

Video Article

Profiling Thiol Redox Proteome Using Isotope Tagging Mass Spectrometry

Jennifer Parker¹, Ning Zhu², Mengmeng Zhu², Sixue Chen^{1,2,3,4}¹Plant Molecular and Cellular Biology Program, University of Florida²Department of Biology, University of Florida³Interdisciplinary Center for Biotechnology Research, University of Florida⁴Genetics Institute, University of FloridaCorrespondence to: Sixue Chen at schen@ufl.eduURL: <http://www.jove.com/video/3766>DOI: [doi:10.3791/3766](https://doi.org/10.3791/3766)Keywords: Genetics, Issue 61, *Pseudomonas syringae* pv. *tomato* (*Pst*), redox proteome, cysteine-reactive tandem mass tag (cysTMT), LTQ-Orbitrap mass spectrometry

Date Published: 3/24/2012

Citation: Parker, J., Zhu, N., Zhu, M., Chen, S. Profiling Thiol Redox Proteome Using Isotope Tagging Mass Spectrometry. *J. Vis. Exp.* (61), e3766, doi:10.3791/3766 (2012).

Abstract

Pseudomonas syringae pv. *tomato* strain DC3000 not only causes bacterial speck disease in *Solanum lycopersicum* but also on *Brassica* species, as well as on *Arabidopsis thaliana*, a genetically tractable host plant^{1,2}. The accumulation of reactive oxygen species (ROS) in cotyledons inoculated with DC3000 indicates a role of ROS in modulating necrotic cell death during bacterial speck disease of tomato³. Hydrogen peroxide, a component of ROS, is produced after inoculation of tomato plants with *Pseudomonas*³. Hydrogen peroxide can be detected using a histochemical stain 3'-3' diaminobenzidine (DAB)⁴. DAB staining reacts with hydrogen peroxide to produce a brown stain on the leaf tissue⁴. ROS has a regulatory role of the cellular redox environment, which can change the redox status of certain proteins⁵. Cysteine is an important amino acid sensitive to redox changes. Under mild oxidation, reversible oxidation of cysteine sulfhydryl groups serves as redox sensors and signal transducers that regulate a variety of physiological processes^{6,7}. Tandem mass tag (TMT) reagents enable concurrent identification and multiplexed quantitation of proteins in different samples using tandem mass spectrometry^{8,9}. The cysteine-reactive TMT (cysTMT) reagents enable selective labeling and relative quantitation of cysteine-containing peptides from up to six biological samples. Each isobaric cysTMT tag has the same nominal parent mass and is composed of a sulfhydryl-reactive group, a MS-neutral spacer arm and an MS/MS reporter¹⁰. After labeling, the samples were subject to protease digestion. The cysteine-labeled peptides were enriched using a resin containing anti-TMT antibody. During MS/MS analysis, a series of reporter ions (i.e., 126-131 Da) emerge in the low mass region, providing information on relative quantitation. The workflow is effective for reducing sample complexity, improving dynamic range and studying cysteine modifications. Here we present redox proteomic analysis of the *Pst* DC3000 treated tomato (Rio Grande) leaves using cysTMT technology. This high-throughput method has the potential to be applied to studying other redox-regulated physiological processes.

Video Link

The video component of this article can be found at <http://www.jove.com/video/3766/>

Protocol

1. Growing Seedlings and Preparing Bacteria

1. Seedlings are germinated in MetroMix 500 soil mixture (BWI Companies) in a growth chamber (160 $\mu\text{mol photons m}^{-2}\text{s}^{-1}$ with a photoperiod of 8 hour light at 22 °C and 16 hour dark at 20 °C for 1 week. Relative humidity was set to 70%. Seedlings were watered with tap water as needed to keep the soil from becoming dry.
2. Two seedlings of similar size are then transplanted to 4" diameter pots containing the MetroMix 500 soil and grown for an additional 3 weeks in the same conditions as 1.1.
3. *Pseudomonas syringae* (*Pst*) is initially T-streaked onto Kings B Medium (KBM) plates (per liter: 20 g Proteose Peptone, 0.1975 g MgSO₄, 1% Glycerol, 1.5 g K₂HPO₄, 18 g Agar) and grown for two days at 28 °C. A single colony is collected, mixed into 300 μl liquid Kings B Medium (20 g Proteose peptone, 0.1975 g MgSO₄, 10 mL Glycerol, 10 mL of 100x K₂HPO₄ stock (1.5 g of K₂HPO₄ in 10 mL of H₂O), and spread onto a Kings B Medium plate. This is cultured overnight at 28 °C.
4. The bacteria inoculum is prepared by scraping the cultured bacteria from the KBM plate into 20 mL H₂O. An OD₆₀₀ of 0.03 (~10⁶ cfu/ml) is prepared in 1 L of inoculation buffer (10 mM MgCl₂ + 400 μL Silwet-70). Inoculation buffer minus bacteria was prepared for the control solution.

2. Inoculation of Tomato with *Pst* and H₂O₂ Histochemical Stain

1. Four week old plants were inoculated by dipping into the above inoculation solution for 30 seconds. Clear plastic bags were put over the plants immediately after inoculation.

2. 3'-3' diaminobenzidine (DAB) stain¹¹ was prepared (1 mg/ml DAB in H₂O, pH 3.8 (HCl)) three days before use to achieve complete solubilization. The solution was rocked to mix at room temperature.
3. Twenty four hours after treatment, the fully expanded leaf was removed, placed in DAB stain epidermal side up, and vacuum infiltrated for 15 min. The leaves were then put in the dark at room temperature over night for staining.
4. Leaves were boiled in 95% ethanol for 10 min, and then kept in 75% ethanol.

3. Protein Extraction

1. After harvesting, tomato leaves were ground to fine powder with liquid nitrogen using mortar and pestle.
2. For 0.5 g of tissue, add 1.25 mL Tris pH 8.8 buffered phenol and 1.25 mL / 0.5 g of extraction buffer (0.1 M Tris-HCl pH 8.8, 10 mM EDTA, 0.9 M sucrose) then continue grinding for a few more min in a fume hood.
3. Transfer the extract to Oakridge centrifuge tube (Thermo Fisher Scientific Nalgene Company) and agitate for 2 hrs at room temperature.
4. Centrifuge at 5,000x g and 15 °C for 10 min. Transfer the top phenol phase to a new clean tube.
5. Back-extract the aqueous phase with equal volume extracted phenol buffer; agitate on a shaker for 30 min. Centrifuge and transfer the extract to a new Falcon tube.
6. Repeat the above step one more time and transfer the extract to new Oakridge tube.
7. Precipitate phenol extracted proteins by adding 5 volumes of 0.1 M ammonium acetate in 100% methanol (stored at -20 °C). Vortex and incubate at -20 °C for overnight.
8. Collect the protein pellet by centrifugation at 20,000x g, 4 °C for 20 min .
9. Wash the pellet 2 times with the cold 0.1M ammonium acetate in methanol and 2 times with 80% cold acetone. Add 1.5 mL cold 70% ethanol to pellet and suspend. Transfer to a 2 mL microcentrifuge tube (USA Scientific) and centrifuge at 14,000 rpm, 4 °C for 20 min. Remove ethanol.
10. Dry the pellet briefly in a SpeedVac concentrator and dissolve in a protein extraction buffer, e.g. ReadyPrep Protein Extraction Kit Reagent 3 (8 M Urea, 4% CHAPS, 40 mM Tris-base, 2 M Thiourea) (Bio-Rad). Centrifuge at 14,000 rpm and 20 °C for 30 min to form a pellet. Collect the supernatant.
11. Measure the protein concentration using a CB-X protein assay according to manufacturer's manual (Geno Technology).

4. Sample Preparation and Peptide Labeling with cystMTs

1. Prepare protein sample at a concentration of 2-5 µg/µL. Here we use 100 µg of protein for each mass tag, 20 µl-50 µL samples. For this experiment all 6 tags were used.
2. Add equal volume of alkylation buffer (100 mM Tris-HCl pH7.5, 200 mM iodoacetamide) to the protein sample to block the free thiol group. Buffer should be prepared fresh. Alkylation is performed at 37 °C for 1 hour.
3. Precipitate the protein by adding 1 mL of cold 80% acetone at -20°C for overnight. Pellet the protein by centrifuging at 14,000 rpm, 4 °C for 20 min. Wash the pellet 3 times with 80% Acetone vortexing each time. Remove the supernatant and allow to air dry while on ice.
4. Resuspend the pellet in 50 µL lysis buffer (6 M Urea, 50 mM Tris base, 1 mM EDTA). 0.5 µL of 100 mM Tris(2-Carboxyethyl) Phosphine (TCEP) is then added to reduce the disulfide bonds by incubating for one hour at room temperature.
5. Prepare tags by adding 20 µL of acetonitrile to the reagent tubes to solubilize the tags. Vortex and spin down to the bottom.
6. Wash out extra TCEP using a Microcon 3KD centrifugal filter device (Millipore). Add 50 µL sample to Microcon 3KD column, and 50 µL lysis buffer to the column. Centrifuge for 15 min at 10,000x g and 4 °C. Repeat this step for a total of three times. Remove column and invert into a clean tube. Centrifuge for 5 min at 1,000x g and 4 °C.
7. Check the pH. If needed, adjust the pH to 7.0-8.0 with 1 M HCl.
8. Add 5 µL of the cystTMT reagent to each sample (The cystTMT reagent kit provides 20 µL tags for up to 500 µg protein. This method uses 100 µg protein, therefore we use one-fifth of the tag.) . Allow the reaction to proceed for 2 hr at room temperature.
9. Samples were combined with Laemmli Sample Buffer (Bio-Rad) in a 1:1 ratio.

5. Removal of Non-reacted Tag Sample Fractionation

1. Samples were boiled for 5 min and separated on a pre-cast 12% polyacrylamide gel (Bio-Rad). The gel was rinsed three times in 5 minute intervals with filtered H₂O and stained with Coomassie Blue (Bio-Rad) for 1 hour. The gel was de-stained overnight with H₂O.
2. Twelve fractions were collected from each gel lane. Fractions were determined by protein band concentrations. Fractions were chopped into 1mm pieces and collected into a 1.5 mL tube.
3. Gel pieces were de-stained using 50% acetonitrile and 0.1 M ammonium bicarbonate in H₂O. This step must be repeated until blue color is removed from gel pieces (~3-4 times, 15 min each time).
4. De-stained gel pieces were dried using 100% acetonitrile (cover gel pieces) for 15 min, supernatant removed, and the pieces dried using a SpeedVac.
5. Dissolve trypsin (Promega) at 1:10 (trypsin: protein) ratio in the same volume (as pooled labeled sample) of 25 mM ammonium bicarbonate. For example, 600 µg protein sample with a volume of 500 µL needs 60 µg trypsin dissolved in 500 µL of 25 mM ammonium bicarbonate.
6. Add the trypsin solution to the gel pieces and set on ice to rehydrate. If pieces are not covered add 50 mM ammonium bicarbonate buffer. After rehydration, incubate at 37 °C for 12-16 hr.
7. Remove the liquid protein extract from the incubated samples.
8. Stop the reaction and remove any remaining protein digest by covering gel pieces with 5% formic acid, 50% acetonitrile.. Shake gel pieces for 20 min at room temperature. Transfer the supernatant to the extracted protein sample tubes from 5.7. Repeat the extraction procedure twice. Dry the extracted peptides using the SpeedVac.

6. Sample Enrichment of cystMT Labeled-peptides

1. Add a gradient of anti-TMT resin to 0.5 mL tubes for a 50% slurry. (Gradient was determined from band concentration in fractionation gel. If a fraction collection consists of a dark band, more resin is needed. Fractions with weaker bands require less resin as there is less protein.)
2. The resin is then washed three times with one column volume of 1x Tris-buffered saline (TBS) (25 mM Tris, 0.15 M NaCl, pH 7.2) (Thermo Scientific Pierce Protein Research Products).
3. Add 200 μ L 1xTBS to each sample. Samples are then added to anti-TMT resin (Thermo Scientific Pierce Protein Research Products) and agitated at room temperature for 2 hr followed by rocking overnight at 4 $^{\circ}$ C.
4. Add sample to column (Thermo Scientific Pierce Protein Research Products).
5. Wash each column three times with 1x 200 μ L TBS. This is then followed with washing three times with 0.05% CHAPs (dissolved in 1x TBS).
6. The column is then washed three times with 4 M urea in 1x TBS. Two hundred microliters of H₂O is then used to wash the column three times.
7. Each sample is eluted three times with 200 μ L 50% elution buffer (50% acetonitrile, 0.4% TFA).
8. Samples are then dried in a vacuum concentrator.

7. Mass Spectrometry Analysis

1. Resuspend the samples in 12 μ L of 3% acetonitrile with 0.1% formic acid and inject 5 μ L directly onto an Eksigent NanoLC-1D high pressure liquid chromatography column (AB SCIEX, USA).
2. Peptides will be separated on a Proteopec II C18 column 75 μ m ID x 20 cm (New Objective, USA) using a 4-60% gradient (A: 3% acetonitrile, 0.1% formic acid; B: 97% acetonitrile, 0.1% formic acid) at 3 μ L/min over 60 min.
3. A Thermo Scientific LTQ Orbitrap XL mass spectrometer was used to detect peptides using a top 2 x 3 experiment consisting of single stage MS followed by acquisition of 3 MS/MS spectra with higher-energy C-trap dissociation (HCD) fragmentation followed by 3 MS/MS with collision induced dissociation (CID) for protein identification. Parameters in this mode were: Isolation width: 3.0 m/z; Collision energy: 50% (10% with two steps). Only doubly- and triply-charged peptides were selected for fragmentation. Dynamic exclusion parameters were set to: Repeat count = 1; Repeat Duration = 60; Exclusion list size = 500; Exclusion duration = 28. Target values are as follows: MS = 5 e⁵; MS/MS (HCD) = 1 e⁵. Ion transfer times were set to 500 for FTMS and 300 for MS/MS (HCD). Two microscans were required for HCD spectra.

8. Database Searching and Quantitation

1. The acquired CID and HCD data were analyzed using Thermo Scientific Proteome Discoverer software 1.2 (Thermo Scientific Pierce Protein Research Products) through a branched workflow which implemented a Reporter Ion Quantizer (20 ppm mass tolerance of fragment ions) to quantify the ratio. A separate segment processed the MS² spectra through the Spectrum Selector, Spectrum Normalizer, and Spectrum Grouper nodes.
2. Data were then searched against the SEQUEST search engine. A 20 ppm mass tolerance was used. Static modifications were set to the cystMT reagent (304.18 Da) and dynamic modifications included phosphorylation and methionine oxidation (15.99 Da). A custom database for *Solanum lycopersicum* was composed using RNA data (with about 350,000 entries from Harvard University)¹³ was used in both cases.

9. Representative Results

A representative image of a control tomato plant leaf and a *Pseudomonas* inoculated leaf is shown in **Figure 1**. A difference between control treated and *Pseudomonas* treated leaves is observed. After the leaves are removed and stained using DAB, the de-staining process allows for the histochemical stain to show signs of ROS in the leaf tissue (**Figure 2**). **Figure 2A** is representative of a control leaf with no staining. **Figure 2B** is representative of a leaf treated with *Pseudomonas* and positive staining for H₂O₂ production. An example of Proteome Discover data output of a differentially redox regulated protein is shown in **Figure 3**. This protein is a known redox regulated protein ferredoxin-1¹⁴ and has been shown to play a role in defense against *Pseudomonas syringae* pv *tomato*¹⁵. Peak intensity between control and inoculated samples is used to obtain relative quantification, which showed significant changes in ferredoxin-1 redox regulation ($p < 0.05$). High intensity peaks suggest that this protein is oxidized in response to the pathogen treatment. **Figure 4** is an example of Proteome Discover data output of a protein that has similar redox regulation of a protein between a control and inoculated sample. Peaks of similar intensity suggest the presence of disulfide bonds not regulated by a change in treatment. The method will revolutionize how scientists detect redox responsive cysteines and disulfides¹⁰.

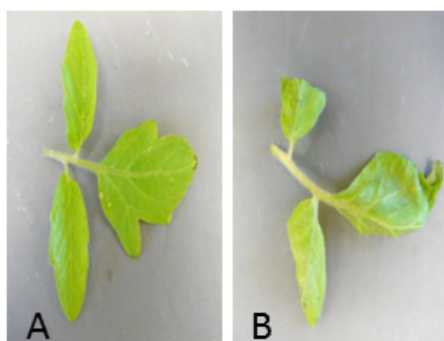


Figure 1. A representative image of inoculated tomato leaves with control solution (A) and *Pseudomonas* (B).

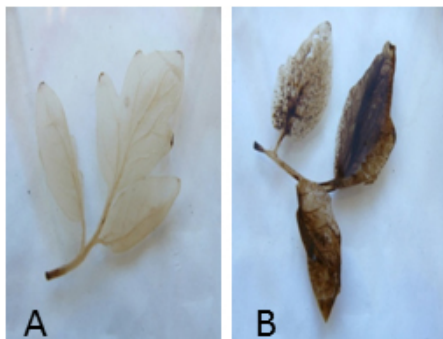


Figure 2. A representative image of DAB staining of inoculated tomato leaves with control solution (A) and *Pseudomonas* (B). Leaves were stained using 3'-3' diaminobenzidine. Chlorophyll was removed from leaves by boiling in 95% ethanol. Dark staining indicates the presence of H₂O₂. Only leaves inoculated with bacteria culture showed dark staining.

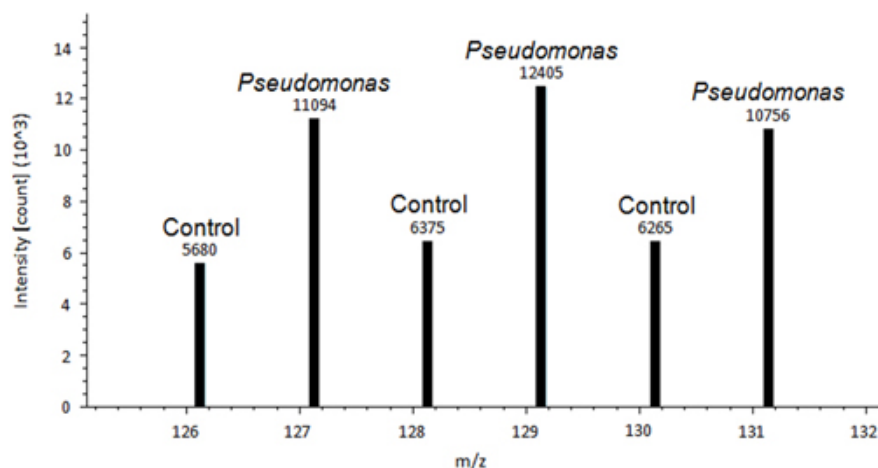


Figure 3. An example of Proteome Discover data output of ferredoxin-1, differentially redox regulated protein¹⁴. Peak intensity above each peak is used for absolute quantification. Peak intensity between control and inoculated samples is used to perform relative quantification.

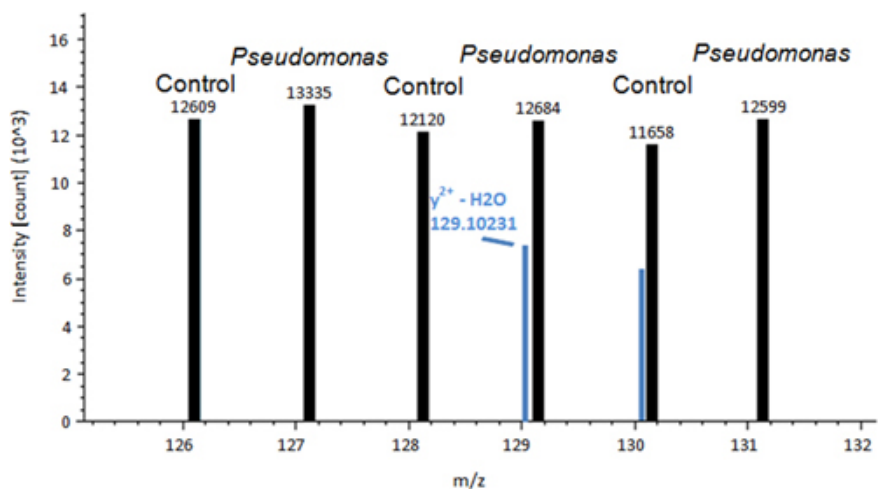


Figure 4. An example of Proteome Discover data output of a protein that has similar peak intensity between a control and inoculated sample. Peaks of similar intensity suggest the presence of disulfide bonds not regulated by a change in treatment.

Discussion

This protocol provides information on performing DAB staining as well as cystTMT labeled redox cysteine quantification. These procedures are beneficial in examining production of ROS as well as the effect on protein regulation when *Solanum lycopersicum* is inoculated with *Pseudomonas syringae*. The methods presented in this protocol provide a way to examine ROS in whole leaf samples in a way that causes the least amount of damage to leaf tissue. The labeling procedure provides a way to examine potentially redox regulated proteins by utilizing a cysteine labeling method. This is beneficial when examining an early stage of stress response.

Methods such as isotope-coded affinity tag (ICAT) and cystTMT can be used in examining potential redox regulated proteins in biological samples. ICAT allows labeling and comparison of two samples¹². Both methods label free cysteines and can be used for protein quantification^{10,12}. However, the cystTMT method allows for a decrease in experimental variation as well as multiplexing¹⁰. The number of tags available allows researchers to include replicates or multiple samples in their experimental design. Having more samples provides the potential for a higher number of proteins identified. A major disadvantage of the cystTMT technique is that it compromises the overall quality of protein identification because of the selective enrichment steps for cystTMT labeled-peptides (6.5-6.6). The number of peptides for the protein identification depends largely on the number of cysteine residues in the protein sequence. This problem can be overcome by submitting part of the tryptic sample before enrichment for mass spectrometry protein identification.

Due to the nature of the experimental design as well as the labeling mechanism that the cystTMT method utilizes, certain steps are critical. While performing protein precipitation and pellet washings (3.9) it is important to keep samples chilled on ice to reduce protein degradation. During cystTMT labeling, removal of the reducing reagent (4.6) is important because samples may undergo reverse labeling. Reverse labeling is possible if reducing reagent remains in the sample. If samples are reduced after labeling, the cystTMT tag can be removed. Once the labels are added to the samples, the pH level must be checked (4.7) in order to have optimal labeling efficiency. In addition, data analysis is dependent on what is required of the researcher and the ultimate goal in using the protocol. It is also dependent on the software being used as each software has different algorithms.

This experiment utilizes pathogen as an elicitor for enhanced production of reactive oxidative species in tomato; however, other redox regulated responses may be measured accordingly. This experimental design is adaptable to other plant and animal systems.

Disclosures

No conflicts of interest declared.

Acknowledgements

The authors would like to thank Dr. Greg Martin (Cornell University) and his group for providing the DC3000 strain, tomato seeds, and advice. They would also like to thank Dr. Zhonglin Mou for help with the DAB protocol and the Proteomics Division at UF Interdisciplinary Center for Biotechnology Research for assistance in the method development. The protocol for protein extraction was modified from Hurkman and Tanaka¹⁶. The protocol on cystTMT labeling, steps 4 to 6 was adapted based on the original Thermo Pierce Fisher Scientific product manual¹⁷. This work was funded by National Science Foundation (MCB 0818051 to S Chen).

References

- Almeida, N.F., et al. A draft genome sequence of *Pseudomonas syringae* pv. *tomato* T1 reveals a type III effector repertoire significantly divergent from that of *Pseudomonas syringae* pv. *tomato* DC3000. *Mol. Plant Microbe Interact.* **22**, 52-62 (2009).
- Preiter, K., et al. Novel virulence gene of *Pseudomonas syringae* pv. *tomato* strain DC3000. *J. Bacteriol.* **187**, 7805-7814 (2005).
- Apostol, I., Heinsteins, P.F., & Low, P.S. Rapid stimulation of an oxidative burst during elicitation of cultured plant cells: role in defense and signal transduction. *Plant Physiol.* **90**, 109-116 (1989).
- Orozco-Cardenas, M. & Ryan, C.A. Hydrogen peroxide is generated systemically in plant leaves by wounding and systemin via the octadecanoid pathway. *Proc. Natl. Acad. Sci. U.S.A.* **96**, 6553-6557 (1999).
- Tonks, N.K. Redox redux: revisiting PTPs and the control of cell signaling. *Cell.* **121**, 667-670 (2005).
- Alvarez, S., Wilson, G.H., & Chen, S. Determination of *in vivo* disulfide-bonded proteins in Arabidopsis. *J. Chromatogr. B.* **877**, 101-104 (2009).
- Apel, K., & Hirt, H. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.* **55**, 373-399 (2004).
- Dayon, L., et al. Relative quantification of proteins in human cerebrospinal fluids by MS/MS using 6-plex isobaric tags. *Anal. Chem.* **80**, 2921-2931 (2008).
- Thompson, A., et al. Tandem mass tags: a novel quantification strategy for comparative analysis of complex protein mixtures by MS/MS. *Anal. Chem.* **75**, 1895-1904 (2003).
- Rosenblatt, M., et al. A novel cysteine-reactive tandem mass tag reagent for subproteome labeling, enrichment and quantitation. The 58th ASMS Conference on Mass Spectrometry. *Thermo Fisher Scientific.*, TP169 (2010).
- Clark, J., et al. Phenotypic analysis of Arabidopsis mutants: diaminobenzidine stain for hydrogen peroxide. *Cold Spring Harb. Protoc.* doi: 10.1101/pdb.prot4981 (2009).
- Sethuraman, M., et al. Isotope-Coded affinity tag (ICAT) approach to redox proteomics: identification and quantification of oxidant-sensitive cysteine thiols in complex protein mixtures. *J. Proteome Res.* **3**, 1228-1233 (2004).
- DFCI- Tomato Gene Index [Internet]. Boston, (MA): Harvard University: Computational Biology and Functional Genomics Laboratory (US); Available from: <http://compbio.dfci.harvard.edu/cgi-bin/tgi/gimain.pl?gudb=tomato>, [updated April 13 2010].

14. Asso, M., *et al.* EPR and redox characterization of ferredoxins I and II from *Desulfovibrio vulgaris* Miyazaki. *Biochem. Biophys. Res. Commun.* **211**, 198-204 (1995).
15. Huang, He, *et al.* Disease resistance to bacterial pathogens affected by the amount of ferredoxin-I proteins in plants. *Mol. Plant Pathol.* **8**, 129-137 (2007).
16. Hurkman W. & Tanaka C. Solubilization of plant membrane proteins for analysis by two-dimensional gel electrophoresis. *Plant Physiol.* **81**, 802-806 (1986).
17. Thermo Scientific [Internet]. Rockford (IL): Thermo Scientific Pierce Biotechnology (US); Available from: <http://www.piercenet.com/instructions/2162220.pdf>, [updated May 31 2011].