

RESEARCH ARTICLE

A Comprehensive Expression Analysis of Mucins in Appendiceal Carcinoma in a Multicenter Study: MUC3 Is a Novel Prognostic Factor

Hiroaki Shibahara¹, Michiyo Higashi^{2*}, Seiya Yokoyama², Karine Rousseau³, Iwao Kitazono², Masahiko Osako⁴, Hiroshi Shirahama⁵, Yukie Tashiro⁵, Yasuhiro Kurumiya⁶, Michihiko Narita⁷, Shingo Kuze⁸, Hiroshi Hasagawa⁹, Takehito Kato¹⁰, Hitoshi Kubota¹¹, Hideaki Suzuki¹², Toshiyuki Arai¹³, Yu Sakai¹⁴, Norihiro Yuasa¹⁵, Masahiko Fujino¹⁶, Shinji Kondo¹⁷, Yoshichika Okamoto¹⁸, Tatsuyoshi Yamamoto¹⁹, Takashi Hiromatsu²⁰, Eiji Sasaki²¹, Kazuhisa Shirai²², Satoru Kawai²³, Koutarou Hattori²⁴, Hideki Tsuji²⁵, Osamu Okochi²⁶, Masaki Sakamoto²⁷, Akinobu Kondo²⁸, Naomi Konishi²⁹, Surinder K. Batra³⁰, Suguru Yonezawa²



CrossMark
click for updates

OPEN ACCESS

Citation: Shibahara H, Higashi M, Yokoyama S, Rousseau K, Kitazono I, et al. (2014) A Comprehensive Expression Analysis of Mucins in Appendiceal Carcinoma in a Multicenter Study: MUC3 Is a Novel Prognostic Factor. PLoS ONE 9(12): e115613. doi:10.1371/journal.pone.0115613

Editor: Joy Marilyn Burchell, King's College London, United Kingdom

Received: June 1, 2014

Accepted: November 28, 2014

Published: December 31, 2014

Copyright: © 2014 Shibahara et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper.

Funding: This study was supported in part by Princess Takamatsu Cancer Research Fund (11-24319) to S. Yonezawa; by JSPS KAKENHI Grants-in-Aid for Scientific Research (B) 26290048 to S. Yonezawa and Scientific Research (C) 24590447 to M. Higashi; by the Kodama Memorial Foundation, Japan to S. Yokoyama; and by USPHS grant CA163120 from the National Institutes of Health to S. K. Batra. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

1. Department of Palliative Care, Toyota Kosei Hospital, Toyota, Japan, 2. Department of Human Pathology, Field of Oncology, Kagoshima University Graduate School of Medical and Dental Sciences, Kagoshima, Japan, 3. Wellcome Trust Centre for Cell-Matrix Research, Faculty of Life Sciences, University of Manchester, United Kingdom, 4. Department of Surgery, Kagoshima Medical Association Hospital, Kagoshima, Japan, 5. Department of Pathology, Imakiire General Hospital, Kagoshima, Japan, 6. Department of Surgery, Toyota Kosei Hospital, Toyota, Japan, 7. Department of Pathology, Toyota Kosei Hospital, Toyota, Japan, 8. Department of Surgery, Chutoen General Medical Center, Kakegawa, Japan, 9. Department of Surgery, Japanese Red Cross Nagoya Daini Hospital, Nagoya, Japan, 10. Department of Surgery, Toyohashi Municipal Hospital, Toyohashi, Japan, 11. Department of Surgery, Handa City Hospital, Handa, Japan, 12. Department of Surgery, Meijo Hospital, Nagoya, Japan, 13. Department of Surgery, Anjo Kosei Hospital, Anjo, Japan, 14. Department of Pathology, Anjo Kosei Hospital, Anjo, Japan, 15. Department of Surgery, Japanese Red Cross Nagoya Daiichi Hospital, Nagoya, Japan, 16. Department of Pathology, Japanese Red Cross Nagoya Daiichi Hospital, Nagoya, Japan, 17. Department of Surgery, Sakashita Hospital, Nakatsugawa, Japan, 18. Department of Surgery, Shizuoka Saiseikai General Hospital, Shizuoka, Japan, 19. Department of Surgery, Tokai Hospital, Nagoya, Japan, 20. Department of Surgery, Kiryu Kosei General Hospital, Kiryu, Japan, 21. Department of Surgery, Kamiida Daiichi General Hospital, Nagoya, Japan, 22. Department of Surgery, Yamashita Hospital, Ichinomiya, Japan, 23. Department of Surgery, Tsushima City Hospital, Tsushima, Japan, 24. Department of Surgery, Minami Seikyo Hospital, Nagoya, Japan, 25. Department of Surgery, Toyota Memorial Hospital, Toyota, Japan, 26. Department of Surgery, Tosei General Hospital, Seto, Japan, 27. Department of Surgery, Nagoya Tokushukai General Hospital, Kasugai, Japan, 28. Department of Surgery, Saiseikai Matsusaka General Hospital, Matsusaka, Japan, 29. Department of Surgery, Mie Prefectural General Medical Center, Yokkaichi, Japan, 30. Departments of Biochemistry and Molecular Biology, Buffett Cancer Center, Eppley Institute for Research in Cancer and Allied Diseases, University of Nebraska Medical Center, Omaha, Nebraska, United States of America

*east@m2.kufm.kagoshima-u.ac.jp

Abstract

Background: Mucins are implicated in survival in various cancers, but there have been no report addressed on survival in appendiceal carcinoma, an uncommon disease with different clinical and pathological features from those of other colon

cancers. We aimed to investigate the clinical implications of expression of mucins in appendiceal carcinoma.

Methods: Expression profiles of MUC1, MUC2, MUC3, MUC4, MUC5AC, MUC6, MUC16 and MUC17 in cancer tissue were examined by immunohistochemistry in 108 cases of surgically resected appendiceal carcinoma.

Results: The following relationships of mucins with clinicopathologic factors were identified: MUC1 with positive lymphatic invasion ($p=0.036$); MUC2 with histological type (mucinous carcinoma, $p<0.001$), superficial invasion depth ($p=0.007$), negative venous invasion ($p=0.003$), and curative resection ($p=0.019$); MUC3 with non-curative resection ($p=0.017$); MUC5AC with histological type (mucinous carcinoma, $p=0.002$), negative lymphatic invasion ($p=0.021$), and negative venous invasion ($p=0.022$); and MUC16 with positive lymph node metastasis ($p=0.035$), positive venous invasion ($p<0.05$), and non-curative resection ($p=0.035$). A poor prognosis was related to positive lymph node metastasis ($p=0.04$), positive lymphatic invasion ($p=0.02$), positive venous invasion ($p<0.001$), non-curative resection ($p<0.001$), and positive expression of MUC3 ($p=0.004$). In multivariate analysis, positive venous invasion (HR: 6.93, 95% CI: 1.93–24.96, $p=0.003$), non-curative resection (HR: 10.19, 95% CI: 3.05–34.07, $p<0.001$) and positive MUC3 expression (HR: 3.37, 95% CI: 1.13–10.03, $p=0.03$) were identified as significant independent prognostic factors in patients with appendiceal carcinoma.

Conclusions: Expression of MUC3 in appendiceal carcinoma is an independent factor for poor prognosis and a useful predictor of outcome in patients with appendiceal carcinoma after surgery.

Introduction

Appendiceal cancer is rare in the United States, with an age-adjusted incidence of 0.12 cases per 1,000,000 people per year [1], and a rate among intestinal cancers of 0.7%, compared to 1.5% for small bowel carcinoma and 97.8% for colon carcinoma in the Surveillance, Epidemiology and End Results (SEER) registry [2]. A similar rarity of appendiceal carcinoma is also found in Japan, with incidences of 0.2% in the Japanese Society for Cancer of the Colon and Rectum Registry and 0.08% in the Japanese Autopsy Annual Database of Colorectal Cancer [3]. The disease differs from cancers at other sites in the colon, with clinical presentation of acute abdominal symptoms suggestive of appendicitis [4, 5] and peritoneal mucinous carcinomatosis. The 5-year survival rate for appendiceal carcinoma after surgery is 46–64% [5–7]. Curative surgical resection is required for improving survival, and the pathological characteristics of the tumor affect prognosis. Among histological types, patients with non-mucinous carcinoma have poorer survival than those with mucinous carcinoma [5, 8], and those with signet

ring cell carcinoma also have poor survival [1]. Cases with a high histological grade have poorer survival than low grade cases [6, 7]. Thus, prognostic factors in appendiceal carcinoma have included curative resection [6], primary tumor status [6], histological type [1, 5, 6, 8], and histological grade [6, 7, 9].

Mucins are high molecular weight glycoproteins having core protein backbones by O-glycosidic linkages with oligosaccharides [10]. Eighteen core proteins for human mucins (MUC1-MUC8, MUC12, MUC13, MUC15-17, MUC19-21) have been identified. The first cloned, MUC1, has been reported to be one of the most important human tumor antigens, namely, the second ranking next to WT1 [11]. Yonezawa et al. showed that MUC1 and/or MUC4 expression is related to a poorer prognosis for various human cancers, whereas MUC2 expression is related to a better prognosis [10, 12]. Aberrant expression of MUC3, MUC4, MUC5AC and MUC6 is found in pancreatic intraepithelial neoplasia [13, 14], and MUC16 and MUC17 are expressed in pancreatobiliary and small intestinal cancers [15–17] and have high prognostic value [15, 17–19]. Mucin expression also occurs in appendiceal carcinoma [20–26], however, there is no study for the relationship between mucin expression and survival of over 100 surgically-treated patients with appendiceal carcinoma.

The aim of this study was to investigate whether expression of mucins (MUC1, MUC2, MUC3, MUC4, MUC5AC, MUC6, MUC16 and MUC17) has prognostic significance in patients with appendiceal carcinoma using surgical specimens collected from multiple centers.

Materials and Methods

Patients and Tissue Specimens

Between 1991 and 2013, 108 resected specimens of appendiceal carcinoma were collected from 23 hospitals in Japan: Toyota Kosei Hospital, Kagoshima Medical Association Hospital, Imakiire General Hospital, Chutoen General Medical Center, Japanese Red Cross Nagoya Daini Hospital, Toyohashi Municipal Hospital, Handa City Hospital, Meijo Hospital, Anjo Kosei Hospital, Japanese Red Cross Nagoya Daiichi Hospital, Sakashita Hospital, Shizuoka Saiseikai General Hospital, Tokai Hospital, Kiryu Kosei General Hospital, Kamiida Daiichi General Hospital, Yamashita Hospital, Tsushima City Hospital, Minami Seikyo Hospital, Toyota Memorial Hospital, Tosei General Hospital, Nagoya Tokushukai General Hospital, Saiseikai Matsusaka General Hospital, and Mie Prefectural General Medical Center.

This study was conducted in accordance with the guiding principles of the Declaration of Helsinki. Informed, written consent was obtained from 10 patients, and was approved by the Ethics Committees of Kagoshima-shi Medical Association Hospital (KMAH 2011-02-02), Japanese Red Cross Nagoya Daini Hospital (IRB20140128-7), Toyota Memorial Hospital (1211-4), and Saiseikai Matsusaka General Hospital (52-2013). For the other patients without informed consent, the Institutional Review Board of Toyota Kosei Hospital (22-ST04), the

Ethics Committees of Imakiire General Hospital (119-2013), Toyohashi Municipal Hospital (43-2011), Japanese Red Cross Nagoya Daiichi Hospital (26-2013), Sakashita Hospital (1-2013), Shizuoka Saiseikai General Hospital (25-3-02), Tsushima City Hospital (2013-06), Toyota Memorial Hospital (1211-4), Tosei General Hospital (420-2013), Chutoen General Medical Center, Handa City Hospital, Kiryu Kosei General Hospital, Kamiida Daiichi General Hospital, and Yamashita Hospital, and the hospital directors of Meijo Hospital, Anjo Kosei Hospital, Tokai Hospital, Minami Seikyo Hospital, Nagoya Tokushukai General Hospital, and Mie Prefectural General Medical Center (no specified number in these eleven hospitals) waived the need for written informed consent from the participants, and gave us their approval for use of the resected specimens, under the strict condition of privacy protection of the personal information of the patients.

Primary appendiceal carcinomas that were clinically and pathologically diagnosed by surgeons and pathologists were included in the study. Possible cecum cancers with invasion of the appendix, metastatic cancer to the appendix, or carcinoid of the appendix were excluded. Samples were collected from 55 males and 53 females with a mean age of 65 years (range 23–95). The surgical procedures are shown as [Table 1](#). Mucinous peritonitis was found in laparotomy in 14 cases. Of the 108 patients, 34 died, and the causes of death were the primary disease in 30, another disease in 3, and an unknown cause in 1. All specimens were fixed in formalin, embedded in paraffin and cut into 4- μ m -thick sections for immunohistochemistry (IHC), in addition to hematoxylin and eosin (HE) staining.

Immunohistochemistry

MUC1 was detected by a monoclonal antibody (MAb) DF3 (mouse IgG, Toray-Fuji Bionics, Tokyo, Japan), MUC2 by MAb Ccp58 (Novocastra Reagents, Leica Biosystems, Newcastle Upon Tyne, UK), MUC3 by MAb mMUC3-1 (generated by K. Rousseau and D. M. Swallow), MUC4 by MAb 8G7 (generated by S. K. Batra), MUC5AC by MAb CLH2 (Novocastra), MUC6 by MAb CLH5 (Novocastra), MUC16 by MAb OC125 (Acris Antibodies GmbH, Herford, Germany), and MUC17 by a polyclonal anti-human MUC17 (rabbit IgG, generated by S. K. Batra).

IHC was performed using the immunoperoxidase method. Antigen retrieval was performed using CC1 antigen retrieval buffer (pH8.5, EDTA, 100°C, 30 min, Ventana Medical Systems, Tucson, AZ, USA). Sections were incubated with a primary antibody (DF3 diluted 1:50, 37°C, 32 min; Ccp58 diluted 1:200, 37°C, 24 min; 8G7 diluted 1:3000, 37°C, 32 min; CLH2 diluted 1:100, 37°C, 24 min; CLH5 diluted 1:100, 37°C, 24 min; OC125 diluted 1: 100, 37°C, 24 min; anti-human MUC17 diluted 1: 100) in phosphate-buffered saline (PBS) pH 7.4 with 1% bovine serum albumin, and stained on a Benchmark XT automated slide stainer using a diaminobenzidine detection kit (ultraView DAB, Ventana Medical Systems). For MUC3 staining, sections were treated at 100°C for 10 min in

Table 1. Surgical Procedure.

Procedure	No. patients
Primary resection only	88
Type of colectomy	
Appendectomy	20
Resection of the cecum	3
Ileocecal resection	56
Right colectomy	3
Right hemicolectomy	6
Combined resection	
Rectosigmoid colon	1
Uterus and adnexa	1
Liver	1
Elective resection ^a	20
Type of colectomy	
Ileocecal resection	15
Right colectomy	1
Right hemicolectomy	3
Mucinous tumor resection	1
Combined resection	
Retroperitoneum, uterus, right adnexa and rectum	1
Lymph node dissection	
Performed	81
Not performed	27
Curability	
Curative resection	64
Non-curative resection	41
Unknown	3

^aElective resection after pathological diagnosis of appendiceal carcinoma using the resected specimen at the first surgery.

doi:10.1371/journal.pone.0115613.t001

0.01 M citrate buffer at pH 6.0, and then reduced with 0.01 M dithiothreitol in 0.1 M Tris/HCl buffer (pH 8.0) for 30 min at room temperature and alkylated with 0.025 M iodoacetamide in 0.1 M Tris/HCl buffer (pH 8.0) for 30 min [13, 17, 27]. They were incubated with mMUC3-1 at 4°C for 16 h and stained by avidin-biotin complex method. Reaction products were not present when hybridoma culture medium, normal mouse serum, normal rabbit serum, or PBS was used instead of primary antibodies.

Evaluation of Staining

The results were evaluated based on the percentage of positively stained carcinoma cells. Staining of the following components was evaluated: membrane and cytoplasm for MUC1 and MUC16; supranuclear area for MUC2; membrane for MUC3, cytoplasm for MUC4, MUC5AC and MUC6; and supranuclear area,

cytoplasm and membrane for MUC17. Carcinoma cells are considered to be stained positively when at least one of the components was positive. A tumor was considered positive if more than 5% of carcinoma cells were stained, based on our previous use of 5% as the cutoff for mucin expression [17, 28–33].

Statistical Analysis

Associations between mucin expression profiles and clinicopathological factors were examined by chi-square test. Postoperative survival was calculated using the Kaplan-Meier method. Differences in survival curves were compared by log-rank test. A Cox proportional hazard analysis was used to estimate hazard ratios (HRs) and corresponding 95% confidence intervals (CIs) in multivariate analysis. $P < 0.05$ was considered significant.

Results

MUC expression in carcinomas

In the 108 cases, the positive expression rates (more than 5% of carcinoma cells stained) of each mucin antigen were MUC1, 47.2% (51/108); MUC2, 71.3% (77/108); MUC3, 18.5% (20/108); MUC4, 93.5% (101/108); MUC5AC, 50.0% (54/108); MUC6, 4.6% (5/108) MUC16, 16.7% (18/108) and MUC17, 86.1% (93/108). Representative mucin expression patterns in cancer tissues are shown in Fig. 1 (MUC3) and Fig. 2 (MUC1, MUC2, MUC4, MUC5AC, MUC6, MUC16 and MUC17). In appendiceal carcinoma cells, MUC3 showed membrane expression in the cell apexes (Fig. 1A–D); MUC1 showed membrane expression (Fig. 2A–C); MUC2 showed supranuclear expression (Fig. 2D–F); MUC4 (Fig. 2G–I), MUC5AC (Fig. 2J–L), and MUC6 (Fig. 2M–O) showed cytoplasmic expression; MUC16 showed membrane expression (Fig. 2P–R), and MUC17 showed supranuclear expression (Fig. 2S–U).

Relationship of MUC Expression in Cancer Cells with Clinicopathological Features

Relationships between mucin expression and clinicopathological features are summarized in Table 2. MUC1 expression was related to lymphatic invasion (higher in positive lymphatic invasion, $p = 0.036$); MUC2 expression was related to histological type (higher for mucinous carcinoma, $p < 0.001$), invasion depth (higher in the superficial area than the muscularis propria, $p = 0.007$), venous invasion (higher for negative venous invasion, $p = 0.003$), and curability (higher in curative resection, $p = 0.019$); MUC3 expression was related to curability (higher in non-curative resection, $p = 0.017$); MUC5AC expression was related to histological type (higher in mucinous carcinoma, $p = 0.002$), lymphatic invasion (higher for negative lymphatic invasion, $p = 0.021$), and venous invasion (higher in negative venous invasion, $p = 0.022$); and MUC16 expression was related to lymph node metastasis (higher in positive lymph node metastasis, $p = 0.035$),

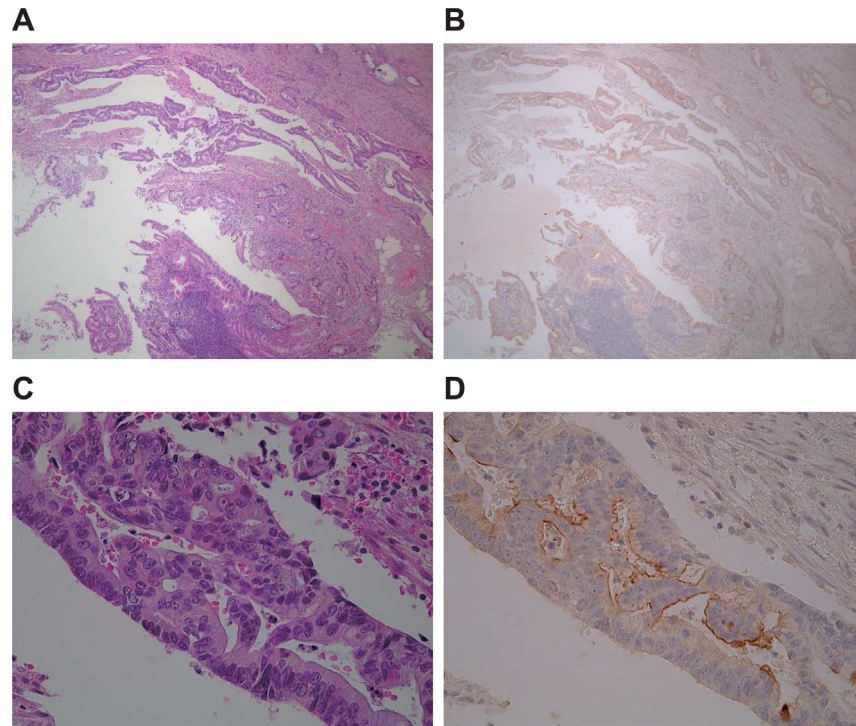


Fig. 1. Histological features of appendiceal carcinoma. (A, C) Hematoxylin and eosin stain. (B, D) Immunohistochemistry. MUC3 showed membrane expression in the cell apices in appendiceal carcinoma.

doi:10.1371/journal.pone.0115613.g001

venous invasion (higher for positive venous invasion, $p < 0.05$), and curability (higher in non-curative resection, $p = 0.035$).

Relationship of Clinicopathological Factors and Mucin Expression with Survival

The 5-year overall survival rate and median survival period were 62.4% and 2.1 years, respectively. Log-rank tests showed that positive lymph node metastasis ($p = 0.04$), positive lymphatic invasion ($p = 0.02$), positive venous invasion ($p < 0.001$), and non-curative resection ($p < 0.001$) were significantly related to a worse prognosis (Table 3). Positive expression of MUC3 ($p = 0.004$) was also significantly related to a worse prognosis (Table 3, Fig. 3), but survival was not correlated with expression of MUC1, MUC2, MUC4, MUC5AC, MUC6, MUC16 and MUC17.

Multivariate Analysis of Prognostic Factors

The above results identified lymph node metastasis, lymphatic invasion, venous invasion, curative resection and MUC3 expression as candidates for prognostic factors. In multivariate analysis using a Cox proportional hazard model, positive venous invasion (HR: 6.93, 95% CI: 1.93–24.96, $p = 0.003$), non-curative resection

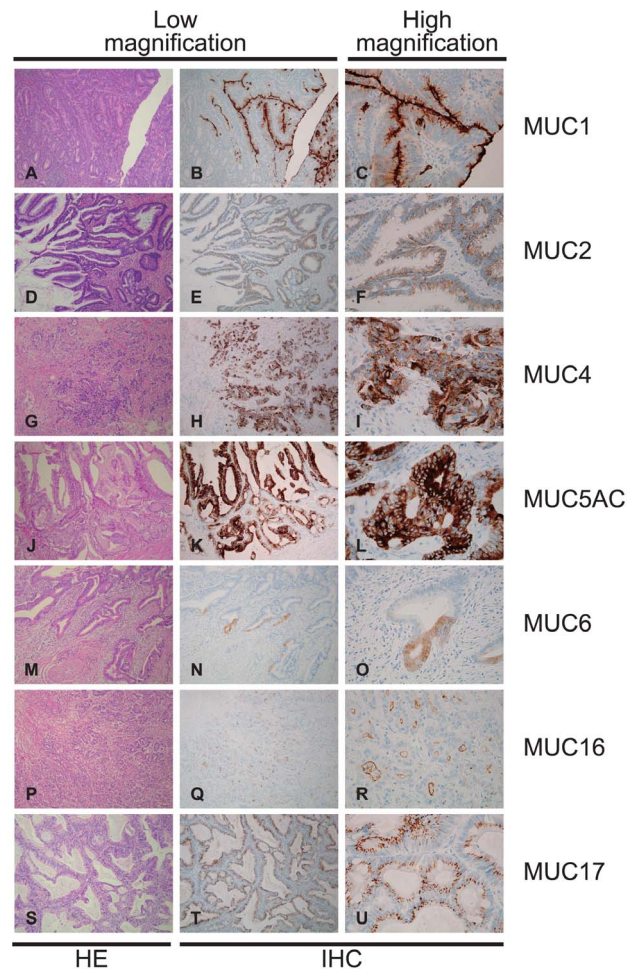


Fig. 2. In appendiceal carcinoma cells (A, D, G, J, M, P and S), MUC1 showed membrane expression (B and C); MUC2 showed supranuclear expression (E and F); MUC4 (H and I), MUC5AC (K and L) and MUC6 (N and O) showed cytoplasmic expression; MUC16 showed membrane expression (Q and R); and MUC17 (T and U) showed supranuclear expression. HE, hematoxylin and eosin stain; IHC, immunohistochemical stain.

doi:10.1371/journal.pone.0115613.g002

(HR: 10.19, 95% CI: 3.05–34.07, $p < 0.001$), and positive MUC3 expression (HR: 3.37, 95% CI: 1.13–10.03, $p = 0.03$) were identified as significant independent prognostic factors in patients with appendiceal carcinoma (Table 4).

Discussion

In this study, the rates of positive expression were MUC1, 47.2%; MUC2, 71.3%; MUC3, 18.5%; MUC4, 93.5%; MUC5AC, 50.0%; MUC6, 4.6%; and MUC16, 16.7% in 108 cases of appendiceal carcinoma. In colorectal carcinoma, these rates are MUC1, 24–32% [10, 34]; MUC2, 38% [10]; MUC3, 74% [34]; MUC4, 94% [35]; MUC5AC, 34–50% [36, 37]; MUC6, 39% [37]; and MUC16, 64% [18]. The MUC17 expression rate in colon cancer is unknown, but is lower than that in

Table 2. Summary of the Data on the Expression of MUC1, MUC2, MUC3, MUC4, MUC5AC, MUC6, MUC16 and MUC17 in Clinicopathological Features of Appendiceal Carcinoma (n = 108).

Category	MUC1			MUC2			MUC3			MUC4			P Value
	No. patients (%)	Positive	P Value	Negative	Positive	P Value	Negative	Positive	P Value	Negative	Positive		
Age (yrs)			0.483			0.132			0.517			0.41	
≥65	61 (56.5)	27 (44.3)		14 (23)	47 (77)		51 (83.6)	10 (16.4)		5 (8.2)	56 (91.8)		
<65	47 (43.5)	24 (51.1)		17 (36.2)	30 (63.8)		37 (78.7)	10 (21.3)		2 (4.3)	45 (95.7)		
Gender			0.126			0.928			0.686			0.659	
Men	55 (50.9)	22 (40)		16 (29.1)	39 (70.9)		44 (80)	11 (20)		3 (5.5)	52 (94.5)		
Women	53 (49.1)	29 (54.7)		15 (28.3)	38 (71.7)		44 (83)	9 (17)		4 (7.5)	49 (92.5)		
Histological type ^a			0.597			<0.001			0.052			0.111	
pap. well. mod	68 (63)	34 (50)		17 (25)	51 (75)		54 (79.4)	14 (20.6)		7 (10.3)	61 (89.7)		
poor. sig	19 (17.6)	7 (36.8)		14 (73.7)	5 (26.3)		19 (100)	0 (0)		0 (0)	19 (100)		
muc	21 (19.4)	10 (47.6)		0 (0)	21 (100)		15 (71.4)	6 (28.6)		0 (0)	21 (100)		
Tumor depth ^b			0.305			0.007			0.103			0.123	
m. sm. mp	26 (24.1)	10 (38.5)		2 (7.7)	24 (92.3)		24 (92.3)	2 (7.7)		0 (0)	26 (100)		
ss. se. si	82 (75.9)	41 (50)		29 (35.4)	53 (64.6)		64 (78)	18 (22)		7 (8.5)	75 (91.5)		
Lymph node metastasis ^c			0.304			0.056			0.756			0.946	
Negative	55 (51.9)	25 (45.5)		12 (21.8)	43 (78.2)		45 (81.8)	10 (18.2)		4 (7.3)	51 (92.7)		
Positive	26 (24.1)	15 (57.7)		11 (42.3)	15 (57.7)		22 (84.6)	4 (15.4)		2 (7.7)	24 (92.3)		
Lymphatic invasion			0.036			0.083			0.854			0.202	
Negative	56 (51.9)	21 (37.5)		12 (21.4)	44 (78.6)		46 (82.1)	10 (17.9)		2 (3.6)	54 (96.4)		
Positive	52 (48.1)	30 (57.7)		19 (36.5)	33 (63.5)		42 (80.8)	10 (19.2)		5 (9.6)	47 (90.4)		
Venous invasion			0.153			0.003			0.256			0.114	
Negative	75 (69.4)	32 (42.7)		15 (20)	60 (80)		59 (78.7)	16 (21.3)		3 (4)	72 (96)		
Positive	33 (30.6)	19 (57.6)		16 (48.5)	17 (51.5)		29 (87.9)	4 (12.1)		4 (12.1)	29 (87.9)		
Curability ^d			0.164			0.019			0.017			0.831	
Curative resection	64 (61)	37 (57.8)		13 (20.3)	51 (79.7)		57 (89.1)	7 (10.9)		4 (6.2)	60 (93.8)		
Non-curative resection	41 (39)	23 (56.1)		17 (41.5)	24 (58.5)		29 (70.7)	12 (29.3)		3 (7.3)	38 (92.7)		
Category	No. patients (%)	Positive	P Value	Negative	Positive	P Value	Negative	Positive	P Value	Negative	Positive	P Value	
Age (yrs)			0.846			0.092			0.931			0.409	
≥65	61 (56.5)	31 (50.8)		60 (98.4)	1 (1.6)		51 (83.6)	10 (16.4)		7 (11.5)	54 (88.5)		
<65	47 (43.5)	23 (48.9)		43 (91.5)	4 (8.5)		39 (83)	8 (17)		8 (17)	39 (83)		
Gender			0.336			0.183			0.263			0.722	
Men	55 (50.9)	30 (54.5)		51 (92.7)	4 (7.3)		48 (87.3)	7 (12.7)		7 (12.7)	48 (87.3)		
Women	53 (49.1)	24 (45.3)		52 (98.1)	1 (1.9)		42 (79.2)	11 (20.8)		8 (15.1)	45 (84.9)		
Histological type ^a			0.002			0.528			0.177			0.105	
pap. well. mod	68 (63)	34 (50)		64 (94.1)	4 (5.9)		56 (82.4)	12 (17.6)		11 (16.2)	57 (83.8)		
poor. sig	19 (17.6)	4 (21.1)		18 (94.7)	1 (5.3)		14 (73.7)	5 (26.3)		4 (21.1)	15 (78.9)		
muc	21 (19.4)	16 (76.2)		21 (100)	0 (0)		20 (95.2)	1 (4.8)		0 (0)	21 (100)		
Tumor depth ^b			0.653			0.827			0.841			0.294	
m. sm. mp	26 (24.1)	14 (53.8)		25 (96.2)	1 (3.8)		22 (84.6)	4 (15.4)		2 (7.7)	24 (92.3)		
ss. se. si	82 (75.9)	40 (48.8)		78 (95.1)	4 (4.9)		68 (82.9)	14 (17.1)		13 (15.9)	69 (84.1)		

Table 2. Cont.

Category	No. patients (%)	MUC5AC		P Value	MUC6		P Value	MUC16		P Value	MUC17		P Value
		Negative	Positive		Negative	Positive		Negative	Positive		Negative	Positive	
Lymph node metastasis ^c				0.295			0.325			0.035			0.067
Negative	55 (67.9)	27 (49.1)	28 (60.9)		53 (96.4)	2 (3.6)		50 (90.9)	5 (9.1)		6 (10.9)	49 (89.1)	
Positive	26 (32.1)	16 (61.5)	10 (38.5)		26 (100)	0 (0)		19 (73.1)	7 (26.9)		7 (26.9)	19 (73.1)	
Lymphatic invasion				0.021			0.709			0.228			0.122
Negative	56 (51.9)	22 (39.3)	34 (60.7)		53 (94.6)	3 (5.4)		49 (87.5)	7 (12.5)		5 (8.9)	51 (91.1)	
Positive	52 (48.1)	32 (61.5)	20 (38.5)		50 (96.2)	2 (3.8)		41 (76.8)	11 (21.2)		10 (19.2)	42 (80.8)	
Venous invasion				0.022			0.639			<0.05			0.392
Negative	75 (69.4)	32 (42.7)	43 (67.3)		72 (96)	3 (4)		66 (88)	9 (12)		9 (12)	66 (88)	
Positive	33 (30.6)	22 (66.7)	11 (33.3)		31 (93.9)	2 (6.1)		24 (72.7)	9 (27.3)		6 (18.2)	27 (81.8)	
Curability ^d				0.781			0.325			0.035			0.288
Curative resection	64 (61)	31 (48.4)	33 (51.6)		62 (96.9)	2 (3.1)		57 (89.1)	7 (10.9)		11 (17.2)	53 (82.8)	
Non-curative resection	41 (39)	21 (51.2)	20 (48.8)		38 (92.7)	3 (7.3)		30 (73.2)	11 (26.8)		4 (9.8)	37 (90.2)	

^apap, papillary adenocarcinoma; well, well differentiated adenocarcinoma; mod, moderately differentiated adenocarcinoma; por, poorly differentiated adenocarcinoma; sig, signet-ring cell carcinoma; muc, mucinous carcinoma.

^bm, mucosa; sm, submucosa; mp, muscularis propria; ss, subserosa, se, serosa; si, invasion to other organ.

^c27 cases without lymph node dissection were excluded.

^d3 cases with unknown details regarding curative or non-curative resection were excluded.

doi:10.1371/journal.pone.0115613.t002

Table 3. Survival in Patients with Appendiceal Carcinoma by the Log-Rank Test (n=108).

Category	No. patients (%)	5-year survival rate (%)	P Value
Age (yrs)			0.054
<65	47 (43.5)	72.4	
≥65	61 (56.5)	53.6	
Gender			0.296
Men	55 (50.9)	57.4	
Women	53 (49.1)	65.9	
Histological type ^a			0.226
pap, well, mod	68 (63)	67.7	
por, sig	19 (17.6)	48.8	
muc	21 (19.4)	60.3	
Tumor depth ^b			0.066
m, sm, mp	26 (24.1)	84	
ss, se, si	82 (75.9)	57.6	
Lymph node metastasis ^c			0.04
Negative	55 (67.9)	76.1	
Positive	26 (32.1)	47.7	
Lymphatic invasion			0.02
Negative	56 (51.9)	77.6	
Positive	52 (48.1)	47	
Venous invasion			<0.001
Negative	75 (69.4)	73.7	
Positive	33 (30.6)	35.4	
Curability ^d			<0.001
Curative resection	64 (61)	83	
Non-curative resection	41 (39)	28.5	
MUC1			0.626
Negative	57 (52.8)	63.1	
Positive	51 (47.2)	62	
MUC2			0.072
Negative	31 (28.7)	51.3	
Positive	77 (71.3)	66.5	
MUC3			0.004
Negative	88 (81.5)	69.1	
Positive	20 (18.5)	38.8	
MUC4			0.467
Negative	7 (6.5)	47.6	
Positive	101 (93.5)	63.4	
MUC5AC			0.433
Negative	54 (50)	59.1	
Positive	54 (50)	66.1	
MUC6			0.698
Negative	103 (95.4)	61.6	
Positive	5 (4.6)	75	

Table 3. Cont.

Category	No. patients (%)	5-year survival rate (%)	P Value
MUC16			0.061
Negative	90 (83.3)	65.4	
Positive	18 (16.7)	48.1	
MUC17			0.5
Negative	15 (13.9)	67.5	
Positive	93 (86.1)	61.9	

^apap, papillary adenocarcinoma; well, well differentiated adenocarcinoma; mod, moderately differentiated adenocarcinoma; por, poorly differentiated adenocarcinoma; sig, signet-ring cell carcinoma; muc, mucinous carcinoma.

^bm, mucosa; sm, submucosa; mp, muscularis propria; ss, subserosa, se, serosa; si, invasion to other organ.

^c27 cases without lymph node dissection were excluded.

^d3 cases with unknown details regarding curative or non-curative resection were excluded.

doi:10.1371/journal.pone.0115613.t003

normal epithelium [38]. In appendiceal carcinoma, MUC3, MUC6 and MUC16 had lower expression, MUC2 expression was markedly higher, and MUC1 expression was higher than the respective rates in colorectal carcinoma. These differences indicate the distinct characteristics of appendiceal carcinoma compared to other colorectal cancers. We also previously examined mucin expression in small intestinal carcinoma, and found positive expression rates of MUC1, 51.7%; MUC2, 26.7%; MUC3, 55.0%; MUC4, 51.7%; MUC5AC, 33.3%; MUC6, 10.0%; and MUC16, 8.3% (MUC17 expression was not examined) [17]. Appendiceal carcinoma was MUC1-positive in about half of the cases, similarly to small intestinal carcinoma, but other mucin profiles were different. Thus, with regard to mucin expression, appendiceal carcinoma may have a different

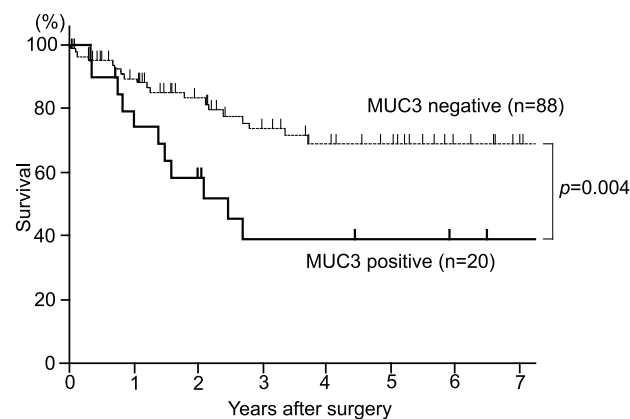


Fig. 3. Correlation between mucin expression and the cumulative survival rate. In the study of the correlation between mucin expression and the cumulative survival rate in patients with appendiceal carcinoma using the Kaplan-Meier method, the survival rate of patients with a positive expression of MUC3 were poorer than those of patients with negative expression of MUC3 ($p=0.004$).

doi:10.1371/journal.pone.0115613.g003

Table 4. Multivariate Analysis of Prognostic Factors.

Category	Hazard Ratio	95% Confidence Interval	P Value
Lymph node metastasis			0.511
Negative	1		
Positive	1.41	0.51–3.91	
Lymphatic invasion			0.488
Negative	1		
Positive	1.67	0.39–7.11	
Venous invasion			0.003
Negative	1		
Positive	6.93	1.93–24.96	
Curability			<0.001
Curative resection	1		
Non-curative resection	10.19	3.05–34.07	
MUC3			0.03
Negative	1		
Positive	3.37	1.13–10.03	

doi:10.1371/journal.pone.0115613.t004

carcinogenesis mechanism compared with other colorectal or small intestinal carcinomas.

Expression of MUC3 has been examined in malignancies of the pancreas, periampullary site, bile duct, kidney, salivary gland, lung, and breast, with examining tumor progression and prognosis [13, 39–44]. Duncan et al. [34] showed that MUC3 did not affect on survival in colorectal cancer. However, in appendiceal carcinoma, we firstly indicated that MUC3 had impact on survival.

MUC3 maps to a mucin cluster on chromosome 7q22 and is a membrane-bound mucin with tandem repeats of 17 amino acids (HSTPSFTS- SITTTETTS) [10]. IHC of MUC3 (mMUC3-1) in formalin-fixed paraffin-embedded specimens has been developed as a specific method for epitope retrieval [13, 17, 27]. We found that clear linear staining of the surface of villi in the normal mucosa of the small intestine is a good positive control for MUC3 staining [17, 27]. Other studies have used different antibodies, including 1143/B7 [34, 39, 44, 45] and M3P [40], and some have evaluated both membranous and cytoplasmic expression [34, 44, 45]. Using the 1143/B7 antibody, Aloysius et al [39] showed that MUC3 membranous expression is an independent prognostic factor in periampullary cancer. The use of different antibodies and evaluation of different expression patterns might give different results for MUC3, and the association of tumor behavior with results from each MUC3 antibody will be an interesting area for future study.

MUC3 is associated with a poor prognosis in appendiceal carcinoma, but the molecular mechanism of MUC3 in carcinogenesis is uncertain. Epigenetically, expression of MUC3A is contributed by promoter hypomethylation [46]. Cysteine-rich domains of MUC3 promote cell migration and inhibit apoptosis

[47], and the MUC3 C-terminal domain undergoes autoproteolysis at its SEA module, which maintains its availability for potentiation of signaling modulated by HER/ErbB2 phosphorylation to promote migration and invasion [48]. Enhanced MUC3 expression by a tetrameric branched peptide with a conserved TFLK motif inhibits bacteria adherence [49], and expression of MUC3 is altered in inflammatory bowel disease and correlated with disease activity and the extent of inflammation [50]. Thus, MUC3 has several potential roles in malignant and inflammatory cells and these effects might be implicated in the poor prognosis of MUC3-positive patients with appendiceal carcinoma.

Expression of MUC1, MUC2, MUC4, MUC5AC, MUC6, MUC16 and MUC17 was not related to survival in appendiceal carcinoma. MUC1 expression is related to a poor prognosis of various human neoplasms and plays an important role in tumor invasion and metastasis [10, 12], but in our cases MUC1 expression was only related to positive lymphatic invasion. Mucinous carcinoma has high MUC2 expression compared to other adenocarcinomas in the pancreas, bile duct, ovary, breast [10, 12] and colorectum [51]. MUC2 expression is also related to a better prognosis of neoplasms in the stomach, pancreas and bile duct [10, 12]. The role of MUC2 in mucinous carcinoma suggests that production of this type of mucin may act as a barrier to cancerous extension, resulting in the indolent nature of many tumors [10]. In the current study, MUC2 expression was associated with mucinous carcinoma, consistent with a previous report [51], and with superficial invasion depth, negative venous invasion, and curative resection, but not with a better prognosis in appendiceal carcinoma.

Shanmugam et al. [35] found that MUC4 expression ($\geq 75\%$) is a poor prognostic factor in colorectal cancer. However, high MUC4 expression ($\geq 75\%$) in appendiceal carcinoma was not significantly related to survival (data not shown). A cut-off value of more than 5% for MUC4 expression detected with antibody 8G7 is significantly related to survival in many tumors [30–32, 52]. Kocer et al. [36] found that MUC5AC expression is associated with a better prognosis in colorectal carcinoma, and we also found that MUC5AC expression was related to favorable clinicopathological factors such as negative lymphatic invasion and negative venous invasion. MUC6 expression is a useful marker of pancreatobiliary neoplasms [53–55], but has no relationship with clinicopathological factors or survival. MUC16 expression is a poor prognostic factor in cholangiocarcinoma and small intestinal cancer [15, 17], and was related to positive lymph node metastasis, positive venous invasion and non-curative resection in appendiceal carcinoma in the current study.

MUC17 expression is related to tumor progression in pancreatic cancer [19], but was not related to clinicopathological factors or survival in appendiceal carcinoma. MUC17 and MUC3 are similarly expressed on the apical surface of intestinal epithelia, are both present in glycocalyx, and are both located on chromosome 7q22 [56, 57]. *MUC17* and *MUC3A* both have promoter methylation sites, but those are different (-179 to $+52$ in *MUC17* and -345 to -75 in *MUC3A*) [58]. Regarding histone modification, histone H3-K9 is more highly acetylated in MUC17-positive cells, whereas H3-K9 does not play a critical role in

MUC3A regulation [58]. In the molecular structures, MUC17 and MUC3 both have an N-terminal large mucin domain, a SEA domain, a transmembrane domain, a cytoplasmic tail, and PDZ-binding motifs [59, 60], but their molecular function is different. In enterocytes, in response to carbachol, MUC17 is relocated from the apical membrane to an intracellular vesicular pool distinct from classical endosomes; this behavior is specific for MUC17, and does not occur for MUC3 [60, 61]. The current study showed a different IHC staining pattern, with MUC17 in the supranuclear area and MUC3 in the membrane, and different clinical significance. The differences in biological behavior between MUC17 and MUC3 may be due to differences in promoters and regulators, or in the structure and domains. Further studies are needed to determine the differences in the roles of these mucins in carcinogenesis.

We emphasize that this study is based on a large collection (n=108) of a very rare appendiceal carcinoma. Furthermore, we prove that MUC3 only affected the survival, while other mucins with prognostic potentials in many malignancies have little importance. These new data would have a significant clinical impact. The patients with positive MUC3 expression of appendiceal carcinoma, should be followed-up carefully after surgery.

In conclusion, we found that expression of MUC3 in appendiceal carcinoma is an independent poor prognostic factor. MUC3 is a useful predictor of outcome in patients after surgery, and the key mucin for tumor progression in this rare tumor.

Acknowledgments

The authors thank Dr. Dallas M. Swallow and Dr. Suzanne Crawley (Galton Laboratory, University College London, London, UK) for providing anti-MUC3 antibody and for valuable discussions. We also thank Mr. Y. Atsuchi, Ms. C. Baba, Mr. S. Matuo, Ms. Y. Nishimura and Ms. S. Yoshimura for their technical assistance, and Ms. Y. Tokura for her assistance with ethics in the institutional review board.

Author Contributions

Conceived and designed the experiments: H. Shibahara MH S. Yonezawa. Performed the experiments: H. Shibahara MH S. Yonezawa. Analyzed the data: H. Shibahara MH S. Yokoyama S. Yonezawa. Contributed reagents/materials/analysis tools: KR IK MO H. Shirahama YT YK MN S. Kuze HH TK HK H. Suzuki TA YS NY MF S. Kondo YO TY TH ES KS S. Kawai KH HT OO MS AK NK SKB S. Yonezawa. Wrote the paper: H. Shibahara MH S. Yonezawa.

References

1. **McCusker ME, Cote TR, Clegg LX, Sobin LH** (2002) Primary malignant neoplasms of the appendix: a population-based study from the surveillance, epidemiology and end-results program, 1973–1998. *Cancer* 94: 3307–3312.
2. **Gustafsson BI, Siddique L, Chan A, Dong M, Drozdov I, et al.** (2008) Uncommon cancers of the small intestine, appendix and colon: an analysis of SEER 1973–2004, and current diagnosis and therapy. *International journal of oncology* 33: 1121–1131.
3. **Ozawa H** (2012) Statistics of appendiceal malignant tumors: data from the JSCCR Registry and the Japan Autopsy Annual Database. *Daichougan Frontier (in Japanese)* 5: 150–153.
4. **Benedix F, Reimer A, Gastinger I, Mroczkowski P, Lippert H, et al.** (2010) Primary appendiceal carcinoma—epidemiology, surgery and survival: results of a German multi-center study. *European journal of surgical oncology : the journal of the European Society of Surgical Oncology and the British Association of Surgical Oncology* 36: 763–771.
5. **Nitecki SS, Wolff BG, Schlinkert R, Sarr MG** (1994) The natural history of surgically treated primary adenocarcinoma of the appendix. *Annals of surgery* 219: 51–57.
6. **Ito H, Osteen RT, Bleday R, Zinner MJ, Ashley SW, et al.** (2004) Appendiceal adenocarcinoma: long-term outcomes after surgical therapy. *Diseases of the colon and rectum* 47: 474–480.
7. **Ko YH, Park SH, Jung CK, Won HS, Hong SH, et al.** (2010) Clinical characteristics and prognostic factors for primary appendiceal carcinoma. *Asia-Pacific journal of clinical oncology* 6: 19–27.
8. **Kabbani W, Houlihan PS, Luthra R, Hamilton SR, Rashid A** (2002) Mucinous and nonmucinous appendiceal adenocarcinomas: different clinicopathological features but similar genetic alterations. *Mod Pathol* 15: 599–605.
9. **Overman MJ, Fournier K, Hu CY, Eng C, Taggart M, et al.** (2013) Improving the AJCC/TNM staging for adenocarcinomas of the appendix: the prognostic impact of histological grade. *Annals of surgery* 257: 1072–1078.
10. **Yonezawa S, Higashi M, Yamada N, Yokoyama S, Kitamoto S, et al.** (2011) Mucins in human neoplasms: clinical pathology, gene expression and diagnostic application. *Pathol Int* 61: 697–716.
11. **Cheever MA, Allison JP, Ferris AS, Finn OJ, Hastings BM, et al.** (2009) The prioritization of cancer antigens: a national cancer institute pilot project for the acceleration of translational research. *Clin Cancer Res* 15: 5323–5337.
12. **Yonezawa S, Goto M, Yamada N, Higashi M, Nomoto M** (2008) Expression profiles of MUC1, MUC2, and MUC4 mucins in human neoplasms and their relationship with biological behavior. *Proteomics* 8: 3329–3341.
13. **Park HU, Kim JW, Kim GE, Bae HI, Crawley SC, et al.** (2003) Aberrant expression of MUC3 and MUC4 membrane-associated mucins and sialyl Le(x) antigen in pancreatic intraepithelial neoplasia. *Pancreas* 26: e48–54.
14. **Kim GE, Bae HI, Park HU, Kuan SF, Crawley SC, et al.** (2002) Aberrant expression of MUC5AC and MUC6 gastric mucins and sialyl Tn antigen in intraepithelial neoplasms of the pancreas. *Gastroenterology* 123: 1052–1060.
15. **Higashi M, Yamada N, Yokoyama S, Kitamoto S, Tabata K, et al.** (2012) Pathobiological implications of MUC16/CA125 expression in intrahepatic cholangiocarcinoma-mass forming type. *Pathobiology* 79: 101–106.
16. **Kitamoto S, Yokoyama S, Higashi M, Yamada N, Matsubara S, et al.** (2012) Expression of MUC17 is regulated by HIF1alpha-mediated hypoxic responses and requires a methylation-free hypoxia responsible element in pancreatic cancer. *PLoS One* 7: e44108.
17. **Shibahara H, Higashi M, Koriyama C, Yokoyama S, Kitazono I, et al.** (2014) Pathobiological implications of mucin (MUC) expression in the outcome of small bowel cancer. *PLoS One* 9: e86111.
18. **Streppel MM, Vincent A, Mukherjee R, Campbell NR, Chen SH, et al.** (2012) Mucin 16 (cancer antigen 125) expression in human tissues and cell lines and correlation with clinical outcome in adenocarcinomas of the pancreas, esophagus, stomach, and colon. *Human pathology* 43: 1755–1763.

19. **Hirono S, Yamaue H, Hoshikawa Y, Ina S, Tani M, et al.** (2010) Molecular markers associated with lymph node metastasis in pancreatic ductal adenocarcinoma by genome-wide expression profiling. *Cancer science* 101: 259–266.
20. **O'Connell JT, Hacker CM, Barsky SH** (2002) MUC2 is a molecular marker for pseudomyxoma peritonei. *Mod Pathol* 15: 958–972.
21. **Yajima N, Wada R, Yamagishi S, Mizukami H, Itabashi C, et al.** (2005) Immunohistochemical expressions of cytokeratins, mucin core proteins, p53, and neuroendocrine cell markers in epithelial neoplasm of appendix. *Human pathology* 36: 1217–1225.
22. **Mall AS, Chirwa N, Govender D, Lotz Z, Tyler M, et al.** (2007) MUC2, MUC5AC and MUC5B in the mucus of a patient with pseudomyxoma peritonei: biochemical and immunohistochemical study. *Pathol Int* 57: 537–547.
23. **Yoon SO, Kim BH, Lee HS, Kang GH, Kim WH, et al.** (2009) Differential protein immunoeexpression profiles in appendiceal mucinous neoplasms: a special reference to classification and predictive factors. *Mod Pathol* 22: 1102–1112.
24. **Suzuki J, Kazama S, Kitayama J, Uozaki H, Miyata T, et al.** (2009) Signet ring cell carcinoma of the appendix manifesting as colonic obstruction and ovarian tumors: report of a case. *Surgery today* 39: 235–240.
25. **Chu PG, Chung L, Weiss LM, Lau SK** (2011) Determining the site of origin of mucinous adenocarcinoma: an immunohistochemical study of 175 cases. *Am J Surg Pathol* 35: 1830–1836.
26. **Chang MS, Byeon SJ, Yoon SO, Kim BH, Lee HS, et al.** (2012) Leptin, MUC2 and mTOR in appendiceal mucinous neoplasms. *Pathobiology* 79: 45–53.
27. **Higashi M, Goto M, Saitou M, Shimizu T, Rousseau K, et al.** (2010) Immunohistochemical study of mucin expression in periampullary adenomyoma. *J Hepatobiliary Pancreat Sci* 17: 275–283.
28. **Higashi M, Yonezawa S, Ho JJ, Tanaka S, Irimura T, et al.** (1999) Expression of MUC1 and MUC2 mucin antigens in intrahepatic bile duct tumors: its relationship with a new morphological classification of cholangiocarcinoma. *Hepatology* 30: 1347–1355.
29. **Tamada S, Goto M, Nomoto M, Nagata K, Shimizu T, et al.** (2002) Expression of MUC1 and MUC2 mucins in extrahepatic bile duct carcinomas: its relationship with tumor progression and prognosis. *Pathol Int* 52: 713–723.
30. **Shibahara H, Tamada S, Higashi M, Goto M, Batra SK, et al.** (2004) MUC4 is a novel prognostic factor of intrahepatic cholangiocarcinoma-mass forming type. *Hepatology* 39: 220–229.
31. **Saitou M, Goto M, Horinouchi M, Tamada S, Nagata K, et al.** (2005) MUC4 expression is a novel prognostic factor in patients with invasive ductal carcinoma of the pancreas. *J Clin Pathol* 58: 845–852.
32. **Tamada S, Shibahara H, Higashi M, Goto M, Batra SK, et al.** (2006) MUC4 is a novel prognostic factor of extrahepatic bile duct carcinoma. *Clin Cancer Res* 12: 4257–4264.
33. **Hamada T, Nomura M, Kamikawa Y, Yamada N, Batra SK, et al.** (2012) DF3 epitope expression on MUC1 mucin is associated with tumor aggressiveness, subsequent lymph node metastasis, and poor prognosis in patients with oral squamous cell carcinoma. *Cancer*.
34. **Duncan TJ, Watson NF, Al-Attar AH, Scholefield JH, Durrant LG** (2007) The role of MUC1 and MUC3 in the biology and prognosis of colorectal cancer. *World journal of surgical oncology* 5: 31.
35. **Shanmugam C, Jhala NC, Katkooori VR, Wan W, Meleth S, et al.** (2010) Prognostic value of mucin 4 expression in colorectal adenocarcinomas. *Cancer* 116: 3577–3586.
36. **Kocer B, Soran A, Erdogan S, Karabeyoglu M, Yildirim O, et al.** (2002) Expression of MUC5AC in colorectal carcinoma and relationship with prognosis. *Pathol Int* 52: 470–477.
37. **Walsh MD, Clendenning M, Williamson E, Pearson SA, Walters RJ, et al.** (2013) Expression of MUC2, MUC5AC, MUC5B, and MUC6 mucins in colorectal cancers and their association with the CpG island methylator phenotype. *Mod Pathol* 26: 1642–1656.
38. **Senapati S, Ho SB, Sharma P, Das S, Chakraborty S, et al.** (2010) Expression of intestinal MUC17 membrane-bound mucin in inflammatory and neoplastic diseases of the colon. *J Clin Pathol* 63: 702–707.
39. **Aloysius MM, Zaitoun AM, Awad S, Ilyas M, Rowlands BJ, et al.** (2010) Mucins and CD56 as markers of tumour invasion and prognosis in periampullary cancer. *Br J Surg* 97: 1269–1278.

40. Mall AS, Tyler MG, Ho SB, Krige JE, Kahn D, et al. (2010) The expression of MUC mucin in cholangiocarcinoma. *Pathology, research and practice* 206: 805–809.
41. Leroy X, Gouyer V, Ballereau C, Zerimech F, Huet G, et al. (2003) Quantitative RT-PCR assay for MUC3 and VEGF mRNA in renal clear cell carcinoma: relationship with nuclear grade and prognosis. *Urology* 62: 771–775.
42. Lee JH, Lee JH, Kim A, Kim I, Chae YS (2005) Unique expression of MUC3, MUC5AC and cytokeratins in salivary gland carcinomas. *Pathol Int* 55: 386–390.
43. Nguyen PL, Niehans GA, Cherwitz DL, Kim YS, Ho SB (1996) Membrane-bound (MUC1) and secretory (MUC2, MUC3, and MUC4) mucin gene expression in human lung cancer. *Tumour Biol* 17: 176–192.
44. Rakha EA, Boyce RW, Abd El-Rehim D, Kurien T, Green AR, et al. (2005) Expression of mucins (MUC1, MUC2, MUC3, MUC4, MUC5AC and MUC6) and their prognostic significance in human breast cancer. *Mod Pathol* 18: 1295–1304.
45. Furuya C, Kawano H, Yamanouchi T, Oga A, Ueda J, et al. (2012) Combined evaluation of CK5/6, ER, p63, and MUC3 for distinguishing breast intraductal papilloma from ductal carcinoma in situ. *Pathol Int* 62: 381–390.
46. Kitamoto S, Yamada N, Yokoyama S, Houjou I, Higashi M, et al. (2010) Promoter hypomethylation contributes to the expression of MUC3A in cancer cells. *Biochemical and biophysical research communications* 397: 333–339.
47. Ho SB, Dvorak LA, Moor RE, Jacobson AC, Frey MR, et al. (2006) Cysteine-rich domains of muc3 intestinal mucin promote cell migration, inhibit apoptosis, and accelerate wound healing. *Gastroenterology* 131: 1501–1517.
48. Peng Z, He Y, Yang Y, Zhu R, Bai J, et al. (2010) Autoproteolysis of the SEA module of rMuc3 C-terminal domain modulates its functional composition. *Archives of biochemistry and biophysics* 503: 238–247.
49. Pan Q, Tian Y, Li X, Ye J, Liu Y, et al. (2013) Enhanced membrane-tethered mucin 3 (MUC3) expression by a tetrameric branched peptide with a conserved TFLK motif inhibits bacteria adherence. *J Biol Chem* 288: 5407–5416.
50. Dorofeyev AE, Vasilenko IV, Rassokhina OA, Kondratiuk RB (2013) Mucosal barrier in ulcerative colitis and Crohn's disease. *Gastroenterology research and practice* 2013: 431231.
51. Li L, Huang PL, Yu XJ, Bu XD (2012) Clinicopathological Significance of Mucin 2 Immunohistochemical Expression in Colorectal Cancer: A Meta-Analysis. *Chinese journal of cancer research = Chung-kuo yen cheng yen chiu* 24: 190–195.
52. Hamada T, Wakamatsu T, Miyahara M, Nagata S, Nomura M, et al. (2012) MUC4: a novel prognostic factor of oral squamous cell carcinoma. *Int J Cancer* 130: 1768–1776.
53. Shibahara H, Tamada S, Goto M, Oda K, Nagino M, et al. (2004) Pathologic features of mucin-producing bile duct tumors: two histopathologic categories as counterparts of pancreatic intraductal papillary-mucinous neoplasms. *Am J Surg Pathol* 28: 327–338.
54. Goto M, Shibahara H, Tamada S, Hamada T, Oda K, et al. (2005) Aberrant expression of pyloric gland-type mucin in mucin-producing bile duct carcinomas: a clear difference between the core peptide and the carbohydrate moiety. *Pathol Int* 55: 464–470.
55. Basturk O, Khayyata S, Klimstra DS, Hruban RH, Zamboni G, et al. (2010) Preferential expression of MUC6 in oncocytic and pancreatobiliary types of intraductal papillary neoplasms highlights a pyloropancreatic pathway, distinct from the intestinal pathway, in pancreatic carcinogenesis. *Am J Surg Pathol* 34: 364–370.
56. Kim YS, Ho SB (2010) Intestinal goblet cells and mucins in health and disease: recent insights and progress. *Current gastroenterology reports* 12: 319–330.
57. Ho SB, Luu Y, Shekels LL, Batra SK, Kandarian B, et al. (2010) Activity of recombinant cysteine-rich domain proteins derived from the membrane-bound MUC17/Muc3 family mucins. *Biochimica et biophysica acta* 1800: 629–638.
58. Yamada N, Kitamoto S, Yokoyama S, Hamada T, Goto M, et al. (2011) Epigenetic regulation of mucin genes in human cancers. *Clinical epigenetics* 2: 85–96.

59. **Pelaseyed T, Hansson GC** (2011) CFTR anion channel modulates expression of human transmembrane mucin MUC3 through the PDZ protein GOPC. *Journal of cell science* 124: 3074–3083.
60. **Pelaseyed T, Bergstrom JH, Gustafsson JK, Ermund A, Birchenough GM, et al.** (2014) The mucus and mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and interact with the immune system. *Immunological reviews* 260: 8–20.
61. **Pelaseyed T, Gustafsson JK, Gustafsson IJ, Ermund A, Hansson GC** (2013) Carbachol-induced MUC17 endocytosis is concomitant with NHE3 internalization and CFTR membrane recruitment in enterocytes. *American journal of physiology Cell physiology* 305: C457–467.