





Draft Genome Sequence of *Providencia* stuartii PS71, a Multidrug-Resistant Strain Associated with Nosocomial Infections in Greece

A. Liakopoulos,^a O. Oikonomou,^b D. W. Wareham^{b,c}

Department of Bacteriology and Epidemiology, CVI of Wageningen University, Lelystad, The Netherlands^a; Antimicrobial Research Group, Blizard Institute, Barts & the London School of Medicine and Dentistry, Queen Mary University of London, London, United Kingdom^b; Division of Infection, Barts Healthcare NHS Trust, London, United Kingdom^c

ABSTRACT Providencia stuartii is frequently associated with nosocomial outbreaks and displays intrinsic resistance to many commonly used antimicrobials. We report here the draft genome sequence of a P. stuartii strain carrying acquired resistance genes conferring panresistance to cephalosporins (bla_{SHV-5} and bla_{VEB-1}), carbapenems (bla_{VIM-1}), and aminoglycosides (rmtB) involved in an outbreak in Greek hospitals.

Providencia stuartii has emerged as an important nosocomial pathogen responsible for urinary tract infections in patients with indwelling urinary catheters, hospital-acquired pneumonia, bloodstream infections, and sepsis (1), with significant impact on patient morbidity, mortality, treatment, and management costs (2).

 $P.\ stuartii$ isolate PS71 was recovered in 2013 from the bloodstream of a patient during a cluster of $P.\ stuartii$ infections in a critical care unit in a tertiary-care hospital in Athens, Greece (3). This outbreak-associated strain exhibited resistance to cephalosporins (first, second, third, and fourth generation), β -lactam–inhibitor combinations (amoxicillin-clavulanate and piperacillin-tazobactam), carbapenems (ertapenem and imipenem), aminoglycosides, fosfomycin, polymyxins (colistin and polymyxin B), quinolones, and tigecycline (3).

Genomic DNA was extracted and subjected to whole-genome sequencing with the Illumina MiSeq platform (Illumina, Inc., San Diego, CA), producing 2×250 -bp pairedend reads, generating a total of 843,412 reads with a 475 bp average length. The Trimmomatic algorithm (version 0.36) (4) was used to trim all the generated reads and their quality assessed with in-house scripts using BedTools (version 2.25.0) (5), BWAmem (version 2) (6), and SAMtools (version 1.3.1) (7) algorithms. High-quality filtered reads were subsequently assembled *de novo* using SPAdes algorithm (version 3.7.1) (8), resulting in 79 scaffolds with an N_{50} of 455,952 bp. The sequence coverage of the *de novo* assemblies was approximately 83 reads per assembled base. The draft genome sequence of PS71 consists of 4,411,042 bp with 41.75% average G+C content.

Provisional annotation carried out using the Prokka algorithm (version 1.11) (9) identified at least 4,026 coding sequences (CDSs), including 76 tRNAs and eight rRNAs. The assembled genome was used to predict the presence of acquired antibiotic resistance genes using ResFinder (10) and confirmed the presence of 24 genes conferring resistance to aminoglycosides [aadA1, aadA2, aadB, aac(6')-Ia, aph(3')-Ia, strA, strB, and rmtB], β -lactams ($bla_{TEM-1b'}$, $bla_{OXA-10'}$, $bla_{SHV-5'}$, $bla_{VEB-1'}$, and bla_{VIM-1}), macrolides, lincosamides, and streptogramin B [mph(A)] and phenicols (cmlA1 and catA3), rifampin (arr-2), sulfonamides (sul1 and sul2), tetracyclines [tet(A), tet(B), and tet(G)], and trimethoprim (dfrA1 and dfrA12). The presence of multidrug efflux pump-mediated resis-

Received 17 January 2017 **Accepted** 24 January 2017 **Published** 23 March 2017

Citation Liakopoulos A, Oikonomou O, Wareham DW. 2017. Draft genome sequence of *Providencia stuartii* PS71, a multidrugresistant strain associated with nosocomial infections in Greece. Genome Announc 5: e00056-17. https://doi.org/10.1128/genomeA.00056-17.

Copyright © 2017 Liakopoulos et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to D. W. Wareham, d.w.wareham@gmul.ac.uk.

tance mechanisms was also inferred from the genome (i.e., acrB, acrR, cmeB, mtrF, mfs, macA, macB, and tolC), as well as resistance to toxic compounds, including arsenic (arsA, arsB, arsC, arsD, and arsR), cadmium, cobalt and zinc (czcB, czcD, trcD, and trMer), and mercury (merA, merC, merD, merE, and merR). PlasmidFinder (11) indicated that PS71 contains plasmids with ColE, A/C, and R replicon types, with the latter located on a multireplicon plasmid (3).

There are increasing reports of multiresistant strains of *Providencia* in the Mediterranean, and enhanced surveillance is needed to identify and track the epidemiology of resistant strains, their impact on human health, and the molecular epidemiology. This is the first draft genome sequence of such a multiresistant *P. stuartii* strain associated with a nosocomial outbreak in Greece.

Accession number(s). The draft genome sequence of *P. stuartii* PS71 has been deposited in the DDBJ/EMBL/GenBank databases under the accession number MSAA00000000. The version described in this paper is the first version, MSAA01000000.

ACKNOWLEDGMENT

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

REFERENCES

- Armbruster CE, Smith SN, Yep A, Mobley HL. 2014. Increased incidence of urolithiasis and bacteremia during *Proteus mirabilis* and *Providencia* stuartii coinfection due to synergistic induction of urease activity. J Infect Dis 209:1524–1532. https://doi.org/10.1093/infdis/jit663.
- Zavascki AP, Carvalhaes CG, da Silva GL, Tavares Soares SP, de Alcântara LR, Elias LS, Sandri AM, Gales AC. 2012. Outbreak of carbapenemresistant *Providencia stuartii* in an intensive care unit. Infect Control Hosp Epidemiol 33:627–630. https://doi.org/10.1086/665730.
- Oikonomou O, Liakopoulos A, Phee LM, Betts J, Mevius D, Wareham DW. 2016. Providencia stuartii isolates from Greece: co-carriage of cephalosporin (bla_{SHV-5}, bla_{VEB-1}), carbapenem (bla_{VIM-1}), and aminoglycoside (rmtB) resistance determinants by a multidrug-resistant outbreak clone. Microb Drug Resist 22:379–386. https://doi.org/10.1089/mdr.2015.0215.
- 4. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10.1093/bioinformatics/btu170.
- Quinlan AR, Hall IM. 2010. BEDTools: a flexible suite of utilities for comparing genomic features. Bioinformatics 26:841–842. https:// doi.org/10.1093/bioinformatics/btq033.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25:1754–1760. https://doi.org/10.1093/bioinformatics/btp324.

- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb .2012.0021.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi.org/10.1093/jac/dks261.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/ AAC.02412-14.