH₂/GCE) were compared. As a result, the peak current of Fe_3O_4 -NH₂/GCE was slightly higher than that of Fe_3O_4 /GCE (1.366 to 1.08, not shown). A possible reason for this increase might be that the amino groups on the surface could attract more target molecules. As far as we know, there has been no previous report regarding the electrochemical oxidation mechanism of CBD. By referring to reported works on the oxidation of Δ^9 -THC, the oxidation process of CBD could be assumed to be a phenol-type oxidation mechanism [34,35].



Figure 4. (a) CV curves of CBD on bare GCE, GN/GCE, Fe₃O₄/GCE, Fe₃O₄-GN/GCE, and Fe₃O₄-NH₂-GN/GCE. CV method: 100 μ mol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range of 0–0.8 V. Scan rate of 0.05 V s⁻¹. (b) Nyquist plots of bare GCE, GN/GCE, Fe₃O₄/GCE, Fe₃O₄-GN/GCE, and Fe₃O₄-NH₂-GN/GCE in 5.0 mmol L⁻¹ of K₃[Fe(CN)₆]/K₄[Fe(CN)₆] and 0.1 mol L⁻¹ of potassium chloride. The amplitude is 0.005 V, with a frequency range of 0.1 to 10⁵ Hz.

In order to optimize the effects of the modifiers, the fabrication sequence of modified sensors was investigated. Through the comparison of $GN/Fe_3O_4-NH_2/GCE$, $Fe_3O_4-NH_2/GN/GCE$, and $Fe_3O_4-NH_2-GN/GCE$, the peak currents of each sensor were obtained, as shown in Table 1. Apparently, the $Fe_3O_4-NH_2-GN/GCE$ showed the best response among these sensors, meaning that the modifiers should first be mixed, and then dropped directly on the surface of the electrode. Based on this finding, different preparation methods of $Fe_3O_4-NH_2$ -GN suspensions were tried (see the ESM). Three kinds of $Fe_3O_4-NH_2$ -GN nanocomposites were compared, and their corresponding peak currents are also shown in Table 1. Although the Fe_3O_4 nanoparticles were directly prepared in the presence of GN via ultrasonication and solvothermal methods, the electrochemical properties obtained were not as good as via the physical mix method. Hence, the $Fe_3O_4-NH_2$ -GN suspension was confirmed as the optimal material in this research.

Table 1. The I_p in different fabrication sequences and process methods of Fe₃O₄-NH₂-GN in modified electrodes.

Fabrication Sequence	<i>I_p</i> (μA)	Process Method	<i>I_p</i> (μA)
GN/Fe ₃ O ₄ -NH ₂ /GCE	1.808	Ultrasonication	3.352
Fe ₃ O ₄ -NH ₂ /GN/GCE	3.388	Solvothermal	4.232
Fe ₃ O ₄ -NH ₂ -GN/GCE	5.327	Mix	5.550

The Nyquist plots from the EIS test reflect the conductivity of the electrodes (Figure 4b). The inset of Figure 4b shows a general equivalent circuit containing the solution resistance (R_s), the electron transfer resistance (R_{et}), the Warburg element (W), and the charge of the constant phase element (C_d) [36]. The value of R_{et} was calculated by fitting the experimental data to the model circuit. As shown in Figure 4b, the Nyquist plot of bare GCE showed a semicircle, with an R_{et} of 1287 Ω . When the GCE was modified with Fe₃O₄ nanoparticles and GN, the R_{et} of Fe₃O₄-OK/GCE and GN/GCE reduced to 141.4 Ω and 28.61 Ω , respectively. Finally, the R_{et} of Fe₃O₄-NH₂-GN/GCE was only 13.73 Ω , which was similar to that of Fe₃O₄-GN/GCE (16.19 Ω). The decreases in resistance could be attributed to the outstanding electric conductivity of GN and magnetic nanoparticles [37]. Consequently, the Fe₃O₄-NH₂-GN/GCE was confirmed as the optimal modified sensor, by reason of its optimal response and conductivity in electrochemical detection.

3.3. Optimization of Electrochemical Conditions

3.3.1. Effect of Composition of Fe₃O₄-NH₂-GN

In order to obtain the optimal mixture composition, the ratios of GN and Fe₃O₄-NH₂ nanoparticles (1:0.5, 1:1, 1:1.25, 1:1.5, 1:2.0 and 1:2.5, *w:w*) were investigated, and are shown

in Figure 5a. The concentration of GN was set at 2.0 mg mL⁻¹, and the concentrations of Fe₃O₄-NH₂ nanoparticles were verified according to the ratios. These illustrated results suggested that the electrochemical signals of CBD were the highest when the ratio was 1:1. When the ratio of Fe₃O₄-NH₂ nanoparticles was higher than 1.0, the response gradually became weaker. Therefore, the material ratio of GN and Fe₃O₄-NH₂ nanoparticles was set to 1:1 as the optimal composition for the fabrication of the electrode.



Figure 5. (a) Effect of compositions of Fe₃O₄-NH₂-GN on I_p in CV. (b) Effect of ultrasonication time of Fe₃O₄-NH₂-GN on I_p in CV. (c) Effect of modification volumes on peak current in CV. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range of 0–0.8 V. Scan rate of 0.05 V s⁻¹.

3.3.2. Effect of Ultrasonication Time of Fe₃O₄-NH₂-GN

To obtain a stable dispersion, various ultrasonication times of the Fe_3O_4 -NH₂-GN suspension were tested, from 1 min to 30 min (1, 5, 10, 20, and 30 min). Then, the materials were used for the fabrication of electrodes. The changes in peak currents using the corresponding modified electrodes are plotted in Figure 5b. It can be seen that the electrochemical response was the highest when the material was treated for 10 min. However, longer ultrasonication time did not make the response better. Hence, the ultrasonication time of Fe_3O_4 -NH₂-GN suspension was confirmed at 10 min.

3.3.3. Effect of Concentration of Fe₃O₄-NH₂-GN

The effect of concentration of Fe₃O₄-NH₂-GN suspension was measured from 1.0 mg mL^{-1} to 8.0 mg mL^{-1} , and the modification volume was fixed at 10.0μ L. As the previous experiment indicated, the ratio of GN and Fe₃O₄-NH₂ nanoparticles was set at 1:1 (*w:w*). It can be seen in Figure 5c that the peak current increased as the concentration increased from 1.0 mg mL^{-1} to 6.0 mg mL^{-1} . However, when the concentrations were more than 6.0 mg mL^{-1} , this trend stopped, and the response began to gradually drop, which was similar to the results of a previous report [38]. Then, the concentration of Fe₃O₄-NH₂-GN suspension was optimized as 6.0 mg mL^{-1} .

3.3.4. Effect of pH

The electrochemical detection using various pH values of the electrolyte (4.0, 5.0, 6.0, 7.0, 8.0, and 9.0) containing CBD as samples was performed with the Fe₃O₄-NH₂-GN/GCE, using the CV method. The trend is shown in Figure 6a, and the peak current of CBD was the highest when the pH of the electrolyte was 5.0. There was a downward trend when the pH of the electrolyte became higher than 5.0, which is consistent with Zanardi's research [39]. Thus, 5.0 was adopted as the optimal electrolyte pH value during the tests.

Moreover, it could be observed that there was a linear shit of the peak potential (Ep) to lower positive values as the pH increased. The linear equation between Ep and pH was expressed as $Ep = -0.053 \text{ pH} + 0.863 (r^2 = 0.984)$ (Figure 6b). The slope of the equation was $\sim -0.053 \text{ V pH}^{-1}$, similar to the theoretical Nernstian slope of 0.059 V pH⁻¹. This parameter corresponded to an oxidation mechanism that included the exchange of an equal number of protons and electrons in the reaction [40].



Figure 6. (a) Effect of pH on I_p in CV. (b) Plot of peak potential (Ep) to pH values. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range of 0–0.8 V. Scan rate of 0.05 V s⁻¹.

3.4. The Influence of the Scan Rate

As an important parameter reflecting the performance of the electrode, the effect of different scan rates (from 5 mV s⁻¹ to 200 mV s⁻¹) on the electrochemical response of CBD in PBs (10 mmol L⁻¹, pH 7.0) was evaluated on the Fe₃O₄-NH₂-GN/GCE, using the CV method. Figure 7a shows the resulting CV curves at a variety of scan rates. It can be seen that the peak currents increased and shifted with the increasing scan rates. A good linear relationship could be obtained between scan rate and peak current, which could be expressed as: $I_p = 164.84 v + 1.73 (r^2 = 0.998)$ (Figure 7b), indicating that the oxidation of CBD was an adsorption-controlled process [41]. However, when the scan rate was increased to more than 200 mV s⁻¹ (250 mV s⁻¹ and 300 mV s⁻¹), the response did not grow proportionately to the former linear trend (lower than former trend). Another linear dependence of the logarithm of the peak current (log I_p) against the logarithm of the scan rate could also be observed, which was fitted as: log $I_p = 0.735 \log v + 2.00 (r^2 = 0.982)$ (Figure 7c). This trend suggests that the electrochemical reaction was controlled by both diffusion and adsorption [42].



Figure 7. (a) CV of Fe₃O₄-NH₂-GN/GCE in CBD solution at different scan rates. (b) The linear graph of I_p and scan rates. (c) The linear graph of log I_p and log (scan rate). (d) Plot of I_p versus concentration of CBD. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range of 0–0.8 V. Scan rate of 0.05 V s⁻¹.

3.5. Quantitative Analysis of CBD

In order to study the quantitative analysis ability of the fabricated Fe₃O₄-NH₂-GN/GCE, the CV curves of CBD at different concentrations from 0.1 μ mol L⁻¹ to 100 μ mol L⁻¹ were observed in PBs (0.01 mol/L, pH 5.0). The results illustrated that the peak current increased with increasing CBD concentrations, and three sections of linear dependences could be found between the peak current and the CBD concentration during this range, with a detection limit of 0.04 μ mol L⁻¹ (S/N = 3), which is consistent with Liu's work [43]. The plot of peak current versus CBD concentration is shown in Figure 7d. The three regression equations could be respectively expressed as: $I_{p1} = 1.284 \text{ C}_1 + 0.528 (0.1-0.974 \,\mu\text{mol L}^{-1})$ $r^2 = 0.984$), $I_{p2} = 0.176 \text{ C}_2 + 1.607 (0.974 - 19.494 \ \mu\text{mol L}^{-1}, r^2 = 0.984$), and $I_{p3} = 0.0617 \text{ C}_3 + 1.607 \text{ C}_3$ 3.836 (19.494–100 μ mol L⁻¹, r^2 = 0.988). It could be found in three regression equations that the slope of the peak current at low concentration was higher than at high concentration. At a lower analyte concentration, the number of active sites on the electrode was relatively higher. However, because of the occupancy of—and decrease in the number of—active sites at higher analyte concentrations, the sensitivity and the slope became lower [44]. This demonstrates that the quantitative analysis of CBD using the Fe₃O₄-NH₂-GN/GCE was interesting and acceptable [45]. The detection abilities of the reported electrochemical sensors for CBD are listed and compared with the Fe₃O₄-NH₂-GN/GCE in Table 2. Through the comparison, the proposed Fe₃O₄-NH₂-GN/GCE exhibited a competitive detection capability and sensitivity for CBD.

Table 2. Comparison of different reported sensors for the electrochemical determination of CBD.

Electrode	Linear Rage (µmol L ⁻¹)	LOD (μ mol L $^{-1}$)	Ref.
GC/CB	0.96-6.37	0.35	[39]
Sonogel-Carbon-PEDOT	1.59–19.1	0.94	[34]
NACE-ED	0.32-31.8	0.064	[46]
GCE	_NM	_NM	[47]
Fe ₃ O ₄ -NH ₂ -GN/GCE	0.1-100.0	0.04	This study

GC/CB: glassy carbon/carbon black; PEDOT: poly-(3,4-ethylenedioxythiophene); NACE–ED: non-aqueous capillary electrophoresis–electrochemical detection; ^{NM}: Not mentioned.

3.6. Practicability of the Fe₃O₄-NH₂-GN/GCE

The anti-interference ability, repeatability, and stability of the Fe_3O_4 -NH₂-GN/GCE were tested, and the results were satisfactory (see the ESM). Moreover, the detection ability of the Fe_3O_4 -NH₂-GN/GCE for CBD was evaluated in the extract of *C. sativa* leaves. In order to verify the results, the standard addition method was employed by spiking different amounts of CBD into samples. The results are shown in Table 3 and compared with those obtained via the HPLC method. The recoveries ranged from 99.1% to 100.4%, indicating that the determination was reliable, and there was consistency between the concentrations of CBD measured by both electrochemical and HPLC methods.

Table 3. Determination of	CBD in real sa	nples. (<i>n</i> =	= 3).	•
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Samples	Added (µmol L^{-1})	Found (µmol L^{-1})	Recovery (%)	RSD (%)	HPLC (μ mol L ⁻¹)
	0	11.95	-	2.23	12.06
Extract of C. sativa	1.0	13.00	100.4	2.05	-
	5.0	16.84	99.1	3.51	-

4. Conclusions

In this study, an electrochemical sensor (Fe₃O₄-NH₂-GN/GCE) was fabricated for the detection of CBD. The applied materials and fabrication conditions were compared and optimized via various characterizations and evaluations. The performance of the Fe₃O₄-NH₂-GN/GCE was investigated for aspects including pH, scan rate, anti-interference

ability, repeatability, and stability. As a result, the proposed Fe_3O_4 - NH_2 -GN/GCE showed an improved electrochemical response compared to a bare GCE. It displayed quantitative analysis ability for CBD, with a linear range of 0.1 µmol L⁻¹ to 100 µmol L⁻¹. The practicability test also showed that the result was in good agreement with that of the HPLC method in the detection of CBD in real samples. Based on these findings, the Fe_3O_4 - NH_2 -GN/GCE could be further utilized for the detection of active compounds in natural extracts.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10 .3390/nano11092227/s1, Figure S1: (a) I_p ratios of Fe₃O₄-NH₂-GN/GCE in CBD solution containing various interfering substances. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range at 0-0.8 V. Scan rate at 0.05 V s⁻¹, Figure S2: Repeatability of Fe₃O₄-NH₂-GN/GCE in CBD solution. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range at 0–0.8 V. Scan rate at 0.05 V s⁻¹, Figure S3: Stability of Fe₃O₄-NH₂-GN/GCE in CBD solution. CV method: 100 µmol L⁻¹ of CBD in 10 mmol L⁻¹ of PBs (pH 5.0, containing 10% methanol). Potential range at 0–0.8 V. Scan rate at 0.05 V s⁻¹.

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