



Draft Genome Sequence of *Mycobacterium interjectum* Strain ATCC 51457^T

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***Mycobacterium interjectum* is a nontuberculosis species rarely responsible for human infection. The draft genome of *M. interjectum* ATCC 51457^T comprises 5,927,979 bp, exhibiting 67.91% G+C content, 5,314 protein-coding genes, and 51 predicted RNA genes.**

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Mycobacterium *interjectum* was delineated as a new species of nontuberculous mycobacteria most closely related to *Mycobacterium simiae* (1, 2), *Mycobacterium saskatchewanense* sp. nov. (3), and *Mycobacterium paraense* sp. nov. (4). *M. interjectum* is an opportunistic pathogen mainly isolated from diseased lymph nodes (1, 5–8) and the respiratory tract (5, 9–11). A few other cases have caused meningoencephalitis (12) and cutaneous infection (13). Also, *M. interjectum* has been isolated from mammals (14–16), fish (17, 18), and clear water (19, 20). It is therefore of medical and general interest to further describe the genome of this species, and we performed whole-genome sequencing of the *M. interjectum* ATCC 51457^T strain. Genomic DNA isolated from *M. interjectum* grown in MGIT Middlebrook liquid culture (Becton, Dickinson, Le Pont-de-Claix, France) was sequenced in two Illumina MiSeq runs (Illumina, Inc., San Diego, CA) using a 5.9-kb mate-paired library. Reads were trimmed using Trimmomatic (21) and assembled using Velvet (version 1.2.03) (22). Contigs were combined by SSPACE version 2 (23), Opera version 2 (24) helped by GapFiller version 1.10 (25), and homemade tools in Python to refine the set. The *M. interjectum* strain ATCC 51457^T draft genome consists of 30 scaffolds and 221 contigs containing 5,927,979 bp and 67.91% G+C content. Noncoding genes and miscellaneous features were predicted using RNAmmer (26), ARAGORN (27), Rfam (28), PFAM (29), and Infernal (30). Coding DNA sequences (CDSs) were predicted using Prodigal (31), and functional annotation was achieved using BLAST+ (32) and HMMER3 (33) against the UniProtKB database (34). The genome was shown to encode at least 51 predicted RNAs, including three rRNAs and 48 tRNAs. A total of 5,314 identified genes yielded a coding capacity of 4,706,163 bp (coding percentage, 79.3%). Among these genes, 4,431 (83.38%) were found to be putative proteins, and 883 (16.6%) were assigned as hypothetical proteins. Moreover, 3,056 genes matched at least one sequence in the Clusters of Orthologous Groups database (35, 36) with BLASTP default parameters. *In silico* DNA-DNA hybridization (DDH) (37) was performed with 16 reference genomes selected on the basis of their 16S rRNA gene proximity with *M. interjectum*. The *M. interjectum* genome was locally aligned 2-by-2 using BLAT algorithm

(38, 39) against each one of the 16 selected genomes, and DDH values were estimated from a generalized linear model (40). The DDH was 27.8% ($\pm 2.43\%$) for *Mycobacterium smegmatis* mc²155, 26.70% ($\pm 2.42\%$) for *Mycobacterium ulcerans* Agy99, 25.9% ($\pm 2.41\%$) for *Mycobacterium avium* subsp. *paratuberculosis* K-10, *Mycobacterium intracellulare* ATCC 13950, and *Mycobacterium indicus pranii* MTCC 9506, 23.6% ($\pm 2.38\%$) for *Mycobacterium kansasii* ATCC 12478, 23.5% ($\pm 2.38\%$) for *Mycobacterium tuberculosis* H37Rv and *Mycobacterium bovis* AF2122/97, 22.4% ($\pm 2.36\%$) for *Mycobacterium marinum* M and *Mycobacterium liflandii* 128FXT, 20.9% ($\pm 2.33\%$) for *Mycobacterium chubuense* NBB4, 20.8% ($\pm 2.33\%$) for *Mycobacterium vanbaalenii* PYR-1, 20.5% ($\pm 2.32\%$) for *Mycobacterium gilvum* PYR-GCK, 20.4% ($\pm 2.32\%$) for *Mycobacterium leprae* TN and *Mycobacterium neaurum* VKM Ac-1815D, and 20.2% ($\pm 2.31\%$) for *Mycobacterium rhodesiae* NBB3.

Nucleotide sequence accession number. The *M. interjectum* strain ATCC 51457^T genome sequence has been deposited at EMBL under the accession no. [FJVQ00000000](#). The version described in this paper is the first version.

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REFERENCES

1. Springer B, Kirschner P, Rost-Meyer G, Schröder KH, Kroppenstedt RM, Böttger EC. 1993. *Mycobacterium interjectum*, a new species isolated from a patient with chronic lymphadenitis. *J Clin Microbiol* 31: 3083–3089.
2. De Zwaan R, van Ingen J, van Soolingen D. 2014. Utility of *rpoB* gene sequencing for identification of nontuberculous mycobacteria in the Netherlands. *J Clin Microbiol* 52:2544–2551. <http://dx.doi.org/10.1128/JCM.00233-14>.
3. Turenne CY, Thibert L, Williams K, Burdz TV, Cook VJ, Wolfe JN, Cockcroft DW, Kabani A. 2004. *Mycobacterium saskatchewanense* sp. nov., a novel slowly growing scotochromogenic species from human clinical isolates related to *Mycobacterium interjectum* and AccuProbe-positive for *Mycobacterium avium* complex. *Int J Syst Evol Microbiol* 54:659–667. <http://dx.doi.org/10.1099/ijss.0.02739-0>.

4. Fusco da Costa AR, Fedrizzi T, Lopes ML, Pecorari M, Oliveira da Costa WL, Giacobazzi E, da Costa Bahia JR, De Sanctis V, Batista Lima KV, Bertorelli R, Grottola A, Fabio A, Mariottini A, Ferretti P, Di Leva F, Fregni Serpini G, Tagliazucchi S, Rumpianesi F, Jousson O, Segata N, Tortoli E. 2015. Characterization of 17 strains belonging to the *Mycobacterium simiae* complex and description of *Mycobacterium paraense* sp. nov. Int J Syst Evol Microbiol 65:656–662. <http://dx.doi.org/10.1099/ij.s.0.068395.0>.
5. Lumb R, Goodwin A, Ratcliff R, Stapledon R, Holland A, Bastian I. 1997. Phenotypic and molecular characterization of three clinical isolates of *Mycobacterium interjectum*. J Clin Microbiol 35:2782–2785.
6. De Baere T, Moerman M, Rigouts L, Dhooge C, Vermeersch H, Verschraegen G, Vaneechoutte M. 2001. *Mycobacterium interjectum* as causative agent of cervical lymphadenitis. J Clin Microbiol 39:725–727. <http://dx.doi.org/10.1128/JCM.39.2.725-727.2001>.
7. Tuerlinckx D, Fauville-Dufaux M, Bodart E, Bogaerts P, Dupont B, Glupeczynski Y. 2010. Submandibular lymphadenitis caused by *Mycobacterium interjectum*: contribution of new diagnostic tools. Eur J Pediatr 169:505–508. <http://dx.doi.org/10.1007/s00431-009-1053-6>.
8. Rose M, Kitz R, Mischke A, Enzensberger R, Schneider V, Zielen S. 2004. Lymphadenitis cervicalis due to *Mycobacterium interjectum* in immunocompetent children. Acta Paediatr 93:424–426. <http://dx.doi.org/10.1111/j.1651-2227.2004.tb02976.x>.
9. Martínez Lacasa J, Cuchi E, Font R. 2009. *Mycobacterium interjectum* as a cause of lung disease mimicking tuberculosis. Int J Tuberc Lung Dis 13:1048.
10. Mirant-Borde MC, Alvarez S, Johnson MM. 2013. *Mycobacterium interjectum* lung infection. Case Rep Pulmonol 2013:193830. <http://dx.doi.org/10.1155/2013/193830>.
11. Villanueva MA, Gullón JA, Álvarez-Navascués F. 2015. *Mycobacterium interjectum* lung infection: a case report. Arch Bronconeumol 51:155–156. <http://dx.doi.org/10.1016/j.arbr.2014.12.016>.
12. O'Dwyer JP, O'Connor JG, McDermott H, Sheehan S, Fanning NF, Corcoran GD, Sweeney B. 2009. Meningoencephalitis associated with non-tuberculous mycobacteria. BMJ Case Rep. bcr03:2009.1696.
13. Fukuoka M, Matsumura Y, Kore-edo S, Iinuma Y, Miyachi Y. 2008. Cutaneous infection due to *Mycobacterium interjectum* in an immunosuppressed patient with microscopic polyangiitis. Br J Dermatol 159: 1382–1384. <http://dx.doi.org/10.1111/j.1365-2133.2008.08867.x>.
14. De Lisle GW, Kawakami RP, Yates GF, Collins DM. 2008. Isolation of *Mycobacterium bovis* and other mycobacterial species from ferrets and stoats. Vet Microbiol 132:402–407. <http://dx.doi.org/10.1016/j.vetmic.2008.05.022>.
15. Zanetti S, Bua A, Molicotti P, Delogu G, Mura A, Ortù S, Sechi LA. 2008. Identification of mycobacterial infections in wild boars in Northern Sardinia, Italy. Acta Vet Hung 56:145–152. <http://dx.doi.org/10.1556/AVet.56.2008.2.1>.
16. Bouts T, Vordermeier M, Flach E, Routh A. 2009. Positive skin and serologic test results of diagnostic assays for bovine tuberculosis and subsequent isolation of *Mycobacterium interjectum* in a pygmy hippopotamus (*Hexaprotodon liberiensis*). J Zoo Wildl Med 40:536–542. <http://dx.doi.org/10.1638/2008-0001.1>.
17. Rhodes MW, Kator H, Kaattari I, Gauthier D, Vogelbein W, Ottlinger CA. 2004. Isolation and characterization of mycobacteria from striped bass *Morone saxatilis* from the Chesapeake Bay. Dis Aquat Organ 61: 41–51. <http://dx.doi.org/10.3354/dao061041>.
18. Zanoni RG, Florio D, Fioravanti ML, Rossi M, Prearo M. 2008. Occurrence of *Mycobacterium* spp. in ornamental fish in Italy. J Fish Dis 31: 433–441. <http://dx.doi.org/10.1111/j.1365-2761.2008.00924.x>.
19. Thomson RM, Carter R, Tolson C, Coulter C, Huygens F, Hargreaves M. 2013. Factors associated with the isolation of nontuberculous mycobacteria (NTM) from a large municipal water system in Brisbane, Australia. BMC Microbiol 13:89. <http://dx.doi.org/10.1186/1471-2180-13-89>.
20. Makovcova J, Slany M, Babak V, Slana I, Kralik P. 2014. The water environment as a source of potentially pathogenic mycobacteria. J Water Health 12:254–263. <http://dx.doi.org/10.2166/wh.2013.102>.
21. Lohse M, Bolger AM, Nagel A, Fernie AR, Lunn JE, Stitt M, Usadel B. 2012. RobiNA: a user-friendly, integrated software solution for RNA-Seq-based transcriptomics. Nucleic Acids Res 40:W622–W627. <http://dx.doi.org/10.1093/nar/gks540>.
22. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
23. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. Bioinformatics 27: 578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
24. Gao S, Sung WK, Nagarajan N. 2011. Opera: reconstructing optimal genomic scaffolds with high-throughput paired-end sequences. J Comput Biol 18:1681–1691. <http://dx.doi.org/10.1089/cmb.2011.0170>.
25. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with Gap-Filler. Genome Biol 13:R56. <http://dx.doi.org/10.1186/gb-2012-13-6-r56>.
26. Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
27. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. <http://dx.doi.org/10.1093/nar/gkh152>.
28. Griffiths-Jones S, Bateman A, Marshall M, Khanna A, Eddy SR. 2003. Rfam: an RNA family database. Nucleic Acids Res 31:439–441. <http://dx.doi.org/10.1093/nar/gkg006>.
29. Punta M, Coggill PC, Eberhardt RY, Mistry J, Tate J, Boursnell C, Pang N, Forslund K, Ceric G, Clements J, Heger A, Holm L, Sonnhammer EL, Eddy SR, Bateman A, Finn RD. 2012. The Pfam protein families database. Nucleic Acids Res 40:D290–D301. <http://dx.doi.org/10.1093/nar/gkr1065>.
30. Nawrocki EP, Kolbe DL, Eddy SR. 2009. Infernal 1.0: inference of RNA alignments. Bioinformatics 25:1335–1337. <http://dx.doi.org/10.1093/bioinformatics/btp157>.
31. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
32. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. <http://dx.doi.org/10.1186/1471-2105-10-421>.
33. Eddy SR. 2011. Accelerated profile HMM searches. PLoS Comput Biol 7:e1002195. <http://dx.doi.org/10.1371/journal.pcbi.1002195>.
34. The UniProt Consortium. 2011. Ongoing and future developments at the universal protein resource. Nucleic Acids Res 39:D214–D219. <http://dx.doi.org/10.1093/nar/gkq1020>.
35. Tatusov RL, Galperin MY, Natale DA, Koonin EV. 2000. The COG database: a tool for genome-scale analysis of protein functions and evolution. Nucleic Acids Res 28:33–36. <http://dx.doi.org/10.1093/nar/28.1.33>.
36. Tatusov RL, Koonin EV, Lipman DJ. 1997. A genomic perspective on protein families. Science 278:631–637. <http://dx.doi.org/10.1126/science.278.5338.631>.
37. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci USA 106: 19126–19131. <http://dx.doi.org/10.1073/pnas.0906412106>.
38. Kent WJ. 2002. BLAT—the blast-like alignment tool. Genome Res 12: 656–664. <http://dx.doi.org/10.1101/gr.229202>. Article published online before March 2002.
39. Auch AF, Von Jan M, Klenk HP, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. Stand Genomic Sci 2:117–134. <http://dx.doi.org/10.4056/sigs.531120>.
40. Meier-Kolthoff JP, Auch AF, Klenk HP, Göker M. 2013. Genome sequence based species delimitation with confidence intervals and improved distance functions. BMC Bioinformatics 14:60. <http://dx.doi.org/10.1186/1471-2105-14-60>.