Marrow angiogenesis-associated factors as prognostic biomarkers in patients with acute myelogenous leukaemia

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Bone marrow (BM) neoangiogenesis plays an important role in acute myelogenous leukaemia (AML), and depends on the interplay of members of the vascular endothelial growth factor (VEGF) and angiopoietin (Ang) families. We determined the marrow levels of seven molecules associated with angiogenesis in 52 AML patients before chemotherapy and 20 healthy controls: VEGF-A, VEGF/PIGF, VEGF-C, VEGF-D, Ang-I, Ang-2, and Tie-2. All the molecules were quantified using enzyme-linked immunosorbent assay (ELISA). Comparing to normal controls, the marrow levels of VEGF/PIGF, Ang-2, and Tie-2 were significantly higher, and those of VEGF-C and Ang-I were significantly lower in the AML patients (P < 0.001). A total of 31 patients were further subjected to survival analysis. Patients with lower Tie-2 (P < 0.001) and Ang-2 levels (P < 0.001) and VEGF-D levels (P < 0.001) and 0.042, respectively), same as patients with higher VEGF/PIGF (P < 0.001) and VEGF-D levels (P < 0.001) and malysis revealed that patients with higher angio-index values (P < 0.001) displayed poor prognosis (hazard ratio 5.91, 95% confidence interval 1.99–17.56; P = 0.001). The angio-index is closely associated with the clinical outcome of AML patients and may be valuable in disease prognosis

British Journal of Cancer (2007) **97,** 877–882. doi:10.1038/sj.bjc.6603966 www.bjcancer.com Published online 11 September 2007 © 2007 Cancer Research UK

Keywords: prognostic biomarker; AML; angiopoietin family; VEGF family

Vascular endothelial growth factor (VEGF) and angiopoietin (Ang) are the two families involved in the regulation of vascular angiogenesis. The VEGF family comprises six members: VEGF-A, -B, -C, -D, -E, and placental growth factor (PlGF). They share three receptors, ie VEGFR-1, -2, and -3 (Korpelainen and Alitalo, 1998; Podar and Anderson, 2005). The Ang family includes four members: Ang-1, -2, -3, and -4. However, they share only a single receptor, ie Tie-2 (Ward and Dumont, 2002).

Both *in vitro* and *in vivo* investigations have revealed that VEGF-A was present in a high concentration in the bone marrow (BM) cells of AML patients and the vast majority of myeloblasts (>90%) could synthesise and secrete VEGF-A (Ghannadan *et al*, 2003). Compared to the levels in the control, increased plasma levels of VEGF-A obtained from peripheral blood (PB) were observed in AML patients (Aguayo *et al*, 2000, 2002). Elevated VEGF-C expression was reported in patients with prostatic carcinoma and oesophageal squamous cell carcinoma (Tsurusaki *et al*, 1999; Kitadai *et al*, 2001) but is controversial with regard to AML patients (Fielder *et al*, 1997; Dias *et al*, 2002; Loges *et al*, 2005). In contrast, decreased VEGF-D expression levels were

Received 8 June 2007; revised 31 July 2007; accepted 8 August 2007; published online 11 September 2007

reported in patients with lung adenocarcinoma and colorectal carcinoma (Niki et al, 2000; Onogawa et al, 2004). However, no previous reports addressed plasma VEGF-C and -D levels.

Both Ang-1 and -2 can only bind to the Tie-2 receptor, but they have opposite effects on Tie-2 activation. Tie-2 is a receptor tyrosine kinase that is expressed on endothelial cells (ECs) and haematopoietic stem cells (HSCs). Ang-1 functions as a stabilising signal for mature vasculature, and Ang-2 can be regarded as a regulator of vessel plasticity (Jones, 2003). In the BM niche, Ang-1/ Tie-2 signalling occurs during interaction between Tie-2-expressing HSCs and Ang-1-expressing osteoblasts (Arai et al, 2004; Arai et al, 2005); this maintains the HSCs in the quiescence and antiapoptotic state. Data from recent studies indicate that Ang-2 expression represents an independent prognostic factor in AML (Loges et al, 2005). However, the limitation of these studies was that plasma samples of the study patients were not available; thus, the investigation was restricted to the analysis of the mRNA expression of angiogenic factors. Therefore, precise data on the protein concentrations are essential.

Neoangiogenesis depends on the interplay of different members of the VEGF and Ang family, and many of these factors that are present in significant amounts in the marrow environment may function in a synchronised fashion in addition to any local autocrine or paracrine effects they may have; therefore, comprehensive analyses on marrow levels of these factors in AML patients and the clinical implications of these levels are important and remain to be explored.

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MATERIALS AND METHODS

Study subjects

Prior to chemotherapy, marrow samples were obtained from 52 newly diagnosed AML patients at National Taiwan University Hospital (Taipei, Taiwan) from 1998 to 1999. This study was approved by the Institutional Review Board of the hospital (serial no 9461712129), and written informed consent was obtained from all patients. The study included 33 males and 19 females. The ages of the patients ranged from 2 to 90 years, with a median age of 45 years. According to the French-American-British (FAB) classification, two patients were classified as M0; 15, M1; 17, M2; 7, M3; 8, M4; 2, M5; and 1, unclassified. Of these, 14 patients did not undergo chemotherapy or were only treated with low-dose cytosine arabinoside due to old age and/or poor performance status; all the other patients with non-M3 AML subtypes underwent conventional induction chemotherapy with one of the anthracyclines (doxorubicin or idarubicin) for 3 days and cytosine arabinoside for 7 days. The patients with acute promyelocytic leukaemia (subtype M3) received all-trans retinoic acid with or without concurrent induction chemotherapy. After the patients achieved complete remission (CR), they underwent consolidation chemotherapy with a conventional dose of cytosine arabinoside and one anthracycline or with high-dose cytosine arabinoside. On the basis of tri-lineage cell regeneration with <5% blasts in the BM, normalisation of the PB cell count and the absence of leukaemia infiltration in the tissue, the patients were considered to have achieved CR. In addition, 20 healthy marrow donors were included as healthy controls in this study; of these, 10 were males and 7 females. The age of the control patients ranged from 11 to 56 years, with a median age of 32 years. After collection, the marrow samples were immediately centrifuged, and the plasma was separated and stored at -20 °C until investigation.

Enzyme-linked immunosorbent assay

One-step sandwich enzyme immunoassay was performed to measure the VEGF-A, VEGF/PIGF, VEGF-C, VEGF-D, Ang-1,

Table I Characteristics of the study groups

Ang-2, and Tie-2 concentrations by using commercially available kits (R&D systems, Minneapolis, MN, USA) according to the manufacturer's instructions.

The results represent the mean values of the duplicate determinations and were calculated using the SoftMax Pro 4.8 software (Molecular Devices Corporation, Sunnyvale, CA, USA).

Statistical methods

Student's t-test and analysis of variance (ANOVA) were used to analyse the angiogenesis factors in the age, sex, FAB subtype, and karyotype subgroups of the AML patients. Spearman's test was used to analyse the correlation between the factors. The receiver operating characteristic (ROC) curve was constructed, and the area under the ROC curve (AUROC) was calculated for each factor to differentiate the patients from the controls. The paired t-test was used for patients who achieved CR to determine the difference between the concentrations of the various factors at presentation and CR. Survival status was investigated by using the Kaplan–Meier survival curve and log-rank test. Univariate and multivariate Cox regression analyses were also used to estimate prognosis. All the statistical analyses were performed by using the MedCalc software (http://www.medcalc.be/) and the SPSS 12.0 statistical software (SPSS Inc., Chicago, IL, USA).

RESULTS

Lack of correlation between plasma levels of seven angiogenesis-related factors in marrow and the egress of leukaemic cells from the BM to PB

The seven angiogenesis-related factors (VEGF-A, VEGF/PlGF, VEGF-C, VEGF-D, Ang-1, Ang-2, and Tie-2) in 52 AML patients at presentation were investigated, and no significant differences were observed in the age and FAB subtypes (Table 1). However, Tie-2 level was higher in males than that in females (P = 0.024),

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	No.	VEGF (pg ml ⁻¹)	VEGF/PIGF (pg ml ⁻¹)	VEGF-C (pg ml ⁻¹)	VEGF-D (pg ml ⁻¹)	Ang-I (pg ml ⁻¹)	Ang-2 (pg ml ⁻¹)	Tie-2 (ng ml ⁻¹)
Age								
<45	25	47.85 ± 43.85	8.52 ± 10.63	378.89 ± 258.09	614.55 ± 381.32	1344.02 ± 939.19	3977.02 ± 2741.20	23.89 ± 8.15
≥45	27	31.49 ± 19.37	19.02 ± 37.01	389.53 ± 247.86	879.69 ± 582.55	1509.71 ± 939.30	2936.34 ± 2742.42	21.81 ± 7.60
P-value		0.096	0.168	0.880	0.057	0.528	0.178	0.346
Sex								
Male	33	41.40 ± 36.56	15.45 ± 31.14	391.74 ± 251.72	844.10 ± 560.19	1279.31 ± 800.31	3192.22 ± 2513.06	24.52 ± 8.36
Female	19	35.81 ± 30.03	11.41 ± 21.78	371.70 ± 254.39	592.64 ± 366.79	1691.86 ± 1103.48	3861.23 ± 3183.72	19.83 ± 6.03
P-value		0.554	0.587	0.785	0.057	0.164	0.438	0.024 ^a
FAB subtype								
M0 //	2	44.13 ± 35.22	9.09 ± 9.20	282.84 ± 21.65	829.40 ± 517.51	1725.07 ± 915.97	1213.66 ± 585.51	24.32 ± 0.74
MI	15	35.48 ± 44.46	3.39 ± 3.68	348.72 ± 186.96	764.07 ± 703.42	1480.47 ± 659.10	3283.47 ± 2327.49	23.40 ± 5.71
M2	17	37.55 ± 25.65	26.76 ± 40.63	417.03 ± 338.62	726.52 ± 462.79	1735.12 ± 1350.26	3362.76 ± 3219.54	20.18 ± 8.67
M3	7	71.88 ± 37.89	7.80 ± 10.92	413.84 ± 131.58	687.09 ± 279.06	783.75 ± 269.15	3951.64 ± 3160.14	24.29 ± 3.90
M4	8	26.14 ± 14.28	17.32 ± 31.84	459.35 ± 263.12	667.53 ± 321.93	1464.84 ± 440.60	4131.93 ± 3134.01	24.75 ± 8.85
M5	2	20.11 ± 1.35	4.35 ± 5.22	89.07 ± 10.65	820.14 ± 272.82	545.18 ± 137.94	3460.07 ± 1308.26	28.80 ± 24.15
P-value		0.133	0.275	0.505	0.996	0.211	0.855	0.583
Cytogenetic subtyp	e							
Favourable	7	55.14 ± 45.01	8.44 ± 10.51	369.97 ± 159.36	702.40 ± 271.53	988.34 ± 848.59	2952.35 ± 2516.61	21.87 ± 5.73
Intermediate	29	37.08 ± 37.03	18.48 ± 35.32	426.92 ± 301.19	726.98 ± 445.84	1735.36 ± 1029.81	3588.60 ± 3129.72	24.55 ± 9.18
Unfavourable	16	36.58 ± 21.30	8.22 ± 13.77	313.69 ± 157.26	819.76 ± 688.43	1069.93 ± 535.84	3373.17 ± 2240.56	20.07 ± 5.06
P-value		0.429	0.434	0.351	0.817	0.026 ^a	0.861	0.179

^aDenotes significant difference. Abbreviations: Ang = angiopoietin; ANOVA = analysis of variance; FAB = French - American - British; PIGF = placental growth factor, VEGF = vascular endothelial growth factor. Age and sex were analysed using Student's t test, and FAB subtypes and cytogenetic subtype were analysed using ANOVA. Favourable cytogenetic subtype: t(15; 17), t(8; 21), and inv(16); unfavourable: -5, -7, +8, and complex; and intermediate: normal cytogenetic subtype and others.

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and the Ang-1 level was significantly different among the karyotype subgroups (P = 0.026).

To clarify whether these seven molecules present in the marrow microenvironment could determine the extent of egress of immature haematopoietic cells from the BM to PB, we correlated the absolute number of immature cells in the PB with the marrow levels of these molecules at disease presentation. We found that the marrow levels did not correlate with the egress of leukaemic cells from the BM to PB (VEGF-A, r = -0.050; VEGF/PlGF, r = 0.229; VEGF-C, r = 0.229; VEGF-D, r = 0.014; Ang-1, r = 0.406; Ang-2, r = 0.169; and Tie-2, r = 0.516).

Significant difference in the levels of VEGF/PlGF, VEGF-C, Ang-1, Ang-2, and Tie-2 between healthy controls and AML patients

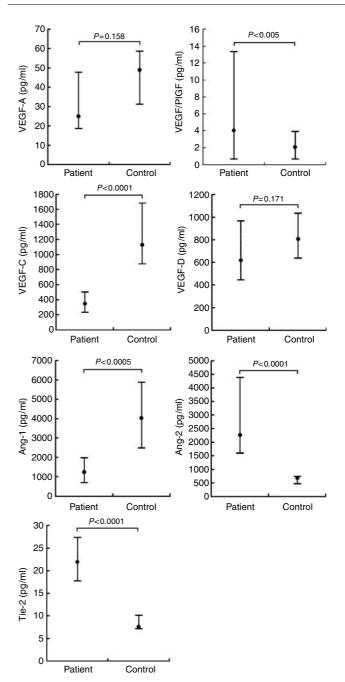
The marrow levels of these seven factors in the 52 AML patients at presentation were compared to those observed in the 20 healthy controls (Figure 1). Significant differences were observed in marrow levels of VEGF/PlGF, VEGF-C, Ang-1, Ang-2, and Tie-2 (P < 0.005, P < 0.0001, P < 0.0005, P < 0.0001, and P < 0.0001,respectively). The ROC curve illustrated the cut-off value for each factor from which we could differentiate AML patients from the healthy controls (Figure 2). Among the seven factors, Ang-2 and Tie-2 displayed excellent results, with cut-off values of $1039.77 \text{ pg ml}^{-1}$ and 11.48 ng ml^{-1} , respectively. The AUROC was 0.993 (95% confidence interval (CI), 0.937-0.995; P < 0.001) and 0.977 (0.910-0.997; P < 0.001), respectively. VEGF-C and Ang-1 displayed good results, with cut-off values of 664.20 and 2556.46 pg ml⁻¹, respectively, and the AUROC for these was 0.859 (0.756-0.929, P < 0.001) and 0.861 (0.759-0.931, P < 0.001), respectively. The remaining three molecules, ie VEGF-A, VEGF/ PIGF, and VEGF-D displayed poor results, with an AUROC of 0.693 (0.573 - 0.796, P = 0.009), 0.631 (0.509 - 0.742, P = 0.063), and 0.667(0.546 - 0.774, P = 0.025), respectively.

Sequential analysis of VEGF/PIGF, Ang-2, Tie-2, and VEGF-C levels in AML patients during the chemotherapy course

Of the 31 AML patients undergoing conventional induction chemotherapy, 22 (71.0%) patients achieved CR after treatment. The marrow levels of VEGF/PIGF, Ang-2, Tie-2, and VEGF-C in the AML patients at presentation were compared to those observed at the CR stage. VEGF/PlGF and Ang-2 levels increased, while Tie-2 and VEGF-C levels decreased significantly (P < 0.005, P < 0.005, P = 0.019, and P = 0.038, respectively). All of these values were trend to normalise at CR stage as we expected, but they did not fit into normal range (Figure 3).

Correlation of marrow levels of VEGF/PlGF, VEGF-D, and Tie-2 with prognosis of AML

Further investigation was performed by using the Kaplan-Meier survival curve and log-rank test to evaluate the suitability of these molecules as prognostic factors. Of 31 AML patients undergoing conventional induction chemotherapy were divided into subgroups with high and low levels based on their distributions. The median follow-up period of the 31 patients was 12.0 months (range, 0.5-66.1 months). A 5-year survival analysis revealed significant differences in VEGF/PlGF, VEGF-D, Ang-2, and Tie-2 levels among the subgroups (Figure 4). Groups that demonstrated marrow VEGF/PlGF levels higher than 1 $pg ml^{-1}$ or VEGF-D levels higher than 350 pg ml⁻¹ demonstrated a greater probability of survival (P = 0.020, 13.0 vs 26.2 months for VEGF/PIGF and P = 0.016, 2.5 vs 22.1 months for VEGF-D). However, groups with marrow Tie-2 levels higher than 26 ng ml⁻¹ or Ang-2 levels higher than $4500 \,\mathrm{pg}\,\mathrm{ml}^{-1}$ demonstrated a poor prognosis (P = 0.037, 26.2vs 17.3 months for Tie-2 and P = 0.042, 18.9 vs 7.2 months for



Marrow levels of vascular endothelial growth factor (VEGF)-A, VEGF/PIGF, VEGF-C, VEGF-D, angiopoietin (Ang)-1, Ang-2, and Tie-2 in 52 AML patients at presentation and in 20 healthy controls. Lower and upper lines indicate the 25th and 75th percentiles, respectively. The dot in each bar denotes the median. The P-value was calculated using the Student's t-test for the AML patient and control groups.

Ang-2). The remaining angiogenesis factors did not reveal any effect on overall survival. Based on the above results, we devised an algorithm known as the angio-index that collectively considered four distinct markers: (Ang-2 in pg ml⁻¹ × Tie-2 in ng ml⁻¹)/ (VEGF/PlGF in pg ml $^{-1}$ × VEGF-D in pg ml $^{-1}$). We discovered that subgroups with an angio-index greater than 50 had substantially poor survival probabilities (P = 0.001, 7.2 vs 26.2 months; Figure 4). In addition, we also analysed the 5-year survival of patients whether or not they achieved CR after standard chemotherapy, and discovered that the subgroups that achieved CR had a considerably better probability of survival (P < 0.001, 26.2 vs 2.0 months; Figure 4). By comparing the probability of CR and the angio-index

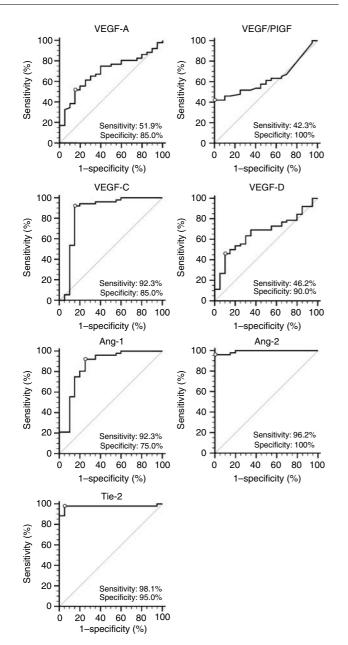


Figure 2 ROC curve of the angiogenesis factors in 52 AML patients at presentation and in 20 healthy controls. The white dots are the cut-off values between the patients and controls: vascular endothelial growth factor (VEGF)-A, 27.00 pg ml $^{-1}$; VEGF/PIGF, 5.87 pg ml $^{-1}$; VEGF-C, 664.20 pg ml $^{-1}$; Ang-I, 2556.46 pg ml $^{-1}$; Ang-2, 1039.77 pg ml $^{-1}$; and Tie-2, 11.48 ng ml $^{-1}$.

at diagnosis, we found that 18 of 20 (90%) patients with an angioindex less than 50 achieved CR after standard treatment; this suggests that the angio-index might be an appropriate indicator for the prediction of disease outcome.

In addition, by using the univariate Cox regression model, the angio-index displayed prominent prognosis value, with a hazard ratio of 4.38 (95% CI, 1.65-11.60, P=0.003); after adjusting for age and sex in a multivariate model, the hazard ratio may reach 5.91 (1.99-17.56, P=0.001).

DISCUSSION

Marrow neoangiogenesis in AML is a complex process involving the interplay of different angiogenic growth factors. Until now,

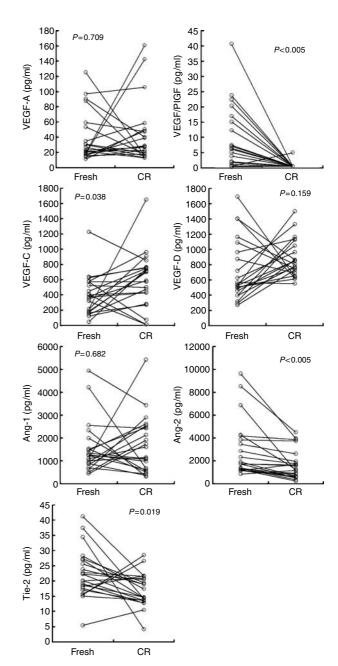


Figure 3 Levels of the angiogenesis factors in 22 patients at diagnosis and at complete remission (CR). Concentrations at the newly diagnosed (fresh) and first CR stages were compared. The P-value was calculated using the paired t-test.

most investigators have attempted to elucidate the impact of a single angiogenic factor on the pathogenesis or prognosis of haematologic malignancies (Aguayo et al, 2002; Litwin et al, 2002; Moehler et al, 2003; Liu et al, 2005; Nowicki et al, 2006). However, HSCs were observed to be in close contact with the marrow stromal cells in vivo (Oostendorp and Dormer, 1997; Tordjman et al, 1999). Furthermore, both fibroblasts and microvascular ECs obtained in normal and leukaemic marrows were reported to produce several angiogenesis molecules (Moehler et al, 2003; De Raeve et al, 2004; Ogawa et al, 2006). Therefore, because of the synchronised action of various angiogenic cytokines, it is important to elucidate the marrow levels of the various angiogenesis factors present in the marrow microenvironment. The current study is the first report on the concentrations of the various angiogenesis factors in the marrow microenvironment of AML



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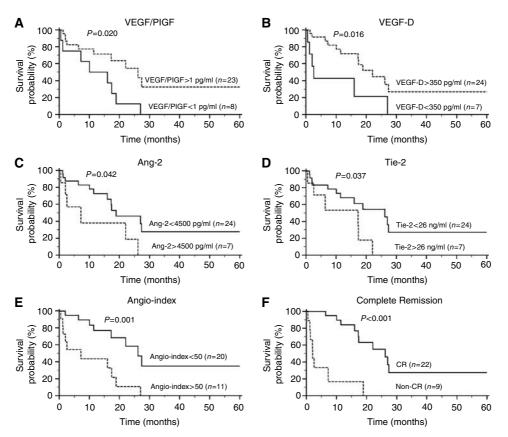


Figure 4 Kaplan—Meier survival curves for the angiogenesis factors, VEGF/PIGF (**A**), VEGF-D (**B**), Ang-2 (**C**), and Tie-2 (**D**), as well as angio-index (**E**) and complete remission achievement (**F**). Each angiogenesis factor was divided into high (dot line) and low (solid line) concentration subgroups based on the distribution of its level. The angio-index was divided into high (dot line) and low (solid line) subgroups with a cut-off value of 50. The *P*-value was calculated using the log-rank test. The angio-index stands for the formula (Ang-2 × Tie-2)(VEGF/PIGF × VEGF-D).

patients. We found that the marrow levels of VEGF/PlGF, Ang-2, and Tie-2 were significantly higher (P < 0.001) and those of VEGF-C and Ang-1 were significantly lower (P < 0.001) in the AML patients as compared to those values in healthy controls. In addition, we found that patients with lower VEGF/PIGF and VEGF-D levels or higher Ang-2 and Tie-2 levels displayed poor prognosis (P = 0.020, 0.016, 0.042, and 0.037, respectively). We used the algorithm angio-index ((Ang-2 \times Tie-2)/(VEGF/PlGF \times VEGF-D)) and achieved the most accurate separation of patients with a highrisk signature from those with a low-risk signature (P = 0.001). Further, the AML patients with higher angio-index values displayed poor prognosis, a hazard ratio of 4.38; suggesting that the combined marrow levels of VEGF/PIGF, VEGF-D, Ang-2, and Tie-2 facilitated prediction of prognosis. To our knowledge, this is the first report that evaluated prognosis by devising an algorithm that combined four distinct angiogenesis-associated factors.

The receptor tyrosine kinase Tie-2 is expressed in the ECs, early HSCs, and the so-called leukaemic cells (Arai et al, 2004; Arai et al, 2005). Ang-1, but not Ang-2, induces autophosphorylation of the Tie-2 receptor upon binding (Korpelainen and Alitalo, 1998; Jones, 2003; Thurston, 2003; Peters et al, 2004). Therefore, Ang-1 and Ang-2 are believed to function as naturally occurring antagonists. The cellular receptor Tie-2 shedding from the cell surface results in the release of a soluble receptor from Tie-2-expressing cells (Reusch et al, 2001). Consequently, the soluble receptor can be detected in human serum and plasma. Recently, Schliemann et al (2006) used an immunohistochemical stain to study BM biopsy samples and discovered that both Ang-2 and Tie-2 were overexpressed in the leukaemic blasts of AML patients; however, Ang-1 was not Loges et al (2005) employed quantitative RT-PCR to study mRNA expression and discovered that the median levels of

VEGF-A, VEGF-C, Ang-1, and Ang-2 were higher in PB blasts than in normal PB AC133⁺ cells. In our study, we also found a similar trend for Ang-2 and Tie-2 in the AML patients; nevertheless, we further revealed significant differences in the Ang-1, VEGF/PlGF, and VEGF-C levels between the AML patients and healthy controls. These reports demonstrated that the Ang-2 and Tie-2 levels displayed a similar trend in AML patients with regard to cellular mRNA and proteins and extracellular proteins. However, similar trends were not observed for VEGF-A, VEGF-C, and Ang-1 levels; this suggests that the distribution of three angiogenesis-associated factors in the cells and microenvironment may vary, and while some display a positive association, others display a negative association. Therefore, the molecules in intracellular and extracellular environments exhibited different implications in prognosis prediction. Schliemann et al (2006) discovered that patients expressing high intracellular Ang-2 levels in leukaemic cells displayed a significantly extended overall survival than that displayed by patients with low intracellular Ang-2 levels Loges et al (2005) discovered that higher Ang-2 mRNA levels in PB blasts tend to indicate a better prognosis in AML patients. However, in our study, high marrow levels of extracellular Ang-2 indicated a poor prognosis. This diversity might be from different sample origin, one was from PB and the other was from marrow.

In the angiogenesis process, VEGF-A is usually present in its homodimeric form. It can also form a heterodimer with PIGF, and this stimulates angiogenesis via the VEGFR-1/VEGFR-2 heterodimer (Autiero et al, 2003; Roy et al, 2006); this is consistent with our finding that VEGF/PIGF was present at substantially higher levels in AML patients than in the healthy controls. Loges et al (2005) reported that the AML patients and healthy donors

demonstrated no significant difference in the level of VEGF-C mRNA expression. Fielder *et al* (1997) reported that leukaemic cells expressed VEGF-C in most of the AML patients. Dias *et al* (2002) found that VEGF-C that was released from the endothelium could induce leukaemic cell proliferation, promote their survival, and protect them from chemotherapy-induced apoptosis. In our study, we observed a difference in the marrow VEGF-C levels between the AML patients and healthy controls, but we could not predict the prognosis of the AML patients based on these levels (P=0.591).

Our previous report (Lin *et al*, 2002) addressed the close relationship between the marrow matrix metalloproteinase (MMP)-9 level and the disease status of AML; the marrow MMP-9 level may be a useful surrogate marker for monitoring the disease status in AML patients. However, the survival times of AML

patients with lower MMP-9 levels were only slightly longer than those of AML patients with higher levels ($P\!=\!0.12$). As observed in this report, the angio-index involving four angiopoiesis-associated factors was an appropriate prognostic marker ($P\!=\!0.001$). Therefore, the estimation of the marrow MMP-9, VEGF/PIGF, VEGF-D, Ang-2, and Tie-2 levels may be helpful in predicting the disease status of AML patients. Further prospective and multi-centre studies would be worthwhile to verify the wide application of these biomarkers.

ACKNOWLEDGEMENTS

This study was supported by a grant NSC95-2320-B-002-084 from National Science Council, Taiwan

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