# Extranodal NK/T-cell Lymphoma, Nasal Type Accompanied by PR3-ANCA-associated Glomerulonephritis

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## Abstract

A 62-year-old man exhibiting nasal obstruction and glomerulonephritis with proteinase 3-antineutrophil cytoplasmic antibodies (PR3-ANCAs) was diagnosed with extranodal NK/T-cell lymphoma, nasal type (ENKL) with infiltration of neutrophils with apoptosis. Chemoradiotherapy reduced the tumor, improved the renal function, and decreased the PR3-ANCA levels. ANCA-positivity is observed in immunoinsufficient diseases, in which neutrophils lead to apoptosis and translocate intracellular granules, such as PR3, to the cell surface, triggering the production of ANCAs. In our case, the PR3-ANCA production was derived from the expression of PR3 on the cell surface of apoptotic neutrophils. This is the first report on ENKL describing the mechanism of ANCA development.

Key words: extranodal NK/T-cell lymphoma, nasal type, proteinase 3 antineutrophil cytoplasmic antibodies, neutrophil, apoptosis

(Intern Med 56: 2007-2012, 2017) (DOI: 10.2169/internalmedicine.56.8365)

## Introduction

Extranodal NK/T-cell lymphoma, nasal type (ENKL) is a rare malignant neoplasm associated with Epstein-Barr virus (EBV) infection (1-4). ENKL is characterized by the diffuse proliferation of lymphoma cells expressing NK cell markers such as CD56 (5-7). In addition, the diffuse infiltration of atypical lymphoid cells, an angiocentric and angiodestructive growth pattern, coagulative necrosis, and admixed apoptotic bodies are seen (8). While the infiltration of inflammatory cells, including small lymphocytes, plasma cells, histiocytes, and eosinophils is common, that of neutrophils is not.

ENKL is located in the nasal cavity in 51% of cases; the initial signs and symptoms of the nasal region are often nasal obstruction and chronic rhinorrhea, and nasal septal perforation occurs in 40% of cases (9, 10). For patients with

ENKL, concurrent chemoradiotherapy (CCRT) comprising radiotherapy of 50 Gy and 3 cycles of dexamethasone, etoposide, ifosfamide, and carboplatin (DeVIC) chemotherapy is recommended as a first-line treatment (11).

We herein report a case of ENKL with proteinase 3antineutrophil cytoplasmic antibody (PR3-ANCA)-associated glomerulonephritis. Although PR3-ANCA-associated vasculitis is considered to be a differential diagnosis of ENKL (12), cases of ENKL complicated with PR3-ANCAassociated glomerulonephritis are quite rare. We also discuss the mechanism of PR3-ANCA production in ENKL patients.

#### **Case Report**

A 62-year-old man presented with a 1-year history of nasal obstruction, chronic rhinorrhea, and frequent epistaxis. He gradually developed a fever and lower extremity edema

Received for publication October 6, 2016; Accepted for publication November 15, 2016

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Peripheral blood			PR3-ANCA	28.5 U/mL
WBC	5100	/µL	MPO-ANCA	Negative
RBC	$258 \times 10^{4}$	/µL	anti-GBM antibody	Negative
Hb	7.6	g/dL	IgG	1508 mg/dL
Ht	22.0	%	IgA	471 mg/dL
Plt	$20.6 \times 10^{4}$	/µL	IgM	50 mg/dL
Blood chemistry			C3	80.3 mg/dL
AST	16	IU/mL	C4	24.9 mg/dL
ALT	14	IU/mL	CH50	51.0 U/mL
LDH	196	IU/mL	Anti-nuclear antibodies	Negative
BUN	28.3	mg/dL	HBs antigen	Negative
Cr	3.05	mg/dL	Anti-HCV antibody	Negative
Na	133	mEq/L	Urinalysis	
K	3.8	mEq/L	Gravity	1.009
Cl	99	mEq/L	pH	6.0
CRP	1.16	mg/dL	Protein	+
			Occult blood	3+
Serological tests			Sediments	
Anti-VCA-IgM antibody		Negative	RBC	50-99/HPF
Anti-VCA-IgG antibody		Positive	RBC casts	+
Anti-EBNA antibody		Positive	Urine chemistry	
EBV-DNA	<40	Copies/µgDNA	Protein	1.28 g/gCr

Table. Laboratory Data on Admission.

VCA: viral capsid antigen, EBNA: Epstein-Barr nuclear antigen.

in the six months prior to presentation. Renal dysfunction with hematuria was detected in a primary hospital, so he visited our department for investigation of rapidly progressive glomerulonephritis (RPGN). Laboratory data revealed severe anemia and renal dysfunction with hematuria and proteinuria, in addition to positivity for PR3-ANCAs (Table). His serum creatinine level had been 0.76 mg/dL 6 months prior, and it had increased from 1.55 to 3.05 mg/dL during the 3 months prior to presentation. On ultrasonography, the bilateral kidneys were mildly enlarged with a longitudinal length of 11 cm. Considering the nasal and renal involvement and PR3-ANCA positivity, we suspected granulomatosis with polyangiitis (GPA). However, he exhibited no symptoms of vasculitis in other target organs, and there were no abnormal findings on chest X-ray or computed tomography.

Nasal endoscopy revealed necrotic lesions on the bilateral nasal mucosa, and a nasal biopsy revealed the diffuse proliferation of CD56-positive lymphoma cells (Fig. 1). Cytotoxic molecules, including granzyme B, perforin, and TIA-1, were detected, and EBV-encoded RNA (EBER) *in situ* hybridization was positive (Fig. 2); he was therefore diagnosed with ENKL. FDG-PET findings revealed the tumor to be limited to the nasal cavity, and he was treated with CCRT (13). A renal biopsy was not performed because of antiplatelet therapy for severe coronary artery disease. CCRT gradually led to a reduction in the tumor size, improvement of the renal function and urine abnormalities, and a decline in the PR3-

ANCA level (Fig. 3).

Further investigation by immunohistochemistry was performed to explore the pathophysiology in this case. Infiltrated neutrophils were detected in an angiocentric and angiodestructive growth pattern by myeloperoxidase (MPO) staining (Fig. 4A). Some of them had become apoptotic (14) (Fig. 4B-D). In addition, it was reported recently that the formation of neutrophil extracellular traps (NETs) contributed to the production of PR3-ANCAs by neutrophils. We therefore performed immunostaining of citrullinated histone H3 (Cit-H3) and lactoferrin, which are component proteins of NETs, to investigate the presence of NET formation and its involvement in the subsequent PR3-ANCA production (15, 16). These stains revealed no formation of NETs (Fig. 4E, F).

### Discussion

Two important points should be mentioned in our case; the positivity of PR3-ANCA and the infiltration of neutrophils with apoptosis. We searched the PubMed database using the search terms "extranodal NK T-cell lymphoma nasal type AND case report." We found 56 case reports with the diagnosis of ENKL written in English. Among them, the ANCA titers were measured in six reports (17-22), only one of which report were positive for ANCAs (21). Furthermore, the infiltration of neutrophils was observed in only three reports (23-25). There were no reports describing the infiltra-



**Figure 1.** The findings of nasal endoscopy and a biopsy. (A) There were necrotic lesions on the bilateral nasal mucosa (arrowheads). (B) Hematoxylin and Eosin staining shows the polymorphous tumor cells (magnification: 400×). (C) Lymphoma cells expressing the NK cell marker CD56 were detected (CD56 stain, 400×). (D) Staining for the T cell marker CD3 was positive (CD3 stain, 400×).

tion of neutrophils in cases with ANCAs, nor were there any reports of ANCAs in the cases with infiltration of neutrophils. This is the first report of ENKL with both positivity for ANCAs and the infiltration of neutrophils.

ANCA-associated vasculitis is characterized by necrotizing small-vessel vasculitis and the presence of ANCAs, comprising microscopic polyangiitis (MPA), eosinophilic granulomatosis with polyangiitis, and GPA (26). GPA, formerly known as Wegener's granulomatosis (WG), features necrotizing granulomatous inflammation involving the upper and/or lower respiratory tract; the nasal lesion is usually considered in the differential diagnosis of ENKL. In our case, ENKL was accompanied by PR3-ANCA-associated glomerulonephritis. Furthermore, the PR3-ANCA levels decreased along with the therapy for ENKL. Therefore, the mechanism of PR3-ANCA production was considered to be associated with the entity of ENKL, although dexamethasone in DeVIC chemotherapy may have influenced the course of glomerulonephritis or PR3-ANCA production. In addition, other types of glomerulonephritis, including IgA nephropathy, cannot be completely ruled out.

The pathogenesis of ANCA-associated vasculitis includes several factors. Many genes are etiologic factors and predispose individuals to the disease (27). For example, mutations in alpha 1-antitrypsin, the natural inhibitor of PR3, are found more frequently in patients with WG (28). Infections

with elements, such as Coxsackie B3 or Staphylococcus aureus, may also predispose individuals to the disease since they are triggers for GPA (29, 30). S. aureus encodes genes of serine proteases, which are considered to be crossreactive with ANCAs (30). Several drugs, including propylthiouracil, hydralazine, and antiuratics, have been reported to induce ANCA positivity (31, 32). Malignancy is also associated with the pathogenesis of ANCA and is increased in patients with MPA and GPA (33). Tatsis et al. performed a retrospective analysis of 477 patients with GPA and found a significant association with malignancy, especially renal cell carcinoma, although the immunopathologic mechanisms remained unclear (34). In hematological disease, Savige first investigated the ANCA positivity in hematological chronic malignancies, particularly myelodysplastic syndrome (MDS) (35, 36). Hamidou also studied ANCA positivity in patients with MDS (37). MDS induces immunological abnormalities, leading to the acceleration of apoptosis of neutrophils and susceptibility to recurrent infections (38, 39). Neutrophil alterations, such as apoptosis, may prime neutrophils and induce the translocation of intracellular granules to the cell surface to interact with circulating ANCAs (40). In addition, Harper reported that the priming and apoptosis of neutrophils may be important triggers for the pathogenesis of ANCAs through the mobilization of granules, such as PR3 and MPO, at the cell surface (41). In



**Figure 2.** The findings of *in situ* hybridization and immunohistochemistry. (A) EBV-encoded RNA (EBER) *in situ* hybridization shows that EBV-infected cells, including large numbers of copies of EBERs, were present in the tissue at 400× magnification. In addition, (B) granzyme B stain, (C) TIA-1 stain, and (D) perforin staining confirmed the diagnosis of nasal type, extranodal NK/T-cell lymphoma, (magnification: 400×). EBER *in situ* hybridization was performed using the EBER PNA probe (Dako) and catalyzed signal amplification system (Dako). The commercially available primary antibodies used were as follows: granzyme B (mouse monoclonal, clone GrB-7, Dako), TIA-1 (mouse monoclonal, clone TIA-1, Coulter Immunology), and perforin (mouse monoclonal, clone 5B10, Novo-castra).



**Figure 3.** Clinical course. With concurrent chemoradiotherapy, the renal function and urine abnormalities were improved. In addition, the PR3-ANCA titer was decreased gradually. PR3-ANCA: proteinase 3-antineutrophil cytoplasmic antibody



**Figure 4.** Immunohistochemical findings exploring the mechanism of PR3-ANCA production. (A) Neutrophils infiltrated in an angiocentric and angiodestructive growth pattern (MPO stain 400×). (B) Apoptotic cells were seen (cleaved caspase 3 stain 400×). (C) (D) Some neutrophils (arrowhead in C, D) were identified as apoptotic using serial sections (C; MPO stain 400×, D; cleaved caspase 3 stain 400×). (E) (F) The components of NETs, such as Cit-H3 and lactoferrin, were not detected (E; Cit-H3 stain 400×, F; lactoferrin stain 400×). The commercially available primary antibodies used were as follows: MPO (rabbit polyclonal, Dako), cleaved caspase 3 (rabbit polyclonal, Cell Signaling Technology), Cit-H3 (rabbit polyclonal, Abcam), and lactoferrin (rabbit polyclonal, GenWay Biotech). PR3-ANCA: proteinase 3-antineutrophil cytoplasmic antibody, MPO: myeloperoxidase, NETs: neutrophil extracellular traps, Cit-H3: citrullinated histone H3

our case, given the detection of apoptotic neutrophils, the mechanism of PR3-ANCA production was deemed to involve the expression of PR3 on the cell surface due to the priming and apoptosis of neutrophils, the subsequent exposure of PR3 to the immune system, and the production of PR3-ANCA. However, it has been reported recently that the formation of NETs, which occur as a type of cell death of neutrophil granulocytes and is characterized by the release of chromatin fibers that trap and kill invading microbes extracellularly, leads to targeting autoantigens PR3 and MPO and triggering ANCA-associated vasculitis (42, 43). In our case, staining for the components that form NETs, such as Cit-H3 and lactoferrin, confirmed that NETs had not been formed. Therefore, the development of ANCAs, in our case, was not associated with NETs.

In conclusion, we herein reported a case of ENKL accompanied by RPGN with PR3-ANCA. The production of PR3-ANCAs in our case may have derived from neutrophil apoptosis in the tumor microenvironment. We therefore must consider GPA as a complication as well as include it in the differential diagnosis of ENKL. The authors state that they have no Conflict of Interest (COI).

#### References

- Lin YN, Chou JW, Chuang PH, Cheng KS, Peng CY, Chiang IP. Primary small intestinal natural killer/T cell lymphoma mimicking tuberculous peritonitis: report of a case and review of the literature. Intern Med 50: 515-518, 2011.
- Grywalska E, Markowicz J, Grabarczyk P, Pasiarski M, Rolinski J. Epstein-Barr virus-associated lymphoproliferative disorders. Postepy Hig Med Dosw (Online) 67: 481-490, 2013.
- Kawa K. Diagnosis and treatment of chronic active EBV infection. Uirusu 52: 257-260, 2002 (in Japanese).
- Kawa K. Epstein-Barr virus: associated diseases in humans. Int J Hematol 71: 108-117, 2000.
- 5. Li X, Babayi A, Sang W, et al. Clinicopathologic, immunophenotypic, and EBER in situ hybridization study of extranodal natural killer/T-cell lymphoma, nasal type in amulti-ethnic groups. Clin Lab 60: 419-425, 2014.
- Suzuki R. Pathogenesis and treatment of extranodal natural killer/ T-cell lymphoma. Semin Hematol 51: 42-51, 2014.
- Suzumiya J, Takeshita M, Kimura N, et al. Expression of adult and fetal natural killer cell markers in sinonasal lymphomas. Blood 83: 2255-2260, 1994.

- Swerdlow S, Campo E, Harris NL, et al. WHO Classification of Tumours of Haematopoietic and Lymphoid Tissue. 4th ed. World Health Organization, 2008.
- **9.** Park S, Ko YH. Epstein-Barr virus-associated T/natural killer-cell lymphoproliferative disorders. J Dermatol **41**: 29-39, 2014.
- 10. Ratech H, Burke JS, Blayney DW, Sheibani K, Rappaport H. A clinicopathologic study of malignant lymphomas of the nose, paranasal sinuses, and hard palate, including cases of lethal midline granuloma. Cancer 64: 2525-2531, 1989.
- Yamaguchi M. Current and future management of NK/T-cell lymphoma based on clinical trials. Int J Hematol 96: 562-571, 2012.
- 12. Bhatt VR, Koirala B, Terjanian T. Extranodal natural killer/T cell lymphoma, nasal type presenting as a palatal perforation and nasooral fistula. BMJ Case Rep 2011 bcr1120103511, 2011.
- 13. Yamaguchi M, Tobinai K, Oguchi M, et al. Phase I/II study of concurrent chemoradiotherapy for localized nasal natural killer/Tcell lymphoma: Japan Clinical Oncology Group Study JCOG0211. J Clin Oncol 27: 5594-5600, 2009.
- Tsutsumi Y, Kanoshida S. Pitfalls and caveats in histochemically demonstrating apoptosis. Acta Histochem Cytochem 36: 271-280, 2003.
- 15. Kazuya S, Takanori O, Yasuyoshi M, Kouhei S, Ken-ichi I, Yutaka T. Visualization of neutrophil extracellular traps and fibrin meshwork in human fibrinopurulent inflammator lesions: I. light microscopic study. Acta Histochem Cytochem 49: 109-116, 2016.
- 16. Onouchi T, Shiogama K, Matsui T, et al. Visualization of neutrophil extracellular traps and fibrin meshwork in human fibrinopurulent inflammatory lesions: II. Ultrastructural Study. Acta Histochem Cytochem 49: 117-123, 2016.
- 17. Fei W, Xiaohong W, Hong Z, Bei H. Pulmonary extranodal natural killer/T-cell lymphoma (nasal type): a case report and radiological image review. Medicine (Baltimore) 94: e1527, 2015.
- 18. Terroso G, Aleixo J, Bernardes M, Mariz E, Fonseca E, Costa L. Nasal type extranodal NK/T cell lymphoma diagnosed in a patient with rheumatoid arthritis under methotrexate. Acta Reumatol Port 39: 77-81, 2014.
- 19. Shimizu I, Hamano Y, Sato S, et al. Neurolymphomatosis in a patient with extranodal NK/T-cell lymphoma, nasal-type: a case report and literature review. Intern Med 53: 471-475, 2014.
- 20. Fujimoto N, Takahashi T, Yamashita M, Nakanishi G, Okabe H, Tanaka T. Extranodal natural killer/T-cell lymphoma, nasal type, with prominent granulomatous reaction. J Dermatol 41: 68-69, 2014.
- Coha B, Vucinic I, Mahovne I, Vukovic-Arar Z. Extranodal lymphomas of head and neck with emphasis on NK/T-cell lymphoma, nasal type. J Craniomaxillofac Surg 42: 149-152, 2014.
- 22. Li Z, Liu C, Huang X, Gao Z. Nasal NK/T cell lymphoma with severe facial disfigurement in a 37-year-old male. Int J Oral Maxillofac Surg 42: 102-105, 2013.
- 23. Aladily TN, Nathwani BN, Miranda RN, et al. Extranodal NK/T-cell lymphoma, nasal type, arising in association with saline breast implant: expanding the spectrum of breast implant-associated lymphomas. Am J Surg Pathol 36: 1729-1734, 2012.
- **24.** Jiang QP, Liu SY, Yang YX, et al. CD20-positive NK/T-cell lymphoma with indolent clinical course: report of case and review of literature. Diagn Pathol **7**: 133, 2012.
- **25.** Rezk SA, Huang Q. Extranodal NK/T-cell lymphoma, nasal type extensively involving the bone marrow. Int J Clin Exp Pathol **4**: 713-717, 2011.
- 26. Lapraik C, Watts R, Bacon P, et al. BSR and BHPR guidelines for the management of adults with ANCA associated vasculitis. Rheu-

matology (Oxford) 46: 1615-1616, 2007.

- 27. Gencik M, Borgmann S, Zahn R, et al. Immunogenetic risk factors for anti-neutrophil cytoplasmic antibody (ANCA)-associated systemic vasculitis. Clin Exp Immunol 117: 412-417, 1999.
- 28. Callea F, Gregorini G, Sinico A, et al. alpha 1-Antitrypsin (AAT) deficiency and ANCA-positive systemic vasculitis: genetic and clinical implications. Eur J Clin Invest 27: 696-702, 1997.
- Barrett TG, Taylor CM, Thomason P, Pall A, Adu D. Environmental trigger for anti-neutrophil cytoplasmic antibodies? Lancet 342: 369-370, 1993.
- 30. Stegeman CA, Tervaert JW, Sluiter WJ, Manson WL, de Jong PE, Kallenberg CG. Association of chronic nasal carriage of *Staphylococcus aureus* and higher relapse rates in Wegener granulomatosis. Ann Intern Med 120: 12-17, 1994.
- 31. Nassberger L, Johansson AC, Bjorck S, Sjoholm AG. Antibodies to neutrophil granulocyte myeloperoxidase and elastase: autoimmune responses in glomerulonephritis due to hydralazine treatment. J Intern Med 229: 261-265, 1991.
- 32. Dolman KM, Gans RO, Vervaat TJ, et al. Vasculitis and antineutrophil cytoplasmic autoantibodies associated with propylthiouracil therapy. Lancet 342: 651-652, 1993.
- Pankhurst T, Savage CO, Gordon C, Harper L. Malignancy is increased in ANCA-associated vasculitis. Rheumatology (Oxford) 43: 1532-1535, 2004.
- 34. Tatsis E, Reinhold-Keller E, Steindorf K, Feller AC, Gross WL. Wegener's granulomatosis associated with renal cell carcinoma. Arthritis Rheum 42: 751-756, 1999.
- 35. Savige JA, Chang L, Smith CL, Duggan JC. Myelodysplasia, vasculitis and anti-neutrophil cytoplasm antibodies. Leuk Lymphoma 9: 49-54, 1993.
- 36. Savige JA, Chang L, Smith CL, Duggan JC. Anti-neutrophil cytoplasmic antibodies (ANCA) in myelodysplasia and other haematological disorders. Aust N Z J Med 24: 282-287, 1994.
- 37. Hamidou MA, Derenne S, Audrain MA, Berthelot JM, Boumalassa A, Grolleau JY. Prevalence of rheumatic manifestations and antineutrophil cytoplasmic antibodies in haematological malignancies. A prospective study. Rheumatology (Oxford) 39: 417-420, 2000.
- Enright H, Miller W. Autoimmune phenomena in patients with myelodysplastic syndromes. Leuk Lymphoma 24: 483-489, 1997.
- 39. Billstrom R, Johansson H, Johansson B, Mitelman F. Immunemediated complications in patients with myelodysplastic syndromes: clinical and cytogenetic features. Eur J Haematol 55: 42-48, 1995.
- 40. Gilligan HM, Bredy B, Brady HR, et al. Antineutrophil cytoplasmic autoantibodies interact with primary granule constituents on the surface of apoptotic neutrophils in the absence of neutrophil priming. J Exp Med 184: 2231-2241, 1996.
- 41. Harper L, Cockwell P, Adu D, Savage CO. Neutrophil priming and apoptosis in anti-neutrophil cytoplasmic autoantibodyassociated vasculitis. Kidney Int 59: 1729-1738, 2001.
- Kessenbrock K, Krumbholz M, Schonermarck U, et al. Netting neutrophils in autoimmune small-vessel vasculitis. Nat Med 15: 623-625, 2009.
- 43. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. Science 303: 1532-1535, 2004.

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