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Expanded View Figures

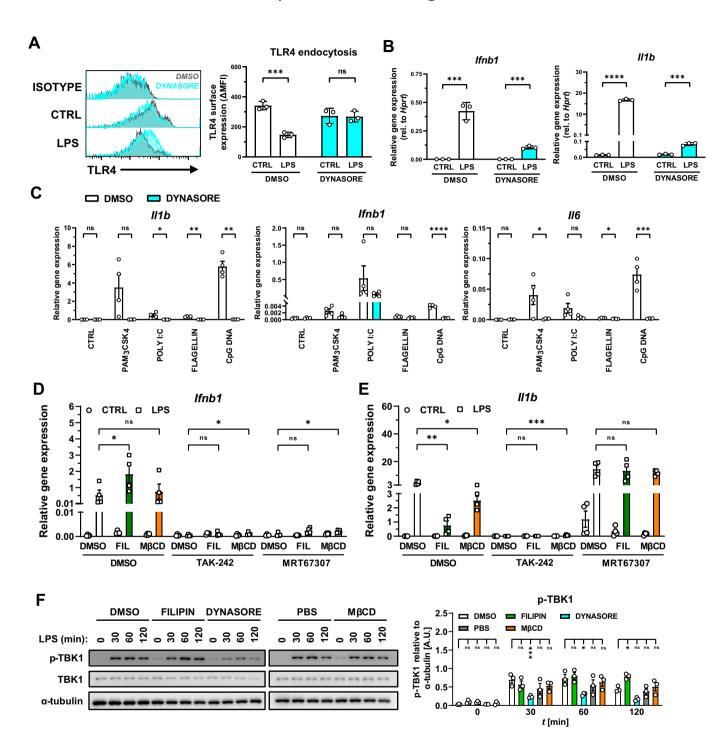


Figure EV1. Impact of dynasore and lipid raft inhibitors on TLR-mediated macrophage activation.

(A) Flow cytometric analysis of TLR4 surface expression in WT BMM treated for 60 min with DMSO or dynasore (80 μ M), followed by stimulation with LPS (100 EU/mL) or left unstimulated (CTRL) for 120 min. (B) qRT-PCR analysis of *Ifnb1* and *If1b* expression in WT BMM treated for 60 min with DMSO or dynasore (80 μ M), followed by stimulation with LPS (100 EU/mL) for 90 min. (C) qRT-PCR analysis of *Iflb1* finb1 and *Il6* mRNA expression in WT BMM treated for 60 min with DMSO or dynasore (80 μ M), followed by stimulation with Pam₃CSK₄ (10 ng/mL), poly I:C (10 µg/mL), flagellin (0.5 µg/mL) or CpG DNA (1 μ M), or left unstimulated (CTRL), for 90 min. (D, E) Impact of TAK-242 (1 μ M) or MRT67307 (5 μ M) on LPS-induced (D) *Iflb1* and (E) *Iflb* expression (90 min) in WT BMM treated with filipin (5 μ M), M β CD (10 mM), or DMSO as control. (F) Time-course of LPS-induced TBK1 phosphorylation in WT BMM treated with filipin, M β CD, dynasore or DMSO as solvent control. Data information: Flow cytometry histograms and immunoblot images depict 1 representative of n=3 biological replicates generated in independent experiments. Bar plots are mean ± SEM of n=3-4 biological replicates generated in independent experiments indicated as data points. Unpaired two-tailed t test performed in (A–C); ordinary two-way ANOVA with Dunnett's multiple comparison test utilized in (D–F); *tP < 0.05; *tP < 0.001, *tP < 0.0001, tP < 0.0001, tP

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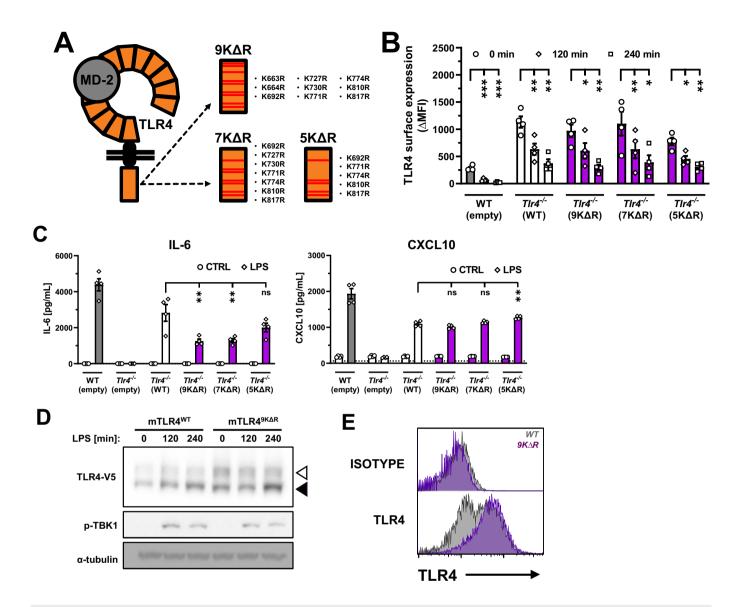


Figure EV2. Lysine residues in the TLR4 TIR domain were not required for LPS-induced TLR4 endocytosis.

(A-C) $TIr4^{-/-}$ BMMs retrovirally reconstituted with (A) mTLR4^{WT}, mTLR4^{9KΔR}, mTLR4^{5KΔR}, mTLR4^{5KΔR} or empty vector were stimulated with LPS (100 EU/mL); (B) TLR4 surface expression assessed at 120 and 240 min post-stimulation and (C) IL-6 and CXCL10 production assessed at 24 h post-stimulation. (D, E) RAW^{TLR4ko} cells were stably transfected with V5-tagged constructs of mTLR4^{WT} or mTLR4^{9KΔR}. (D) Total cellular expression of TLR4-V5 during LPS stimulation (100 EU/mL, 0, 120, 1240 min) was assessed via immunoblot. Phospho-TBK1 served as control for cellular activation; a-tubulin served as loading control. Black arrow head indicates low glycosylated intracellular TLR4, white arrow head indicates highly-glycosylated cell surface-expressed TLR4. (E) TLR4 surface expression was assessed in resting cells via flow cytometry. Data information: Immunoblot images depict 1 representative of n = 3 biological replicates generated in independent experiments. Flow cytometry histogram represents cells post sorting. Bar plots are mean ± SEM of n = 3-4 biological replicates generated in independent experiments indicated as data points. RM two-way ANOVA with Dunnett's multiple comparisons test utilized in (B). Ordinary one-way ANOVA with Dunnett's multiple comparisons test utilized in (C). *P < 0.05; **P < 0

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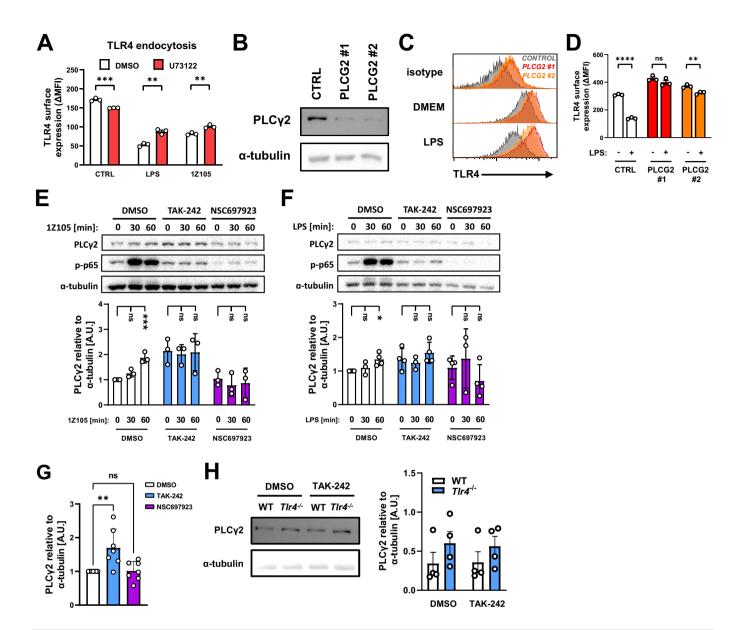


Figure EV3. PLCγ2 regulates TLR4 endocytosis.

(A) Impact of PLC inhibitor (U73122, 10 μ M) on LPS- (100 EU/mL) and 1Z105-induced (10 μ M) TLR4 endocytosis (120 min) in WT BMM. (B-D) CRISPR/Cas9-mediated knockdown of PLC γ 2 expression in WT BMM (B) was verified by immunoblot analysis, and (C, D) shown to impair LPS-induced (100 EU/mL, 120 min) TLR4 endocytosis as assessed by flow cytometry. (E, F) Immunoblot analysis of the impact of TAK-242 (1 μ M) or NSC694923 (10 μ M) on total PLC γ 2, phosphorylated NF-κB p65 and α -tubulin in WT BMM upon stimulation with (C) 1Z105 (10 μ M) or (D) LPS (100 EU/mL) for the indicated times. (G) Comparison of PLC γ 2 expression in unstimulated cells presented in (E, F). (H) Immunoblot analysis of PLC γ 2 cellular expression in WT and $T l r 4^{-/-}$ BMM treated with DMSO or TAK-242 (1 μ M) for 120 min. Data information: Flow cytometry histograms and immunoblot images depict 1 representative of n = 3-4 biological replicates generated in independent experiments. Bar plots are mean ± SEM of n = 3-7 biological replicates generated in independent experiments indicated as data points. Unpaired two-tailed t test utilized in (A, D). Ordinary one-way ANOVA with Dunnett's multiple comparisons test utilized in (E-G). *P<0.05; * *P <0.01; * *P <0.001, * *T <0.001, * *T <0.0001, *T <0.0001, *T <0.0001, *T <0.0001, *T

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