



Review Article

Carotid Disease and Ageing: A Literature Review on the Pathogenesis of Vascular Senescence in Older Subjects

Eleni Sertedaki ^{1,2} **Dimitris Veroutis**,² **Flora Zagouri**,³ **George Galyfos** ¹,
Konstadinos Filis,¹ **Alexandros Papalambros**,⁴ **Konstantina Aggeli**,⁵ **Panagiota Tsioli**,⁶
George Charalambous,¹ **George Zografos**,¹ and **Fragiska Sigala**¹

¹First Department of Propaedeutic Surgery, Hippocraton General Hospital, School of Medicine, National Kapodistrian University of Athens, Athens, Greece

²Molecular Carcinogenesis Group, Department of Histology and Embryology, School of Medicine, National Kapodistrian University of Athens, Athens, Greece

³Clinical Therapeutics Department, Alexandra General Hospital, School of Medicine, National Kapodistrian University of Athens, Athens, Greece

⁴First Department of Surgery, Laikon General Hospital, School of Medicine, National Kapodistrian University of Athens, Athens, Greece

⁵First Department of Cardiology, University of Athens Medical School, Hippocraton Hospital, Athens, Greece

⁶First Department of Pathology, Laikon General Hospital, School of Medicine, National Kapodistrian University of Athens, Athens, Greece

Correspondence should be addressed to Eleni Sertedaki; elenasertedaki@gmail.com

Received 4 December 2019; Revised 11 May 2020; Accepted 16 May 2020; Published 13 June 2020

Academic Editor: Jacek Witkowski

Copyright © 2020 Eleni Sertedaki et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Aging is a natural process that affects all systems of the human organism, leading to its inability to adapt to environmental changes. Advancing age has been correlated with various pathological conditions, especially cardiovascular and cerebrovascular diseases. Carotid artery (CA) is mainly affected by age-induced functional and morphological alterations causing atheromatous disease. The evolution of biomedical sciences has allowed the elucidation of many aspects of this condition. Symptomatic carotid disease (CD) derives from critical luminal stenosis or eruption of an atheromatous plaque due to structural modifications of the vessels, such as carotid intima-media thickening. At a histologic level, the aforementioned changes are mediated by elastin fragmentation, collagen deposition, immune cell infiltration, and accumulation of cytokines and vasoconstrictors. Underlying mechanisms include chronic inflammation and oxidative stress, dysregulation of cellular homeostatic systems, and senescence. Thus, there is an imbalance in components of the vessel wall, which fails to counteract exterior stress stimuli. Consequently, arterial relaxation is impaired and atherosclerotic lesions progress. This is a review of current evidence regarding the relationship of aging with vascular senescence and CD. A deeper understanding of these mechanisms can contribute to the production of efficient prevention methods and targeted therapeutic strategies.

1. Introduction

Carotid disease (CD) is one of the most common age-related diseases. It refers to a number of macroscopic morphological changes and to alterations at cytopathological level in common carotid artery (CCA) and internal carotid artery (ICA) that lead to cerebrovascular events such as strokes and transient ischemic attacks (TIAs) that comprise a major

cause of mortality and morbidity, especially amongst the elderly [1].

Occurrence of CD is strongly associated with cardiovascular disease, atherosclerosis, and arterial stiffening induced by aging as well as by various atherogenic risk factors, including metabolic syndrome, hypertension, dyslipidemia, especially high low-density lipoprotein (LDL) levels, smoking, and diabetes mellitus (DM). Particularly, aging

itself has been identified actually as an independent and unmodifiable risk factor for arterial stiffening and CD irrespective of patient comorbidities, whilst carotid atherosclerotic disease is considered a typical manifestation of vascular aging or vascular senescence [1, 2].

Vascular senescence refers to the establishment of clinical and subclinical structural and functional changes at a histological and cellular level, involving cells and the extracellular matrix of vessel walls [3] that are related with the loss of its homeostatic potential to adapt to environmental stress stimuli. It is a separate entity from aging, provoked by ageing itself, but also by the aforementioned atherogenic factors [2]. It is also correlated with “inflammaging,” a sterile, low-grade, chronic inflammation induced by advancing age, leading to a number of diseases, mostly cardiovascular and cerebrovascular ones [4].

In fact, aging is a natural process that causes multiple changes to the human organism decreasing its ability to maintain its homeostasis and make it more vulnerable to threats. It is associated with the impairment of vascular function and onset of cardiovascular disease [5, 6] even in the absence of other risk factors [7, 8]. There seems to be a linear relationship between arterial stiffness, age [9], and accelerated arterial stiffening between 50 and 60 years of age [10].

Multiple macrovascular and microvascular complications are initiated due to the formation of atherosclerotic lesions, under the impact of oxidized LDLs (oxLDLs), endothelial dysfunction, and impaired arterial wall homeostasis, which in turn exacerbate media thickening and fibrosis, leading to arterial stiffness and cerebrovascular disease [5, 6, 11–19]. As atherosclerosis is a disease of aging that involves cellular senescence, a deep understanding of the underlying mechanisms that regulate cell survival and onset of atherosclerotic disease with increasing age is a valuable tool in prevention and management of the progression of CD [4, 6, 10].

The purpose of this review is to summarize existing findings regarding aging and progression of carotid atherosclerotic disease. Thus, we have collected data through a comprehensive literature search online. We believe that deciphering molecular mechanisms involved in the pathogenesis of the disease will give us new perspectives in the prevention, diagnosis, and treatment of vascular disease in older ages.

1.1. Macroscopic Changes in Carotid Artery with Advancing Age. Anatomical and geometrical features of the carotid artery (CA) play an important role in the onset of ICA stenosis and occurrence of CD, manifested as transient ischemic attack, stroke, or syncope [8, 20]. Such morphological changes take place with advancing age, including increases in vessel volume, diameter, and rotation of the CCA and ICA, in the bifurcation and ICA angles. A higher anatomical position of ICA has also been detected in patients over 60 years old. Moreover, the ICA angle seems to affect ICA radius and degree of stenosis, while the bifurcation angle seems to influence carotid circulation, constituting one

of the most common sites for the formation of atherosclerotic plaques [20]. The aforementioned changes alter the hemodynamic forces that affect crucially the progression of atherosclerosis.

However, prior to the establishment of these morphological changes, functional deterioration of vascular wall takes place. Arterial stiffness is known to progress with age and elastic properties of CA which tend to differentiate [21]. Vascular functionality is reduced significantly by arterial stiffness, as the vessel wall fails to adapt to pressure fluctuations [8]. According to ultrasound findings, the carotid β -stiffness index (ultrasound parameter-assessing arterial stiffening) and vessel diameter increase, while arterial strain (ability to adjust with pressure changes with mechanical deformation) reduces with advancing age [22]. Consistently, carotid intima-media thickness (cIMT), a common measure of atherosclerosis and useful tool for the prediction of cardiovascular and cerebrovascular events, displays a linear increase along with age irrespectively of other factors [23, 24]. Furthermore, complicated carotid atheromas with intraplaque hemorrhage are related with older age as confirmed in multiple studies with use of ultrasound, magnetic resonance, and histological assessment of carotid plaques [25, 26].

At a histologic level, it is known that the hallmark of atherosclerosis is the formation of an atherosclerotic lesion. Hemodynamic shear stress results in endothelial injury and expression of cell adhesion molecules (CAMs) and other cell surface receptors that lead to increased endothelial permeability and accumulation of vascular smooth muscle cells (VSMCs) in the subendothelial space. Circulating LDL molecules, especially when at high serum concentrations, enter into subendothelial spaces, bind to glycoproteins, and are oxidized by free endothelial cell receptors. OxLDLs promote transendothelial migration of monocytes into the subendothelial spaces and their differentiation to macrophages under the impact of cytokines. These macrophages absorb oxLDLs becoming foam cells that stimulate a local inflammatory reaction, which further attracts inflammatory cells initiating lesion formation. This atherosclerotic lesion is made up of a lipid core and a fibrous cap consisting of macrophages, VSMCs, collagen, elastin, and proteoglycans [27].

Stable plaques that are at a solid state have a thick fibrous cap, while vulnerable or unstable ones are at a rather liquid state having a large lipid core and a thin fibrous cap related with inflammatory infiltrates [25–27].

Evidently, the progression of atherosclerotic plaques at a histologic level seems to be enhanced by advancing age, as their morphology is altered towards instability and vulnerability to rupture. Plaques become more atherosclerotic, inflammatory, and less fibrous, with a decrease in elastin fibers, VSMCs, and overall cellularity. Lipid accumulation, calcification, hemorrhage, and deficient tissue repair that make plaques more prone to injury are identified. Macrophage infiltrates are also detected mainly in plaques of symptomatic old patients rather than asymptomatic ones [26]. Moreover, plaques from elderly patients contain less fractalkine, interferon-gamma (IFN- γ), and tumor necrosis

factor- α (TNF- α), a finding suggesting that infiltrating macrophages in elderly are probably less functional. They also have a high content of the proinflammatory and prothrombotic molecule sCD40L and vascular endothelial growth factor (VEGF), which is a stimulus for plaque neovascularization at conditions of inflammation and poor blood supply. The concurrent reduction of endothelial shear stress and increase of arterial stiffness and wall tension promote atherosclerosis. Consequently, the age-associated plaque morphology seems to change towards more vulnerable plaques, [25, 28] the rupture of which leads to acute cerebrovascular and cardiovascular events, which increase in incidence with age.

2. Underlying Mechanisms

2.1. Endothelial Dysfunction. Endothelial cells (ECs) form a thin layer between circulating blood and the rest of the vessel wall. Endothelium is a strategically placed barrier with an active role on vascular homeostasis [29]. In response to physical and chemical signals, ECs produce multiple substances that regulate nutrient supply, cellular adhesion, coagulation, and fibrinolysis, as well as VSMC proliferation, vessel wall inflammation, and vascular tone [30]. In particular, ECs have autocrine, paracrine, and endocrine functions. They produce prothrombotic and antithrombotic substances, as well as both fibrinolytic and antifibrinolytic ones. They regulate VSMC migration to the intima as well as leucocyte adhesion and activation [27, 28]. In addition, the regulation of vascular tone is mediated by endothelium-derived factors (vasodilators and vasoconstrictors) that affect vascular structure and end-organ perfusion in the long term. Especially, nitric oxide (NO) is a critical vasorelaxant factor generated by the endothelial NO synthase (eNOS) [31, 32]. However, when ECs are activated by prolonged stress stimuli, such as aging and atherogenic factors, they become senescent or undergo apoptosis that leads to the loss of integrity of the endothelial layer. They lose their capacity to maintain homeostasis, and they are invaded by lipids, platelets, and leucocytes that stimulate plaque formation [27, 28].

2.2. Endothelial Dysfunction due to Nitric Oxide (NO) Depletion. Endothelial dysfunction refers to the inability of endothelium to maintain vascular homeostasis, enhancing vasoconstriction, oxidative, and inflammatory reactions that lead to various cardiovascular diseases. Impaired vasodilation in hypercholesterolemia results mainly from diminishment of NO bioavailability [33]. Moreover, many of the aforementioned protective endothelial functions including inhibition of platelet and leucocyte adhesion and aggregation, VSMC migration, and inhibition of LDL clustering are dysregulated due to NO reduced bioavailability, resulting in the evolution of atheromatous disease [27, 28, 31, 32]. According to experimental and clinical data, increasing age is related with reduced levels of the endothelial NO synthase (eNOS), thus reduced bioavailability of NO, and increased levels of the inducible NO synthase (iNOS), which is

upregulated under inflammatory conditions producing excessive amounts of NO that rapidly interact with oxidative species forming ONOO- that exacerbates nitrosative stress and deteriorates endothelial homeostasis [16, 27, 33]. This imbalance in synthesis and degradation of NO ends up to its depletion and loss of its beneficial effects. Thus, atheromatous disease evolves further due to the diminished NO generation and impaired NO diffusion from endothelium to VSMCs [16, 27, 33]. Consistently, iNOS inhibition could improve endothelium-dependent responses according to experimental data [33, 34].

2.3. Endothelial Dysfunction due to Increased Production of Vasoconstrictors. Another enzyme involved in ageing-induced endothelial dysfunction is cyclooxygenase (COX). It leads to generation of vasoconstrictor substances, such as thromboxane A₂ (TXA₂) and prostaglandins (PGs), such as PGH₂, PGF_{2 α} , and PGE₂, which deteriorate endothelial function [35–37]. In addition, activation of endothelin-1 is reinforced with increasing age, enhancing vasoconstriction and inflammation [38]. Endothelial vasoconstrictors have been shown to cause chemotactic accumulation of leukocytes and platelets mediated by the expression of surface adhesion molecules (selectins, integrins, immunoglobulins, intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), etc.). Thus, monocytes adhere to the endothelial surface and migrate into the intima [27, 28, 38]. COX inhibitors, such as indomethacin, have been shown to conserve vascular functionality in older adults and aged animals [39]. Evidence remains inconsistent, though, as some studies show no effect with the use of COX-2 inhibitors [40]. Nevertheless, the lack of prostaglandin I₂ (PGI₂)-mediated vasodilatation with aging has been documented both in vivo and in vitro [41].

2.4. Chronic Vascular Inflammation. The major underlying factor is low-grade chronic inflammation detected in aged vasculature. It acts as a continuous stress stimulus that promotes oxidative strain, cellular senescence, and impairment of vascular function [42].

In this context, multiple cytokines are expressed composing networks that get differentiated with increasing age irrespectively of other risk factors [43]. Blood samples from older adults reveal prevalence of C-reactive protein (CRP) as well as of proinflammatory cytokines and adhesion molecules such as tumor necrosis factor- α (TNF- α), interleukin (IL)-1 β , interleukin-6 (IL-6), and VCAM-1 [43]. Overexpression of these molecules is associated with aggravation of endothelium impairment and arterial stiffening [44–46] that lead to cerebrovascular and cardiovascular events, and increased mortality and morbidity amongst the elderly [46, 47]. It is worth mentioning that inflammatory mediators, such as TNF- α that regulates eNOS and iNOS expression, are produced by ECs too [48, 49].

On the contrary, anti-inflammatory cytokines, such as interleukin-10 (IL-10) and adiponectin, which decelerate senescence and protect arteries against cardiovascular disease, seem to be downregulated with increasing age [50–54].

In particular, inhibition of IL-10, known to protect from carotid atheromatous disease and coronary calcification, [50] led to endothelial dysfunction and vascular inflammation of carotid arteries in IL-10 knockout aged mice according to experimental findings [55, 56].

Moreover, immune cell infiltrates with macrophages and T lymphocytes have been detected in histologic carotid lesions conserving inflammatory phenotype in aged subjects [57, 58]. Consistently, macrophage and leukocyte infiltrates have been detected in aortic adventitia and perivascular fat tissue of aged mice [44, 58, 59].

Nuclear factor kappa-B (NF- κ B), a thoroughly studied protein complex, is considered to have a critical role in the initiation and conservation of inflammation as well as in the induction of senescence [60, 61]. It remains inactive linked to protein I κ B- α in the cytoplasm; when activated by stress stimuli, NF- κ B is transferred to nucleus, where it controls expression of numerous target genes [60–64] activating TNF- α , interleukins (IL-1 β and IL-6), chemokines (IL-8 and RANTES), adhesion molecules (intercellular adhesion molecule and vascular cell adhesion molecule), and enzymes (iNOS and COX-2) [60, 61, 64, 65]. Through a vicious cycle, NF- κ B expression is reinforced by the products of its target-signaling cascades [61, 64, 65]. Thus, it constitutes a major regulator of immune vascular responses to aging and its inhibition could delay ageing process [66–71].

2.5. Oxidative Stress. In addition to chronic inflammation, chronic oxidative stress is a main feature of the aging process [16, 69, 72]. Reactive oxygen species (ROS) (superoxide anion and hydrogen peroxide) generation is mediated by various oxidant enzymes (e.g., NADPH oxidase (NOX), xanthine oxidase, cytochromes, and uncoupled eNOS) that are upregulated with advancing age; mitochondrial hydrogen peroxide is an important source of ROS in aging vasculature too [73–77]. This increase though would stimulate cellular antioxidant defensive mechanisms. However, there is a significant decrease in critical antioxidant enzymes (dismutases, catalases, and deacetylases) under the impact of numerous inflammatory factors, such as NF- κ B [78]. Thus, antioxidant capacity is reduced in the elderly, leading to an imbalance in favor of oxidants on the vascular wall [72, 79]. In support of this, elimination of oxidative stress could improve or reverse aging-induced vascular dysfunction according to findings of multiple experimental studies, constituting a feasible targeted therapeutic approach [78–80].

Furthermore, although native LDL molecules are inactive, oxidative stress involved in atherosclerosis process, turns them into the highly immunogenic oxLDLs, which trigger the production of endothelial adhesion molecules, attract monocytes, and enhance proinflammatory gene activity [16, 27, 74]. In fact, this disequilibrium between pro and antioxidant factors increases endothelial permeability and promotes foam cell formation [73, 74, 81].

3. Cell Regulatory Systems

3.1. Role of microRNAs. Cell modulatory systems include microRNAs, small noncoding RNAs that bind to

3' untranslated regions of mRNAs regulating their stability and translation [82]. Due to their ability to interact with multiple mRNAs, miRNAs have a critical role in controlling signaling pathways related with inflammation, senescence, and aging [82–90] miRNAs with inflammatory potential, such as miR-34a (the inducer of vascular smooth muscle senescence and arterial dysfunction), [91] miR-146a (the senescence and inflammation inducer interacting with telomeres and telomerase) [90, 92], and miR-125a-5p (the dysregulator of eNOS and VEGF) are upregulated in vascular cells of older subjects and involved in different signaling pathways (e.g., NF- κ B activation) [63, 88]. Conversely, miRNAs with anti-inflammatory effects such as miR-155 and miR-27a are downregulated [93–95].

3.2. Other Cell Regulatory Systems. Additional modulatory systems with critical cellular homeostatic functions are involved in vascular inflammation and cellular senescence that evolves with ageing [95–98].

To begin with, SIRT1, a NAD-dependent histone deacetylase, a member of the sirtuins family, plays a crucial role in cell survival and metabolism [97–99]. It is mostly expressed in the nucleus where it regulates transcription of multiple genes. It inhibits NF- κ B with a negative impact on senescence [96, 99]. Its levels though are diminished in aged subjects, with an enhancement of ROS production by NADPH oxidases that remain unrestrained. As a result, inflammation, endothelial, and smooth muscle cell dysfunction and arterial stiffness progress [100–102]. Restoration of SIRT1 levels (e.g., with resveratrol) is a promising therapeutic target to reverse vascular inflammation and endothelial dysfunction [101–105].

Likewise, SIRT6, another nuclear deacetylase, has similar effects. It suppresses NF- κ B activity, inhibits oxidative stress, ageing phenotype, and senescence, expanding lifespan, according to preclinical data on mouse models and human ECs [106–109]. SIRT6 expression is downregulated by inflammatory stimuli though, including ageing [110, 111].

Another regulatory molecule is nuclear-related factor-2 (Nrf2) that controls cell responses to oxidative stress in ageing process [112–114]. It represses NF- κ B activation and nuclear translocation, [67, 68] while NF- κ B antagonizes Nrf2 signaling at the transcription level, according to experimental studies on aorta and carotid arteries from aged animals [70, 72, 114]. Furthermore, Nrf-2 stimulates a cytoprotective kind of angiogenesis through enhancement of angiogenic potential of endothelial cells, which is impaired with advancing age along with the reduction of Nrf-2 levels, while inflammation and ischemia-related neovascularization is amplified [16, 115].

Finally, AMPK (AMP-activated kinase) and mTOR (mammalian target of rapamycin) are additional modulatory factors. They are crucial energy sensors with important roles on cell metabolism and survival. Moreover, they have been identified to antagonize each other; AMPK can directly inhibit mTOR while high levels of AMPK and low levels of mTOR are associated with longevity. Hence, AMPK, which affects intracellular pathways critical for cell cycle and

metabolism, is downregulated in arteries of old subjects. On the other hand, mTOR signaling is enhanced in aging and related diseases and its inhibition seems to exert antioxidant and protective effects on endothelium [116–119].

3.3. Autophagy and Senescence. Autophagy refers to deterioration of nonfunctional proteins and organelles [44, 120, 121] and apoptosis to the programmed death of damaged cells in order to ensure overall cellularity survival and tissue homeostasis [122, 123]. However, with age, complex changes in the immune system take place and both procedures are found reduced [57, 58]. Accumulating evidence indicates that advancing age entails a dysregulation of all homeostatic systems enhancing inflammatory and oxidative phenotype [60, 65, 124] genomic instability, telomere damage, and stress-related p53/p21 and p16/pRB cellular pathways. In consequence, cellular senescence, a state of long-term cycle arrest that eliminates cellular functionality and regenerative potential, occurs [125–129].

Senescent cells (endothelial and smooth muscle cells) are directly related with pathogenic mechanisms of vascular dysfunction in multiple ways. They display reduced proliferative potential, impaired microangiogenesis, and multiple autocrine and paracrine actions known as SASP (senescence-associated secretory phenotype). The generated substances have proinflammatory and oxidant effects on neighboring cells deteriorating vascular functionality. Consistently, senolytic therapies seem to decelerate aging complications and endothelial dysfunction [130–134].

3.4. Extracellular Matrix Changes. All mechanisms, described above to affect intracellular and cellular components of the arterial wall, equally affect the extracellular matrix, which displays characteristic changes with advancing age, such as elastin fragmentation, collagen accumulation, and matrix protein cross-linking. Elastolysis and fibrosis derive from the actions of metalloproteinases (MMP-2 and MMP-9) and cytokine cascades (tumor growth factor β 1) as well as from other senescence-related pathways [135–137]. Metalloproteinases' imbalance then results in smooth muscle cells' migration, arterial remodelling, and formation of atherosclerotic lesions [135–137].

This stiffening of the extracellular matrix contributes to the disruption of normal cell-to-cell junctions within the endothelial monolayer. As a result, endothelial permeability is increased, allowing lipoproteins and immune cells to infiltrate the intima. These inflammatory components in combination with endothelial cellular debris initiate the formation of an atherosclerotic lesion, establishing the starting point of atheromatous disease [27, 28, 74, 81].

4. Conclusions and Perspectives

Carotid atheromatous disease and stenosis is a major factor of mortality among the elderly [3, 20, 138]. Advancing age is linearly associated with arterial stiffness and strongly associated with thickening of the vascular wall as assessed with cIMT and pulse wave velocity [21–25]. Alterations involving

CA diameter and tortuosity increase along with age, while histological findings reveal consistent structural deformation [21–26, 139, 140]. Moreover, atheromatous plaques develop with their score increasing along age [25]. The aforementioned changes are explained by various processes at the cellular level including loss of elastic vessel wall components, chronic inflammation, oxidative stress, senescence, and loss of cellular homeostasis. Multiple regulatory molecules are implicated interacting with each other [25, 27, 138, 140]. Moreover, coexistence of aging with other risk factors, such as DM, has cumulative deteriorating effects on the vascular wall [141–145]. Elucidation of aging-induced pathways has expanded our perspective on the relationship between aging and CD [141–143]. The immersion of the field of senescence has also added valuable knowledge towards a deeper understanding of the pathogenic mechanisms [146–149]. Deciphering “vascular senescence” has led to the identification of intracellular targets for the development of novel therapies and new prevention and diagnostic strategies [150–154].

Disclosure

All authors are responsible for all aspects of the work.

Conflicts of Interest

The authors declare that no conflicts of interest exist.

Authors' Contributions

All authors approved the final version of the manuscript.

Supplementary Materials

This includes a figure summarizing vascular ageing mechanisms and procedures that are described in the manuscript. (*Supplementary Materials*)

References

- [1] T. Hirata, Y. Arai, M. Takayama, Y. Abe, K. Ohkuma, and T. Takebayashi, “Carotid plaque score and risk of cardiovascular mortality in the oldest old: results from the TOOTH study,” *Journal of Atherosclerosis and Thrombosis*, vol. 25, no. 1, pp. 55–64, 2018.
- [2] C. Carallo, C. Tripolino, M. S. De Franceschi, C. Irace, X. Y. Xu, and A. Gnasso, “Carotid endothelial shear stress reduction with aging is associated with plaque development in twelve years,” *Atherosclerosis*, vol. 251, pp. 63–69, 2016.
- [3] M. E. Rubio-Ruiz, I. Pérez-Torres, M. E. Soto, G. Pastelín, and V. Guarner-Lans, “Aging in blood vessels. Medicinal agents for systemic arterial hypertension in the elderly,” *Ageing Research Reviews*, vol. 18, pp. 132–147, 2014.
- [4] C. Franceschi, P. Garagnani, P. Parini, C. Giuliani, and A. Santoro, “Inflammaging: a new immune-metabolic viewpoint for age-related diseases,” *Nature Reviews Endocrinology*, vol. 14, no. 10, pp. 576–590, 2018.
- [5] E. G. Lakatta and D. Levy, “Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises,” *Circulation*, vol. 107, no. 1, pp. 139–146, 2003.

- [6] F. Paneni, F. Cosentino, and F. Cosentino, "Risk stratification," *Diabetes and Cardiovascular Disease*, vol. 128, no. 2, pp. 69–83, 2015.
- [7] J. Wu, S. Xia, B. Kalionis, W. Wan, and T. Sun, "The role of oxidative stress and inflammation in cardiovascular aging," *BioMed Research International*, vol. 2014, Article ID 615312, 13 pages, 2014.
- [8] V. Kotsis, S. Stabouli, I. Karafillis, and P. Nilsson, "Early vascular aging and the role of central blood pressure," *Journal of Hypertension*, vol. 29, no. 10, pp. 1847–1853, 2011.
- [9] W. Wen, R. Luo, X. Tang et al., "Age-related progression of arterial stiffness and its elevated positive association with blood pressure in healthy people," *Atherosclerosis*, vol. 238, no. 1, pp. 147–152, 2015.
- [10] C. M. McEniery, H. I. R. Yasmin, I. R. Hall, A. Qasem, I. B. Wilkinson, and J. R. Cockcroft, "Normal vascular aging: differential effects on wave reflection and aortic pulse wave velocity," *Journal of the American College of Cardiology*, vol. 46, no. 9, pp. 1753–1760, 2005.
- [11] A. Scuteri, M. Tesouro, and S. Rizza, "Endothelial function and arterial stiffness in normotensive normoglycemic first-degree relatives of diabetic patients are independent of the metabolic syndrome," *Nutrition, Metabolism and Cardiovascular Diseases*, vol. 18, no. 5, pp. 349–356, 2008.
- [12] C. Steyers and F. Miller, "Endothelial dysfunction in chronic inflammatory diseases," *International Journal of Molecular Sciences*, vol. 15, no. 7, pp. 11324–11349, 2014.
- [13] R. L. Matz and R. Andriantsitohaina, "Age-related endothelial dysfunction," *Drugs & Aging*, vol. 20, no. 7, pp. 527–550, 2003.
- [14] S. Dal-Ros, C. Bronner, C. Auger, and V. B. Schini-Kerth, "Red wine polyphenols improve an established aging-related endothelial dysfunction in the mesenteric artery of middle-aged rats: role of oxidative stress," *Biochemical and Biophysical Research Communications*, vol. 419, no. 2, pp. 381–387, 2012.
- [15] R. Brandes, I. Fleming, and R. Busse, "Endothelial aging," *Cardiovascular Research*, vol. 66, no. 2, pp. 286–294, 2005.
- [16] M. E. Assar, J. Angulo, and L. Rodríguez-Mañás, "Diabetes and ageing-induced vascular inflammation," *The Journal of Physiology*, vol. 594, no. 8, pp. 2125–2146, 2016.
- [17] N. Toda, "Age-related changes in endothelial function and blood flow regulation," *Pharmacology & Therapeutics*, vol. 133, no. 2, pp. 159–176, 2012.
- [18] A. E. Walker, R. E. Kaplon, G. L. Pierce, M. J. Nowlan, and D. R. Seals, "Prevention of age-related endothelial dysfunction by habitual aerobic exercise in healthy humans: possible role of nuclear factor κ B," *Clinical Science*, vol. 127, no. 11, pp. 645–654, 2014.
- [19] A. Viridis, L. Ghiadoni, C. Giannarelli, and S. Taddei, "Endothelial dysfunction and vascular disease in later life," *Maturitas*, vol. 67, no. 1, pp. 20–24, 2010.
- [20] S. J. Jeon, H. S. Kwak, and G. H. Chung, "Widening and rotation of carotid artery with age: geometric approach," *Journal of Stroke and Cerebrovascular Diseases*, vol. 27, no. 4, pp. 865–870, 2018.
- [21] O. van Oostrom, E. Velema, A. H. Schoneveld et al., "Age-related changes in plaque composition," *Cardiovascular Pathology*, vol. 14, no. 3, pp. 126–134, 2005.
- [22] A. V. Kamenskiy, I. I. Pipinos, J. S. Carson, J. N. MacTaggart, and B. T. Baxter, "Age and disease-related geometric and structural remodeling of the carotid artery," *Journal of Vascular Surgery*, vol. 62, no. 6, pp. 1521–1528, 2015.
- [23] I. C. L. van den Munckhof, H. Jones, M. T. E. Hopman et al., "Relation between age and carotid artery intima-medial thickness: a systematic review," *Clinical Cardiology*, vol. 41, no. 5, pp. 698–704, 2018.
- [24] A. J. Rosenberg, A. D. Lane-Cordova, S. O. Wee et al., "Healthy aging and carotid performance: strain measures and β -stiffness index," *Hypertension Research*, vol. 41, no. 9, pp. 748–755, 2018.
- [25] J. Pelisek, H. Wendorff, C. Wendorff, A. Kuehnl, and H.-H. Eckstein, "Age-associated changes in human carotid atherosclerotic plaques," *Annals of Medicine*, vol. 48, no. 7, pp. 541–551, 2016.
- [26] H. Grufman, A. Schiopu, A. Edsfieldt et al., "Evidence for altered inflammatory and repair responses in symptomatic carotid plaques from elderly patients," *Atherosclerosis*, vol. 237, no. 1, pp. 177–182, 2014.
- [27] R. B. Singh, S. A. Mengi, Y. J. Xu, A. S. Arneja, and N. S. Dhalla, "Pathogenesis of atherosclerosis: a multifactorial process," *Experimental and Clinical Cardiology*, vol. 7, no. 1, pp. 40–53, 2002.
- [28] J. E. Deanfield, J. P. Halcox, and T. J. Rabelink, "Endothelial function and dysfunction," *Circulation*, vol. 115, no. 10, pp. 1285–1295, 2007.
- [29] S. Godo and H. Shimokawa, "Endothelial functions," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 37, no. 9, pp. e108–e114, 2017.
- [30] Z. S. Katusic, "Mechanisms of endothelial dysfunction induced by aging," *Circulation Research*, vol. 101, no. 7, pp. 640–641, 2007.
- [31] M. Tesouro, A. Mauriello, V. Rovella et al., "Arterial ageing: from endothelial dysfunction to vascular calcification," *Journal of Internal Medicine*, vol. 281, no. 5, pp. 471–482, 2017.
- [32] M. H. M. Bach, E. Sadoun, and M. J. Reed, "Defects in activation of nitric oxide synthases occur during delayed angiogenesis in aging," *Mechanisms of Ageing and Development*, vol. 126, no. 4, pp. 467–473, 2005.
- [33] B. van der Loo, R. Labugger, and J. N. Skepper, "Enhanced peroxynitrite formation is associated with vascular aging," *Journal of Experimental Medicine*, vol. 192, no. 12, pp. 1731–1744, 2000.
- [34] J. Tian, Z. Yan, and Y. Wu, "Inhibition of iNOS protects endothelial-dependent vasodilation in aged rats," *Acta Pharmacologica Sinica*, vol. 31, no. 10, pp. 1324–1328, 2010.
- [35] J. H. Kim, L. J. Bugaj, Y. J. Oh et al., "Arginase inhibition restores NOS coupling and reverses endothelial dysfunction and vascular stiffness in old rats," *Journal of Applied Physiology*, vol. 107, no. 4, pp. 1249–1257, 2009.
- [36] Y. Shi, R. Y. Man, and P. M. Vanhoutte, "Two isoforms of cyclooxygenase contribute to augmented endothelium-dependent contractions in femoral arteries of 1-year-old rats," *Acta Pharmacologica Sinica*, vol. 29, no. 2, pp. 185–192, 2008.
- [37] M. Félétou, Y. Huang, and P. M. Vanhoutte, "Endothelium-mediated control of vascular tone: COX-1 and COX-2 products," *British Journal of Pharmacology*, vol. 164, no. 3, pp. 894–912, 2011.
- [38] Z. Barrett-O'Keefe, S. J. Ives, J. D. Trinity, G. Morgan, and M. J. Rossman, "Endothelin-A-mediated vasoconstriction during exercise with advancing age," *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 70, no. 5, pp. 554–565, 2015.
- [39] M. Á. de Sotomayor, C. Pérez-Guerrero, and M. D. Herrera, "Improvement of age-related endothelial dysfunction by

- simvastatin: effect on NO and COX pathways," *British Journal of Pharmacology*, vol. 146, no. 8, pp. 1130–1138, 2005.
- [40] R. L. Matz, M. A. de Sotomayor, C. Schott, J.-C. Stoclet, and R. Andriantsitohaina, "Vascular bed heterogeneity in age-related endothelial dysfunction with respect to NO and eicosanoids," *British Journal of Pharmacology*, vol. 131, no. 2, pp. 303–311, 2000.
- [41] W. G. Schrage, J. H. Eisenach, and M. J. Joyner, "Ageing reduces nitric-oxide- and prostaglandin-mediated vasodilatation in exercising humans," *The Journal of Physiology*, vol. 579, no. 1, pp. 227–236, 2007.
- [42] E. Cevenini, D. Monti, and C. Franceschi, "Inflamm-aging," *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 16, no. 1, pp. 14–20, 2013.
- [43] L. Ferrucci, A. Corsi, F. Lauretani et al., "The origins of age-related proinflammatory state," *Blood*, vol. 105, no. 6, pp. 2294–2299, 2005.
- [44] L. A. Lesniewski, J. R. Durrant, M. L. Connell et al., "Aerobic exercise reverses arterial inflammation with aging in mice," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 301, no. 3, pp. H1025–H1032, 2011.
- [45] T. J. LaRocca, G. D. Henson, A. Thorburn, A. L. Sindler, G. L. Pierce, and D. R. Seals, "Translational evidence that impaired autophagy contributes to arterial ageing," *The Journal of Physiology*, vol. 590, no. 14, pp. 3305–3316, 2012.
- [46] M. Michaud, L. Balardy, G. Moulis et al., "Proinflammatory cytokines, aging, and age-related diseases," *Journal of the American Medical Directors Association*, vol. 14, no. 12, pp. 877–882, 2013.
- [47] E. A. Miles, D. Rees, T. Banerjee et al., "Age-related increases in circulating inflammatory markers in men are independent of BMI, blood pressure and blood lipid concentrations," *Atherosclerosis*, vol. 196, no. 1, pp. 298–305, 2008.
- [48] H. Zhang, Y. Park, J. Wu et al., "Role of TNF- α in vascular dysfunction," *Clinical Science*, vol. 116, no. 3, pp. 219–230, 2009.
- [49] K. L. MacNaul and N. I. Hutchinson, "Differential expression of iNOS and cNOS mRNA in human vascular smooth muscle cells and endothelial cells under normal and inflammatory conditions," *Biochemical and Biophysical Research Communications*, vol. 196, no. 3, pp. 1330–1334, 1993.
- [50] W. M. Freitas, L. A. Quaglia, S. N. Santos et al., "Association of systemic inflammatory activity with coronary and carotid atherosclerosis in the very elderly," *Atherosclerosis*, vol. 216, no. 1, pp. 212–216, 2011.
- [51] R. Kalani, S. Judge, C. Carter, M. Pahor, and C. Leeuwenburgh, "Effects of caloric restriction and exercise on age-related, chronic inflammation assessed by c-reactive protein and interleukin-6," *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 61, no. 3, pp. 211–217, 2006.
- [52] T. Pischon, C. J. Girman, G. S. Hotamisligil, N. Rifai, F. B. Hu, and E. B. Rimm, "Plasma adiponectin levels and risk of myocardial infarction in men," *JAMA*, vol. 291, no. 14, pp. 1730–1737, 2004.
- [53] N. Sattar, G. Wannamethee, N. Sarwar et al., "Adiponectin and coronary heart disease," *Circulation*, vol. 114, no. 7, pp. 623–629, 2006.
- [54] J. R. Kizer, D. Benkeser, A. M. Arnold et al., "Associations of total and high-molecular-weight adiponectin with all-cause and cardiovascular mortality in older persons," *Circulation*, vol. 126, no. 25, pp. 2951–2961, 2012.
- [55] D. A. Kinzenbaw, Y. Chu, R. A. Peña Silva, S. P. Didion, and F. M. Faraci, "Interleukin-10 protects against aging-induced endothelial dysfunction," *Physiological Reports*, vol. 1, no. 6, p. e00149, 2013.
- [56] S. K. Verma, V. N. S. Garikipati, P. Krishnamurthy et al., "IL-10 accelerates re-endothelialization and inhibits post-injury intimal hyperplasia following carotid artery denudation," *PLoS One*, vol. 11, no. 1, Article ID e0147615, 2016.
- [57] C. Weber, A. Zernecke, and P. Libby, "The multifaceted contributions of leukocyte subsets to atherosclerosis: lessons from mouse models," *Nature Reviews Immunology*, vol. 8, no. 10, pp. 802–815, 2008.
- [58] N. Alberts-Grill, A. Rezvan, D. J. Son et al., "Dynamic immune cell accumulation during flow-induced atherogenesis in mouse carotid artery," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 32, no. 3, pp. 623–632, 2012.
- [59] Y. Zou, S. Yoon, K. J. Jung et al., "Upregulation of aortic adhesion molecules during aging," *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 61, no. 3, pp. 232–244, 2006.
- [60] A. Csiszar, M. Wang, E. G. Lakatta, and Z. Ungvari, "Inflammation and endothelial dysfunction during aging: role of NF- κ B," *Journal of Applied Physiology*, vol. 105, no. 4, pp. 1333–1341, 2008.
- [61] T. Collins and M. I. Cybulsky, "NF- κ B: pivotal mediator or innocent bystander in atherogenesis?" *Journal of Clinical Investigation*, vol. 107, no. 3, pp. 255–264, 2001.
- [62] A. J. Donato, A. D. Black, K. L. Jablonski, L. B. Gano, and D. R. Seals, "Aging is associated with greater nuclear NF κ B, reduced I κ B α , and increased expression of proinflammatory cytokines in vascular endothelial cells of healthy humans," *Aging Cell*, vol. 7, no. 6, pp. 805–812, 2008.
- [63] L. Song, C. Lin, H. Gong et al., "miR-486 sustains NF- κ B activity by disrupting multiple NF- κ B-negative feedback loops," *Cell Research*, vol. 23, no. 2, pp. 274–289, 2013.
- [64] A. Salminen, J. Huuskonen, J. Ojala, A. Kauppinen, K. Kaarniranta, and T. Suuronen, "Activation of innate immunity system during aging: NF- κ B signaling is the molecular culprit of inflamm-aging," *Ageing Research Reviews*, vol. 7, no. 2, pp. 83–105, 2008.
- [65] A. Csiszar, Z. Ungvari, A. Koller, J. G. Edwards, and G. Kaley, "Aging-induced proinflammatory shift in cytokine expression profile in rat coronary arteries," *The FASEB Journal*, vol. 17, no. 9, pp. 1183–1185, 2003.
- [66] Z. Ungvari, A. Csiszar, and G. Kaley, "Vascular inflammation in aging," *Herz*, vol. 29, no. 8, pp. 733–740, 2004.
- [67] W.-S. Jeong, I.-W. Kim, R. Hu, and A.-N. T. Kong, "Modulatory properties of various natural chemopreventive agents on the activation of NF- κ B signaling pathway," *Pharmaceutical Research*, vol. 21, no. 4, pp. 661–670, 2004.
- [68] C. Xu, G. Shen, C. Chen, C. Gélinas, and A.-N. T. Kong, "Suppression of NF- κ B and NF- κ B-regulated gene expression by sulforaphane and PEITC through I κ B α , IKK pathway in human prostate cancer PC-3 cells," *Oncogene*, vol. 24, no. 28, pp. 4486–4495, 2005.
- [69] M. El Assar, J. Angulo, and L. Rodríguez-Mañas, "Oxidative stress and vascular inflammation in aging," *Free Radical Biology and Medicine*, vol. 65, pp. 380–401, 2013.
- [70] J. S. Tilstra, A. R. Robinson, J. Wang et al., "NF- κ B inhibition delays DNA damage-induced senescence and aging in mice," *Journal of Clinical Investigation*, vol. 122, no. 7, pp. 2601–2612, 2012.
- [71] Z. Ungvari, L. Bailey-Downs, T. Gautam et al., "Age-associated vascular oxidative stress, Nrf2 dysfunction, and NF- κ B activation in the nonhuman primate *Macaca mulatta*," *The*

- Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 66A, no. 8, pp. 866–875, 2011.
- [72] F. Mármol, J. Sánchez, D. López et al., “Loss of adaptation to oxidative stress as a mechanism for aortic damage in aging rats,” *Journal of Physiology and Biochemistry*, vol. 63, no. 3, pp. 239–247, 2007.
- [73] H. Cai and D. G. Harrison, “Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress,” *Circulation Research*, vol. 87, no. 10, pp. 840–844, 2000.
- [74] I. Peluso, G. Morabito, L. Urban, F. Ioannone, and M. Serafi, “Oxidative stress in atherosclerosis development: the central role of LDL and oxidative burst,” *Endocrine, Metabolic & Immune Disorders-Drug Targets*, vol. 12, no. 4, pp. 351–360, 2012.
- [75] K. K. Griendling, D. Sorescu, and M. Ushio-Fukai, “NAD(P)H oxidase,” *Circulation Research*, vol. 86, no. 5, pp. 494–501, 2000.
- [76] A. V. R. Santhanam, L. V. d’Uscio, L. A. Smith, and Z. S. Katusic, “Uncoupling of eNOS causes superoxide anion production and impairs NO signaling in the cerebral microvessels of hph-1 mice,” *Journal of Neurochemistry*, vol. 122, no. 6, pp. 1211–1218, 2012.
- [77] A. E. Vendrov, K. C. Vendrov, A. Smith et al., “NOX4 NADPH oxidase-dependent mitochondrial oxidative stress in aging-associated cardiovascular disease,” *Antioxidants & Redox Signaling*, vol. 23, no. 18, pp. 1389–1409, 2015.
- [78] D. D. Lund, Y. Chu, J. D. Miller, and D. D. Heistad, “Protective effect of extracellular superoxide dismutase on endothelial function during aging,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 296, no. 6, pp. H1920–H1925, 2009.
- [79] X. Gong, Y. Ma, Y. Ruan, G. Fu, and S. Wu, “Long-term atorvastatin improves age-related endothelial dysfunction by ameliorating oxidative stress and normalizing eNOS/iNOS imbalance in rat aorta,” *Experimental Gerontology*, vol. 52, pp. 9–17, 2014.
- [80] D. N. Meijles and P. J. Pagano, “Nox and inflammation in the vascular adventitia,” *Hypertension*, vol. 67, no. 1, pp. 14–19, 2011.
- [81] J. Huynh, N. Nishimura, K. Rana et al., “Age-related intimal stiffening enhances endothelial permeability and leukocyte transmigration,” *Science Translational Medicine*, vol. 3, no. 112, Article ID 112ra122, 2011.
- [82] S. L. Ameres and P. D. Zamore, “Diversifying microRNA sequence and function,” *Nature Reviews Molecular Cell Biology*, vol. 14, no. 8, pp. 475–488, 2013.
- [83] L.-H. Chen, G.-Y. Chiou, Y.-W. Chen, H.-Y. Li, and S.-H. Chiou, “MicroRNA and aging: a novel modulator in regulating the aging network,” *Ageing Research Reviews*, vol. 9, no. 1, pp. S59–S66, 2010.
- [84] F. Olivieri, R. Lazzarini, R. Recchioni et al., “MiR-146a as marker of senescence-associated pro-inflammatory status in cells involved in vascular remodelling,” *Age*, vol. 35, no. 4, pp. 1157–1172, 2013.
- [85] F. Olivieri, M. R. Rippo, A. D. Procopio, and F. Fazioli, “Circulating inflamma-miRs in aging and age-related diseases,” *Frontiers in Genetics*, vol. 4, p. 2013.
- [86] R. A. Boon, T. Seeger, S. Heydt et al., “MicroRNA-29 in aortic dilation: implications for aneurysm formation,” *Circulation Research*, vol. 109, no. 10, pp. 1115–1119, 2011.
- [87] R. Menghini, R. Stöhr, and M. Federici, “MicroRNAs in vascular aging and atherosclerosis,” *Ageing Research Reviews*, vol. 17, pp. 68–78, 2014.
- [88] M. P. Gantier, H. J. Stunden, C. E. McCoy et al., “A miR-19 regulon that controls NF- κ B signaling,” *Nucleic Acids Research*, vol. 40, no. 16, pp. 8048–8058, 2012.
- [89] N. Noren Hooten, K. Abdelmohsen, M. Gorospe, N. Ejiogu, A. B. Zonderman, and M. K. Evans, “microRNA expression patterns reveal differential expression of target genes with age,” *PLoS One*, vol. 5, no. 5, Article ID e10724, 2010.
- [90] Q. Wang, S. N. Bozack, Y. Yan, M. E. Boulton, M. B. Grant, and J. V. Busik, “Regulation of retinal inflammation by rhythmic expression of MiR-146a in diabetic retina,” *Investigative Ophthalmology & Visual Science*, vol. 55, no. 6, pp. 3986–3994, 2014.
- [91] I. Badi, I. Burba, C. Ruggeri et al., “MicroRNA-34a induces vascular smooth muscle cells senescence by SIRT1 down-regulation and promotes the expression of age-associated pro-inflammatory secretory factors,” *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 70, no. 11, pp. 1304–1311, 2015.
- [92] D. Bhaumik, G. K. Scott, S. Schokrpur et al., “MicroRNAs miR-146a/b negatively modulate the senescence-associated inflammatory mediators IL-6 and IL-8,” *Ageing*, vol. 1, no. 4, pp. 402–411, 2009.
- [93] H. S. Cheng, M.-S. b. Njock, N. Khyzha, L. T. Dang, and J. E. Fish, “Noncoding RNAs regulate NF- κ B signaling to modulate blood vessel inflammation,” *Frontiers in Genetics*, vol. 5, p. 422, 2014.
- [94] T. Staszal, B. Zapała, A. Polus et al., “Role of microRNAs in endothelial cell pathophysiology,” *Polish Archives of Internal Medicine*, vol. 121, no. 10, pp. 361–367, 2011.
- [95] Y. Zhao, K. K. Ting, J. Li et al., “Targeting vascular endothelial-cadherin in tumor-associated blood vessels promotes t-cell-mediated immunotherapy,” *Cancer Research*, vol. 77, no. 16, pp. 4434–4447, 2017.
- [96] N. Kourtis and N. Tavernarakis, “Cellular stress response pathways and ageing: intricate molecular relationships,” *The EMBO Journal*, vol. 30, no. 13, pp. 2520–2531, 2011.
- [97] S. Rahman and R. Islam, “Mammalian sirt1: insights on its biological functions,” *Cell Communication and Signalling*, vol. 9, no. 1, p. 11, 2011.
- [98] A. M. Thompson, R. Wagner, and E. M. Rzczidlo, “Age-related loss of SirT1 expression results in dysregulated human vascular smooth muscle cell function,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 307, no. 4, pp. H533–H541, 2014.
- [99] H.-C. Chang and L. Guarente, “Sirt1 and other sirtuins in metabolism,” *Trends in Endocrinology & Metabolism*, vol. 25, no. 3, pp. 138–145, 2014.
- [100] F. Yeung, J. E. Hoberg, C. S. Ramsey et al., “Modulation of NF- κ B-dependent transcription and cell survival by the SIRT1 deacetylase,” *The EMBO Journal*, vol. 23, no. 12, pp. 2369–2380, 2004.
- [101] A. J. Donato, K. A. Magerko, B. R. Lawson, J. R. Durrant, L. A. Lesniewski, and D. R. Seals, “SIRT-1 and vascular endothelial dysfunction with ageing in mice and humans,” *The Journal of Physiology*, vol. 589, no. 18, pp. 4545–4554, 2011.
- [102] S. Pillarisetti, “A review of sirt1 and sirt1 modulators in cardiovascular and metabolic diseases,” *Recent Patents on Cardiovascular Drug Discovery*, vol. 3, no. 3, pp. 156–164, 2008.
- [103] G. Boily, E. L. Seifert, L. Bevilacqua et al., “Sirt1 regulates energy metabolism and response to caloric restriction in mice,” *PLoS One*, vol. 3, no. 3, Article ID e1759, 2008.

- [104] Y. Tang, J. Xu, W. Qu et al., "Resveratrol reduces vascular cell senescence through attenuation of oxidative stress by SIRT1/NADPH oxidase-dependent mechanisms," *The Journal of Nutritional Biochemistry*, vol. 23, no. 11, pp. 1410–1416, 2012.
- [105] L. B. Gano, A. J. Donato, H. M. Pasha, C. M. Hearon, A. L. Sindler, and D. R. Seals, "The SIRT1 activator SRT1720 reverses vascular endothelial dysfunction, excessive superoxide production, and inflammation with aging in mice," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 307, no. 12, pp. H1754–H1763, 2014.
- [106] R. Liu, H. Liu, Y. Ha, R. G. Tilton, and W. Zhang, "Oxidative stress induces endothelial cell senescence via downregulation of Sirt6," *BioMed Research International*, vol. 2014, Article ID 902842, 13 pages, 2014.
- [107] Y. Kanfi, S. Naiman, G. Amir et al., "The sirtuin SIRT6 regulates lifespan in male mice," *Nature*, vol. 483, no. 7388, pp. 218–221, 2012.
- [108] Y. Kanfi, R. Shalman, V. Peshti et al., "Regulation of SIRT6 protein levels by nutrient availability," *FEBS Letters*, vol. 582, no. 5, pp. 543–548, 2008.
- [109] S. Xu, M. Yin, M. Koroleva et al., "SIRT6 protects against endothelial dysfunction and atherosclerosis in mice," *Aging*, vol. 8, no. 5, pp. 1064–1078, 2016.
- [110] D. B. Lombard, B. Schwer, F. W. Alt, and R. Mostoslavsky, "SIRT6 in DNA repair, metabolism and ageing," *Journal of Internal Medicine*, vol. 263, no. 2, pp. 128–141, 2008.
- [111] A. Cardus, A. K. Uryga, G. Walters, and J. D. Erusalimsky, "SIRT6 protects human endothelial cells from DNA damage, telomere dysfunction, and senescence," *Cardiovascular Research*, vol. 97, no. 3, pp. 571–579, 2013.
- [112] K. N. Lewis, J. Mele, J. D. Hayes, and R. Buffenstein, "Nrf2, a guardian of healthspan and gatekeeper of species longevity," *Integrative and Comparative Biology*, vol. 50, no. 5, pp. 829–843, 2010.
- [113] K. N. Lewis, E. Wason, Y. H. Edrey, D. M. Kristan, E. Nevo, and R. Buffenstein, "Regulation of Nrf2 signaling and longevity in naturally long-lived rodents," *Proceedings of the National Academy of Sciences*, vol. 112, no. 12, pp. 3722–3727, 2015.
- [114] N. Wakabayashi, S. L. Slocum, J. J. Skoko, S. Shin, and T. W. Kensler, "When NRF2 talks, who's listening?" *Antioxidants & Redox Signaling*, vol. 13, no. 11, pp. 1649–1663, 2010.
- [115] L. Kuang, J. Feng, G. He, and T. Jing, "Knockdown of Nrf2 inhibits the angiogenesis of rat cardiac micro-vascular endothelial cells under hypoxic conditions," *International Journal of Biological Sciences*, vol. 9, no. 7, pp. 656–665, 2013.
- [116] Z. Chen, I.-C. Peng, W. Sun et al., "AMP-activated protein kinase functionally phosphorylates endothelial nitric oxide synthase ser633," *Circulation Research*, vol. 104, no. 4, pp. 496–505, 2009.
- [117] L. A. Lesniewski, M. C. Zigler, J. R. Durrant, A. J. Donato, and D. R. Seals, "Sustained activation of AMPK ameliorates age-associated vascular endothelial dysfunction via a nitric oxide-independent mechanism," *Mechanisms of Ageing and Development*, vol. 133, no. 5, pp. 368–371, 2012.
- [118] R. Watanabe, L. Wei, and J. Huang, "mTOR signaling, function, novel inhibitors, and therapeutic targets," *Journal of Nuclear Medicine*, vol. 52, no. 4, pp. 497–500, 2011.
- [119] Y. Zhao, M.-M. Zhao, Y. Cai et al., "Mammalian target of rapamycin signaling inhibition ameliorates vascular calcification via Klotho upregulation," *Kidney International*, vol. 88, no. 4, pp. 711–721, 2015.
- [120] T. J. LaRocca, R. A. Gioscia-Ryan, C. M. Hearon, and D. R. Seals, "The autophagy enhancer spermidine reverses arterial aging," *Mechanisms of Ageing and Development*, vol. 134, no. 7–8, pp. 314–320, 2013.
- [121] E. Morselli, M. C. Maiuri, M. Markaki et al., "Caloric restriction and resveratrol promote longevity through the sirtuin-1-dependent induction of autophagy," *Cell Death & Disease*, vol. 1, no. 1, p. e10, 2010.
- [122] S. Elmore, "Apoptosis: a review of programmed cell death," *Toxicologic Pathology*, vol. 35, no. 4, pp. 495–516, 2007.
- [123] V. O. Kaminsky and B. Zhivotovsky, "Free radicals in cross talk between autophagy and apoptosis," *Antioxidants & Redox Signaling*, vol. 21, no. 1, pp. 86–102, 2014.
- [124] R. Mostoslavsky, K. F. Chua, D. B. Lombard et al., "Genomic instability and aging-like phenotype in the absence of mammalian SIRT6," *Cell*, vol. 124, no. 2, pp. 315–329, 2006.
- [125] V. G. Gorgoulis, H. Pratsinis, P. Zacharatos et al., "p53-dependent ICAM-1 overexpression in senescent human cells identified in atherosclerotic lesions," *Laboratory Investigation*, vol. 85, no. 4, pp. 502–511, 2005.
- [126] D. J. Baker, B. G. Childs, M. Durik et al., "Naturally occurring p16Ink4a-positive cells shorten healthy lifespan," *Nature*, vol. 530, no. 7589, pp. 184–189, 2016.
- [127] C. López-Otin, M. A. Blasco, L. Partridge, M. Serrano, and G. Kroemer, "The hallmarks of aging," *Cell*, vol. 153, no. 6, pp. 1194–1217, 2013.
- [128] A. Hernandez-Segura, J. Nehme, and M. Demaria, "Hallmarks of cellular senescence," *Trends in Cell Biology*, vol. 28, no. 6, pp. 436–453, 2018.
- [129] Y. Liu, S. I. Bloom, and A. J. Donato, "The role of senescence, telomere dysfunction and shelterin in vascular aging," *Microcirculation*, vol. 26, no. 2, p. e12487, 2019.
- [130] P. Lacolley, V. Regnault, and A. P. Avolio, "Smooth muscle cell and arterial aging: basic and clinical aspects," *Cardiovascular Research*, vol. 114, no. 4, pp. 513–528, 2018.
- [131] Z. Ungvari, G. Kaley, R. de Cabo, W. E. Sonntag, and A. Csizsar, "Mechanisms of vascular aging: new perspectives," *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 65A, no. 10, pp. 1028–1041, 2010.
- [132] M. Durik, M. Kavousi, I. van der Pluijm et al., "Nucleotide excision DNA repair is associated with age-related vascular dysfunction," *Circulation*, vol. 126, no. 4, pp. 468–478, 2012.
- [133] F. Rodier, "Detection of the senescence-associated secretory phenotype (SASP)," *Methods in Molecular Biology*, vol. 965, pp. 165–173, 2013.
- [134] C. M. Roos, B. Zhang, A. K. Palmer et al., "Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice," *Aging Cell*, vol. 15, no. 5, pp. 973–977, 2016.
- [135] J. P. Spiers, E. J. Kelso, W. F. Siah et al., "Alterations in vascular matrix metalloproteinase due to ageing and chronic hypertension: effects of endothelin receptor blockade," *Journal of Hypertension*, vol. 23, no. 9, pp. 1717–1724, 2005.
- [136] M. Wang and E. G. Lakatta, "Altered regulation of matrix metalloproteinase-2 in aortic remodeling during aging," *Hypertension*, vol. 39, no. 4, pp. 865–873, 2002.
- [137] M. Milkiewicz, C. Kelland, S. Colgan, and T. L. Haas, "Nitric oxide and p38 MAP kinase mediate shear stress-dependent inhibition of MMP-2 production in microvascular endothelial cells," *Journal of Cellular Physiology*, vol. 208, no. 1, pp. 229–237, 2006.
- [138] K. Watanabe, M. Ouchi, M. Ohara et al., "Change of carotid intima-media thickness is associated with age in elderly

- Japanese patients without a history of cardiovascular disease,” *Geriatrics & Gerontology International*, vol. 15, no. 8, pp. 1023–1030, 2015.
- [139] M. Wang, J. Zhang, L.-Q. Jiang et al., “Proinflammatory profile within the grossly normal aged human aortic wall,” *Hypertension*, vol. 50, no. 1, pp. 219–227, 2007.
- [140] R. Schnabel, M. G. Larson, and J. Dupuis, “Relations of inflammatory biomarkers and common genetic variants with arterial stiffness and wave reflection,” *Hypertension*, vol. 51, no. 6, pp. 1651–1657, 2010.
- [141] S. Shinozaki, K. Chang, M. Sakai et al., “Inflammatory stimuli induce inhibitory S-nitrosylation of the deacetylase SIRT1 to increase acetylation and activation of p53 and p65,” *Science Signaling*, vol. 7, no. 351, p. ra106, 2014.
- [142] B. C. van Bussel, R. M. Henry, I. Ferreira et al., “A healthy diet is associated with less endothelial dysfunction and less low-grade inflammation over a 7-year period in adults at risk of cardiovascular disease,” *The Journal of Nutrition*, vol. 145, no. 3, pp. 532–540, 2015.
- [143] P. M. Vanhoutte and E. H. C. Tang, “Endothelium-dependent contractions: when a good guy turns bad!” *The Journal of Physiology*, vol. 586, no. 22, pp. 5295–5304, 2008.
- [144] C. M. Roos, M. Hagler, B. Zhang, E. A. Oehler, A. Arghami, and J. D. Miller, “Transcriptional and phenotypic changes in aorta and aortic valve with aging and MnSOD deficiency in mice,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 305, no. 10, pp. H1428–H1439, 2013.
- [145] F. Antonicelli, G. Bellon, L. Debelle, and W. Hornebeck, “Elastin-elastases and inflamm-aging,” *Current Topics in Developmental Biology*, vol. 79, pp. 99–155, 2007.
- [146] Y. Mikhed, A. Daiber, and S. Steven, “Mitochondrial oxidative stress, mitochondrial DNA damage and their role in age-related vascular dysfunction,” *International Journal of Molecular Sciences*, vol. 16, no. 7, pp. 15918–15953, 2015.
- [147] S. Novella, A. P. Dantas, G. Segarra et al., “Aging-related endothelial dysfunction in the aorta from female senescence-accelerated mice is associated with decreased nitric oxide synthase expression,” *Experimental Gerontology*, vol. 48, no. 11, pp. 1329–1337, 2013.
- [148] A. Csiszar, N. Labinsky, K. Smith, A. Rivera, Z. Orosz, and Z. Ungvari, “Vasculoprotective effects of anti-tumor necrosis factor- α treatment in aging,” *The American Journal of Pathology*, vol. 170, no. 1, pp. 388–398, 2007.
- [149] R. Marfella, C. D. Filippa, M. T. Laieta et al., “Effects of ubiquitin-proteasome system deregulation on the vascular senescence and atherosclerosis process in elderly patients,” *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, vol. 63, no. 2, pp. 200–203, 2008.
- [150] T. Hillen, R. Nieczaj, H. Münzberg, R. Schaub, M. Borchelt, and E. Steinhagen-Thiessen, “Carotid atherosclerosis, vascular risk profile and mortality in a population-based sample of functionally healthy elderly subjects: the Berlin ageing study,” *Journal of Internal Medicine*, vol. 247, no. 6, pp. 679–688, 2000.
- [151] M. L. Lucas, C. C. Carraro, A. Belló-Klein, A. N. Kalil, and N. Aerts, “Oxidative stress in carotid arteries of patients submitted to carotid endarterectomy. The role of aging process,” *Acta Cirurgica Brasileira*, vol. 31, no. 8, pp. 564–568, 2016.
- [152] B. S. Fleenor, K. D. Marshall, J. R. Durrant, L. A. Lesniewski, and D. R. Seals, “Arterial stiffening with ageing is associated with transforming growth factor- β 1-related changes in adventitial collagen: reversal by aerobic exercise,” *The Journal of Physiology*, vol. 588, no. 20, pp. 3971–3982, 2010.
- [153] D. R. Seals, R. E. Kaplon, R. A. Gioscia-Ryan, and T. J. LaRocca, “You’re only as old as your arteries: translational strategies for preserving vascular endothelial function with aging,” *Physiology*, vol. 29, no. 4, pp. 250–264, 2014.
- [154] M. J. Zarzuelo, R. López-Sepúlveda, M. Sánchez et al., “SIRT1 inhibits NADPH oxidase activation and protects endothelial function in the rat aorta: implications for vascular aging,” *Biochemical Pharmacology*, vol. 85, no. 9, pp. 1288–1296, 2013.