

Guest editorial:

EXPRESSION CLASSIFIERS FOR DEVELOPMENTAL TOXICANTS

Raymond Reif

Leibniz Research Centre for Working Environment and Human Factors at TU Dortmund (IfADo), Ardeystrasse 67, 44139 Dortmund, Germany; reif@ifado.de

<http://dx.doi.org/10.17179/excli2015-765>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>).

In its recent edition the Archives of Toxicology have published a transcriptome-based classifier that identifies developmental toxicants acting as histone deacetylase inhibitors. The authors, Eugen Rempel and colleagues from Dortmund Technical University, demonstrate that based on only eight genes the gene expression pattern of developmental toxicants acting as histone deacetylase inhibitors can be identified. They used a test system that recapitulates generation of neuroectoderm from pluripotent stem cells (Balmer et al., 2014; Zimmer et al., 2014; Leist et al., 2013; Stöber et al., 2014; Reif, 2014). Exposure of the test cells for six days to six histone deacetylase inhibitors (HDACi) caused alterations in expression profiles that could be clearly differentiated from other compound classes, such as a heterogeneous group of ‘mercurials’.

Currently, classification and grouping represents a cutting-edge topic in toxicological research (Gocht et al., 2015; Godoy et al., 2015; 2013; Grinberg et al., 2014; Shinde et al., 2015; Meganathan et al., 2015; Weng et al., 2014; Krause et al., 2013; Gebel et al., 2014). To reach this goal in the field of developmental toxicity, stem cell based test systems have been developed that recapitulate different phases of human development (Krug et al., 2013; Leist et al., 2013; Bolt, 2013; Hoelting et al., 2013). Using these systems it has been shown that three concentra-

tion ranges can be differentiated, namely tolerated concentrations where no gene expression changes occur, the teratogenic range which leads to deregulation of critical developmental genes and the cytotoxic concentration range where additional genes associated with cell death and catabolic metabolism are observed (Waldmann et al., 2014). Importantly, the ‘teratogenic concentration range’ *in vitro* overlapped with concentrations known to cause teratogenic effects in humans *in vivo*. A next challenge in test system development is to develop expression signatures which indicate that specific toxic mechanisms are active. In this respect the present work of Rempel et al., using a set of structurally not related compounds acting by a similar mechanism, represents an important proof of concept (Rempel et al., 2015). Next open questions to be addressed are whether the test systems correctly identify further classes of developmental compounds and differentiate them from compounds acting by unspecific toxic mechanisms or substances that preferentially cause other types of toxicities. In the past and also presently developmental toxicity has mostly been tested *in vivo* (Lee et al., 2011; 2007; Liu et al., 2010; Moss et al., 2009; Xi et al., 2009; Oesch et al., 2008; Stapleton and Chan, 2009; Ejaz and Woong, 2006). Although some first important steps have been achieved in this field of *in vitro* research there seems to be a long

way to go until animal experiments in developmental toxicity can be fully replaced by alternative methods.

REFERENCES

- Balmer NV, Klima S, Rempel E, Ivanova VN, Kolde R, Weng MK, et al. From transient transcriptome responses to disturbed neurodevelopment: role of histone acetylation and methylation as epigenetic switch between reversible and irreversible drug effects. *Arch Toxicol.* 2014;88:1451-68.
- Bolt HM. Developmental neurotoxicity testing with human embryonic stem cell-derived in vitro systems: the novel FP7 ESNATS tests are available. *Arch Toxicol.* 2013;87:5-6.
- Ejaz S, Woong LC. Diminished embryonic movements of developing embryo by direct exposure of sidestream whole smoke solutions. *Arch Toxicol.* 2006;80:107-14.
- Gebel T, Foth H, Damm G, Freyberger A, Kramer PJ, Lilienblum W, et al. Manufactured nanomaterials: categorization and approaches to hazard assessment. *Arch Toxicol.* 2014;88:2191-211.
- Gocht T, Berggren E, Ahr HJ, Cotgreave I, Cronin MT, Daston G, et al. The SEURAT-1 approach towards animal free human safety assessment. *ALTEX.* 2015;32:9-24.
- Godoy P, Hewitt NJ, Albrecht U, Andersen ME, Ansari N, Bhattacharya S, et al. Recent advances in 2D and 3D in vitro systems using primary hepatocytes, alternative hepatocyte sources and non-parenchymal liver cells and their use in investigating mechanisms of hepatotoxicity, cell signaling and ADME. *Arch Toxicol.* 2013;87:1315-530.
- Godoy P, Schmidt-Heck W, Natarajan K, Lucendo-Villarin B, Szkolnicka D, Asplund A, et al. Gene networks and transcription factor motifs defining the differentiation of stem cells into hepatocyte-like cells. *J Hepatol.* 2015;63:934-42.
- Grinberg M, Stöber RM, Edlund K, Rempel E, Godoy P, Reif R, et al. Toxicogenomics directory of chemically exposed human hepatocytes. *Arch Toxicol.* 2014;88:2261-87.
- Hoelting L, Scheinhardt B, Bondarenko O, Schildknecht S, Kapitzka M, Tanavde V, et al. A 3-dimensional human embryonic stem cell (hESC)-derived model to detect developmental neurotoxicity of nanoparticles. *Arch Toxicol.* 2013;87:721-33.
- Krause KH, van Thriel C, De Sousa PA, Leist M, Hengstler JG. Monocrotophos in Gandaman village: India school lunch deaths and need for improved toxicity testing. *Arch Toxicol.* 2013;87:1877-81.
- Krug AK, Kolde R, Gaspar JA, Rempel E, Balmer NV, Meganathan K, et al. Human embryonic stem cell-derived test systems for developmental neurotoxicity: a transcriptomics approach. *Arch Toxicol.* 2013;87:123-43.
- Lee SK, Ou YC, Andersen ME, Yang RS. A physiologically based pharmacokinetic model for lactational transfer of PCB 153 with or without PCB 126 in mice. *Arch Toxicol.* 2007;81:101-11.
- Lee YJ, Ahn MY, Kim HS, Kwack SJ, Park KL, Yoon S, et al. Role of phospholipase D in regulation of testicular Leydig cell hyperplasia in Sprague-Dawley rats treated with di(2-ethylhexyl) phthalate. *Arch Toxicol.* 2011;85:975-85.
- Leist M, Ringwald A, Kolde R, Bremer S, van Thriel C, Krause KH, et al. Test systems of developmental toxicity: state-of-the art and future perspectives. *Arch Toxicol.* 2013;87:2037-42.
- Liu X, Liu W, Jin Y, Yu W, Liu L, Yu H. Effects of subchronic perfluorooctane sulfonate exposure of rats on calcium-dependent signaling molecules in the brain tissue. *Arch Toxicol.* 2010;84:471-9.
- Meganathan K, Jagtap S, Srinivasan SP, Wagh V, Hescheler J, Hengstler J, et al. Neuronal developmental gene and miRNA signatures induced by histone deacetylase inhibitors in human embryonic stem cells. *Cell Death Dis.* 2015;6:e1756.
- Moss JI, Pontes E, Hansen PJ. Insulin-like growth factor-1 protects preimplantation embryos from anti-developmental actions of menadione. *Arch Toxicol.* 2009;83:1001-7.
- Oesch F, Dietrich C, Naegeli H, Schwarz M, van der Horst G, Zanger U, et al. New aspects on mechanisms of chemical carcinogenesis: emphasis on species and gender/sex differences and developmental/aging determinants. *Arch Toxicol.* 2008;82:875-80.
- Reif R. The body-on-a-chip concept: possibilities and limitations. *EXCLI J.* 2014;13:1283-5.
- Rempel E, Hoelting L, Waldmann T, Balmer NV, Schildknecht S, Grinberg M, et al. A transcriptome-based classifier to identify developmental toxicants by stem cell testing: design, validation and optimization for histone deacetylase inhibitors. *Arch Toxicol.* 2015;89:1599-618.

Shinde V, Stöber R, Nemade H, Sotiriadou I, Hescheler J, Hengstler J, et al. Transcriptomics of hepatocytes treated with toxicants for investigating molecular mechanisms underlying hepatotoxicity. *Methods Mol Biol.* 2015;1250:225-40.

Stapleton AR, Chan VT. Subtoxic chlorpyrifos treatment resulted in differential expression of genes implicated in neurological functions and development. *Arch Toxicol.* 2009;83:319-33.

Stöber R. Transcriptome based differentiation of harmless, teratogenic and cytotoxic concentration ranges of valproic acid. *EXCLI J.* 2014;13:1281-2.

Waldmann T, Rempel E, Balmer NV, König A, Kolde R, Gaspar JA, et al. Design principles of concentration-dependent transcriptome deviations in drug-exposed differentiating stem cells. *Chem Res Toxicol.* 2014;27:408-20.

Weng Z, Ohtani K, Suda M, Yanagiba Y, Kawamoto T, Nakajima T, et al. Assessment of the reproductive toxicity of inhalation exposure to ethyl tertiary butyl ether in male mice with normal, low active and inactive ALDH2. *Arch Toxicol.* 2014;88:1007-21.

Xi S, Sun W, Wang F, Jin Y, Sun G. Transplacental and early life exposure to inorganic arsenic affected development and behavior in offspring rats. *Arch Toxicol.* 2009;83:549-56.

Zimmer B, Pallocca G, Dreser N, Foerster S, Waldmann T, Westerhout J, et al. Profiling of drugs and environmental chemicals for functional impairment of neural crest migration in a novel stem cell-based test battery. *Arch Toxicol.* 2014;88:1109-26.