



Association of blood lead level with neurobehavior and neurotransmitter expressions in Indian children

Malavika L.¹, Prasenjit Mitra^{*,1}, Taru Goyal, Abhilasha, Shailja Sharma, Purvi Purohit, Praveen Sharma^{*}

Department of Biochemistry, All India Institute of Medical Sciences, Jodhpur, India

ARTICLE INFO

Edited by Dr. A.M. Tsatsaka

Keywords:

Blood lead level
Neurobehavior
Neurotransmitter
Serotonin
Dopamine

ABSTRACT

Present study aimed to assess the alterations in neurotransmitter expression and its association with Blood Lead Level (BLL) and neurobehavioral pattern in children. 72 school going children were recruited. Blood lead levels were determined by Atomic Absorption Spectrophotometer. Neurobehavioral state was assessed by means of population specific scale i.e. CPMS (Childhood Psychopathological measurement Schedule). Serum serotonin and dopamine were estimated by ELISA, receptor and transporter gene expressions were assessed by quantitative real time PCR. Significant positive correlation was observed between Total CPMS score (i.e. adverse neurobehaviour) and BLL. Further, serum serotonin levels and dopamine receptor expression showed a negative and positive association with BLL, respectively. In similarity, serum serotonin levels showed a negative correlation and dopamine receptor expression had a significant positive correlation with total CPMS score. Environmental exposure to Lead (Pb) may result in significant alterations in the neurotransmitter levels which may be associated with neurobehavioral changes in the children exposed to Pb.

1. Introduction

Lead (Pb), is a heavy metal that is commonly used in industries manufacturing paints, dyes, batteries, toys, jewellery etc. Irrespective of the advantage offered by Pb for industrial use, its adverse effects on health has led to tremendous reduction in its use. Occupational exposure remains to be the major source for Pb exposure in adults, whereas, ingestion of Pb from contaminated food, water and inhalation of Pb in dusts are chief source of Pb in children. Further, use of ceramic cookware and unfiltered water have also been reported as potential sources for Pb. Pb in water could be a result of environmental pollution from anthropogenic activities or from erosion of pipes used in water distribution system. In addition, certain lifestyle habits such as consumption of roadside food, owning pets, use of cosmetics, residing in old houses or residing close to vehicular traffic may also increase the risk of Pb exposure [1–4].

Pb is a multi-system toxicant that can cause acute encephalopathy at very high levels (>40 µg/dL), while increase incidence of hypertension, renal disorder, and neurodevelopmental defects at lower concentrations

[5]. 63 % of idiopathic intellectual development failure and 10 % global burden of hypertensive heart disease have been attributed to Pb toxicity [37]. Children, in particular, are vulnerable to neurotoxic effects of Pb. Early studies have reported neurotoxicity in children at very high Blood Lead Levels (BLLs) (>40 µg/dL), however, later studies have identified Pb to be neurotoxic at much lower concentrations (<10 µg/dL) [6]. The US Centre for Disease Control and Prevention (CDC) defines BLL >5 µg/dL as toxic cut off level for children, with no threshold being safe ([7]). About 74 % of children residing in low- and middle-income countries of South East Asia and 64 % of children in Kathmandu valley of Nepal have BLL >5 µg/dL [8]. Childhood Pb toxicity is of major concern due to the irreversible nature of neurological damage incurred on the developing nervous system. Various aspects of neuronal function, such as, cognition, behaviour, and motor functions, are affected by Pb neurotoxicity [9]. A decline in overall IQ, in addition to, poor reading, math and spelling capabilities have been reported in children with high BLL [10]. Pb causes nearly 6 lakh (600,000) new cases of childhood intellectual disability every year with substantial proportion residing in developing countries [11]. Further, longitudinal studies report poor

^{*} Corresponding authors at: Department of Biochemistry, All India Institute of Medical Sciences, Jodhpur, Rajasthan, India.

E-mail addresses: malavika.eswar@gmail.com (M. L.), mitrap@aiimsjodhpur.edu.in (P. Mitra), tarugoyal123@gmail.com (T. Goyal), abhilashabankul@gmail.com (Abhilasha), shailjachambial@yahoo.com (S. Sharma), purohitp@aiimsjodhpur.edu.in (P. Purohit), sharmapr@aiimsjodhpur.edu.in (P. Sharma).

¹ Co- first Authors: These authors has contributed equally as first author in this work.

executive function and decision making in adults with high childhood BLL, suggestive of irreversible effect on cognitive development [12]. Behavioural changes of both internalising (depression, anxiety) and externalising (aggressiveness, impulsivity) nature have been observed in children with high BLL [13]. However, a characteristic neuropathological signature associated with Pb neurotoxicity is yet undefined.

Mechanisms such as oxidative stress, direct neural damage, imbalance in calcium and neurotransmitter homeostasis have been implicated in Pb neurotoxicity [14–16]. The chief neurotransmitter systems involved in Pb neurotoxicity include monoaminergic, cholinergic, and GABAergic systems. Serotonin (5 H T) and dopamine (DA) monoaminergic neurotransmitters mediate cognition, memory, learning, attention, mood, and reward behaviour. Pb is proposed to affect neurotransmitter availability by altering its presynaptic and post-synaptic factors such as synaptosome formation, release, uptake, receptors, and transporters, in addition to, altering levels of second and third messengers such as PKC, Ca, transcription factors etc. [17]. Studies have reported a decrease in DA levels, with increased expression of Dopamine Transporter (DAT) and a possible compensatory increase in Dopamine Receptor (DR) levels. However, these changes have been region specific and varied with duration and concentration of Pb exposure. Likewise, changes in 5 H T levels, its receptor, and, transporter expression have also been reported. However, majority of these studies have been based on animal models and lack verification on human subjects. Irrespective of extensive studies on mechanism underlying Pb neurotoxicity, precise understanding of its pathogenesis remains unclear. Further, the infeasibility to conduct CNS related studies in live subjects adds to the deficiency in human studies. However, mRNA expression in peripheral lymphocytes and serum levels of neurotransmitters have shown to be an effective representatives of CNS levels [18]. Therefore, the present study has been designed to assess neurotransmitter expression in relation to BLL and neurobehavioral pattern in children.

2. Materials and methods

2.1. Study population

Study subjects consisted of school going children of Jodhpur, between 6–15 years of age ($N = 72$). The sample size for present study was calculated to be 73, based on 5% prevalence of High BLL ($>5 \mu\text{g/dL}$) in Jodhpur, as reported previously [19]. The power of study estimated to be 90 with the present sample size of 72. Ethical consent from Institutional Ethics Committee (Reference no. AIIMS/IEC/2018/640) and signed consent from parent/guardian was obtained prior to sample recruitment. Demographic details of the study participants were obtained by means of self-made questionnaire. Additionally, information regarding potential sources for Pb exposure as reported in previous studies, such as distance of residence from vehicular traffic, use of cosmetics, consumption of roadside food etc. were also included in the questionnaire [20]. In addition to demographic details, neurobehavioral assessment and venous blood sample were collected at the time of sample recruitment.

2.2. Sample collection

Venous blood sample was collected from each subject by means of venepuncture under aseptic conditions. 2 mL of whole blood sample was used for blood lead and mRNA expression analysis and 2 mL of serum sample was used for ELISA.

2.3. Estimation of blood lead level (BLL)

Blood lead levels (BLL) were estimated by Graphite Furnace Atomic Absorption Spectrometry (GFAAS) using Zeeman correction in an ICE 3000 system (Thermo Fisher scientific, Waltham, MA) ensuring

appropriate quality control. Analysis was carried out by a five-step programme ($130^\circ\text{C}/20 \text{ s}$, $200^\circ\text{C}/30 \text{ s}$, $600^\circ\text{C}/10 \text{ s}$, $2000^\circ\text{C}/3 \text{ s}$, $2450^\circ\text{C}/3 \text{ s}$) followed by measurement of absorbance at 282.3 nm. A 5-point calibration curve was obtained using standard solutions of Pb (Pb concentrations: 2.5 ppb, 5 ppb, 10 ppb, 20 ppb and 40 ppb) prepared from serial dilutions of 1000 ppm stock Pb solution. Samples and standards were prepared in 1:20 dilution using matrix modifier (0.1 % diammonium hydrogen phosphate, 0.5 % Triton X-100 and 0.2 % HNO_3) as diluent. Clin check whole blood control (Recipe, Munich, Germany) were used for quality check. Results are expressed in $\mu\text{g/dL}$.

2.4. Peripheral blood neurotransmitter levels

Serum Serotonin (5 H T) and Dopamine (DA) levels were estimated by commercially available ELISA kits (Elab sciences) following manufacturer instructions. Absorbance was measured at 450 nm. Standard curve was obtained using serial dilution of known concentrate. The results for 5 H T and DA are expressed as ng/mL and pg/mL respectively.

2.5. RNA isolation and relative mRNA expression

RNA was isolated from 750 μL whole blood sample by Trizol method using Trizol LS reagent (Sigma-Aldrich, Merck, Germany) as per manufacturer's instructions. Quality and quantity of isolated RNA were determined using Thermo Scientific nanodrop One^c. Isolated RNA was stored at -80°C till further use. Subsequently, relative mRNA expression was performed in two-step reverse transcriptase PCR. Firstly, cDNA was synthesized from isolated RNA in 20 μL volume reactions using thermo scientific verso cDNA synthesis kit (Waltham, Massachusetts, USA) following manufacturer instructions. Following cDNA synthesis, quantitative real time PCR (qPCR) was carried out in Bio-Rad CFX96 real time thermal cycler (Bio-Rad, USA) using SYBR green master mix (Thermo Scientific, Massachusetts, USA). Pre-validated primer sequences from Sigma-Aldrich for Serotonin Receptor (5 H TR2A, 5 H TR3A), Serotonin Transporter (ST), Dopamine Receptor (DRD2, DRD3, DRD4) and Dopamine Transporter (DT) gene were used. GAPDH was used as housekeeping gene for normalization of target gene expressions. The Ct of the target genes were normalized to Ct of the GAPDH gene.

2.6. Neurobehavioral assessment

Neurobehavioral state of the children was evaluated by means of Childhood Psychopathological Measurement Schedule (CPMS). CPMS is an adaptation of Achenbach Child Behaviour Check List (CBCL) standardized for Indian population [21]. It is a widely used screening tool for assessing psychopathological abnormalities in children between 4–14 years in both boys and girls. It is a one-time assessment questionnaire comprising of 75 questions assessing eight factors i.e. Factor I- low intelligence with behavioural problems, Factor II- Conduct disorder, Factor III- Anxiety, Factor IV- Depression, Factor V- Psychotic symptoms, Factor VI- Special symptoms, Factor VII -Physical illness with emotional problems and Factor VIII- Somatization. Each question evokes a 'Yes' or 'No' response carrying 1 (in presence of symptom) or 0 (in absence of symptom) score, respectively. A score of more than 10 indicates a high probability of psychopathological disorder in children. Questions were directed to the parent/guardian of the child and responses were recorded.

2.7. Statistical analysis

The data collected was tabulated and analysed by Microsoft Excel 2013. Statistical tests were performed using GraphPad Prism 8.3 and SPSS version 23. Descriptive statistics were carried out to determine mean, standard deviation (SD), median and range. To assess normal distribution of the data, Shapiro Wilk Test was carried out. Parametric

data was compared using *t*-test and non-parametric data using Mann Whitney *U* test *P*-value <0.05 was considered statistically significant. To assess the correlation, Pearson or Spearman correlation test was performed.

3. Results

3.1. Demographic data

The median age of children was 14 years with a male to female ratio of 0.84. BLL was found to be significantly greater in older age group and female children. Children living in older houses had significantly higher BLL than children living in newly constructed houses i.e houses constructed in the recent 5 years. BLL did not differ significantly between children differing in other lifestyle factors such as habit of washing of hands before food, frequency of roadside food consumption, usage of kohl/surma, traditional medicine and distance of residence from vehicular traffic prone areas. Details of socio-demographic and lifestyle characteristics are provided in Table 1.

3.2. Correlation of BLL and 5 H T

Serum serotonin levels showed significant negative association with

Table 1
Sociodemographic details of the study participants.

Risk factor	N	BLL (µg/dL) Mean ±SD	P value
Age			0.01
9–13 years	20	3.8 ± 1.83	
13–15 years	52	7.41 ± 7.55	
Sex			0.03
Male	33	6.22 ± 3.71	
Female	39	6.55 ± 8.46	
Habit of washing hands before food			0.88
No	16	6.06 ± 4.77	
Yes	56	6.5 ± 7.15	
Habit of eating roadside food frequently (> twice in a week)			0.56
No	44	6.88 ± 7.72	
Yes	28	5.66 ± 4.61	
Habit of using kohl/surma routinely (>5 days in a week)			0.22
No	60	6.7 ± 1.11	
Yes	12	4.49 ± 3.36	
Habit of using traditional medicine			0.99
No	44	5.3 ± 3.23	
Yes	28	8 ± 9.79	
Maternal education			0.02
No formal education	30	4.22 ± 2.4	
Any formal education	42	8.06 ± 8.36	
Paternal education			0.29
No formal education	8	6.9 ± 5.14	
Any formal education	64	6.37 ± 6.93	
Residing in new house (constructed within last 5 years)			0.03
No	56	6.8 ± 7.35	
Yes	16	4.66 ± 3.1	
History of recent painting (painted atleast once in last 2 years)			0.36
No	43	5.79 ± 7.71	
Yes	29	7.29 ± 5.14	
Residence close to traffic (located <2 km from vehicular traffic prone areas)			0.47
No	15	6.07 ± 3.69	
Yes	57	6.5 ± 7.41	

P value < 0.05 was considered significant.

BLL. Whereas serotonin receptor or transporter gene expression did not have any significant association with BLL (Table 2).

3.3. Correlation of BLL and DA

Unlike serum 5 H T levels, serum DA did not have any significant association with BLL. However, dopamine receptor (DRD2 and DRD3) gene expression had significant mild positive correlation with BLL (Table 3).

3.4. Neurotransmitter and BLL groups

When the study participants were divided into High and Low BLL groups based on median BLL of 4.9 µg/dL, serum 5 H T levels and serotonin receptor (5 H TR3A) expression were found to be significantly different between the groups. Serum 5 H T levels were significantly lower with increased expression of serotonin receptor (5 H TR3A) in High BLL group (Table 4). Further, dopamine receptor (DRD2 and DRD3) expression was also significantly higher in the High BLL group (Table 4). Additionally, a correlation matrix of BLL, serum 5 H T, its receptor and transporter expression showed a significant positive relation of 5 H TR2A (*P* < 0.05, *r* = 0.30) and 5 H TR3A (*P* < 0.01, *r* = 0.36) with ST gene expression, respectively (data not shown).

3.5. BLL and neurobehavior

Spearman correlation analysis revealed a significant positive correlation between BLL and total CPMS score. Further, except for Factor II (Conduct Disorder), all other factors had significant association with BLL, independently (Table 5). Between the High and Low BLL groups, significant difference was observed with Total CPMS score, as well as Factors I, III, IV, V and VII (Table 6). A total of 16 children had CPMS score more than 10 (Data not shown) among which 15 children belong High BLL group and only 1 child in Low BLL group.

3.6. Neurotransmitter levels and neurobehavior

Serum 5 H T levels had significant negative association with Total CPMS score, whereas serotonin receptor or transporter expression showed no significant association with total CPMS score. Additionally, individual factors such as Factor I, II, V and VIII showed mild negative association with serum 5 H T. Although ST did not show any significant association with total CPMS score, there was mild association between ST and Factor VIII.

Unlike 5 H T, serum DA did not show any significant association with total CPMS score or with scores of individual factors and only dopamine receptor (DRD3) expression had significant positive association with Total CPMS score, as well as with Factor III, IV, V and VI (Table 7).

4. Discussion

Pb is one of the most studied heavy metal toxicants known to cause adverse effects in children [22]. Recent studies report toxic effects of Pb at concentrations lower than present cut off value (<5 µg/dL) [6]. The median BLL of children in present study was 4.95 µg/dL, with 50 % of

Table 2
Association between BLL and 5 H T in study population (N = 72).

	BLL (µg/dL)		
	<i>r</i>	<i>P</i> value	95 % CI
Serum 5 H T (ng/mL)	−0.41	0.0003	−0.59 to −0.19
5 H TR2A (2'dct)	−0.12	0.30	−0.35 to 0.11
5 H TR3A (2'dct)	−0.15	0.19	−0.37 to 0.08
ST (2'dct)	−0.08	0.51	−0.32 to 0.16

P value < 0.05 was considered significant.

Table 3

Association between BLL and DA in study population (N = 72).

BLL (µg/dL)	r	P value	95 % CI
Serum DA (pg/mL)	0.11	0.35	−0.13 to 0.34
DRD2 (2'dct)	0.33	0.005	0.09 to 0.52
DRD3 (2'dct)	0.26	0.02	0.02 to 0.46
DRD4 (2'dct)	0.05	0.67	−0.19 to 0.28
DT (2'dct)	0.11	0.37	−0.13 to 0.34

P value < 0.05 was considered significant.

Table 4

Difference in neurotransmitter protein and gene expression between High BLL and Low BLL groups.

Parameter	High BLL (>4.9 µg/dL) (N = 36) Median (IQR)	Low BLL (<4.9 µg/dL) (N = 36) Median (IQR)	P value
Serum Serotonin (ng/mL)	123.3 (162.6)	303.1 (239.7)	0.0008
5 H TR2A (2'dct)	0.06 (0.1)	0.11 (0.4)	0.49
5 H TR3A (2'dct)	13.15 (15.8)	0.17 (4.1)	<0.0001
ST (2'dct)	0.81 (0.5)	0.69 (1.0)	0.66
Serum Dopamine (pg/mL)	770.2 (65.6)	774.6 (64.6)	0.77
DRD2 (2'dct)	2.53 (7.5)	0.32 (3.1)	0.003
DRD3 (2'dct)	13.1 (15.9)	6.85 (6.3)	0.0036
DRD4 (2'dct)	8.70 (12.0)	5.57 (20.7)	0.56
DT (2'dct)	0.1 (0.3)	0.08 (0.2)	0.14

P value < 0.05 was considered significant.

Table 5

Association between BLL and CPMS scores in total study subjects (N = 72).

BLL	r	P value	95 % CI
Factor 1 Low intelligence with behaviour problems	0.35	0.02	0.12 to 0.54
Factor 2 Conduct disorder	0.20	0.13	−0.06 to 0.39
Factor 3 Anxiety	0.37	0.0001	0.21 to 0.60
Factor 4 Depression	0.49	0.007	0.26 to 0.64
Factor 5 Psychotic symptoms	0.37	0.01	0.14 to 0.56
Factor 6 Speical symptoms	0.34	0.009	0.07 to 0.50
Factor 7 Physical illness with emotional problems	0.44	0.0004	0.17 to 0.58
Factor 8 Somatization	0.23	0.01	0.006 to 0.45
Total CPMS	0.68	0.001	0.51 to 0.78

Data is expressed as r i.e. spearman's correlation coefficient; P value < 0.05 was considered significant.

Table 6

Comparison of CPMS score between High and Low BLL groups.

CPMS Factors	High BLL (>4.9 µg/dL) (N = 36)		Low BLL (<4.9 µg/dL) (N = 36)		P value
	Mean ± SD	Median (IQR)	Mean ± SD	Median (IQR)	
Factor 1	2.58 ± 1.48	3 (3)	1.47 ± 1.38	1.5 (3)	0.002
Factor 2	1.70 ± 2.36	1.5 (2)	0.75 ± 1.02	0 (1.75)	0.060
Factor 3	0.97 ± 1.05	1(2)	0.3 ± 0.70	0 (0)	0.001
Factor 4	1.91 ± 1.81	2(3)	0.55 ± 1.05	0 (0.75)	<0.001
Factor 5	0.26 ± 0.51	0 (0.75)	0.02 ± 0.16	0	0.012
Factor 6	0.17 ± 0.45	0	0.02 ± 0.16	0	0.092
Factor 7	0.52 ± 0.66	0 (1)	0.27 ± 0.51	0 (0.75)	0.042
Factor 8	1.14 ± 1.25	1 (2)	0.75 ± 1.02	0 (1)	0.109
Total	9.29 ± 4.3	9 (6.75)	4.16 ± 2.19	5 (4.75)	<0.001

P value < 0.05 was considered significant.

them having BLL above present cut off limit. Survey of BLL in children in US and major cities of Europe indicated a drastic fall in BLL, with majority of children having BLL <5 µg/dL, however, although limited, BLL survey in Indian children indicate prevalence of higher BLL than in other countries ([36] CDC, 2002; [23]). A survey conducted by George foundation in six major cities of India, namely, Bangalore, Hyderabad, Chennai, Calcutta, Delhi, and Mumbai reported nearly 50 % of the children to have BLL > 10 µg/dL [24]. Whereas, study by Roy et al. carried out in 756 children residing in Chennai, also reported mean BLL of 11.4 µg/dL [13]. Lower median BLL in this study, in comparison to other Indian studies, could be attributed to the difference in study region, as these surveys were carried out in heavily polluted or densely populated regions. Jodhpur, although not a metropolitan city housing dense population or heavy pollution, harbours several small-scale industries using Pb for dye making, textile making, welding works (handicrafts) etc. Further, BLL was significantly affected by both age and sex, with older age and female children having higher BLLs. Higher BLL in older children could also be due to high proportion of children belonging to older age group in our study, however, higher BLL in girls could not be attributed to kohl/ surma use, as reported by another study [25]. Additionally, Roy et al. reported an inverse relationship between BLL, socio-economic and educational status of the parents [13]. Nevertheless, educational status did not significantly affect BLL in current population and effect of socio-economic status could not be evaluated owing to lack of relevant data.

In present study, neurobehavioral changes evaluated by CPMS showed significant positive correlation between total CPMS scores and BLL (P = 0.002, r = 0.35). Further, it was found to be higher (i.e. worse outcome) in children with High BLL. Factor I of CPMS relating to Intelligence and behavioural problems showed significant direct relation to BLL (P = 0.002, r = 0.35), which was in correspondence to findings in other studies that showed low IQ and ADHD like symptoms in association to BLL [26]. National Health and Nutrition Examination Survey of 2006 reported a 4.1 times increased risk of ADHD among children in highest quintile of BLL when compared to children in lowest quintile [27].

Imbalance in neurotransmitter signalling pathways have been implicated in Pb neurotoxicity in children. At lower concentrations, Pb is known to suppress neurotransmitter levels, whereas, at higher concentrations and prolonged exposure, it is known to mimic Ca and enhance neurotransmitter release, thereby, increasing their concentrations. A differential expression of dopamine levels, its receptor and transporter proteins have been reported in rats exposed to Pb. In the present study, no significant correlation was observed between BLL and serum dopamine levels, whereas a significant positive association was found between dopamine receptor expression (DRD2 and DRD3) and BLL. Also, when compared between High and Low BLL groups, DRD2 and DRD3 were significantly higher in High BLL group (P < 0.05). Moresco et al. reported a similar finding of significant increase in DRD2 in striatum of Pb treated rats, with significant reduction in striatal DA levels [28]. However, there was only mild insignificant decrease in serum DA levels in High BLL group (P = 0.77) in our study. When serum DA levels, DA receptor and transporter expression were compared with neurobehavioral assessment scores, DRD3 (P < 0.001, r = 0.44), and not DRD2 (P > 0.05, r = 0.21), showed significant positive association with total CPMS scores. Suggesting an increased expression of DRD3 with an increase in behavioural impairment. DRD3 also had significant positive association with Factor 4 (P < 0.05, r = 0.42) of CPMS, that evaluates for depressive symptoms in children. Increase in striatal DRD2,3 receptor levels have been reported in cases of Major Depressive Disorder (MDD) when compared to healthy controls [29]. Decline in DA functionality has also been attributed to increase in DA uptake from increased DAT expression pre-synaptically. Chang et al. reported an increase in DAT expression in Pb exposed rats and intracellular DA levels with no significant change in DRD2 levels [30]. However, DAT expression was not found to be significantly altered in present study.

Table 7

Correlation between CPMS factors and neurotransmitter protein and gene expressions.

	Spearman's Correlation coefficient (r)								Total
	Factor 1	Factor 2	Factor 3	Factor 4	Factor 5	Factor 6	Factor 7	Factor 8	
S.5 H T (ng/mL)	−0.26*	−0.28*	−0.09	−0.31	−0.28**	−0.12	−0.07	−0.28*	−0.37**
5 H TR2A (2 ^o dct)	0.06	−0.21	−0.09	0.02	−0.14	−0.01	−0.06	0.15	−0.07
5 H TR2A (2 ^o dct)	−0.05	−0.25*	−0.004	0.11	−0.11	−0.01	0.16	0.07	−0.03
ST (2 ^o dct)	0.07	0.02	−0.05	0.07	0.12	−0.11	0.00	0.26*	0.13
S.DA (pg/mL)	0.20	0.00	−0.11	0.20	0.13	0.27*	0.18	−0.02	0.15
DRD2 (2 ^o dct)	0.03	−0.12	0.20	0.23	0.06*	0.16	0.22	0.13	0.21
DRD3 (2 ^o dct)	0.16	0.12	0.29*	0.45*	0.23**	0.23*	0.13	0.18	0.44***
DRD4 (2 ^o dct)	0.14	−0.18	0.002	0.35	0.10**	0.07	0.02	0.06	0.13
DT (2 ^o dct)	−0.08	−0.19	0.07	0.33	0.06**	0.20	0.05	0.04	0.10

Gene expressions expressed as 2^o-dct values. S. BDNF- Serum BDNF, S.5HT- Serum Serotonin, S. DA-Serum Dopamine.

* P<0.05.

** P<0.01.

*** P<0.001.

5 H T levels, like DA, has been observed to be altered by Pb [31]. In present study, serum 5 H T levels had a significant negative correlation with BLL ($P < 0.001$, $r = -0.41$). Likewise, a serum 5 H T was significantly lower in High BLL group when compared to Low BLL group ($P < 0.001$). A decrease in CNS 5 H T levels and serum 5 H T levels have been reported in mice and adult humans with high BLL, respectively [31]. Overexpression of ST can decrease 5 H T availability, however, no significant difference was observed in ST expression in present study. Also, significant positive correlation was also observed between ST and serotonin receptors (5 H TR2A and 5 H TR3A). Further, 5 H TR3A gene expression was significantly greater in High BLL group ($P < 0.001$) with significant positive correlation between serum ST and its receptors gene expressions (5 H TR2A and 5 H TR3A). Further, 5 H T levels had significant negative association with total CPMS scores, Factor 1, 2 and 8 evaluating for low intelligence, behavioural problems, conduct disorder, and somatization, respectively. Cumulatively, these findings suggest a Pb induced increase in ST with resultant decrease in serum 5 H T levels and possible compensatory increase in serotonin receptor expression, that may underlie Pb induced cognitive and behavioural defects. The changes in serotonergic and dopaminergic system that were observed in the present study were found to be concurrent with findings of Mansour et al., who also reported a decrease 5 H T levels with no significant change in DA levels in Pb treated rats with evident motor defects [32].

Different methods for assessment of Pb exposure and body burden of Pb have evolved over time. Although measurement of BLL has been the most widely accepted method for identification of Pb exposure, it is only an indicator of recent Pb exposure. Teeth and bone Pb measurements would serve as better indicators for long term Pb exposure, as Pb tends to accumulate in the skeletal system with chronic exposure. Further, blood Pb measurements may not reflect Pb concentrations at target tissue level and measurement of Pb at CNS tissue or CSF would be the preferable method to assess Pb neurotoxicity [33]. However, due to the practical difficulties associated with acquiring CNS samples from live subjects, BLL was used as a means to measure Pb exposure. Further, measurement of Pb levels in urine samples have also been reported in previous studies, however, the spontaneous excretion of Pb in urine may not be a reliable marker as it may have high inter-individual variability depending on hydration status, renal function etc [34]. A study carried out by Sullivan et al. [35] to compare the gene expression in blood and brain samples, suggested a significant similarity in gene expression between blood and brain tissues [35]. However, they have also suggested to retain caution while using peripheral cell mRNA expression as a surrogate for CNS evaluation. Therefore, the findings of the present study should be interpreted with caution as changes in peripheral mRNA expression to evaluate CNS expression of the genes might be a possible limitation of our study and requires further affirmation by means of CNS specific evaluation.

In conclusion, studies on Pb neurotoxicity and alteration in

neurotransmitter expression have been conducted mainly on animal models, lacking verification by human studies, existing humans studies have also been limited to one neurotransmitter. Therefore, the results from our study may add to the existing knowledge on Pb neuropathogenesis by affirming the findings of animal studies, as well as, by depicting the gene expression of receptors and transporters of both serotonergic and dopaminergic system. However, the probability of long term Pb exposure since early childhood in our study population should be kept in mind, as exposure to lead (even at low concentration) as early as 2 years of age could critically impair intellectual development. Since our study is a cross-sectional study, the effect of early childhood lead exposure could not be evaluated in this study, nevertheless, there was substantial finding indicative of definitive psychopathological changes with increasing BLL. Additionally, the alterations in serotonergic and dopaminergic system could underlie the neurobehavioral changes induced by Pb.

Conflict of Interest

The authors declare no conflict of interest.

Compliance with ethical standards

This study involves participations of human subjects with the approval from Institutional Ethical Committee (IEC), All India Institute of Medical Sciences, Jodhpur, Rajasthan India. The study was performed in accordance with down in the 1964 Declaration of Helsinki and its later amendments or comparable ethical standards.

Informed consent

Prior written informed consent was obtained from all subjects recruited in this study.

Funding

The study was funded by All India Institute of Medical Sciences, Jodhpur, Rajasthan.

Author's contributions

Conceptualization: PM; **Methodology:** ML, PM, TG **Formal analysis and investigation:** ML, PM, TG, AA, PS **Software-** ML, PM, TG; **Writing - original draft preparation:** ML, PM; **Writing - review and editing:** ML, PM, TG, AA, SS, PSh, PP; **Funding acquisition:** PSh; **Resources:** PSh; **Supervision:** PM, SS, PSh, PP.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.toxrep.2021.05.002>.

References

- [1] M. Cabral, G. Garçon, A. Touré, F. Bah, D. Dewaele, S. Bouhsina, F. Cazier, A. Faye, M. Fall, D. Courcot, A. Verdin, Renal impairment assessment on adults living nearby a landfill: early kidney dysfunction biomarkers linked to the environmental exposure to heavy metals, *Toxicol. Rep.* 8 (2021) 386–394, <https://doi.org/10.1016/j.toxrep.2021.02.009>.
- [2] N. González, J. Calderón, A. Rúbies, I. Timoner, V. Castell, J.L. Domingo, M. Nadal, Dietary intake of arsenic, cadmium, mercury and lead by the population of Catalonia, Spain: analysis of the temporal trend, *Food Chem. Toxicol.* 132 (2019), 110721, <https://doi.org/10.1016/j.fct.2019.110721>.
- [3] Y. Li, J. Zhang, Exposure to lead and cadmium of the Belgian consumers from ceramic food contact articles, *Toxicol. Rep.* 8 (2021) 548–556, <https://doi.org/10.1016/j.toxrep.2021.02.015>.
- [4] M. Malavolti, S.J. Fairweather-Tait, C. Malagoli, L. Vescovi, M. Vinceti, T. Filippini, Lead exposure in an Italian population: food content, dietary intake and risk assessment, *Food Res. Int.* 137 (2020), 109370, <https://doi.org/10.1016/j.foodres.2020.109370>.
- [5] A.-M. Aloizou, V. Siokas, C. Vogiatzi, E. Peristeri, A.O. Docea, D. Petrakis, A. Provatas, V. Folia, C. Chalkia, M. Vinceti, M. Wilks, B.N. Izotov, A. Tsatsakis, D. P. Bogdanos, E. Dardiotis, Pesticides, cognitive functions and dementia: a review, *Toxicol. Lett.* 326 (2020) 31–51, <https://doi.org/10.1016/j.toxlet.2020.03.005>.
- [6] D.C. Bellinger, Very low lead exposures and children's neurodevelopment, *Curr. Opin. Pediatr.* 20 (2008) 172–177, <https://doi.org/10.1097/MOP.0b013e3282f497b>.
- [7] Blood Lead Levels in Children | Lead | CDC [WWW Document], 2019. URL <https://www.cdc.gov/nceh/lead/prevention/blood-lead-levels.htm> (accessed 4.4.20).
- [8] M. Dhimal, K.B. Karki, K.K. Aryal, B. Dhimal, H.D. Joshi, S. Puri, A.R. Pandey, P. Dhakal, A.K. Sharma, G.B. Raya, I. Ansari, D.A. Groneberg, R. Müller, U. Kuch, High blood levels of lead in children aged 6–36 months in Kathmandu Valley, Nepal: a cross-sectional study of associated factors, *PLoS One* 12 (2017), e0179233, <https://doi.org/10.1371/journal.pone.0179233>.
- [9] T.I. Lidsky, J.S. Schneider, Lead neurotoxicity in children: basic mechanisms and clinical correlates, *Brain* 126 (2003) 5–19, <https://doi.org/10.1093/brain/awg014>.
- [10] B.P. Lanphear, Cognitive Deficits Associated with Blood Lead Concentrations <10 microg/dL in US Children and Adolescents, *Public Health Rep.* 115 (2000) 521–529, <https://doi.org/10.1093/phr/115.6.521>.
- [11] A. Prüss-Ustün, C. Vickers, P. Haefliger, R. Bertollini, Knowns and unknowns on burden of disease due to chemicals: a systematic review, *Environ. Health A Glob. Access Sci. Source* 10 (2011) 9, <https://doi.org/10.1186/1476-069X-10-9>.
- [12] M. Mazumdar, D.C. Bellinger, M. Gregas, K. Abanilla, J. Bacic, H.L. Needleman, Low-level environmental lead exposure in childhood and adult intellectual function: a follow-up study, *Environ. Health* 10 (2011) 24, <https://doi.org/10.1186/1476-069X-10-24>.
- [13] A. Roy, D. Bellinger, H. Hu, J. Schwartz, A.S. Ettinger, R.O. Wright, M. Bouchard, K. Palaniappan, K. Balakrishnan, Lead exposure and behavior among young children in Chennai, India, *Environ. Health Perspect.* 117 (2009) 1607–1611, <https://doi.org/10.1289/ehp.0900625>.
- [14] P. Mitra, S. Sharma, P. Purohit, P. Sharma, Clinical and molecular aspects of lead toxicity: an update, *Crit. Rev. Clin. Lab. Sci.* 54 (2017) 506–528, <https://doi.org/10.1080/10408363.2017.1408562>.
- [15] N. Priyanka, N. Geetha, T. Manish, S.V. Sahi, P. Venkatachalam, Zinc oxide nanocatalyst mediates cadmium and lead toxicity tolerance mechanism by differential regulation of photosynthetic machinery and antioxidant enzymes level in cotton seedlings, *Toxicol. Rep.* 8 (2021) 295–302, <https://doi.org/10.1016/j.toxrep.2021.01.016>.
- [16] J.C. Oyem, L.E. Chris-Ozoko, M.T. Enaohwo, F.O. Otabor, V.A. Okudayo, O.A. Udi, Antioxidative properties of Ocimum gratissimum alters Lead acetate induced oxidative damage in lymphoid tissues and hematological parameters of adult Wistar rats, *Toxicol. Rep.* 8 (2021) 215–222, <https://doi.org/10.1016/j.toxrep.2021.01.003>.
- [17] S. Sadiq, Z. Ghazala, A. Chowdhury, D. Büsselberg, Metal toxicity at the synapse: presynaptic, postsynaptic, and long-term effects, *J. Toxicol.* 2012 (2012) 1–42, <https://doi.org/10.1155/2012/132671>.
- [18] M. Amidfar, Y.-K. Kim, L. Colic, M. Arbabi, G. Mobaraki, G. Hassanzadeh, M. Walter, Increased levels of 5HT2A receptor mRNA expression in peripheral blood mononuclear cells of patients with major depression: correlations with severity and duration of illness, *Nord. J. Psychiatry* 71 (2017) 282–288, <https://doi.org/10.1080/08039488.2016.1276624>.
- [19] S. Chambial, K.K. Shukla, S. Dwivedi, P. Bhardwaj, P. Sharma, Blood lead level (BLL) in the adult population of Jodhpur: a pilot study, *Indian J. Clin. Biochem.* 30 (2015) 357–359, <https://doi.org/10.1007/s12291-015-0496-y>.
- [20] V. Kalra, K.T. Chitralkha, T. Dua, R.M. Pandey, Y. Gupta, Blood lead levels and risk factors for lead toxicity in children from schools and an urban slum in Delhi, *J. Trop. Pediatr.* 49 (2003) 121–123, <https://doi.org/10.1093/tropej/49.2.121>.
- [21] S. Malhotra, V.K. Varma, S.K. Verma, A. Malhotra, Childhood psychopathology measurement schedule: development and standardization, *Indian J. Psychiatry* 30 (1988) 325–331.
- [22] A.A. Moncrieff, O.P. Koumides, B.E. Clayton, A.D. Patrick, A.G.C. Renwick, G. E. Roberts, Lead poisoning in children, *Arch. Dis. Child.* 39 (1964) 1–13.
- [23] V.R. Mohan, S. Sharma, K. Ramanujam, S. Babji, B. Koshy, J.D. Bondu, S.M. John, G. Kang, Effects of elevated blood lead levels in preschool children in urban Vellore, *Indian Pediatr.* 51 (2014) 621–625, <https://doi.org/10.1007/s13312-014-0464-2>.
- [24] N.B. Jain, H. Hu, Childhood correlates of blood lead levels in Mumbai and Delhi, *Environ. Health Perspect.* 114 (2006) 466–470, <https://doi.org/10.1289/ehp.8399>.
- [25] S.T. Gogte, N. Basu, S. Sinclair, O.P. Ghai, N.K. Bhide, Blood lead levels of children with pica and surma use, *Indian J. Pediatr.* 58 (1991) 513–519, <https://doi.org/10.1007/BF02750933>.
- [26] R. Nicolescu, C. Petcu, A. Cordeanu, K. Fabritius, M. Schlumpf, R. Krebs, U. Krämer, G. Winneke, Environmental exposure to lead, but not other neurotoxic metals, relates to core elements of ADHD in Romanian children: performance and questionnaire data*, ***Study approval: this study has been reviewed and approved by the Ethics Committee of the “Romanian College of Physicians”, *Environ. Res.* 110 (2010) 476–483, <https://doi.org/10.1016/j.envres.2010.04.002>.
- [27] J.M. Braun, R.S. Kahn, T. Froehlich, P. Auinger, B.P. Lanphear, Exposures to environmental toxicants and attention deficit hyperactivity disorder in U.S. Children, *Environ. Health Perspect.* 114 (2006) 1904–1909, <https://doi.org/10.1289/ehp.9478>.
- [28] R.M. Moresco, R. Dall'Olio, O. Gandolfi, S. Govoni, S. Di Giovine, M. Trabucchi, Lead neurotoxicity: a role for dopamine receptors, *Toxicology* 53 (1988) 315–322, [https://doi.org/10.1016/0300-483X\(88\)90223-5](https://doi.org/10.1016/0300-483X(88)90223-5).
- [29] M. Pecina, M. Sikora, E.T. Avery, J. Heffernan, S. Pecina, B.J. Mickey, J.-K. Zubieta, Striatal dopamine D2/3 receptor-mediated neurotransmission in major depression: implications for anhedonia, anxiety and treatment response, *Eur. Neuropsychopharmacol.* 27 (2017) 977–986, <https://doi.org/10.1016/j.euroneuro.2017.08.427>.
- [30] J. Chang, C. Kueon, J. Kim, Influence of lead on repetitive behavior and dopamine metabolism in a mouse model of Iron overload, *Toxicol. Res.* 30 (2014) 267–276, <https://doi.org/10.5487/TR.2014.30.4.267>.
- [31] A. Mariani, A. Anies, H.R.S. Abdurachim, Causality Pattern of the Blood Lead, Monoamine Oxidase a, and Serotonin Levels in Brass Home Industry Workers Chronically Exposed to Lead 7, 2016.
- [32] M.T. Mansouri, B. Naghizadeh, P. López-Larrubia, O. Cauli, Behavioral deficits induced by lead exposure are accompanied by serotonergic and cholinergic alterations in the prefrontal cortex, *Neurochem. Int.* 62 (2013) 232–239, <https://doi.org/10.1016/j.neuint.2012.12.009>.
- [33] Measuring Lead Exposure in Infants, Children, and Other Sensitive Populations, National Academies Press, Washington, D.C., 1993, <https://doi.org/10.17226/2232>.
- [34] J.R. Tesman, A. Hills, Developmental effects of lead exposure in children, *Soc. Policy Rep.* 8 (1994) 1–20, <https://doi.org/10.1002/j.2379-3988.1994.tb00031.x>.
- [35] P.F. Sullivan, C. Fan, C.M. Perou, Evaluating the comparability of gene expression in blood and brain, *Am. J. Med. Genet.* 141B (2006) 261–268, <https://doi.org/10.1002/ajmg.b.30272>.
- [36] Blood Lead Levels — United States, 1999–2002 [WWW Document], n.d. URL <https://www.cdc.gov/mmwr/preview/mmwrhtml/mm5420a5.htm> (accessed 4.5.20).
- [37] Lead poisoning and health [WWW Document], n.d. URL <https://www.who.int/news-room/fact-sheets/detail/lead-poisoning-and-health> (accessed 4.4.20).