

Review

Cocoa Shell: A By-Product with Great Potential for Wide Application

Jelena Panak Balentić ^{1,*} , Đurđica Ačkar ^{1,*} , Stela Jokić ¹ , Antun Jozinović ¹, Jurislav Babić ¹, Borislav Miličević ¹, Drago Šubarić ¹ and Nika Pavlović ²

¹ Faculty of Food Technology Osijek, Josip Juraj Strossmayer University of Osijek, Kuhačeva 20, 31000 Osijek, Croatia; jelena.panak@ptfos.hr (J.P.B.); stela.jokic@ptfos.hr (S.J.); antun.jozinovic@ptfos.hr (A.J.); jbabic@ptfos.hr (J.B.); borislav.milicevic@ptfos.hr (B.M.); dsubaric@ptfos.hr (D.Š.)

² Faculty of Medicine, Josip Juraj Strossmayer University of Osijek, Cara Hadrijana 10E, 31000 Osijek, Croatia; nika.felicita@gmail.com

* Correspondence: dackar@ptfos.hr; Tel.: +385-31-224-391

Received: 28 May 2018; Accepted: 7 June 2018; Published: 9 June 2018



Abstract: Solving the problem of large quantities of organic waste, which represents an enormous ecological and financial burden for all aspects of the process industry, is a necessity. Therefore, there is an emerged need to find specific solutions to utilize raw materials as efficiently as possible in the production process. The cocoa shell is a valuable by-product obtained from the chocolate industry. It is rich in protein, dietary fiber, and ash, as well as in some other valuable bioactive compounds, such as methylxanthines and phenolics. This paper gives an overview of published results related to the cocoa shell, mostly on important bioactive compounds and possible applications of the cocoa shell in different areas. The cocoa shell, due to its nutritional value and high-value bioactive compounds, could become a desirable raw material in a large spectrum of functional, pharmaceutical, or cosmetic products, as well as in the production of energy or biofuels in the near future.

Keywords: cocoa shell; cocoa by-product; bioactive compounds; reuse

1. Introduction

Food industry waste often consists of inedible parts, so-called by-products. Today, there are huge quantities of by-products that are discarded, causing enormous economic problems by polluting the environment. Considering the growing world population and disappearing raw materials, and a real threat of reduced food sources, it is not surprising that awareness about the needs of preservation and re-use of materials that are treated as a waste is rising [1]. Cocoa shells are just one of the examples of by-products with high-value bioactive components and interesting nutritional value that have been discarded, although they could be re-used in many ways.

The main raw material for the production of all kinds of cocoa products is dried and fermented cocoa beans, and cocoa shells are one of the by-products of cocoa beans obtained in the chocolate industry. Approximately twenty types of cocoa (*Theobroma cacao*) are known, and the three most popular types (*Criollo*, *Forastero*, and *Tritinario*) make up 95% of the world's total cocoa production. World production of cocoa beans reached almost 3.7 million tons in 2007/2008 [2]. When cocoa is processed, there are three types of co-products: cocoa pod husk, cocoa bean shells (Figure 1), and cocoa mucilage.

These by-products are usually considered as “waste” and left to rot on the cocoa plantation, which can cause environmental problems, such as producing foul odors or propagate diseases (e.g., pod rot, because they are not composted) [3–5].

Figure 2 shows statistical data for the value of cocoa shells, pod husks, skins, and other cocoa waste exported from the United Kingdom (U.K.) annually from 2001 to 2015. In 2014, exports of cocoa shells, pod husks, skins, and other cocoa waste were valued at approximately 13,000 British Pounds [6]. Countries in Central and West Africa account for 71.4% of total production, and generate an estimated 6.7 million metric tons of cocoa pod husk, as well as cocoa bean shells and cocoa bean cakes [7].



Figure 1. Cocoa shells obtained after roasting beans.

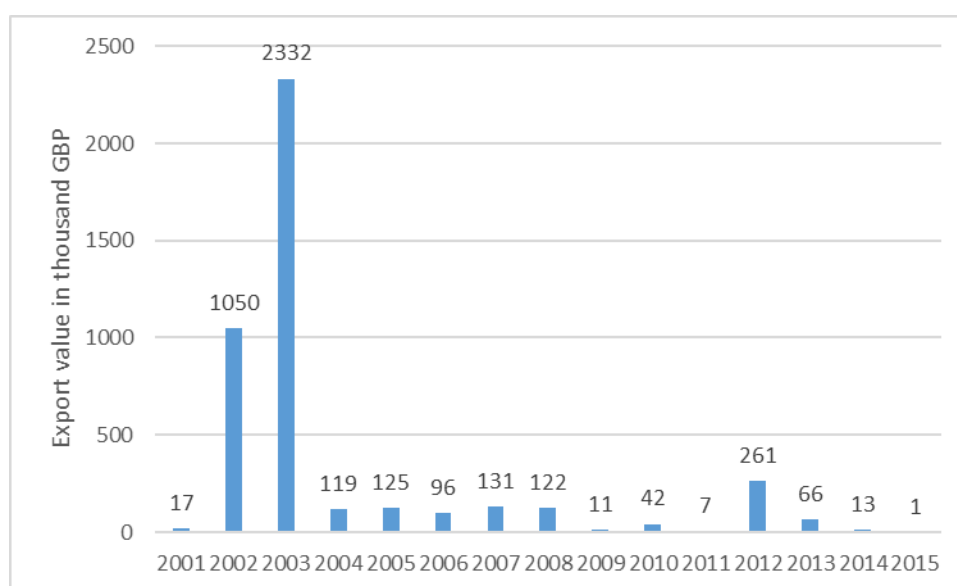


Figure 2. Value of cocoa shells, pod husks, skins, and other cocoa waste exported from the United Kingdom (U.K.) from 2001 to 2015 (in 1000 GBP) [7].

Cocoa shells are the main by-product of cocoa, separated from the cotyledons during the pre-roasting process or after the roasting process [8].

In the Republic of Croatia, there are several chocolate factories and cocoa shells, as a by-product of these factories, are a readily available raw material. Detailed reviews of cocoa shell composition and properties of the most important cocoa shell compounds (polyphenolics, methylxantines, lipids, and fiber) are given by Okiyama et al. [9], thus this paper will focus on further usage of cocoa shells in different areas.

2. Cocoa Shell Application

It is possible to use milled cocoa shells, without any modifications, as well as to alkalize cocoa shells, and then use them as a food additive [10].

Relatively high values of dietary fiber [11], together with phenolic compounds, imply that this by-product is interesting to the food industry (in the manufacturing of confectionery products and bakery products, or in the preparation of low calorie dietetic and fiber-rich products, etc.) [12,13]. However, the most common use is still for feedstuff.

2.1. Use in Feedstuff

A number of studies explored the potential of cocoa shells to replace a part of a usual animal diet and investigated their influence on animals, because it contains theobromine, which may have a negative effect on some species [14–16]. Specifically, cocoa beans contain approximately 2–3% of theobromine, which crosses from seed into shell during fermentation [17].

The toxicity of a cocoa shell meal to broilers was examined by Day & Dilworth [18]. They added cocoa shell in amounts of 1, 2, 4, and 6% to the meal and concluded that 4 and 6% had a significant influence on the decrease of body weight of broilers. In a subsequent experiment, they added exactly the same amount of pure theobromine as there was in cocoa shells that were in the previous meals, but the broilers' weight was drastically decreased. Pure theobromine was more toxic than that furnished by the cocoa shell meal. Emiola, Ojebiyi, & Akande [19] confirmed that increasing the intake of sun-dried cocoa shells from 0 to 30% resulted in decreasing average daily feed intake and egg production, together with decreased weight of spleen, kidney, and ovary in hens fed with a diet containing 25 and 30% cocoa shell, because of increased theobromine intake. Olubamiwa et al. [20], however, claimed that cocoa shells that were boiled for 15 min could be used in laying hen feed up to 20% without an influence on egg production and feed conversion.

Adeloye [21] assessed the ability of the goat to utilize some lignocellulosic materials, composed of *Leucaena* leaf meal, yam peels, and cocoa bean shells, together making up 92% of the total diet. This experimental diet had impressive results on weight gain of the goats; 122 and 139 g/day. The feed conversion was 170 g weight gain per kg of feed intake for the female and intact goats, and 200 g weight gain per kg of feed for the castrates.

Adebowale & Olubamiwa [22] investigated the influence of cocoa shells on growth of *Clarias gariepinus* Burchell, 1822 juvenile. The results indicated that replacement of up to 20% of maize with cocoa shells can favorably support the growth performance of the above-mentioned fish. However, the bitter taste of cocoa shells was one of the major factors that limited its utilization.

There are other published articles about the usage of cocoa shells for fish diets [23–26]. In total, there are few other conclusions for juvenile Nile tilapia (*Oreochromis niloticus*) fed with cocoa shells. A cost analysis of diets showed a considerable reduction in the production cost of one kilogram of fish, and feed and nutrient retention efficiencies show that cocoa husk appears to be a viable partial dietary protein source [27].

The economics of using cocoa shells as a food supplement for rabbits was examined by Ayinde et al. [28]. The authors concluded that untreated cocoa shells can be used at 100 g/kg inclusion in rabbit feed, while cocoa shells treated hot water can be included up to 200 g/kg in rabbit feed, for optimum growth performance and the highest cost–benefit ratio.

Recent studies were oriented on growing pigs. Magistrelli et al. [29] have shown that the use of cocoa shells in pig nutrition may have a positive effect on the balance of intestinal microbial ecosystem. Cocoa shell feeding for three weeks increased microbial populations of the *Bacteroides-Prevotella* group and *Faecalibacterium prausnitzii*, which produce short chain fatty acids, in particular butyrate, which positively influences growth and differentiation of enterocytes, and exerts anti-inflammatory effects, thereby reducing the incidence of a wide range of intestinal inflammatory diseases. Despite a reduction of *Lactobacilli*, cocoa shell feeding improved the proportion between the main phyla of the intestinal ecosystem, which may help to reduce the risk of excessive fattening, which is considered to be detrimental to the quality of the end products [29].

Ogunsipe et al. [30] also examined the addition of cocoa shells into pigs' meals and found that 20% was the optimal biological level of cocoa shells as an energy substitute for maize in a pig diet.

2.2. Use in Agriculture

It is possible to use cocoa husk mulch to suppress weed in perennial fruit crops, gardens, urban landscapes, and occasionally in vegetable crops in organic production systems [31,32].

Arentoft et al. [33] examined the difference between cocoa mulch and bark mulch in suppressing weed growth. Cocoa mulch was more effective, because when compared with bark mulch, a thinner layer of cocoa mulch was needed to reduce the percentage of green pixels by 50% or 90% in relation to control plots.

Agricultural productivity is impaired in many areas of the world as a result of poor natural conditions and loss of added nutrients that are necessary for plant growth. Hale et al. [34] investigated sorption and desorption of phosphate-P, ammonium-N, and nitrate-N in cocoa shell and corn cob biochars. The authors confirmed that biochar can add and slowly release essential nutrients to soil in order to improve agricultural properties, as the real world biochars used here were able to release $\text{PO}_4^{3-}\text{-P}$ and weakly exchange $\text{NH}_4^+\text{-N}$.

2.3. Use in Biofuels

Since fossil fuels present a problem for future generations, there is need for some alternative sources of fuel. Ethanol from lignocellulosic biomass, such as agricultural residue, is one of the important alternatives. In their study, Awolu & Oyeyemi [35] examined ethanol production from cocoa shells using acid hydrolysis and *Saccharomyces cerevisiae*. The result showed that pH had the highest effect on the cocoa shell ethanol yield, followed by fermentation time and yeast concentration; that cocoa shells are an excellent source for such production; and that response surface methodology is a promising tool in the optimization of ethanol production. Additionally, cocoa shells showed good potential for biogas production, with cumulative methane yields [36].

Malatak & Bradna [37] investigated the use of waste material mixtures for energy purposes in small combustion devices. They assessed the energy use of solid biofuels (wheat and rape straw) and their blends with suitable additives (cocoa shells, brown coal, and coal sludge). The results of thermal emission measurements show that all samples meet the requirements of Directive No. 13-2006 [38] for carbon monoxide, but the average nitrogen oxides emission concentrations exceed emission limits. This is because of the high temperature in the combustion chamber and the increasing excess air coefficient.

2.4. Use as an Adsorbent

For the process of adsorption, agricultural waste products are used as natural or modified products through the activation process [39]. Fiorese et al. [40] investigated the grafting of aryl diazonium salt on cocoa shells. The authors concluded that modified cocoa shells can be used as a low-cost adsorbent to entrap pollutants such as heavy metal ions, gases, or industrial dye.

Bernaet & Ruysscher [41] patented the production of cocoa shell powder free from heavy metals. The cocoa shells have good potential for the treatment of agro-industry wastewaters. They are less effective in removing organic pollutants than the polyethylene material used as a bacterial support. The treatment using cocoa shells produced sludge residues (made up of a mixture of cocoa shells and biomass) containing high amounts of nutrients such as nitrogen, and can be potentially re-used in agriculture as a compost [42].

One way to increase the value of cocoa by-products is by converting the waste into activated carbon. It is suggested as an opportunity to replace the commercial coal-based activated carbon, as it is an eco-friendly product [8]. There are a few different studies about this subject published by Ahmad et al. [43–45], Kalaivani et al. [46], Ribas et al. [47], Saucier et al. [48], and Plaza-Recobert et al. [49].

Ahmad et al. [43–45] showed that cocoa shell-based activated carbons have the potential to be used as an adsorbent for 4-nitrophenol and methylene blue (MB) dye in water or wastewater treatments, and

that acid treatment at a higher temperature and higher acid concentration resulted in the development of a new structure of cocoa shell-based activated carbon, which, through the elimination of carbonates and formation of amorphous silica, is highly mesoporous.

Kalaivani et al. [46] reported that activated cocoa shell carbon (TCAC1 and TCAC2) prepared at two different temperatures (30 and 350 °C) show considerable potential for the removal of Ni(II) ion from an aqueous solution. The heat treatment of the adsorbent has resulted in a smaller particle size and larger surface area and, as a result of the increase in adsorption capacity, shows almost 62% higher efficiency in Ni (II) removal, when compared with TCAC1.

The application of microwave-assisted activated carbon from cocoa shells as an adsorbent for removal of sodium diclofenac and nimesulide from aqueous effluents was investigated by Saucier et al. [48]. It effectively removed approximately 95% of a mixture of different organic compounds in a medium with high salinity and sugar contents. Plaza-Recobert et al. [49] examined preparation of binderless activated carbon monoliths from cocoa bean husk. The results prove that an adequate combination of the macromolecular components of the cocoa bean husk (lignocellulosic molecules, pectin, gums, and fats), together with a laminate macromolecular microstructure, made it more suitable for obtaining binderless carbon monoliths than other lignocellulosic precursors. Activation of these carbon monoliths gives activated carbon, with larger micropore volume and good mechanical performance.

2.5. Use as a Dye

In Tran et al. [50], biofilaments based on cocoa shell waste and biodegradable poly (ϵ -caprolactone) (PCL) have been prepared using a single-screw extruder. Using this simple and solvent-free fabrication technique, uniformly structured cocoa shell waste biofilaments can be produced in a very reproducible manner, and used in 3D printing of diverse objects with potential household and biomedical applications.

There is also a study that shows that cocoa shell pigment has potential applications as natural dye for fabric dyeing and in the production of UV protective cotton fabric [51].

2.6. Use in Food Products

Jozinović et al. [52] produced corn snack products enriched with cocoa shells. They added milled shells to corn grits in 5%, 10%, and 15% d.m., and extruded in a laboratory single-screw extruder (Figure 3). The authors concluded that it can be successfully employed as nutritional fortification agent. Sanchez Mundo et al. [53,54] used cocoa shell flour for production of muffins and biscuits.



Figure 3. Corn extrudates enriched with cocoa bean shells.

3. Cocoa Shell Extracts

Cocoa shells can also be used as a raw material for the production of extracts rich in fibers, polyphenols, antioxidants, and so on, which can then be used for further applications [55–57].

The most abundant bioactive compounds in cocoa shell, according to some authors, are shown in Table 1, together with the applied extraction technique.

Table 1. The most abundant bioactive components in cocoa shells. n.d.—not determined in the study.

Extraction method	Hernández-Hernández et al. [58]			Barbosa-Pereira et al. [59]		Arlorio et al. (2001) [60]	
	Methanol-water extraction	Ethanol-acidified water extraction	Water extraction	Methanol-acidified water extraction	Acidified water extraction	Pulsed Electric Field Extraction	Supercritical CO ₂ extraction
Total phenols (mg/g)	14.64	49.46	5.77	20.39	9.40	24.93 - 32.30	18.2
Theobromine (mg/g)	10.20	11.00	8.47	11.62	6.6	4.64 - 10.92	12.9
Caffeine (mg/g)	n.d.	n.d.	n.d.	n.d.	n.d.	1.59 - 4.21	n.d.
Catechin (mg/g)	1.02	1.97	1.65	4.00	6.16	n.d.	n.d.
Epicatechin (mg/g)	15.84	9.00	6.93	17.70	7.04	0.21 - 2.12	n.d.

3.1. Polyphenol-Rich Cocoa Shell Extracts

As cocoa shells contain a certain proportion of phenolic components, which are stored in cocoa seed cotyledons, it is believed that they migrate from cotyledon cocoa beans in different chocolate production processes, such as fermentation, roasting, and alkalizing. This reduces the amount of polyphenols in cocoa beans and gives polyphenol-enriched cocoa shells. The most common compounds are flavanols: epicatechin, catechin, and procyanidins. There are a few methods to obtain polyphenol-enriched extracts from cocoa shells. Generally, higher polyphenolic yields are expected from unfermented shells when compared with fermented ones, as well as from roasted shells when compared with unroasted shells.

The majority of published papers are related to classic extraction of cocoa shells using different organic solvents. For example, Hernández-Hernández et al. [58] compared five different extraction methods of cocoa shells, given in Table 1. In the first method, cocoa shells were extracted with a methanol/water (80:20) solution at 70 °C for 1 h. The second extraction was made with ethanol/acidified water with HCl at pH 3 (30:70) *v/v* by stirring for 2 h at room temperature, followed by filtration and extraction once again, but with a different solution (acetone/water, 70:30, *v/v*). The third extraction was done twice with distilled water, by stirring at 70 °C for 1 h. The fourth extraction was done with methanol/acidified water, similar to second extraction method, and the fifth one was done with acidified water extraction, similar to third extraction method. The methanol/acidified water extraction showed the highest results for flavonoid content.

Usually, traditional extraction techniques are very time consuming and require high amounts of solvents and heating, and as a result, there is increasing demand for novel environmentally friendly extraction techniques that give better extract quality and reduce extraction time and solvent consumption. Barbosa-Pererira et al. [59] used pulsed electric field-assisted extraction to obtain extracts with 20% higher recovery of polyphenols (with epicatechin as the main phenolic compound) and methylxanthines than conventional extraction. Mazzutti et al. [61] used integrated green-based processes, combining supercritical CO₂ and pressurized liquid extraction with ethanol. The extracts with the highest phenolic content were obtained when cocoa shells were previously defatted using supercritical CO₂ at a pressure of 20 MPa and temperature of 40 °C, and then submitted to pressurized liquid extraction at 10 MPa and 70 °C. Supercritical CO₂ as a solvent recovers mainly lipids, similar to soxhlet with hexane, and the phenolic content increased in defatted samples. Therefore, the combination of these two extractions, supercritical CO₂ and pressurized liquid extraction, with ethanol, can be a promising technology to provide two important fractions from cocoa shells, first lipid enriched extracts from SC-CO₂ and phenolic rich extracts using pressurized liquid extraction.

There are a few studies showing that cocoa shell extracts rich in polyphenols can be used in oral care, because these polyphenols exhibit anti-glucosyltransferase activity. Because of this, the influence of cocoa shells on caries-inducing properties of *mutans streptococci* in vitro, and on experimental dental caries in specific pathogen-free rats infected with *mutans streptococci*, were examined. The results indicate that cocoa shell extract possesses powerful anti-cariogenic potential [62]. Osawa et al. [63] isolated cariostatic substances from the cocoa shell; higher-molecular-weight polyphenolic compounds; and unsaturated free fatty acids, such as oleic and linoleic acids. Higher-molecular-weight polyphenolic compounds showed anti-glucosyltransferase (GTF) activities, and unsaturated free fatty acids showed antibacterial activity against *S. mutans*.

Kim et al. [64] have patented manufacturing process of glucosyltransferase inhibitors from cocoa shells. The same group of authors [65] found optimal conditions for the recovery of cocoa shells with a high anti-GTF activity, and a high polyphenol content was determined in extracts obtained with 50% aqueous acetone solution at 60 °C for 4 h, followed by fractionation with 50% aqueous ethanol solution using styrene-based resin.

Matsumoto et al. [66] examined the inhibitory effects of cocoa shell extract on plaque formation in vitro and in vivo. They showed that cocoa shells significantly reduced the adherence of *Streptococcus mutans* to saliva-coated hydroxyapatite, artificial dental plaque formation by *S. mutans*, and the number

of *S. mutans* in plaque in vitro. Furthermore, when used as a mouth rinse, the extract significantly inhibited plaque depositions on the tooth surfaces of human subjects. The authors concluded that cocoa shells may be useful for controlling dental plaque formation and subsequent dental caries development in humans.

The data from Percival et al. [67] show that cocoa polyphenols can inhibit biofilm formation and acid production by *S. mutans*. Venkatesh Babu et al. [68] compared the antimicrobial efficiency of chlorhexidine and cocoa shell extract mouth rinses in children. There was a significant reduction in *Streptococcus mutans* counts in saliva at all follow-up intervals for both mouth rinse groups, but no significant difference in reduction of *Streptococcus mutans* counts in saliva between the chlorhexidine mouth rinse group and cocoa shell extract mouth rinse group. Consequently, it can be concluded that cocoa shell extract mouth rinse can be used in children as an alternative to chlorhexidine mouth rinse, as it has similar antimicrobial properties and evades the side effects of the latter.

Unten et al. [69] investigated whether cocoa shell flavonoids can inhibit cytopathic effects of human immunodeficiency virus (HIV) in cell culture. They have demonstrated that cocoa shell pigment inhibited the replication of HIV in two different assay systems, but the activity of cocoa shell pigment was expressed maximally when cocoa shell pigment was added at the same time as virus adsorption. Cocoa shell pigment is stable against pH and heat, and has no toxicity after oral administration, which are reasons why it has medicinal potential, especially as an antiviral drug [69].

3.2. Methylxanthine-Rich Cocoa Shell Extracts

During the processing of cocoa beans, in the fermentation stage, methylxanthines migrate from the bean into the shell [17]. Theobromine is the most abundant methylxanthine in cocoa shells, followed by caffeine and theophylline. Theobromine is a white powder; a harsh and odorless component that can be stimulant in moderate amounts, while it may be poisonous in larger amounts. It is characterized by important pharmacological functions, such as anticancer, diuretic, smooth-muscle relaxant, and cardiac stimulants [70]. Theobromine gives bitterness to cocoa and chocolate products [71]. Theobromine also has some antioxidant properties [9,72–74]. Arlorio et al. [75] reported 13 g/kg theobromine in dried cocoa shells. According to most studies, theophylline is detected at such low levels that its presence can be ignored.

Using cocoa shells for animal feed has become questionable because of the high theobromine content [9,71]. This component can have harmful effects on animals if consumed in large quantities, as described earlier in the text. Theobromine can be removed from the cocoa shell by extraction techniques, such as supercritical CO₂ extraction. It is possible to completely remove theobromine from shells using supercritical CO₂ and to obtain extracts rich in this component. Hot water treatment has also proved to be capable in reducing the theobromine content [28].

Bradbury & Kopp [57] patented the production of two different extracts from cocoa shells; theobromine fraction and polyphenol enriched fraction. The defatted cocoa shells can be extracted with a solution of acetone/water, after which the acetone needs to be separated, leaving only the aqueous solution. The material is then concentrated, followed by the gel filtration. The theobromine fraction can be rinsed with water, after which a polyphenolic fraction can be rinsed through a column with a low molecular weight solvent.

3.3. Fiber-Rich Cocoa Shell Extracts

The proportion of fiber in cocoa shells depends on whether they are roasted or not. It has been reported that, in roasted seeds and shells, formation of Maillard compounds increases the fiber content [76]. The optimal technique for extraction of pectins was given by Mollea et al. [77]. They recommended hot acid extraction in terms of extraction yield, with pH 2.5 and an extraction time of 1 h. Another study shows that the highest yield of pectin (7.62%) was obtained using citric acid at pH 2.5 [1:25 (w/v)] at 95 °C for 3 h. The highest uronic acid content (65.20%) in the pectin was obtained using water [1:25 (w/v)] at 95 °C for 3 h [78].

Martin-Cabrejas et al. [79] reported a value of 50% for total dietary fiber, while Bonvehi and Beneria [80] determined that total dietary fiber was 57%. Each of these papers reported that the main constituents of insoluble fiber are glucose and uronic acid, with lesser amounts of galactose, arabinose, xylose, and mannose. These sugars indicate that the cell wall polysaccharides in cocoa shell are predominantly cellulose, with lesser amounts of pectin and hemicellulose also present. Redgwell et al. [76] published that total dietary fiber content was approximately 40%, not as high as previous reports. Yeoup Chung et al. [81] found that 100 g d.m. cocoa shell has 26.38 g of lignin, 24.24 g of cellulose, and 8.72 g of hemicellulose.

Although a diet rich in fiber is recommended for preventing and treating constipation, the efficacy of fiber supplements has not been tested sufficiently. A study by Castillejo et al. [82] confirms the beneficial effect of a supplement of cocoa shells that are rich in dietary fiber (39.6 g of total fiber and 13.6 g of β -fructosans per 100 g of product) on chronic idiopathic constipation in children.

Cocoa shells have potential health effects on high cholesterol levels as well. This is confirmed by Ramos et al. [83], who reported that the cocoa product obtained after enzymatic treatment of cocoa shells, rich in soluble dietary fiber and with appreciable amounts of antioxidant polyphenols, brought about remarkable hypocholesterolemic and hypotriglyceridemic responses in rats that were fed an atherogenic diet. It also decreased lipid peroxidation, thus diminishing several risk factors for cardiovascular diseases.

It is also showed that the cocoa shell has nutritional effects, reducing food intake and body weight gain. Another study that shows good potential of soluble dietary fiber is presented by Sanchez et al. [84]. They have been experimenting on rats, where they supplemented their diet with soluble cocoa fiber (SCF) (5%). The results indicate that SCF may modulate parameters that appear altered in the metabolic syndrome, such as body weight, glycemia, insulinemia, lipids, and blood pressure. All of these results show that development of a new source of natural fiber from a waste product from the chocolate industry, such as cocoa shells, could offer a valuable and cheap source of dietary fiber and allow for extensive applications in the food industry.

4. Conclusions

During the processing of raw materials in chocolate industry, a certain amount of by-products is produced, which does not necessarily have to be “waste”, but a by-product or high-value raw material for the development of new products. This review shows that the cocoa shell represents a valuable food industry by-product. Cocoa shells are a rich source of dietary fiber and protein, as well as valuable bioactive compounds (theobromine, caffeine, flavonoids, etc.), and because of their composition, they can be used for further applications as an ingredient in food processing—or in other industries such as pharmaceutical, cosmetic, or agricultural industries—with a constant increase in new applications. In addition, cocoa shell recovery has high economical value, because it is a cheap raw material for the extraction of different components and can be used as biofuel. However, safety of the cocoa shell should be explored more extensively because it is treated with different pesticides, and may contain heavy metals and aflatoxins.

Author Contributions: Conceptualization, Đ.A., S.J.; Investigation, J.P.B., A.J., J.B., B.M., D.Š., N.P.; Resources, J.P.B., N.P.; Writing—original draft preparation, J.P.B., N.P.; Writing—review and editing, J.P.B., Đ.A., S.J., A.J., J.B., B.M., D.Š.; Visualization, J.P.B., Đ.A., S.J.; Funding acquisition, Đ.A., S.J.

Funding: This research received no external funding.

Acknowledgments: This work has been supported in part by the Croatian Science Foundation under the projects “Application of innovative techniques of the extraction of bioactive components from by-products of plant origin” (UIP-2017-05-9909) and “Application of cocoa husk in production of chocolate and chocolate-like products” (UIP-2017-05-8709).

Conflicts of Interest: The authors declare no conflicts of interest

References

1. Ravindran, R.; Jaiswal, A.K. Exploitation of Food Industry Waste for High-Value Products. *Trends Biotechnol.* **2016**, *34*, 58–69. [[CrossRef](#)] [[PubMed](#)]
2. ICCO-International Cocoa Organization Annual Report 2007/2008. Available online: https://www.icco.org/about-us/international-cocoa-agreements/cat_view/1-annual-report/23-icco-annual-report-in-english.html (accessed on 19 September 2017).
3. Martínez, R.; Torres, P.; Meneses, M.A.; Figueroa, J.G.; Pérez-Álvarez, J.A.; Viuda-Martos, M. Chemical, technological and in vitro antioxidant properties of cocoa (*Theobroma cacao* L.) co-products. *Food Res. Int.* **2012**, *49*, 39–45. [[CrossRef](#)]
4. Barazarte, H.; Sangronis, E.; Unai, E. La cascara de cacao (*Theobroma cacao* L.): Una posible fuente comercial de pectinas. *Arch. Latinoam. Nutr.* **2008**, *58*, 64–70. [[PubMed](#)]
5. Sukha, D.A. Potential value added products from Trinidad and Tobago cocoa. In Proceedings of the Revitalisation of the Trinidad and Tobago cocoa industry—Targets, Problems and Options, St. Augustine, FL, USA, 20 September 2003; pp. 69–73.
6. Statista—The Statistics Portal. Value of Cocoa Shells, Husks, Skins and Other Cocoa Waste Exported from the United Kingdom (UK) from 2001 to 2015 (in 1000 GBP). 2017. Available online: <https://www.statista.com/statistics/520014/cocoa-shells-husks-skins-and-other-cocoa-waste-export-value-united-kingdom-uk/> (accessed on 19 September 2017).
7. Adamafio, N.A. Theobromine Toxicity and Remediation of Cocoa By-products: An Overview. *J. Biol. Sci.* **2013**, *37*, 570–576. [[CrossRef](#)]
8. Mylsamy, S. Studies on Synthesis And Characterization of Activated Carbon Prepared from Cocoa (*Theobroma cacao*) Shell And Its Adsorption Modeling Of Dissolved Organic Pollutants. PhD Thesis, Faculty of Science and Humanities, Anna University, Chennai, India, 14 September 2012.
9. Okiyama, D.C.G.; Navarro, S.L.B.; Rodrigues, C.E.C. Cocoa shell and its compounds: Applications in the food industry. *Trends Food Sci. Technol.* **2017**, *63*, 103–112. [[CrossRef](#)]
10. Chronopoulos, D.; Zuurbier, R.; Brandstetter, B.; Jung, C. Food Comprising Alkalized Cocoa Shells and Method Therefor. U.S. Patent 2011/0151098 A1, 23 June 2011.
11. Nsor-Atindana, J.; Zhong, F.; Mothibe, K.J. In vitro hypoglycemic and cholesterol lowering effects of dietary fiber prepared from cocoa (*Theobroma cacao* L.) shells. *Food Funct.* **2012**, *3*, 1044–1050. [[CrossRef](#)] [[PubMed](#)]
12. Nsor-Atindana, J.; Zhong, F.; Mothibe, K.J.; Bangoura, M.L.; Lagnika, C. Quantification of total polyphenolic content and antimicrobial activity of cocoa (*Theobroma cacao* L.) bean shells. *Pak. J. Nutr.* **2012**, *11*, 672–677. [[CrossRef](#)]
13. Vītola, V.; Ciproviča, I. The effect of cocoa beans heavy and trace elements on safety and stability of confectionery products. *Rural Sustain. Res.* **2016**, *35*, 19–23. [[CrossRef](#)]
14. Silva, H.G.; Pires, A.J.V.; Silva, F.F.; Veloso, C.M.; Cravalho, G.G.P.; Cezário, A.S.; Santos, C.C. Farelo de Cacau (*Theobroma cacao* L.) e Torta de Dendê (*Elaeis guineensis*, Jacq) na Alimentação de Cabras em Lactação : Consumo e Produção de Leite. *Rev. Bras. Zootecn.* **2005**, *34*, 1786–1794. [[CrossRef](#)]
15. Andrade, I.V.O.; Pires, A.J.V.; Carvalho, G.G.P.; de Veloso, C.M.; Bonomo, P. Perdas da características fermentativas e valor nutritivo da silagem de capim—elefante contendo subprodutos agrícolas—Losses, fermentation characteristics and nutritional. *Rev. Bras. Zootecn.* **2010**, *39*, 2578–2588. [[CrossRef](#)]
16. Carvalho Junior, J.N.; Pires, A.J.V.; Veloso, C.M.; Silva, F.F.; Reis, R.A.; Carvalho, G.G.P. Digestibilidade aparente da dieta com capim-elefante ensilado com diferentes aditivos. *Arq. Bras. Med. Vet. Zootec.* **2010**, *62*, 889–897. [[CrossRef](#)]
17. Beckett, S.T. *Industrial Chocolate Manufacture and Use*, 4th ed.; Wiley-Blackwell: Chichester, West Sussex, York, UK, 2009; ISBN 978-1-405-13949-6.
18. Day, E.J.; Dilworth, B.C. Toxicity of jimson weed seed and cocoa shell meal to broiler. *Poult. Sci.* **1984**, *63*, 466–468. [[CrossRef](#)] [[PubMed](#)]
19. Emiola, I.A.; Ojebiyi, O.O.; Akande, T.O. Performance and Organ Weights of Laying Hens Fed Diets Containing Graded Levels of Sun-dried Cocoa Bean Shell (CBS). *Int. J. Poult. Sci.* **2011**, *10*, 987–990. [[CrossRef](#)]
20. Olubamiwa, O.; Ikyo, S.M.; Adebowale, B.A.; Omojola, A.B.; Hamzat, R.A. Effect of Boiling Time on the Utilization of Cocoa Bean Shell in Laying Hen Feeds. *Int. J. Poult. Sci.* **2006**, *5*, 1137–1139. [[CrossRef](#)]

21. Adeloye, A. Efficiencies of Conversion of Some Lignocellulosic Waste Materials by Goats. *Bioresour. Technol.* **1992**, *40*, 167–169. [[CrossRef](#)]
22. Adebowale, B.A.; Olubamiwa, O. Growth Response of *Clarias gariepinus* juvenile to Cocoa Husk Endocarp Based Diets. *Agric. J.* **2008**, *3*, 425–428.
23. Falaye, A.E. Utilization of Cocoa Husk in the Nutrition of tilapia (*Oreochromis niloticus*). Ph.D. Thesis, University of Ibadan, Ibadan, Nigeria, 1988.
24. Falay, A.E. Evaluation of the Chemical and Nutrient composition of Cocoa husk (*Theobroma cacao*) and its potential as a fish feed ingredient. *Niger. J. Lasic Appl. Sci.* **1990**, *4*, 157–164.
25. Falaye, A.E. Utilization of agro-industrial waste as fish feedstuffs of Nigeria. In Proceedings of the 10th Annual Conference of the Fisheries Society of Nigeria (FISON), Abeokuta, Nigeria, 16–20 November 1992; pp. 47–57.
26. Falaye, A.E.; Jauncey, K. Acceptability and digestibility by tilapia *Oreochromis niloticus* of feeds containing cocoa husk. *Aquac. Nutr.* **1999**, *5*, 157–161. [[CrossRef](#)]
27. Pouomogne, V.; Gabriel, T.; Pouemegne, J.B. A preliminary evaluation of cacao husks diets for juvenile Nile tilapia (*Oreochromis niloticus*). *Aquaculture* **1997**, *156*, 211–219. [[CrossRef](#)]
28. Ayinde, O.E.; Ojo, V.; Adeyina, A.A.; Adesoye, O. Economics of Using Cocoa Bean Shell as Feed Supplement for Rabbits. *Pak. J. Nutr.* **2010**, *9*, 195–197. [[CrossRef](#)]
29. Magistrelli, D.; Zanchi, R.; Malagutti, L.; Galassi, G.; Canzi, E.; Rosi, F. Effects of Cocoa Husk Feeding on the Composition of Swine Intestinal Microbiota. *J. Agric. Food Chem.* **2016**, *64*, 2046–2052. [[CrossRef](#)] [[PubMed](#)]
30. Ogunsipe, M.H.; Ibadapo, I.; Oloruntola, O.D.; Agbede, J.O. Growth performance of pigs on dietary cocoa bean shell meal. *Livest. Res. Rural Dev.* **2017**, *29*, 1.
31. Billeaud, L.A.; Zajicek, J.M. Influence of mulches on weed control, soil pH, soil nitrogen content, and growth of *Ligustrum japonicum*. *J. Environ. Hortic.* **1989**, *7*, 155–157.
32. Bond, B.; Grundy, A.C. Non-chemical weed management in organic farming systems. *Weed Res.* **2001**, *41*, 383–405. [[CrossRef](#)]
33. Arentoft, B.W.; Ali, A.; Streibig, J.C.; Andreassen, C. A new method to evaluate the weed-suppressing effect of mulches: A comparison between spruce bark and cocoa husk mulches. *Weed Res.* **2013**, *53*, 169–175. [[CrossRef](#)]
34. Hale, S.; Alling, V.; Martinsen, V.; Mulder, J.; Breedveld, G.; Cornelissen, G. The sorption and desorption of phosphate-P, ammonium-N and nitrate-N in cacao shell and corn cob biochars. *Chemosphere* **2013**, *91*, 1612–1619. [[CrossRef](#)] [[PubMed](#)]
35. Awolu, O.O.; Oyeyemi, S.O. Optimization of bioethanol production from cocoa (*Theobroma cacao*) bean shell. *Int. J. Curr. Microbiol. App. Sci.* **2015**, *4*, 506–514.
36. Mancini, G.; Papirio, S.; Lens Piet, N.L.; Esposito, G. Solvent Pretreatments of Lignocellulosic Materials to Enhance Biogas Production: A Review. *Environ. Eng. Sci.* **2016**, *33*, 843–850. [[CrossRef](#)]
37. Malaták, J.; Bradna, J. Use of waste material mixtures for energy purposes in small combustion devices. *Res. Agric. Eng.* **2014**, *60*, 50–59. [[CrossRef](#)]
38. Directive of Ministry of the Environment of the Czech Republic to the Requirements for Awarding Marks—Hot Water Boilers for Central Heating Systems for Biomass; Directive No. 13; Ministry of the Environment of the Czech Republic: Prague, Czech Republic, 2006.
39. Rangabhashiyam, S.; Anu, N.; Selvaraju, N. Sequestration of dye from textile industry wastewater using agricultural waste products as adsorbents. *J. Environ. Chem. Eng.* **2013**, *1*, 629–641. [[CrossRef](#)]
40. Fiorese, F.; Vieillard, J.; Bargougui, R.; Bouazizi, N.; Nkuigie Fotsing, P.; Djoufac Woumfo, E.; Brun, N.; Mofaddel, N.; Le Derf, F. Chemical modification of the cocoa shell surface using diazonium salts. *J. Colloid Interface Sci.* **2017**, *494*, 92–97. [[CrossRef](#)] [[PubMed](#)]
41. Bernaet, H.; Ruyscher, I. Methods for Extraction Cocoa. U.S. Patent 2013/0302473 A1, 14 November 2013.
42. Turcotte, V.; Blais, J.-F.; Mercier, G.; Drogui, P. Use of cocoa shells as biofiltration support for the treatment of effluents from the food industry. *Can. J. Civ. Eng.* **2009**, *36*, 1059–1070. [[CrossRef](#)]
43. Ahmad, F.; Daud, W.M.A.W.; Ahmad, M.A.; Radzi, R. Using cocoa (*Theobroma cacao*) shell-based activated carbon to remove 4-nitrophenol from aqueous solution: Kinetics and equilibrium studies. *Chem. Eng. J.* **2011**, *178*, 461–467. [[CrossRef](#)]

44. Ahmad, F.; Daud, W.M.A.W.; Ahmad, M.A.; Radzi, R. Cocoa (*Theobroma cacao*) shell-based activated carbon by CO₂ activation in removing of Cationic dye from aqueous solution: Kinetics and equilibrium studies. *Chem. Eng. Res. Des.* **2012**, *90*, 1480–1490. [[CrossRef](#)]
45. Ahmad, F.; Daud, W.M.A.W.; Ahmad, M.A.; Radzi, R. The effects of acid leaching on porosity and surface functional groups of cocoa (*Theobroma cacao*)-shell based activated carbon. *Chem. Eng. Res. Des.* **2013**, *91*, 1028–1038. [[CrossRef](#)]
46. Kalaivani, S.S.; Vidhyadevi, T.; Murugesan, A.; Baskaralingam, P.; Anuradha, C.D.; Ravikumar, L.; Sivanesan, S. Equilibrium and kinetic studies on the adsorption of Ni(II) ion from an aqueous solution using activated carbon prepared from *Theobroma cacao* (cocoa) shell. *Desalin. Water Treat.* **2014**, *54*, 1629–1641. [[CrossRef](#)]
47. Ribas, M.C.; Adebayo, M.A.; Prola, L.D.T.; Lima, E.C.; Cataluña, R.; Feris, L.A.; Calvete, T. Comparison of a homemade cocoa shell activated carbon with commercial activated carbon for the removal of reactive violet 5 dye from aqueous solutions. *Chem. Eng. J.* **2014**, *248*, 315–326. [[CrossRef](#)]
48. Saucier, C.; Adebayo, M.A.; Lima, E.C.; Cataluña, R.; Thue, P.S.; Prola, L.D.; Puchana-Rosero, M.J.; Machado, F.M.; Pavan, F.A.; Dotto, G.L. Microwave-assisted activated carbon from cocoa shell as adsorbent for removal of sodium diclofenac and nimesulide from aqueous effluents. *J. Hazard. Mater.* **2015**, *289*, 18–27. [[CrossRef](#)] [[PubMed](#)]
49. Plaza-Recobert, M.; Trautwein, G.; Perez-Cadenas, M.; Alca-niz-Monge, J. Preparation of binderless activated carbon monoliths from cocoa bean husk. *Microporous Mesoporous Mater.* **2017**, *243*, 28–38. [[CrossRef](#)]
50. Tran, T.N.; Bayer, I.S.; Heredia-Guerrero, J.A.; Frugone, M.; Lagomarsino, M.; Maggio, F.; Athanassiou, A. Cocoa Shell Waste Biofilaments for 3D Printing Applications. *Macromol. Mater. Eng.* **2017**, *302*, 1700219. [[CrossRef](#)]
51. Tu, C. Study about Stability of Cacao Husk Pigment and Its Dyeing Properties on Cotton. *Key Eng. Mater.* **2016**, *671*, 133–138. [[CrossRef](#)]
52. Jozinović, A.; Panak Balentić, J.; Ačkar, Đ.; Babić, J.; Pajin, B.; Miličević, B.; Guberac, S.; Šubarić, D. Cocoa husk application in enrichment of extruded snack products. In Proceedings of the Fourth International Congress on Cocoa Coffee and Tea, Torino, Italy, 25–28 June 2017; p. 68.
53. Sanchez, D.; Moulay, L.; Muguerza, B.; Quiñones, M.; Miguel, M.; Aleixandre, A. Effect of a Soluble Cocoa Fiber-Enriched Diet in Zucker Fatty Rats. *J. Med. Food* **2010**, *13*, 621–628. [[CrossRef](#)] [[PubMed](#)]
54. Sanchez Mundo, M.L.; Jaramillo Flores, M.E.; Espinosa Solis, V.; Chávez-Reyes, Y.; Díaz Ramírez, M.; Salgado Cruz, M.P.; Calderón Domínguez, G. Muffins enriched with cocoa shell fiber. In Proceedings of the Fourth International Congress on Cocoa Coffee and Tea, Torino, Italy, 25–28 June 2017; p. 124.
55. Martínez-Cervera, S.; Salvador, A.; Muguerza, B.; Moulay, L.; Fiszman, S.M. Cocoa fibre and its application as a fat replacer in chocolate muffins. *LWT Food Sci. Technol.* **2011**, *44*, 729–736. [[CrossRef](#)]
56. Collar, C.; Rosell, C.M.; Muguerza, B.; Moulay, L. Breadmaking performance and keeping behavior of cocoa-soluble fiber-enriched wheat breads. *Food Sci. Technol. Int.* **2009**, *15*, 79–87. [[CrossRef](#)]
57. Bradbury, A.; Kopp, G. Polyphenol-Enriched Composition from Cocoa Shell Extraction. U.S. Patent 0879946A2, 30 November 2006.
58. Hernández-Hernández, C.; Viera-Alcaide, I.; Sillero, A.M.M.; Fernández-Bolaños, J.; Rodríguez-Gutiérrez, G. Bioactive compounds in Mexican genotypes of cocoa cotyledon and husk. *Food Chem.* **2018**, *240*, 831–839. [[CrossRef](#)] [[PubMed](#)]
59. Barbosa-Pereira, L.; Guglielmetti, A.; Zeppa, G. Pulsed Electric Field Assisted Extraction of Bioactive Compounds from Cocoa Bean Shell and Coffee Silverskin. *Food Bioproc. Technol.* **2018**, *11*, 818–835. [[CrossRef](#)]
60. Arlorio, M.; Coisson, J.D.; Restani, P.; Martelli, A. Characterization of Pectins and Some Secondary Compounds from *Theobroma cacao* Hulls. *J. Food Sci.* **2001**, *66*, 653–656. [[CrossRef](#)]
61. Mazzutti, S.; Goncalves Rodrigues, L.G.; Mezzomo, N.; Venturi, V.; Ferreira, S.R.S. Integrated green-based processes using supercritical CO₂ and pressurized ethanol applied to recover antioxidant compounds from cocoa (*Theobroma cacao*) bean hulls. *J. Supercrit. Fluids* **2018**, *135*, 52–59. [[CrossRef](#)]
62. Ooshima, T.; Osaka, Y.; Sasaki, H.; Osawa, K.; Yasuda, H.; Matsumura, M.; Sobue, S.; Matsumoto, M. Caries inhibitory activity of cacao bean husk extract in in-vitro and animal experiments. *Arch. Oral Biol.* **2000**, *45*, 639–645. [[CrossRef](#)]

63. Osawa, K.; Miyazaki, K.; Shimura, S.; Okuda, J.; Matsumoto, M.; Ooshima, T. Identification of Cariostatic Substances in the Cacao Bean Husk: Their Anti-glycosyltransferase and Antibacterial Activities. *J. Dent. Res.* **2001**, *80*, 2000–2004. [[CrossRef](#)] [[PubMed](#)]
64. Kim, D.Y.; Park, H.J.; Park, H.H.; Kim, H.S.; Kwon, I.B. Manufacturing process of glycosyltransferase inhibitors from cacao bean husk. U.S. Patent US006159451A, 12 December 2000.
65. Kim, K.H.; Lee, K.W.; Kim, D.Y.; Park, H.H.; Kwon, I.B.; Lee, H.J. Extraction and fractionation of glycosyltransferase inhibitors from cacao bean husk. *Process Biochem.* **2004**, *39*, 2043–2046. [[CrossRef](#)]
66. Matsumoto, M.; Tsuji, M.; Okuda, J.; Sasaki, H.; Nakano, K.; Osawa, K.; Shimura, S.; Ooshima, T. Inhibitory effects of cacao bean husk extract on plaque formation in vitro and in vivo. *Eur. J. Oral Sci.* **2004**, *112*, 249–252. [[CrossRef](#)] [[PubMed](#)]
67. Percival, R.S.; Devine, D.A.; Duggal, M.S.; Chartron, S.; Marsh, P.D. The effect of cocoa polyphenols on the growth, metabolism and biofilm formation by *Streptococcus mutans* and *Streptococcus sanguinis*. *Eur. J. Oral Sci.* **2006**, *114*, 343–348. [[CrossRef](#)] [[PubMed](#)]
68. Venkatesh Babu, N.S.; Vivek, D.K.; Ambika, G. Comparative evaluation of chlorhexidine mouthrinse versus cacao bean husk extract mouthrinse as antimicrobial agents in children. *Eur. Arch. Paediatr. Dent.* **2011**, *12*, 245–249. [[CrossRef](#)] [[PubMed](#)]
69. Unten, S.; Ushijima, H.; Shimizu, H.; Tsuchie, H.; Kitamura, T.; Moritome, N.; Sakacamb, I. Effect of cacao husk extract on human immunodeficiency virus infection. *Let. Appl. Microbiol.* **1991**, *14*, 251–254. [[CrossRef](#)]
70. Hartati, I. Hydrotropic Extraction of Theobromine from Cocoa Bean Shell. *Momentum* **2010**, *6*, 17–20.
71. Bentil, J.A.; Dzugbefia, V.B.; Alemawor, F. Enhancement of the nutritive value of cocoa (*Theobroma cacao*) bean shells for use as feed for animals through a two-stage solid state fermentation with *Pleurotus ostreatus* and *Aspergillus niger*. *Int. J. Appl. Microbiol. Biotechnol. Res.* **2015**, *3*, 20–30.
72. Timbie, D.J.; Sechrist, L.; Keeney, P.G. Application of high-pressure liquid chromatography to the study of variables affecting theobromine and caffeine concentrations in cocoa beans. *J. Food Sci.* **1978**, *43*, 560–565. [[CrossRef](#)]
73. Azam, S.; Hadi, N.; Khan, N.U.; Hadi, S.M. Antioxidant and prooxidant properties of caffeine, theobromine and xanthine. *Med. Sci. Monit.* **2003**, *9*, 325–330.
74. Fredholm, B.B. Methylxanthines. In *Handbook of Experimental Pharmacology*; Fredholm, B.B., Ed.; Springer: Berlin/Heidelberg, Germany, 2011; Volume 200, ISBN 978 3 642 13442 5.
75. Arlorio, M.; Coisson, J.D.; Travaglia, F.; Varsaldi, F.; Miglio, G.; Lombardi, G.; Martelli, A. Antioxidant and biological activity of phenolic pigments from *Theobroma cacao* hulls extracted with supercritical CO₂. *Food Res. Int.* **2005**, *38*, 1009–1014. [[CrossRef](#)]
76. Redgwell, R.; Trovato, V.; Merinat, S.; Curti, D.; Hediger, S.; Manez, A. Dietary fibre in cocoa shell: Characterisation of component polysaccharides. *Food Chem.* **2003**, *81*, 103–112. [[CrossRef](#)]
77. Mollea, C.; Chiampo, F.; Conti, R. Extraction and characterization of pectins from cocoa husks: A preliminary study. *Food Chem.* **2008**, *107*, 1353–1356. [[CrossRef](#)]
78. Chan, S.-Y.; Choo, W.-S. Effect of extraction conditions on the yield and chemical properties of pectin from cocoa husks. *Food Chem.* **2013**, *141*, 3752–3758. [[CrossRef](#)] [[PubMed](#)]
79. Martin-Cabrejas, M.A.; Valiente, C.; Esteban, R.M.; Molla, E.; Waldron, K. Cocoa hull: A potential source of dietary fibre. *J. Sci. Food Agric.* **1994**, *66*, 307–311. [[CrossRef](#)]
80. Bonvehi, J.S.; Beneria, M.A. Composition of dietary fibre in cocoa husk. *Zeitschrift für Lebensmitteluntersuchung und -Forschung A* **1998**, *207*, 105–109.
81. Chung, B.Y.; Iiyama, K.; Han, K.-W. Compositional characterization of cacao (*Theobroma cacao* L.) hull. *Agric. Chem. Biotechnol.* **2003**, *46*, 12–16.
82. Castillejo, G.; Bulló, M.; Anguera, A.; Escribano, J.; Salas-Salvadó, J. A controlled, randomized, double-blind trial to evaluate the effect of a supplement of cocoa husk that is rich in dietary fiber on colonic transit in constipated pediatric patients. *Pediatrics* **2006**, *118*, 641–648. [[CrossRef](#)] [[PubMed](#)]

83. Ramos, S.; Moulay, L.; Granado-Serrano, A.B.; Vilanova, O.; Muguerza, B.; Goya, L.; Bravo, L. Hypolipidemic Effect in Cholesterol-Fed Rats of a Soluble Fiber-Rich Product Obtained from Cocoa Husks. *J. Agric. Food Chem.* **2008**, *56*, 6985–6993. [[CrossRef](#)] [[PubMed](#)]
84. Sanchez Mundo, M.L.; Martínez Mendez, D.; Chávez-Reyes, Y.; Espinosa-Solis, V.; Jaramillo-Flores, M.-E.; Torruco-Uco, J.G. Chemical and nutritional characteristics of biscuits added with cocoa shell powder. In Proceedings of the Fourth International Congress on Cocoa Coffee and Tea, Torino, Italy, 25–28 June 2017; p. 147.



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).