Phenotypic Drift as a Cause for Intratumoral Morphological Heterogeneity of Invasive Ductal Breast Carcinoma Not Otherwise Specified

Marina V. Zavyalova,^{1,2} Evgeny V. Denisov,³ Lubov A. Tashireva,¹ Tatiana S. Gerashchenko,^{3,4} Nikolay V. Litviakov,³ Nikolay A. Skryabin,⁵ Sergey V. Vtorushin,^{1,2} Nadezhda S. Telegina,² Elena M. Slonimskaya,^{6,7} Nadezhda V. Cherdyntseva,^{3,7} and Vladimir M. Perelmuter^{1,2}

Abstract

Invasive ductal carcinoma (IDC) not otherwise specified (NOS), the most common type of breast cancer, demonstrates great intratumoral morphological heterogeneity, which encompasses the presence of different types of morphological structures—tubular, trabecular, solid, and alveolar structures and discrete groups of tumor cells, the origins of which remain unclear at present. In this study of 162 IDC NOS patients, we investigated whether the distribution of different types of morphological structures is related to the basic clinicopathological parameters of IDC NOS. Our results showed that in patients with only one type of tumor structure, the presence of any one of the five types was equally probable; however, cases with two types of structures were more likely to contain trabecular structures than the other four types. The development of intratumoral morphological heterogeneity was not associated with menopausal status, tumor size, histological grade, hematogenic metastasis, or recurrence. However, the number of different types of morphological structures was significantly higher in luminal tumors than in triple-negative tumors. An increase in the frequency of lymph node metastasis correlated with the increased number of different types of structures in breast tumors; however, in contrast to premenopausal patients, this association was explained by the presence of alveolar structures in postmenopausal women. In addition, we showed a significant decrease in the numbers of positive lymph nodes in tumors with high numbers of morphological variants. The frequency of lymph node metastases and the number of positive nodes were generally independent features and formed by different mechanisms. Based on the evidence, the term "phenotypic drift" has been designated as the basis for the development of intratumoral morphological heterogeneity of IDC NOS.

Key words: breast cancer; lymph node metastasis; menopause; phenotypic drift; tumor heterogeneity

Introduction

INVASIVE DUCTAL CARCINOMA (IDC) not otherwise specified (NOS) or no special type is the most common histological type of breast cancer (up to 75% of all invasive lesions). It is described as a heterogeneous group of tumors with extremely diverse morphological "portraits"^{1,2} that make it difficult to predict the individual prognoses of IDC NOS patients. IDC NOS tumors often contain minor components with a special type of histology, including architectural features of the invasive process, which may vary widely both within a single tumor and from case to case.³ Previously, we described five different morphological types of invasive components or morphological structures in IDC NOS tumors—tubular, trabecular, solid, and alveolar structures and discrete groups of tumor cells. We also reported an association between the presence of alveolar structures and an increased risk of lymph node metastasis, which was more

¹Department of Pathological Anatomy and Cytology, ³Department of Experimental Oncology, ⁶Department of General Oncology, Cancer Research Institute, Siberian Branch of the Russian Academy of Medical Sciences, Tomsk, Russian Federation.

⁵Laboratory of Cytogenetics, The Russian Institute of Medical Genetics, Siberian Branch of the Russian Academy of Medical Sciences, Tomsk, Russian Federation.

²Department of Pathological Anatomy, ⁴Department of Biochemistry and Molecular Biology, ⁷Department of Oncology, Siberian State Medical University, Tomsk, Russian Federation.

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significant in postmenopausal patients than in premenopausal patients.⁴ However, it is not clear so far whether such intratumoral morphological heterogeneity is an initial characteristic of an IDC NOS tumor or a later phenomenon formed during tumor development. In this context, there is great interest in elucidating a potential link between the clinicopathological parameters, which reflect tumor progression potential, and the development of intratumoral morphological heterogeneity. In a study by Wilson and Evans,⁵ phenotypic drift was described as the phenomenon of dedifferentiation of tubular cancer, a prognostically favorable type of breast cancer, into high-grade tumors of mixed tubular and ductal nonspecial type morphologies with elevated lifethreatening potential. In the present study, the term "phenotypic drift" was defined as the change in the morphological phenotype of an IDC NOS tumor. In other words, any alteration in the distribution of various morphological structures, namely their numbers, during tumor development was attributed to phenotypic drift.

Based on the definition for phenotypic drift, we aimed to investigate the patterns of development of intratumoral morphological heterogeneity in IDC NOS, particularly focusing on whether the distribution of different types of morphological structures is dependent on the basic clinicopathological parameters of breast tumors.

Materials and Methods

In total, 162 patients (mean age of 43.3 ± 6.7 years for premenopausal women and 61.5 ± 8.0 years for postmenopausal women) with IDC NOS diagnosed in the Cancer Research Institute (Tomsk, Russia) between 1999 and 2006 and not treated by neoadjuvant chemotherapies were included (Table 1). The distinction of IDC NOS from other histological types was performed according to the World Health Organization Classification of Tumors.⁶

Tumors were staged according to the tumor-node-metastasis (TNM) classification of malignant tumors (T1–3 N0–3 M0). Fresh tumor tissues were fixed in 10% neutral formalin (Karbolit) for 24 h, rinsed with a mixture of isopropanol (Biovitrum), and embedded in paraffin (Biovitrum). Morphological and immunohistochemical analyses were performed by light microscope (Jenamed, Carl Zeiss). All tumor slides (five from each sample) were reviewed by three experienced pathologists. Intra-observer variability was 5%. In case of discrepancy, results were discussed between the pathologists, and a consensus was reached.

Morphological analysis included determination of histological grade and calculation of different types of morphological structures in breast tumors. Sections of $5-6 \mu m$ thickness were stained by hematoxylin (Dako) and eosin (Dako). Histological grade was scored according to the modified Scarff-Bloom-Richardson grading system,⁷ taking into account the number of tubular and ductal structures, mitoses, and nuclear atypia/pleomorphism. The presence of different types of morphological structures was evaluated in tumor slides stained by hematoxylin and eosin. Tubular structures used in tumor grading were presented by rows of tiny tube-shaped cell aggregations. Trabecular structures were formed by two or more rows of cells located within the stroma. Solid structures represented groups of hundreds of cells with different sizes and shapes. Discrete groups of

TABLE 1. CHARACTERISTICS OF STUDY GROUP OF BREAST CANCER PATIENTS

	n		
Tumor characteristic	Premenopausal patients N=53	Postmenopausal patients N = 109	p-Value
Tumor size < 20 mm 20–50 mm > 50 mm	30 (56.7) 21 (39.6) 2 (3.7)	37 (33.8) 66 (60.5) 6 (5.7)	0.006 0.01 0.72
Grade Low Medium High	4 (7.5) 49 (92.5) 0 (0.0)	7 (6.4) 97 (89.0) 5 (4.6)	1.00 0.58
ER status Positive Negative Unknown	31 (68.9) 14 (31.1) 8	59 (67.0) 29 (33.0) 21	0.82
PR status Positive Negative Unknown	33 (73.3) 12 (26.7) 8	48 (55.2) 39 (44.8) 22	0.04
HER2/neu status Negative Positive Unknown	31 (81.6) 7 (18.4) 15	61 (81.3) 14 (18.7) 34	0.82
Molecular subtypes Luminal HER2-enriched Triple-negative Unknown	30 (81.1) 3 (8.1) 4 (10.8) 16	51 (68.9) 7 (9.5) 16 (21.6) 35	0.26 1.00 0.19
Lymph node metast _ +	ases 36 (67.9) 17 (32.1)	66 (60.6) 43 (39.4)	0.36
Hematogenic metas - +	tases 41 (77.4) 12 (22.6)	96 (88.1) 13 (11.9)	0.08
Recurrence - +	46 (86.8) 7 (13.2)	102 (93.6) 7 (6.4)	0.23

n, number of patients presenting with a specific tumor characteristic; *N*, total number of patients in group of pre- or postmenopausal patients; ER, estrogen receptor; PR, progesterone receptor; –, absent; +, present.

tumor cells were detected as single cells or as groups of up to five cells. Alveolar structures with rounded shapes contained 5–25 cells.⁴ It is important to note that tumors from different patients could have different types of morphological structures.

Immunohistochemical analysis was performed using antibodies directed against estrogen receptors (ER, ID5 clone, ready-to-use, nuclear expression, Dako), progesterone receptors (PR, PgR636 clone, ready-to-use, nuclear expression, Dako), and c-erbB2 (also known as HER2/neu, polyclonal antibody, 1:500 dilution, membrane expression, Dako). Five- to six-micrometer-thick sections were deparaffinized in three changes of 96% ethanol, rinsed with distilled water for 5 min, placed in plastic racks and subjected to high temperature antigen unmasking technique in 0.01 M citrate buffer, pH 6.0 (Dako). Antigen retrieval was carried out in three steps: heating for 7 min at 60 W, cooling-down for 1 min, and heating 7 min at 40 W. After cooling the sections to room temperature and rinsing in two changes of phosphate buffer (Biolot), incubation was performed with peroxidase blocking reagent (Dako) for 10 min. Further, the sections were rinsed with distilled water for 5 min and phosphate buffer for 5 min, and incubated with antibodies for 60 min at 25°C. After washing, the sections were treated with biotinylated antibodies (ready-to-use, Dako) for 10 min. Following another buffer wash, the sections were labeled with streptavidin-biotin complex (Dako) for 10 min. To visualize the immunoreactions, the sections were incubated with diaminobenzidine (Dako) for 10 min. The sections were counterstained with hematoxylin (Dako) and mounted in balsam (Panreac, Spain). Immunohistochemical visualization was performed with LSAB2 system-HRP (Dako).

Molecular classification of breast cancer cases was performed based on the ER, PR, and HER2/neu status. In this study, we focused luminal subtype, including A and B (ER, PR positive, absence or overexpression of HER2/neu), HER2-enriched (ER, PR negative, overexpression of HER2/ neu), and triple-negative (ER, PR negative, absence of HER2/neu).⁸ Long-term results, namely the presence of hematogenic metastases and recurrences, analyzed throughout 5 years by patient records. This study was approved by the institutional review board, and all patients signed an informed consent for voluntary participation.

Statistical processing was performed by Pearson's chisquared test and regression analysis. Yates' corrected chisquare test (when expected frequencies <10 were found)⁹ and Fisher's exact test (when expected frequencies <6 were found)¹⁰ were also utilized. All analyses were performed using STATISTICA 8.0 software (StatSoft) and 2x2 Contingency Table (http://vassarstats.net/odds2x2.html). Results were considered statistically significant at p < 0.05, and p-values were two-sided.

Results

This study was conducted to reveal whether the distribution of different types of morphological structures is related to basic clinicopathological parameters of breast cancer, including menopausal status, tumor size, histological grade, molecular subtypes, lymph node involvement, hematogenic metastasis, and recurrence. We did not find any significant association between the distribution of different structural types and menopausal status, tumor size, histological grade, hematogenic metastasis, or recurrence. However, intratumoral morphological heterogeneity of IDC NOS was strongly associated with molecular subtypes and lymph node involvement, including the number of positive nodes.

To understand which types of morphological structures originate first and which ones occur later, we performed comparisons of the frequencies of their combinations in the study group (Fig. 1). Regardless of menopausal status, there was a nearly equal distribution of each structural variant in tumors with only one type of structure. Nonetheless, in individuals with any two types of structures, trabecular structures were detected in a majority (75%–80%) of cases.

With regard to the dependence on molecular classification, tumors of postmenopausal patients with any single type of morphological structure were more frequently found in the triple-negative subtype than in the luminal breast cancers (18.7% vs. 3.9%; p=0.04; Table 2). In contrast, tumors with all variants of structures more frequently occurred in the luminal cancers than in the triple-negative ones (23.5% vs. 6.3%; p=0.04; Table 2). Similar tendencies were also observed in premenopausal women; however, the study group was too



FIG. 1. Distribution of different types of morphological structures in **(A)** pre- and **(B)** postmenopausal patients. The *x*-axis designates the number of different types of morphological structures. The *y*-axis designates the percentage of cases with a particular variant of structures. Regardless of menopausal status, there was a nearly equal distribution of each structural variant in tumors with only one type of structure. Nonetheless, in individuals with any two types of structures, trabecular structures were detected in a majority (75%–80%) of cases. In patients with two to four types of structural variants, there were insignificant differences between the numbers of tubular, solid, and alveolar structures and of discrete groups of tumor cells. Cases with five types logically contain all variants of morphological structures.

No. of different types of morphological structures	Premenopausal, n (%)		Postmenopausal, n (%)			
	Luminal	HER2-enriched	Triple-negative	Luminal	HER2-enriched	Triple-negative
1	1 (3.3)	1 (33.3)	1 (25.0)	2 (3.9)	0 (0.0)	$3(18.7)^{a}$
2	9 (30.0)	0 (0.0)	1 (25.0)	9 (17.6)	3 (42.9)	3 (18.7)
3	11 (36.7)	0 (0.0)	0 (0.0)	16 (31.5)	3 (42.9)	5 (31.3)
4	7 (23.3)	2 (66.7)	2 (50.0)	12 (23.5)	1 (14.2)	4 (25.0)
5	2 (6.7)	0 (0.0)	0 (0.0)	$12(23.5)^{a}$	0 (0.0)	1 (6.3)
Total	30	3	`4 ´	51	7	16

 TABLE 2. NUMBER OF DIFFERENT TYPES OF MORPHOLOGICAL STRUCTURES

 IN THE DIFFERENT MOLECULAR SUBTYPES OF BREAST TUMORS

^a*p*=0.04 (Fisher's exact test, differences between patients with luminal and triple-negative subtype).

n, number of patients with molecular subtype.

small to reach statistical significance. The disparity between the number of ER, PR positive (Table 1), and luminal (Table 2) patients, as well as between the number of HER2/neu positive and HER2-enriched patients, may be explained by the following reasons. First, in ER and PR expression, some of the HER2/ neu positive tumors were considered as luminal B. Second, the simultaneous assessment of the ER, PR, and HER2/neu status that was not included in this study would be needed to define molecular subtype of breast cancer.

Previously, we reported the association between the presence of alveolar structures in breast tumors and an increased risk of lymph node metastasis, which was more significant in postmenopausal than in premenopausal women.⁴ Based on the present results, it is interesting to note that the percentage of premenopausal lymph node positive subjects, who had alveolar structures in tumors, increased with an increasing number of structural variants (β =0.97; p=0.02; Table 3). In contrast, only the presence of alveolar structures, but not the number of structural variants, was associated with a high likelihood of lymph node involvement in postmenopausal patients (β =0.69; p=0.30; Table 3).

Next, we studied how the number of different types of morphological structures in breast tumors is related to the frequency of lymph node metastasis and the number of involved lymph nodes. In both pre- and postmenopausal patients, the frequency of lymph node involvement was elevated with an increasing number of structural variants; however, the differences reached statistical significance only in postmenopausal patients (0.02 ; Table 4). In addition, in postmenopausal women, the number of affected lymph nodes decreased with an increasing number of structural variants (<math>0.04 ; Table 4).

Discussion

Breast cancer shows significant heterogeneity at both interand intratumoral levels.^{11–13} Intratumoral heterogeneity, which refers to the presence of different clones of tumor cells with different malignancy potential and responsiveness to treatment, is a primary obstacle in oncology, confounding the molecular classification, disease prognostication, and therapeutic intervention of breast cancers.^{14–18} Several hypotheses have been advanced to explain the origin of intratumoral heterogeneity in breast cancer, beginning with the cancer stem cell and clonal evolution models and ending with the mutator phenotype model.^{14,19–21} Mathematical approaches have been applied to study the evolutionary dynamics of intratumoral heterogeneity. Calculations have shown that old but not young tumors, and the accumulation of neutral but not advantageous mutations, are associated with high levels of intratumoral diversity.^{22,23} In other

Table 3. Percentage of Lymph Node Positive Patients with Presence and Absence of Alveolar Structures in Breast Tumor

No. of different types of morphological structures	Lymph node positive patients with alveolar and other morphological structures, n/N (%)			
	Premenopausal		Postmenopausal	
	Alveolar	Other	Alveolar	Other
1	1/1 (100.0)	0/3 (0.0)	0/0 (0.0)	0/0 (0.0)
2	1/7 (14.2)	2/9 (22.2)	$4/8(50.0)^{a}$	1/17(5.9)
3	3/8 (37.5)	1/6 (16.7)	$15/26(58.0)^{a}$	1/10 (10.0)
4	7/16 (43.7)	0/0 (0.0)	13/22 (59.0)	0/2(0.0)
5	2/3 (66.7)	0/0(0.0)	8/14 (57.1)	0/0(0.0)
Regression coefficient (β)	0.97		0.69	
<i>p</i> -Value	0.02		0.30	

Regression coefficient and *p*-value are based on four observations: one, two, three, and four types.

 ${}^{a}p = 0.02$ (Fisher's exact test, differences between postmenopausal patients with alveolar and other structures).

n, number of patients with alveolar/other structures and lymph node metastases; N, number of patients in sample.

No. of different types of morphological structures	Premenope	ausal patients	Postmenopausal patients		
	Frequency of lymph node metastasis, n/N (%)	Number of lymph nodes with metastases, mean±SD (n)	Frequency of lymph node metastasis, n/N (%)	Number of lymph nodes with metastases, mean $\pm SD$ (n)	
1	1/4 (25.0)	5.0 (1)	1/10 (10.0)	3.0 (1)	
2	3/16 (18.7)	4.0 ± 2.8 (3)	5/25 (20.0)	10.5 ± 8.8 (5)	
3	4/14 (28.5)	5.5 ± 8.3 (4)	16/36 (44.4)*	$5.0 \pm 4.4 (16)^{\dagger}$	
4	7/16 (43.7)	4.8 ± 4.8 (7)	13/24 (54.2)**	$4.2 \pm 3.3 (13)^{\dagger\dagger}$	
5	2/3 (66.7)	1.0 ± 0.0 (3)	8/14 (57.1)***	$3.5 \pm 3.1 (8)^{\dagger \dagger \dagger}$	

 TABLE 4. Association Between Parameters of Lymph Node Metastasis and the Number of Different Types
 OF Morphological Structures in Breast Tumors

*p = 0.06 (differences between patients with three types and one or two types of morphological structures). **p = 0.02 (differences between patients with four types and one or two types of morphological structures).

***p = 0.03 (differences between patients with five types and one or two types of morphological structures).

p=0.071 (differences between patients with three types and two types of morphological structures).

 $^{\dagger\dagger}p$ = 0.036 (differences between patients with four types and two types of morphological structures).

 $\dagger\dagger\dagger p = 0.059$ (differences between patients with five types and two types of morphological structures).

n, number of patients with lymph node metastases; *N*, number of patients in sample.

words, tumors take a long time to become phenotypically diverse, while advantageous variations should probably be absent because their presence would eventually result in the formation of dominant clones. Therefore, the development of intratumoral heterogeneity is a time-dependent and mutation effect–controlled process.

In this study, we evaluated whether intratumoral morphological heterogeneity in invasive ductal carcinoma, the most common histological type of breast cancer, is dependent on clinicopathological parameters of the disease. Specifically, five different types of morphological structures were examined: tubular, trabecular, solid, and alveolar structures and discrete groups of tumor cells. In patients with only one type of tumor structure, the presence of any one of the five types was equally probable; however, cases with two types of structures were more likely to contain trabecular structures than the other four types. The mechanisms of origin and evolution of different types of morphological structures in breast tumors remain unclear. However, using laser capture microdissection (PALM, Carl Zeiss) our preliminary data demonstrate that there are differences in genotypes of some cancer genes²⁴ and in expression of multidrug resistance genes²⁵ between different morphological variants.

From the clinicopathological parameters we studied, only molecular subtypes and lymph node metastasis were linked with the development of intratumoral morphological heterogeneity. In particular, tumors with all of the structural variants were often detected in the luminal subtype, whereas cases with any single type of structure were mostly triple negative. There are two possible reasons for this observation: the expression of hormone receptors and the effect of tumor progression time. Regarding the first reason, previous studies have demonstrated an association between estrogen receptor expression and increased tumor differentiation, mediated by the ERlinked up-regulation of adherens junction proteins.^{26,27} Such data suggest that estrogen receptor expression should be linked to low intratumoral heterogeneity, which conflicts with our data showing high intratumoral diversity in estrogen receptor positive (luminal) breast cancer. The second reason suggests that due to a favorable prognosis of luminal breast cancer, there is more time to allow the formation of all morphological variants in this type of tumor than in the triple-negative cases.

Despite comparable distributions of different types of structures in breast tumors of pre- and postmenopausal women, there are observations to suggest that menopausal status influences the contribution of intratumoral morphological heterogeneity to lymph node metastasis.⁴ From this study, the percentage of premenopausal lymph node positive subjects, who had alveolar structures in tumors, increased with an increasing number of different types of morphological structures (Table 3). However, in postmenopausal group (Table 3), the high risk of lymphogenic metastasis was linked to the presence of alveolar structures, as was previously suggested.⁴ In premenopausal patients, the nature of the association between a high frequency of lymph node involvement and a large number of morphological variants could be related to the occurrence of diverse subpopulations of tumor cells with various biological characteristics, including the ability to form lymph node metastases. In contrast, the significance of alveolar structures in lymph node involvement in postmenopausal patients could be linked to menopause-specific factors^{28–30} that provoke tumor cells within these structures to metastasize. However, further studies are needed to understand the mechanisms of these associations.

An important observation in this study was an inversely proportional correlation between the number of lymph nodes with metastases and the number of different types of morphological structures in postmenopausal breast tumors; however, it is interesting that the frequency of lymph node involvement increased with increasing intratumoral diversity (Table 4). At first sight, it would appear that our data are contradictory. However, the frequency of lymph node metastasis and the number of positive nodes are generally independent features formed by different mechanisms. Otherwise, the values of these two prognostic factors would be equal. The number of positive lymph nodes has a greater prognostic significance for assessment of risk of hematogenic metastasis. Meanwhile, metastasis involving four or more lymph nodes denotes a high probability of development of hematogenic metastases. In addition, high variability in the numbers of positive nodes (from 1 to 20 and more) in the origins of lymph node metastases indicates that the two features are independent from each other.³¹

The importance of our data is highlighted by the fact that there are no known factors that can determine the number of involved lymph nodes. Based on the numbers of morphological structure variants in breast tumors, it is possible to estimate the numbers of lymph nodes, which would facilitate the diagnosis and treatment of tumors with high metastatic potential.

To designate changes in the distribution of various types of morphological structures, namely in their numbers, during tumor development, the term "phenotypic drift" was applied. Previously, phenotypic drift³² was proposed to explain rapid shifts in the metastatic behaviors of tumor cells and spontaneous and reproducible "waves" in their sensitivities to chemotherapeutical drugs.^{33,34} Later, Wilson and Evans⁵ used the term to describe dedifferentiation of tubular cancers into high-grade tumors of mixed tubular and ductal nonspecial type morphologies. Recently, phenotypic drift was illustrated as the natural evolution of the more aggressive cells in a heterogeneous tumor to become the dominant histological type.³⁵ In any case, phenotypic drift or "dynamic heterogeneity," as it is called by Hill and colleagues,³⁶ reflects specific fluctuations in the functional status of tumor cells, owing to the time-dependent continuous generation of diverse cell subpopulations and maintained by different genetic alterations and environmental factors.^{32,37,38} In the present study, and in the context of breast cancer development dynamics, phenotypic drift could be considered as a shift of the morphological phenotype of IDC NOS or change of the distribution of different types of morphological structures containing tumor cells with specific biological features. Collectively, data from previous reports and the current study confirm that phenotypic drift is an original source of intratumoral heterogeneity, as has been suggested by Nicolson.³⁴ However, we demonstrated that intratumoral morphological heterogeneity of IDC NOS has functional manifestations in terms of its association with lymph node metastasis and chemotherapy efficiency, as has been found earlier.³⁹ Accordingly, in this study phenotypic drift was observed for not only morphological features, but also for functional features of tumor cells during breast cancer development. Interestingly, the drift of morphological phenotype of IDC NOS appears to be nonrandom, because it is strongly associated with tumor behavior.

It is interesting to note that phenotypic drift was also placed in lymph node involvement of IDC NOS. With an increase in the degree of replacement of lymphoid tissue by cancer cells, the morphological diversity of metastases grew. In up to 10% of replacement of lymphoid tissue, the number of different types of morphological structures was 1.8 ± 0.9 (mean±standard deviation), from 10% to 50% (2.7 ± 0.9), from 50% to 75% (3.2 ± 1.0), and more than 75% (2.9 ± 1.0 ; data not shown).

Therefore, our data provide evidence that phenotypic drift is a cause for the development of intratumoral morphological heterogeneity in invasive ductal carcinoma NOS, represented by different types of morphological structures within tumors. In particular, intratumoral diversity was found to be dependent on the molecular subtype of breast cancer and associated with the frequency of lymph node metastasis and the number of positive nodes. Furthermore, the association between alveolar structures and lymph node metastasis was exclusively found in postmenopausal patients.

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Author Disclosure Statement

All authors declare that no competing financial interests exist.

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Address correspondence to: Evgeny Denisov, PhD Department of Experimental Oncology Cancer Research Institute Siberian Branch of the Russian Academy of Medical Sciences Tomsk 634050 Russian Federation

E-mail: d_evgeniy@oncology.tomsk.ru