

Taxonomy of the *Cryptopygus* complex. II. Affinity of austral *Cryptopygus* s.s. and *Folsomia*, with the description of two new *Folsomia* species (Collembola, Isotomidae)

Mikhail Potapov¹, Charlene Janion-Scheepers², Louis Deharveng³

1 Moscow State Pedagogical University, Kibalchicha str., 6, korp.3, Moscow, 129278, Russia **2** School of Biological Sciences, Monash University, Victoria 3800, Australia **3** Institut de Systématique, Evolution, Biodiversité, ISYEB – UMR 7205 – CNRS, MNHN, UPMC, EPHE, Museum national d'Histoire naturelle, Sorbonne Universités, 45 rue Buffon, CP50, F-75005 Paris, France

Corresponding author: Charlene Janion-Scheepers (cjanion@gmail.com)

Academic editor: W. M. Weiner | Received 24 November 2016 | Accepted 31 January 2017 | Published 28 February 2017

<http://zoobank.org/233A4BBC-3BC6-4094-BA60-9DC210E4640C>

Citation: Potapov M, Janion-Scheepers C, Deharveng L (2017) Taxonomy of the *Cryptopygus* complex. II. Affinity of austral *Cryptopygus* s.s. and *Folsomia*, with the description of two new *Folsomia* species (Collembola, Isotomidae). ZooKeys 658: 131–146. <https://doi.org/10.3897/zookeys.658.11227>

Abstract

Folsomia minorae sp. n. and *F. australica* sp. n. are described from New Zealand and Australia, respectively. Their possible affinity to two different groups of *Cryptopygus sensu stricto* is discussed. Attention is paid to the variability of sensillary patterns of the genital segment in *Cryptopygus*: mainly, all s-chaetae are subequal, but in more advanced forms the dorsal triplet, lateral duplet or either of them become macrochaeta-like in length. *Cryptopygus ulrikeae* (= *Folsomia ulrikeae* Najt & Thibaud, 1987), **comb. n.** is given a new generic position.

Keywords

Australia, New Zealand, taxonomy

Introduction

The genus *Cryptopygus* Willem, 1902 *sensu stricto* has not received its modern generic diagnosis and most of its “austral” species need to be revised (Rusek 2002, Deharveng et al. 2005, Jordana et al. 2009, Potapov et al. 2013, Greenslade 2015). Several species related to *C. antarcticus* Willem, 1902 (the type species of *Cryptopygus*) are common in the high latitudes of the Southern Hemisphere and form a clear taxonomic group. They clearly differ from other genera of the *Cryptopygus* complex that are found in more northern areas (tropical and further into the Northern Hemisphere). In this paper we temporarily define *Cryptopygus s.s.* as having 3 and 5 s-chaetae on Abd IV and V respectively, s-chaetae in mid-tergal position on body tergites, and having no differentiated foil-chaetae at the end of the abdomen. Whilst hemispheric distribution patterns of certain Collembola genera have been well documented (e.g. Holarctic for *Tetracanthella* Schött, 1891, see Deharveng 1987), this group of “austral” *Cryptopygus* (*Cryptopygus s.s.* below) can be considered a geographical equivalent of the mostly Holarctic genus *Folsomia* Willem, 1902. Although occurring in different hemispheres, both genera show an increasing diversity towards the poles, a similar set of life forms and play similar ecological roles in collembolan communities, which are more evident in polar zones. This geographical segregation is even further complicated by the movement of a few *Cryptopygus s.s.* and *Folsomia* to the opposite hemisphere. Until now, at least two “apparent” members of *Cryptopygus s.s.*, *C. clavatus* (Schött, 1893) and *C. roberti* (Fjellberg, 1991) are known from areas of the North Atlantic. Despite the strong similarity between these two forms and the main group of “austral” *Cryptopygus* the final generic position of *clavatus* and *roberti* remains unresolved. The state of a few native “austral” members of *Folsomia* is more obscure as important characters are lacking. Here we present two apparently native and new species from the Australian region. They are described and compared with “austral” *Cryptopygus s.s.*, the potential ancestors of “austral” *Folsomia*. These two species indicate that the latter genus possibly resulted from the convergent evolution of several species of the *Cryptopygus s.s.*

Materials and methods

The notation system accp-as-al (Szeptycki 1972) was used to describe the set of s-chaetae. To establish the links between s-chaetae the approach by Potapov and Greenslade (2010) was applied to the genus *Folsomia*

Abbreviations

Abd. I–VI	abdominal segments I–VI
accp	accessory p-row s-chaeta
Ant. I–IV	antennal segments I–IV
as	anterosubmedial s-chaeta

a.s.l.	above sea level
bms	basal micro s-chaeta on antennal segments
e7	'guard' of labial papilla E
Leg I, II, III	first, second and third pairs of legs
M	macrochaeta
ms	micro s-chaeta(e) (=microsensillum(a) auct.)
s	macro s-chaeta or s-chaetae (=macrosensillum(a) or sensillum(a) auct.)
s.s.	<i>sensu stricto</i>
PAO	postantennal organ
Th.II–III	thoracic segments II and III
U3	inner edge of unguis

Institutional acronyms

MNZTPT	Museum of New Zealand Te Papa Tongarewa, Wellington
SAMA	South Australian Museum, Adelaide
MSPU	Moscow State Pedagogical University, Russia.

Results

Descriptions of new species

Folsomia minorae sp. n.

<http://zoobank.org/08169B5D-CB65-4F2C-9C35-93A8EE6F0C63>

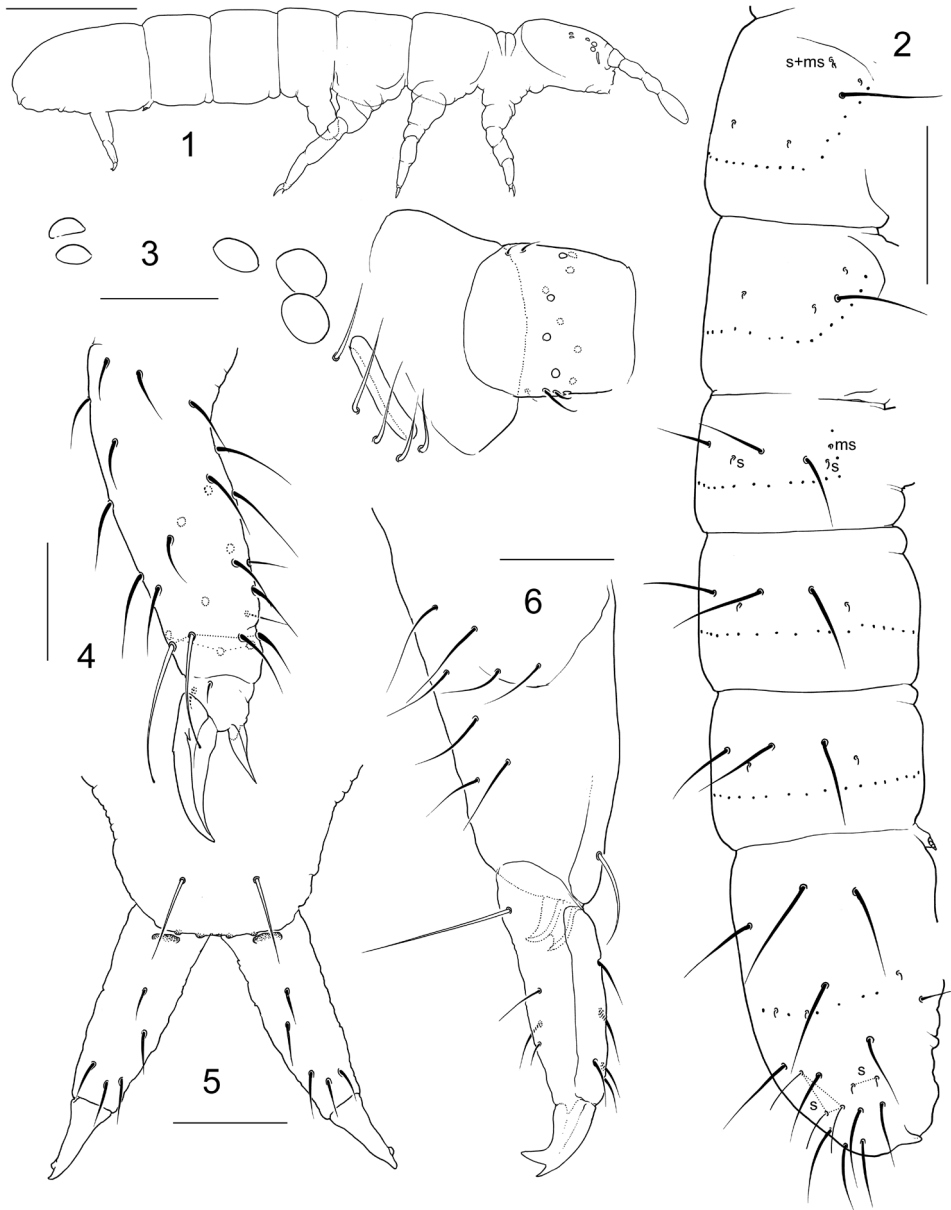
Figs 1–13

Type material. Holotype: adult ♀. New Zealand, southern South Island, Central Otago, Pisa Range, 44°52'03"S, 169°9'33"E, 1700 m a.s.l., in soil and debris under *Dracophyllum muscoides* cushion, 18.ii.2014, coll. M. Minor (on slide). Paratypes. 10 paratypes, subadult ♀♀ and ♂♂ of similar size with holotype, 7 of which from the same locality (and in close proximity), and 3 from Central Otago, The Remarkables Mts, 45°3'42"S, 168°48'40"E, 1829 m a.s.l., herbaceous snowbank, in soil, 19.ii.2014, coll. M. Minor (all on slides). Holotype and 4 paratypes kept in MNZTPT; 5 paratypes in MSPU.

Other material. One ♀ identified in all details as *F. minorae* sp. n. by A. Fjellberg (not seen by us): New Zealand, South Island (northern part), Avalanche Peak trail above Arthur Pass, 42°56'26"S, 171°33'29"E, forest litter, 23.i.2004, coll. A. Fjellberg.

Diagnosis. *Folsomia* species with 5+5 ocelli; slender subapical organite of Ant.IV; clavate tibiotarsal hairs; outer teeth on claws; stout dens with few chaetae and a large mucro; and characteristic '3+2' sensillary pattern of s-chaetae on Abd.V.

Description. Body size of the only adult female 1.75 mm. Dark blue, appendages paler. Body cylindrical (Fig. 1). Abd.IV, V and VI clearly fused dorsally, Abd.IV and



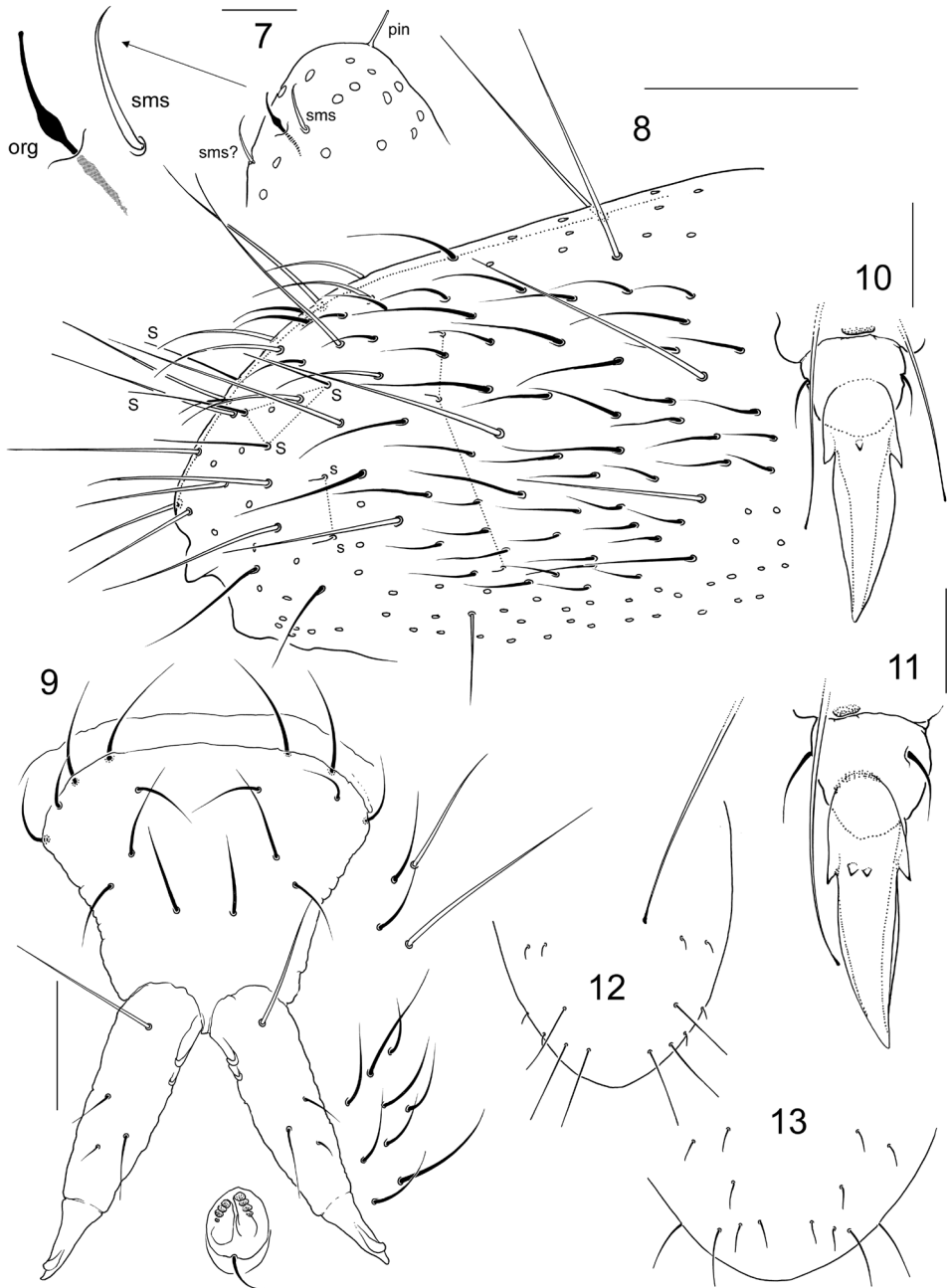
Figures 1–6. *Folsomia minorae* sp. n. **1** habitus **2** macrochaetae and s and ms-chaetae on body **3** anterior part of head **4** distal part of Leg III **5–6** furca, anterior (**5**) and lateral (**6**) view. Scale bar 0.3 mm in **1**; 0.15 mm in **2**, 0.03 mm in others.

III well separated. Cuticle reticulated, with roundish polygons, the largest of which almost as large as chaetae bases. Ocelli 5+5, arranged in two groups: three anterior and two posterior. PAO slender, not constricted, almost as long as width of Ant I (0.8–1.0) and 1.1–1.4 as long as inner unguis length (Fig. 3). Maxillary outer lobe with four

sublobal hairs, maxillary palp bifurcate. Labral formula as 4/5,5,4. Labium with five papillae (A–E) and full set of ‘guards’, ‘guard’ e7 present, with three proximal, four basomedian, and five basolateral chaetae. Ventral side of head with 4+4 postlabial chaetae. Ant.I with two ventral s-chaetae (s) and three small basal ms-chaetae (bms), two dorsal and one ventral (Fig. 3), Ant.II with three bms and a latero-distal s, one of bms enlarged, Ant.III with one bms and with six distal s (including two lateral), without additional s-chaetae. S-chaetae on Ant.IV weakly differentiated. Apex of Ant. IV with two subapical ms (sms) both set at a distance from very long organite (org). Both sms of normal shape, organite with swelling in proximal part chili-shaped. The second subapical ms subequal to the first one, located more dorsally (Fig. 7). S-chaetae formula as common for the genus, 4,3/2,2,2,3,5 (s) and 1,0/1,0,0 (ms) (Fig. 2). Tergal s-chaetae much shorter than common chaetae and distinct. Medial s-chaetae on Th.II–Abd.III in mid-tergal position, on Abd.I–III between Mac1 and Mac2 (Fig. 2). Abd.V with five s-chaetae: three dorsal ones (al, accp1, accp2) long and slender, and two lateral short (‘3+2’ pattern) (Figs 2, 8, 12). Macrochaetae very long, stout and smooth, 1,1/3,3,3 in number, medial ones on Abd.V more than twice as long as dens (2.0–2.4) and 4.7–5.5 times longer than mucro. Foil chaetae at the tip of abdomen absent. Axial chaetotaxy of Th.II–Abd.III as 6–8,6–7/4,4,4. Thorax without ventral chaetae. Unguis stout, without inner teeth, with one (two on Leg III) outer and two large lateral teeth (Figs 4, 10, 11). The doubling of outer tooth on Unguis 3 well visible only in anterior position (Fig. 11). Empodial appendage about half as long as unguis (empodial appendage length: $U3 = 0.46–0.53$). Upper and lower subcoxae of Leg I, II, III with 1,1; 2–3,6; 4–5,6–9 chaetae. Tibiotarsi without additional chaetae on Leg I and II (21 chaetae), and with several additional chaetae on Leg III (more than 26 at whole). Tibiotarsal tenent chaetae clavate, long (1.3–1.6 longer than inner edge of U3), in number 1, 2, 2 on Leg I, II, III. VT with 4+4 laterodistal and 6 posterior chaetae, anteriorly without chaetae. Laterodistal chaetae arranged almost in a line, posterior chaetae in two rows, proximal (2) and distal (4). Tenaculum with 4+4 teeth and one chaeta. Basal tooth smaller than others (Fig. 9). Anterior furcal subcoxae with 8–9 chaetae, posterior one with four chaetae. Anterior side of manubrium with a pair of chaetae (Fig. 5). Posterior side of manubrium with 4+4 laterobasal and 4+4 on main part, without apical and lateral chaetae. To describe chaetae on main part the notation system of Fjellberg (2007) can be somewhat applied: chaetae M1, M2, pr and ml1 present (Fig. 9). Dens stout, with five anterior chaetae arranged as 1,1,3, the second single chaeta positioned more medially than the first (Figs 5, 6). Posterior side of dens almost smooth, with four chaetae of which one strong basal and three in central part (two of normal size and one small) (Fig. 9). Very large, chitinized, bidentate (Figs 6, 9). Ratio of manubrium : dens : mucro = 3.6–4.2 : 2.0–2.4 : 1.

Etymology. The name is given after Maria Minor, who kindly provided some of the material on the new species.

Discussion. To date eight species of *Folsomia* are known from New Zealand (Greenslade 1994; 2012). In addition, three species, *F. parasitica* Salmon, 1942, *F. novaezealandiae* Salmon, 1943, and *F. lunata* Salmon, 1943, were removed from the list



Figures 7–13. *Folsomia minorae* sp. n. (**7–12**) and *F. australica* sp. n. (**13**) **7** apex of Ant. IV, lateral view **8** chaetotaxy of posterior part of Abd. IV, Abd. V and VI **9** furca, tenaculum, and furcal subcoxae, posterior view **10–11** apical part of Leg II (**10**) and III (**11**) **12–13** s-patterns chaetae of Abd. IV–V (lateral s of Abd. IV not shown). org—organite, sms—subapical ms, pin—pin-chaeta. Scal bar 0.1 mm in **8**, 0.03 mm in **9**, others, 0.01 mm.

as synonyms or were moved to the genus *Cryptopygus* (Bellinger et al. 2016). Among the valid species, five are blind, while others show different number of ocelli (8, 2 and 1, vs. 5 in *F. minorae* sp. n.). Very little morphological data are available for endemic New Zealand *Folsomia* species (*F. miradentata* Salmon, 1943, *F. pusilla* Salmon, 1944, *F. salmoni* Stach, 1947, and *F. sedecimoculata* Salmon, 1943). Particularly, figures of the furca are known only for *F. sedecimoculata* and *F. pusilla*. Both species show a more common structure of the dens (typical of the genus), which is slender and continuously narrowed, unlike in *F. minorae* sp. n. Clavate tibiotarsal hairs were not figured or mentioned in descriptions of New Zealand forms (present in the new species). A comparison between *F. minorae* sp. n. and *Cryptopygus* s.s. is given below.

Differentiating characters of the new species are: five ocelli, unique subapical organite of Ant.IV, clavate tibiotarsal hairs, presence of outer teeth on claws, stout dens with few chaetae, and a very large mucro. Well differentiated '3+2' sensillary pattern of s-chaetae on Abd.V is also characteristic (see below). Three long and slender dorsal s-chaetae of Abd.V are found in species of several groups of *Folsomia* of the Holarctic (i.e. *F. penicula* Bagnall, 1939, *F. quadrioculata* (Tullberg, 1871) and *F. sensibilis* Kseneman, 1936), which belong to either '3+2' or '3+1+1' patterns.

Distribution and ecology. *F. minorae* sp. n. is known from three localities in South Island, New Zealand. It is probably a species restricted to mountainous areas.

***Folsomia australica* sp. n.**

<http://zoobank.org/C9C7BCD0-5200-4701-A90A-7F2763C9A550>

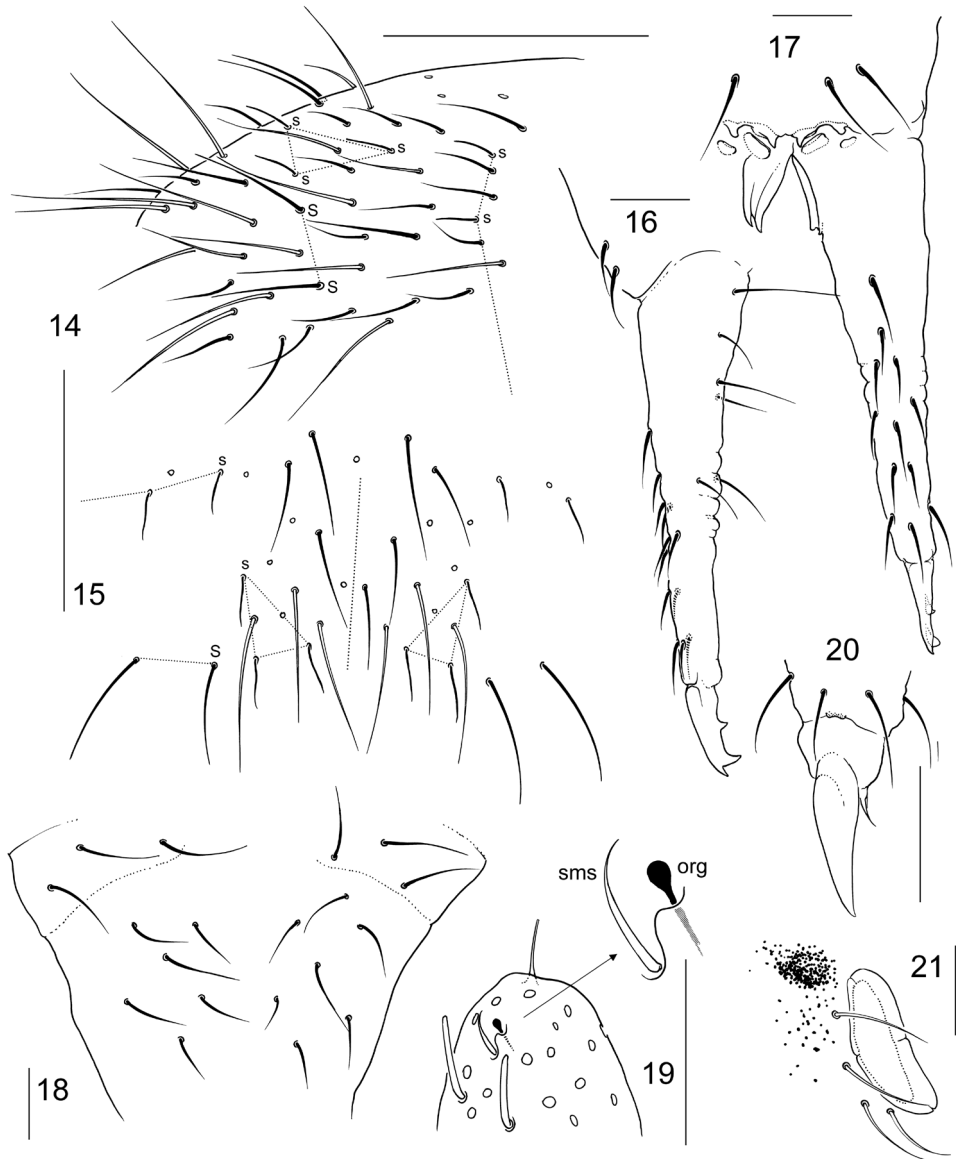
Figs 14–21

Type material. Holotype: adult ♀. Australia, Chiltern National Park, Victoria, heathy dry forest, mostly native, 36°7'53"S, 146°36'20"E, 12.iv.2015, M. Lythe leg. Four paratypes, one adult ♀, one adult ♂ and two sub-adult ♀♀ of the same size as adults. Holotype and two paratypes in SAMA, two paratypes in MSPU.

Other material. 10 specimens in ethanol (SAMA). Australia, Victoria, Mt Pilot National Park, 36°18'45"S, 146°33'16"E. 20.vii.2015, M. Lythe leg.

Diagnosis. *Folsomia* species with 1+1 ocelli; chaetotaxy of dens 12/6; tridentate mucro; 2 lateral s-chaetae on Abd.V clearly longer than 3 dorsal ones; 2+2 chaetae on anterior side of manubrium.

Description. Body size from 0.60 (adult male) to 0.75 mm (one of sub-adult females). White, with one black ocellus on each side of the head (Fig. 21). Body of normal shape for the genus. Abd IV, V and VI clearly fused dorsally, Abd.IV and III well separated. Cuticle "smooth", with fine orthogonal granulation, granules much smaller than chaetae bases. Ocelli 1+1, well-marked only by pigmentation, cuticular cornea weak. PAO wide, constricted, smaller (ca. 0.8) than width of Ant I, about 1.5 as long as inner unguis length (Fig. 21). Maxillary outer lobe with four sublobal hairs, maxillary palp bifurcate. Labral formula as 4/5,5,4. Labium with five papillae (A–E), 'guard' e7 present (whole number of 'guards' hard to estimate), with three proximal,



Figures 14–21. *Folsomia australica* sp. n. **14–15** chaetotaxy of posterior part of abdomen, lateral (**14**) and dorsal (**15**) views **16–17** furca, lateral (**16**) and anterior (**17**) view **18** manubrium, posterior view **19** apex of Ant.IV, dorsal view, left antenna **20** distal part of Leg III **21** ocellus and PAO. org—organite, sms—subapical ms. Scale bar 0.05 mm in **14**, **15**, others, 0.01 mm.

four basomedian, and five basolateral chaetae. Ventral side of head with 4+4 postlabial chaetae. Ant.I with three ventral s-chaetae (s) and two small basal ms-chaetae (bms), dorsal and ventral, Ant.II with three bms and one latero-distal s. Ant.III with one bms and with five distal s (including one lateral), without additional s-chaetae. Ant.IV

with several tubular s-chaetae. Subapical organite large and roundish, set together with subapical ms, as common for family (Fig. 19). S-chaetae formula 4,3/2,2,2,3,5 (s) and 1,0/0,0,0 (ms). Tergal s-chaetae shorter than common chaetae. Medial s-chaetae on Th.II–Abd.III in mid-tergal position, on Abd.I–III between Mac1 and Mac2. Abd.V with five s-chaetae with three dorsal ones (al, accp1, accp2), almost as long as common chaetae, and two lateral long, macrochaetae-like ('3+2' pattern) (Figs 14–15). Two lateral s-chaetae often slightly thickened on proximal 2/3 that makes them more distinct. Macrochaetae smooth, 1,1/3,3,3 in number, medial ones on Abd.V shorter than dens (0.6–0.8) and 2.6–3.0 times longer than mucro. Foil chaetae at the tip of abdomen absent. Axial chaetotaxy of Th.II–Abd.III as 9–10,6–8/4–5,4–5,4. Th.III with 1+1 ventral chaetae. Unguis without teeth (Fig 20). Empodial appendage about 0.6 as long as U3. Upper and lower subcoxae of Leg I, II, III with 1,1; 3,6; 5–6,6–7 chaetae. Tibiotarsi without additional chaetae on Leg I and II (21 chaetae), and with several additional chaetae on Leg III. Tibiotarsal tenent chaetae pointed, shorter than U3 (0.8–1.0). VT with 3+3 laterodistal and five posterior chaetae, of which four in transversal row, anteriorly without chaetae. Tenaculum with 4+4 teeth and a chaeta. Anterior furcal subcoxae with 8–12, posterior one with five chaetae. Anterior side of manubrium with two pair of chaetae, 2+2 (rarely 1+2) (Figs 16, 17). Posterior side of manubrium with 3+3 laterobasal, 6-7+6-7 on main part, without apical and lateral chaetae (Fig. 18) (shown in the only variant seen). Dens slender, with 12 anterior chaetae arranged as 1,1,2,3,2,3 (Figs 16, 17). Posterior side of dens with few distinct crenulations at the middle, four chaetae on proximal half and two medially. Mucro tridentate. Ratio of manubrium : dens : mucro = 2.9–3.4 : 3.6–4.2 : 1.

Etymology. The name is given after the geographical distribution of the new species.

Discussion. *Folsomia australica* sp. n. resembles the only other native Australian species of the genus, i.e. *F. loftyensis* (Womersley, 1934) (after the redescription of Potapov and Greenslade 2010) by chaetotaxy of dens 12/6, tridentate mucro, 1+1 ocelli, ms-formula of body 10/000, differentiation of s-chaetae on Abd.V, and other characters. It differs in having 2+2 chaetae (vs. 4–5+4–5 in *F. loftyensis*) on the anterior side of manubrium. Juvenile specimens of the two species are probably hard to distinguish. The new species was recorded by Potapov and Greenslade (2010) as "*Folsomia* sp. aff. *loftyensis*". *Folsomia australica* sp. n. and *F. minorae* sp. n. are dissimilar indicating that the "austral" members of the genus *Folsomia* can also be heterogeneous, as in the Northern Hemisphere.

Morphological features of the furca of *F. australica* and *F. loftyensis*, especially the tridentate mucro, are shared with several species of *Cryptopygus*: *C. tricuspis* Enderlein, 1909 (sub-Antarctic), *C. insignis* Massoud and Rapoport, 1968 (South America), *C. patagonicus* Izarra, 1972 (South America), and three unnamed species from South Africa (*Cryptopygus* sp. 5, *C. sp. 6*, and *C. sp. 7*, see below). These species probably represent another group of *Cryptopygus* s.s., dissimilar to *C. antarcticus*, which could be ancestral to the "Australian" species of *Folsomia*.

Distribution. *Folsomia australica* sp. n. is known from two localities in south-eastern part of Australia (Victoria and New South Wales).

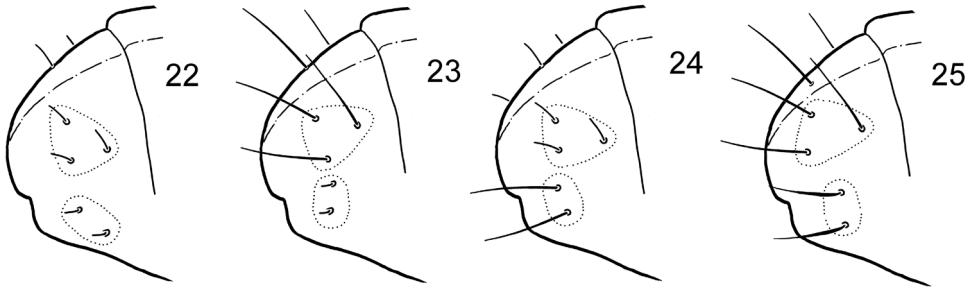
General discussion

The fifth abdominal segment of all species of the genus *Cryptopygus*, as in *Folsomia*, has five s-chaetae on each side: accp1, accp2, accp3, accp4 and as. The s-pattern of *C. antarcticus* and several other species is probably the most primitive as it consists of regularly scattered sub-equal short and thin s-chaetae (Fig. 26). Weak differentiation is observed: s-chaetae of dorsal triplet (as+accp1+accp2) slightly longer and sometimes thinner than s-chaetae of lateral duplet (accp1+accp2) (Figs 22, 26). Such arrangement and differentiation of s-chaetae can be called as a weak ‘3+2’ pattern (for terminology, see Potapov and Greenslade 2010). Most species of *Cryptopygus s.s.* show this pattern, sometimes the difference between the length of “triplet” and “duplet” s-chaetae is hardly evident. Subsequent evolution is expressed by stronger differentiation: s-chaetae of dorsal triplet, lateral duplet or either of them become macrochaeta-like (Figs 23–25). Only one species for each of these three apomorphic conditions was found in the material studied here, all only currently known from South Africa. The representation of s-patterns in *Cryptopygus s.s.* for Abd.V is as follows:

1. Weakly differentiated “3+2” pattern (Figs 22, 26). S-chaetae shorter than common chaetae. In a few short-haired species, (*C. badasa* Greenslade, 1995, *C. binoculatus* Deharveng, 1981, *C. lawrencei* Deharveng, 1981, *C. sp.4*) s-chaetae are almost as long as common chaetae.

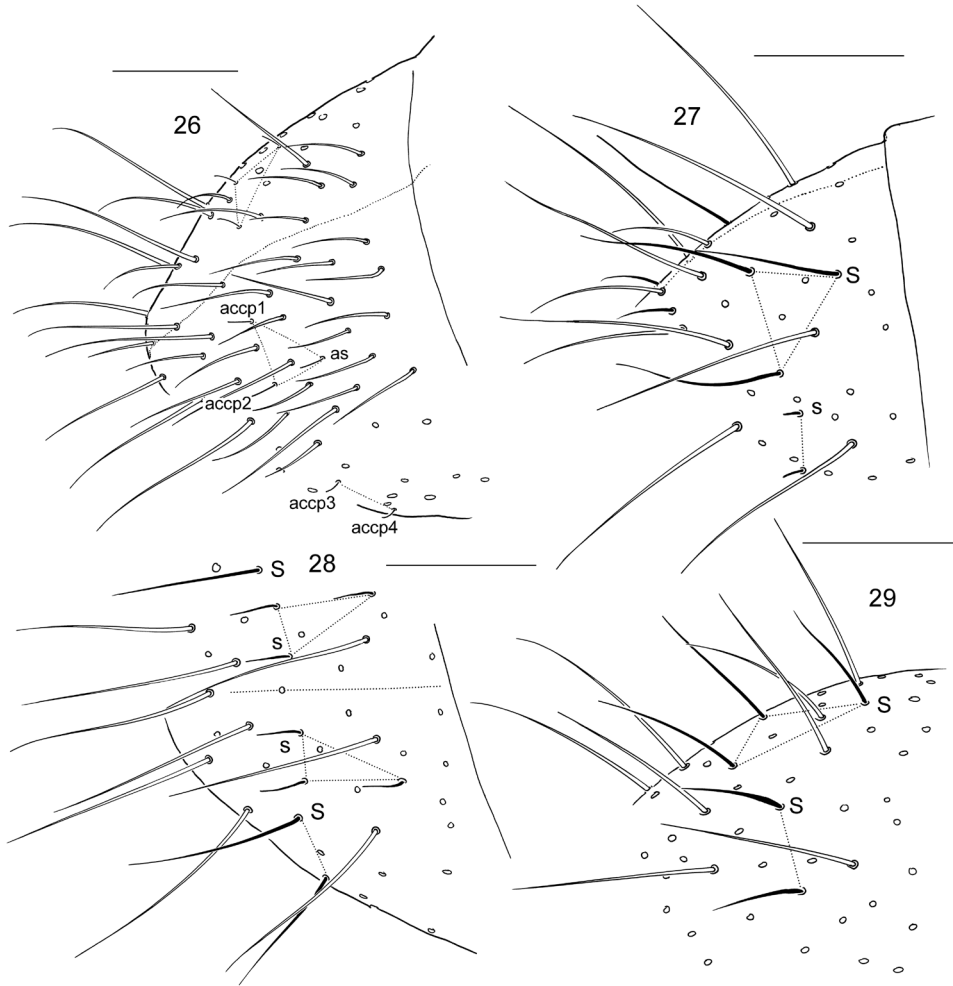
Representatives:

- *C. antarcticus antarcticus* Willem, 1902. Material: several locations in Antarctic Peninsula (King George Isl., Deception Isl., Devil Isl.) leg. D. Russell.
- *C. antarcticus maximus* Deharveng, 1981. S-chaetae as in nominate subspecies. Material: Kerguelen Isl. leg. J. Travé.
- *C. antarcticus reagens* (Enderlein, 1909). S-chaetae of dorsal triplet almost twice longer than s-chaetae of duplet. Material: Crozet Isl. (sub-Antarctic), leg. J. Travé.
- *C. antarcticus travei* Deharveng, 1981. S-chaetae of dorsal triplet almost as long as those of duplet. Material: Marion Isl. (sub-Antarctic), leg. J. Travé.
- *C. araucanus* Massoud & Rapoport, 1968. S-chaetae of dorsal triplet slightly longer than s-chaetae of duplet. Material: syntypes, several locations in Argentina (Futalaufquen, Lago Curruhé, Lago Menendez). Collections of the Museum national d’Histoire naturelle (Paris, France).
- *C. badasa* Greenslade, 1995. All s-chaetae very short, “triplet” s-chaetae slightly longer. Material: Antarctic Peninsula (Devils Isl.), leg. D. Russell; South Georgia (sub-Antarctic), leg. V. Bulavintsev.
- *C. binoculatus* Deharveng, 1981. S-chaetae subequal. Material: holotype, Crozet Isl. (sub-Antarctic).
- *C. insignis* Massoud & Rapoport, 1968. S-chaetae subequal. Material: syntypes, Lago Menendez (Argentina). Collections of the Museum national d’Histoire naturelle (Paris, France).



Figures 22–25. “3+2” s-patterns of austral *Cryptopygus*. **22** *C. antarcticus* **23–25** *Cryptopygus*: sp. 5 (**23**) sp. 6 (**24**) sp. 7 (**25**) (all three species from South Africa).

- *C. hirsutus* (Denis, 1931). S-chaetae subequal. Material: possible syntypes, Costa Rica. Collections of the Museum national d’Histoire naturelle (Paris, France).
 - *C. lawrencei* Deharveng, 1981. “Triplet” s-chaetae slightly longer. Material: Kerguelen Isl. (sub-Antarctic), leg. J. Travé.
 - *C. pilosus* (Womersley, 1934). S-chaetae as in *C. antarcticus*. Material: South Australia, Lofty Ranges, leg. P. Greenslade.
 - *C. tricuspis* Enderlein, 1909. S-chaetae as in *C. antarcticus*. Material: Kerguelen Isl. (sub-Antarctic), leg. J. Travé.
 - *C. ulrikeae* (Najt & Thibaud, 1987), **comb. n.** S-chaetae sub-equal. Separation of Abd.IV and V as in other species of the genus *Cryptopygus*. Primarily, it was described as *Folsomia ulrikeae* (Najt and Thibaud, 1987). Material: holotype, Ecuador. Collections of the Museum national d’Histoire naturelle (Paris, France).
 - *Cryptopygus* sp. 1 (complex ‘antarcticus’). S-chaetae as in *C. antarcticus*. Characters common with the nominotypic subspecies of *C. antarcticus* but body more slender. Material: New Zealand (South Island), leg. M. Minor.
 - *Cryptopygus* sp. 2 (complex ‘antarcticus’). S-chaetae as in *C. antarcticus*. With the characters of *C. antarcticus* but ms formula of body tergites as 10/000. Material: New Zealand (North Island).
 - *Cryptopygus* sp. 3. S-chaetae as in *C. antarcticus*. With 6+6 ocelli. Manubrium without anterior chaetae Dens with one anterior chaeta, mucro bidentate. Material: South Africa (Jonkershoek), leg. C. Janion-Scheepers.
 - *Cryptopygus* sp. 4. S-chaetae as in *C. antarcticus*. With 8+8 ocelli. Dens rather long, with nine anterior and 5 posterior chaetae, mucro bidentate. Common chaetae and macrochaetae on body short. Material: South Africa, (Sutherland), leg. C. Janion-Scheepers.
2. “3+2” pattern with development of dorsal triplet (Figs 23, 27). Three dorsal s-chaetae (as, accp1, accp2) almost as long as macrochaetae, two lateral (accp3, accp4) short.
- *Cryptopygus* sp. 5. Dens with 9-10 anterior and 6 posterior chaeta, mucro tridentate. Material: South Africa (Table Mountain), leg. L. Deharveng and A. Bedos.



Figures 26–29. S-chetae on Abd.V in austral *Cryptopygus*. **26** *C. antarcticus* **27–29** *Cryptopygus*: sp.5 (**27**) sp.6 (**28**) sp.7 (**29**) (all three species from South Africa). Scale bar 0.05 mm.

3. “3+2” pattern with development of lateral duplet (Figs 24, 28). Three dorsal s-chetae (as, accp1, accp2) short, two lateral (accp3, accp4) long.
 - *Cryptopygus* sp.6. With 4+4 or 5+5 ocelli. PAO with strong inner denticles. Dens with ten anterior chaetae, mucro tridentate. Material: South Africa (Little Switzerland), leg. E. Krzemińska.
4. “3+2” pattern with development of all s-chetae (Figs 25, 29). Three dorsal s-chetae (as, accp1, accp2) thin, two lateral (accp3, accp4) also long, somewhat shorter than dorsal, somewhat flame-shaped.
 - *Cryptopygus* sp.7. With 4+4 ocelli. Dens with 11-12 anterior and five posterior chaeta, mucro tridentate. Material: South Africa (Sutherland), leg. C. Janion-Scheepers.

S-chaetae patterns of *Cryptopygus* are probably more diverse than shown above: *Cryptopygus yosii* Izarra, 1965 (Argentina, after our study of a syntype) shows “3+1+1” pattern in which accp3 is thick and tubular and accp4 is short and moved to the latero-ventral position. More material on less primitive species needs to be studied to complete the generic overview. Nevertheless, s-patterns of Abd.V in *Cryptopygus* seem to be less divergent than in the larger genus *Folsomia* (Potapov and Greenslade 2010), while “austral” variant 4 (see above) has not been discovered in the latter genus.

The dorsal fusion or separation of genital (Abd.V) and pre-genital segment (Abd.IV) is traditionally considered to be of great taxonomic value in the classification of the sub-family Anurophorinae s.l., and the genus *Folsomia* is defined by the apomorphic condition of this character (fusion). Based on the available literature and our own observations, the s-chaetotaxy of “austral” *Cryptopygus* s.s. shows principally the same characteristics as in *Folsomia*, particularly 4,3/2,2,2,3,5 set and arrangement of s-chaetae on Abd.V. The more adaptive characters (furca, ocelli, etc.) vary considerably within both genera. Therefore Abd.IV–V fusion seems to be the only apomorphic character that separates *Folsomia* from *Cryptopygus* and the former genus can be easily derived from the latter. This key character can potentially show a high level of homoplasy and *Folsomia* is probably a polyphyletic or paraphyletic group. In the Northern Hemisphere, the high diversity of *Folsomia* makes it difficult to find an appropriate ancestor or even ancestors among known taxa. In contrast, at least three “austral” native *Folsomia* mentioned above show much in common with certain species of *Cryptopygus* s.s. Thus, all the main characters of *F. minorae* sp. n. (ocelli, clavate tibiotarsal hairs, outer teeth on claws, dens and mucro) indicate its close relationship to a group of species similar to *Cryptopygus antarcticus* (Wise 1967; Massoud and Rapoport 1968, Deharveng 1981), while the characters of *F. australica* sp. n. are shared with several *Cryptopygus* species with a slender dens and tridentate mucro (see the remarks to the species). S-chaetae patterns of Abd. V in *F. minorae* is the same as in *Cryptopygus* sp. 5 from South Africa (Figs 12, 23), to which this new species is, however, less similar than to the “antarcticus” group. S-pattern of *F. australica* and *F. loftyensis* is identical to *Cryptopygus* sp. 6 from South Africa (Figs 14, 24).

The generic position of both lineages, *F. minorae* and *F. australica-loftyensis*, can be modified in the future, depending on the increase of knowledge on the generic groups “*Cryptopygus*” and “*Folsomia*”. The genus *Folsomia* is also very diverse in the Holarctic and consists of several species groups of which several differ in characters of great taxonomical value and may justify the status of new separate genera.

Acknowledgements

We thank Maria Minor (Massey University, New Zealand) and Arne Fjellberg (Norway) who provided material on *F. minorae* sp. n. The material belonging to the following persons was also used: Valery Bulavintsev (Moscow), Penelope Greenslade (Ballarat), Morgan Lythe (Melbourne), David Russell (Görlitz) and Wanda M. Weiner (Kraków). We thank the New Zealand Department of Conservation for the collection

permit (national authorization # 38116-GEO to M. Minor), the staff of the Snow Farm for help in providing access, Dr. Alastair Robertson (Institute of Agriculture & Environment, Massey University) for help with fieldwork, CapeNature and SanParks (South Africa) and Parks Victoria (Australia) for collection permits. The work in South Africa was supported by the Protea I and II South Africa-France bilateral grants (no. 68652 for L. Deharveng). This work was partly supported by Museum national d'Histoire Naturelle (Paris, France) and RFBR research project № 14-04-01140 (Russia) for MP. Jean-Marc Thibaud (Paris) lent some material of *Cryptopygus*. This work would not have been possible without creative initiative of Anatoly Babenko (Moscow). We are grateful to Arne Fjellberg, Wanda Weiner and three anonymous reviewers for their comments. The collections in Antarctic Peninsula were supported by Federal Environment Agency and funded by the German Federal Ministry for the Environment, Nature Conservation and Nuclear Safety under the project nr. FKZ 3709 85 157. CJS was supported by Australian Research Council grant DP140102815.

References

- Bagnall RS (1939) Notes on British Collembola. III-VII. The Entomologist's Monthly Magazine 75: 56–63.
- Bellinger PF, Christiansen KA, Janssens F (1996-2016) Checklist of the Collembola of the World. <http://www.collembola.org>
- Deharveng L (1981) Collembolles des Iles Subantarctiques de l'Océan Indien. Biologie des sols. Comité National Français des Recherches Antarctiques 48: 33–109.
- Deharveng L, Potapov M, Bedos A (2005) *Cylindropygus ferox* gen. n., sp. n.: A new member of the *Cryptopygus* complex (Collembola, Isotomidae) from central France. Journal of Natural History 39: 2179–2185. <https://doi.org/10.1080/00222930500061213>
- Deharveng L (1987) Révision taxonomique du genre *Tetracanthella* Schött, 891 (Collembola: Isotomidae). Travaux du Laboratoire d'Écobiologie des Arthropodes édaphiques, Toulouse 5(3): 1–151.
- Denis JR (1931) Contributo alla conoscenza del Microgenton di Costa Rica II. Collembolles de Costa Rica avec une contribution aux espèces de l'ordre. Bolletino del Laboratorio di Zoologia Generale e Agraria Portici 25: 69–170.
- Enderlein G (1909) Die Insekten des antarktischen Gebiets. Deutsche Südpolar Expedition, 1901–1903. 10(4): 361–528.
- Fjellberg A (1991) *Proisotoma roberti* n. sp. from Greenland, and redescription of *P. ripicola* Linnaniemi, 1912 (Collembola, Isotomidae). Entomologiske Meddelelser 59: 81–83.
- Fjellberg A (2007) The Collembola of Fennoscandia and Denmark. Part II: Entomobryomorpha and Symphypleona. Fauna Entomologica Scandinavica 42: 1–264. <https://doi.org/10.1163/ej.9789004157705.i-265>
- Greenslade P (1994) Collembola. In: Houston WWK (Ed.) Zoological catalogue of Australia. Volume 22. Protura, Collembola, Diplura. CSIRO, Melbourne, 19–138.

- Greenslade P (1995) Collembola from the Scotia Arc and Antarctic Peninsula including descriptions of two new species and notes on biogeography. *Polskie Pismo Entomologiczne* 64: 305–319.
- Greenslade P (2012) Collembola. In: Gordon DP (Ed.) *The New Zealand inventory of biodiversity: a species 2000 symposium review*. Volume 1, Animalia. Canterbury University Press, Christchurch, 237–243.
- Greenslade P (2015) Synonymy of two monobasic Anurophorinae genera (Collembola: Isotomidae) from the Antarctic Continent. *New Zealand Entomologist* 38: 134–141. <https://doi.org/10.1080/00779962.2015.1033810>
- Izarra DC (1965) Fauna Colembologica de Sierra de la Ventana (Provincia de Buenos Aires, Argentina). *Physis* 25(70): 263–276
- Izarra DC (1972) Fauna Colembologica de Isla Victoria (Provincia de Neuquen, Argentina). III. Familias Isotomidae y Entomobryidae. *Physis* 31(83): 373–382.
- Jordana R, Hamra-Kroua S, Baquero E (2009) Redescription of *Isotominella geophila* Delamare Deboutteville, 1948 from Algeria (Collembola, Entomobryomorpha, Isotomidae), a second world record for an Ivory Coast species. *Zootaxa* 2169: 63–68.
- Kseneman M (1936) Diagnosen neuer Collembolenarten aus Mitteleuropa. *Annalen der Tschechoslowakischen Akademie der Landwirtschaft* 11: 101–109.
- Massoud Z, Rapoport EH (1968) Collemboles Isotomides d’Amérique du sud et de l’Antarctique. *Biologie d’Amérique Australe* 4: 307–337.
- Massoud Z, Rapoport EH (1968) Collemboles Isotomides d’Amérique du sud et de l’Antarctique. *Biologie d’Amérique Australe* 4: 307–337.
- Najt J, Thibaud J-M (1987) Collemboles (Insecta) de l’Équateur. I. Hypogastruridae, Neaururinae et Isotomidae. *Bulletin du Muséum national d’Histoire naturelle*, 4^e série, section A, 9(1): 201–209.
- Potapov M, Gao Y, Deharveng L (2013) Taxonomy of the *Cryptopygus* complex. I. *Pauropygus* – a new worldwide littoral genus (Collembola, Isotomidae). *ZooKeys* 304: 1–16. <https://doi.org/10.3897/zookeys.304.4083>
- Potapov MB, Greenslade P (2010) Redescription of *Folsomia loftyensis* Womersley with notes on the sensillary arrangement of the genital segment in the genus (Collembola: Isotomidae). *Zoologischer Anzeiger* 249: 13–20. <https://doi.org/10.1016/j.jcz.2010.01.004>
- Rusek J (2002) Do we have *Cryptopygus* representatives (Collembola: Isotomidae) in Europe? *Pedobiologia* 46: 302–310. <https://doi.org/10.1078/0031-4056-00137>
- Salmon JT (1942) New records of Collembola from New Zealand, with descriptions of new genera and species. Part I. Arthropleona. *Transactions and Proceedings of the Royal Society of New Zealand* 72: 373–388.
- Salmon JT (1943) The Genus *Folsomia* (Collembola) in New Zealand. *Transactions and Proceedings of the Royal Society of New Zealand* 73: 73–75.
- Salmon JT (1944) New genera, species and records of New Zealand Collembola and a discussion of *Entomobrya atrocincta* Schött. *Records of the Dominion Museum* 1(2): 123–182.
- Schött H (1891) Nya nordiska Collembola. *Entomologisk Tidskrift*, 12: 191–192.
- Schött H (1893) Zur Systematik und Verbreitung palaarktischer Collembola. *Kongliga Svenska vetenskaps-akademiens handlingar* (B) 25(11): 1–100.

- Stach J (1947) The Apterygotan Fauna of Poland in Relation to the World-Fauna of this Group of Insects. Family: Isotomidae. Polska Akademia Umiejętności, Acta monographica Musei Historiae Naturalis, Kraków, 488 pp.
- Szeptycki A (1972) Morpho-systematic studies of Collembola. III. Body chaetotaxy in first instars of several genera of Entomobryomorpha. Acta Zoologica Cracoviensia 17: 341–372.
- Tullberg T (1871) Förteckning öfver svenska Podurider. Öfversigt af Kongliga Vetenskaps-Akademiens Förhandlingar 28: 143–155.
- Willem V (1902) Les collemboles recueillis par l'expédition antarctique belge. Annals of the society for Entomology of Belgium 45: 260–262.
- Willem V (1902) Note préliminaire sur les Collemboles des Grottes de Han et de Rochefort. Annales de la Société Entomologique de Belgique 46: 275–83.
- Wise KAJ (1967) Collembola (Springtails). Antarctic Research Series 10: 123–148. <https://doi.org/10.1029/ar010p0123>
- Womersley H (1934) A preliminary account of the Collembola – Arthropleona of Australia. II. – superfamily Entomobryoidea. Transactions of the Royal Society of South Australia 58: 86–138.