

Intestinal transport mechanism and in vivo anticancer efficacy of a solid oral formulation incorporating an ion-pairing complex of pemetrexed with deoxycholic acid derivative

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Objective: The rational combination of immunotherapy with standard chemotherapy shows synergistic clinical activities in cancer treatment. In the present study, an oral powder formulation of pemetrexed (PMX) was developed to enhance intestinal membrane permeability and investigate its application in metronomic chemotherapy in combination with immunotherapy.

Methods: PMX was ionically complexed with a bile acid derivative (N^α-deoxycholyll-lysyl-methylester; DCK) as a permeation enhancer and mixed with dispersing agents, such as poloxamer 188 (P188) and Labrasol, to form an amorphous oral powder formulation of PMX/DCK (PMX/DCK-OP).

Results: The apparent permeability (P_{app}) of PMX/DCK-OP across a Caco-2 cell monolayer was 2.46- and 8.26-fold greater than that of PMX/DCK and free PMX, respectively, which may have been due to the specific interaction of DCK with bile acid transporters, as well as the alteration of membrane fluidity due to Labrasol and P188. Furthermore, inhibition of bile acid transporters by actinomycin D in Caco-2 cell monolayers decreased the P_{app} of PMX/DCK-OP by 75.4%, suggesting a predominant role of bile acid transporters in the intestinal absorption of PMX/DCK-OP. In addition, caveola/lipid raft-dependent endocytosis, macropinocytosis, passive diffusion, and paracellular transport mechanisms significantly influenced the permeation of PMX/DCK-OP through the intestinal membrane. Therefore, the oral bioavailability of PMX/DCK-OP in rats was 19.8%±6.93%, which was 294% higher than that of oral PMX. Moreover, an in vivo anticancer efficacy study in B16F10 cell-bearing mice treated with a combination of oral PMX/DCK-OP and intraperitoneal anti-PD1 exhibited significant suppression of tumor growth, and the tumor volume was maximally inhibited by 2.03- and 3.16-fold compared to the oral PMX/DCK-OP and control groups, respectively.

Conclusion: These findings indicated the therapeutic potential of a combination of low-dose oral chemotherapy and immunotherapy for synergistic anticancer efficacy.

Keywords: pemetrexed, bile acid derivative, oral powder formulation, intestinal membrane permeability, oral delivery, oral metronomic chemotherapy

Introduction

Intravenous administration of the maximum tolerated dose of an antineoplastic chemotherapeutic agent has been widely accepted as a clinical strategy to obtain complete remission or cure of cancer. However, this strategy is associated with the development of chemoresistance and both long- and short-term toxicity, including

myelosuppression, neutropenia, thrombocytopenia, and gastrointestinal dysfunction, along with the depletion of both innate and adaptive defense mechanisms. To address these issues related to conventional chemotherapy, a novel approach involving frequent low-dose oral administration, also known as metronomic chemotherapy, has been suggested, which maintains uniform serum levels of drugs, overcomes drug resistance by targeting the tumor vasculature, and avoids toxic effects.¹ Moreover, metronomic chemotherapy inhibits tumor angiogenesis, promotes activation of innate and adaptive immunity, and induces tumor dormancy.² However, metronomic scheduling for improved patient quality of life can be preferentially achieved by oral drug administration. In addition, oral administration is considered one of the safest and most convenient routes to maintaining prolonged exposure of cancerous cells to chemotherapeutic agents. However, oral delivery of these anticancer drugs is a major challenge due to their physicochemical properties, variable rate of degradation in gastrointestinal fluid, and high drug efflux through P-glycoprotein (P-gp).^{3,4} Furthermore, recent studies suggested the use of low-dose chemotherapeutics in combination with immunotherapy to overcome the immunosuppression induced by cancer, intensify host immunity to destroy tumor cells, produce highly active tumor-specific T cells, and synergistically optimize anti-tumor effects.^{5,6} Therefore, the rational combination of immunotherapy with oral low-dose chemotherapy is expected to control tumor growth and address the immunosuppressive tumor environment.

Pemetrexed (N-[4-[2-(2-amino-3,4-dihydro-4-oxo-7H-pyrrolo[2,3-d] pyrimidin-5-yl)-ethyl]-benzoyl]-L-glutamic acid; PMX) is a novel multi-targeted antineoplastic agent that disrupts metabolic processes related to the *de novo* biosynthesis of thymidine and purine nucleotides. Clinically, PMX in combination with cisplatin has been approved by the US Food and Drug Administration as the first-line treatment of patients with non-squamous and non-small cell lung cancer (NSCLC), as well as to treat relapsed or refractory NLCLC after chemotherapy with platinum-based regimens. Moreover, treatment of advanced NSCLC with a combination of PMX and immunotherapeutic agents, such as bevacizumab, has been recommended as maintenance therapy, where both drugs are highly effective, well tolerated, and exhibit low toxicity.⁷ PMX is a polar-charged compound with a relatively small distribution volume and rapid clearance.

Moreover, PMX exhibits low oral bioavailability, which may be due to its poor intestinal absorption.^{8,9}

To overcome these obstacles, various approaches, including coadministration of therapeutic agents with functional excipients (P-gp inhibitors) or use of permeation enhancers, have been developed.^{10–12} In addition, polymeric or lipid-based drug carriers, such as polymeric nanoparticles, polymeric micelles, microemulsions, self-emulsifying drug delivery systems, lipid-drug conjugates, and nanocrystals have been used to improve the oral bioavailability of drugs.^{13,14}

There is increasing interest in the design of potential prodrug targets for carrier-mediated uptake via bile acid transporters to improve intestinal drug absorption.^{15,16} The prodrug approach includes chemical modification of drugs to improve their charge and lipophilicity, to in turn promote intestinal permeation. Prodrugs targeting endogenous transporters present on enterocytes can facilitate site-specific transport and represent a novel strategy for intestinal delivery of drugs with very low oral bioavailability.¹⁵ Various approaches to exploit the prodrug concept have already been applied to increase the central nervous system (CNS) delivery of several drugs. Bonina et al reported greater reserpine-induced hypolocomotion effects in rats when composites were linked via the C-3 glucose position.¹⁷ Bilsky et al demonstrated increased uptake of opioid peptide agonist through the blood-brain barrier (BBB) after conjugation with D-glucose.¹⁸ In both cases, the prodrug was targeted to glucose transporter (GLUT) expressed in the BBB. Similarly, terminal ileum of the intestine is the major site of bile acid reabsorption in humans, and bile acid is actively transported to the terminal ileum by the well-characterized apical sodium-dependent bile acid transporter (ASBT).¹⁹ The use of prodrugs to target ASBT and increase intestinal drug permeability is widely practiced. For example, acyclovir showed an unambiguous increase in oral bioavailability via a prodrug targeting ASBT, ie, acyclovir valylchenodeoxycholate.²⁰

In our previous studies, we prepared an ionic complex of hydrophilic drugs with a bile acid derivative (N^α-deoxycholyll-L-lysyl-methylester; DCK) as a permeation enhancer.^{16,21,22} DCK was prepared by chemical linkage of deoxycholic acid (DOCA) with positively charged L-lysine, which showed increased intestinal permeability of hydrophilic drugs, such as bisphosphonates, insulin, and oxaliplatin. The ion-paired complex is known to be less stable than the chemically conjugated prodrug form, and a major concern when utilizing the ion-pairing approach is the possibility of premature complex

dissociation in the biological fluids prior to absorption.²³ However, chemical modification by attaching a functional molecule to the drug may result in loss of biological activity; new tests to assess safety and efficacy are required in accordance with the development of novel drug therapies.²⁴ Furthermore, the ion-paired DCK can more effectively deliver specific target molecules through the intestinal membrane via ASBT-mediated endocytosis and/or an increase in concentration gradients through the intestinal membrane compared to other non-specific and physically mixed absorption enhancers, such as surfactants.²⁵ Recently, we also prepared an ion-pairing complex of PMX and DCK in combination with dispersants, such as 2-hydroxypropyl- β -cyclodextrin (HP- β -CD) and poloxamer 188 (P188), forming the HP- β -CD/PMX/DCK/P188 complex. Furthermore, we incorporated the ion-pairing complex into the inner aqueous phase of multiple water-in-oil-in-water nanoemulsions in a supersaturated state. Both the HP- β -CD/PMX/DCK/P188 complex and its nanoemulsion showed significant increases in the oral bioavailability of PMX, which may have been due to increased lipophilicity of the complex caused by DCK, disruption of the intestinal barrier function by DOCA, and surfactant-induced membrane fluidity. However, incorporation of HP- β -CD/PMX/DCK/P188 complex into the aqueous phase of the nanoemulsions resulted in stability problems during long-term storage. The oral powder formulation of PMX effectively enhanced oral bioavailability and drug stability.

The main objective of the present study was to design an oral powder formulation of PMX for application in metronomic chemotherapy, as well as for combination use with immunotherapy. PMX was ionically complexed with DCK in combination with a dispersing agent, such as P188 or Labrasol, forming an oral powder formulation of PMX/DCK (PMX/DCK-OP). This can be effectively absorbed across the intestinal membrane due to increased lipophilicity of PMX achieved by ion-pairing complex with DCK and interaction with bile acid transporters. In addition, stable formation of micelles in the aqueous solution, dispersant activity of P188, opening of tight junctions (TJs), and surfactant activity of Labrasol can also enhance the intestinal membrane permeability of PMX. Therefore, to confirm the increase in oral absorption of PMX after preparation of an oral powder formulation, we determined the artificial membrane and Caco-2 cell monolayer permeability of PMX/DCK and PMX/DCK-OP. The intestinal transport pathways of PMX/DCK-OP through a Caco-2 cell monolayer were then assessed using specific pharmacological inhibitors. Finally, the oral bioavailability of

PMX/DCK-OP in rats, and its antitumor efficacy in combination with intraperitoneal anti-PD1 antibody in tumor-bearing mice, was evaluated.

Materials and methods

Materials

The PMX disodium hemipentahydrate was obtained from Shipla Medicare (Karnataka, India). Caprylocaproyl macrogol-8-glyceride (Labrasol) was provided by Gattefossé (Saint Priest, France). Polyoxyethylene (160) polyoxypropylene (30) glycol (P188) was purchased from BASF (Ludwigshafen, Germany). Chlorpromazine, methyl- β -cyclodextrin (M β CD), brefeldin A, genistein, actinomycin D (Act D), cyclosporine A, ethylene glycol-bis-(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA), DOCA, and 4-[N-(2,4-diamino-6-pteridinylmethyl)-N-methylamino]benzoic acid hemihydrochloride hydrate (DAMPA) were obtained from Sigma-Aldrich Inc. (St. Louis, MO). Anti-mouse PD-1 antibody (anti-PD1) was purchased from Bio X Cell (West Lebanon, NH). Solvents for high-performance liquid chromatography (HPLC) were obtained from Merck Millipore (Billerica, MA) and Thermo Fisher Scientific Inc. (Waltham, MA).

Animals

Sprague-Dawley rats (males, 6–7 weeks old, 200–250 g) and BALB/c mice (females, 6–7 weeks old, 20–25 g) were provided by Orient Bio (Gwangju, Republic of Korea). The animals were housed under environmentally controlled conditions of temperature (23 °C \pm 2 °C), relative humidity (55% \pm 10%), and light (12/12 hr light/dark cycle). The animals were fed a standard laboratory diet (Nestlé Purina, St. Louis, MO) and ion-sterilized tap water. Ethical approval for this study was obtained from the Institutional Animal Care and Use Committee (IACUC) of Mokpo National University (Jeonnam, Republic of Korea). All animal experiments were performed in accordance with the NIH Guidelines for the Care and Use of Laboratory Animals and the guidelines of the IACUC.

Preparation and characterization of PMX/DCK-OP

We synthesized DCK as an oral permeation enhancer, as described previously.¹⁶ Briefly, H-Lys(Boc)-OMe-HCl (20 g) dissolved in *N*-methyl morpholine (7.4 mL) and ethyl chloroformate (6.4 mL) were mixed with DOCA (26 g) in tetrahydrofuran (800 mL) with continuous stirring for 30 mins, followed by 2 hr reflux. The precipitates

obtained after stirring of the reaction mixture overnight at room temperature were collected by filtration and the residual solvents in the precipitates were removed by evaporation. Purified Lys(Boc)DOCA was obtained by column chromatography (chloroform/methanol) of the dried precipitates, which was further dissolved in a mixture of acetyl chloride and methanol in an ice bath. After complete removal of solvent, the residue was dissolved in water and washed three times with chloroform. Then, the aqueous layer was collected and freeze-dried to obtain DCK powder. Next, ion-pairing complex of PMX with DCK (PMX/DCK) was prepared by dissolving 50 mg of PMX in 10 mL of deionized water. Then, 10 mL of DCK solution in deionized water (6.68 mg/mL) was prepared and added dropwise into the PMX solution at a molar ratio of 1:1, with vortex mixing after each addition. The solution mixture of PMX and DCK was freeze-dried at -70°C to obtain PMX/DCK complex powder.

To enhance the intestinal membrane permeability of PMX, PMX/DCK-OP was prepared using the solid dispersion technique. Briefly, PMX (50 mg) was dissolved in P188 solution prepared by dissolving 500 mg of P188 in 10 mL of deionized water containing 375 μL of Labrasol. DCK solution was prepared in another tube by dissolving 66.8 mg of DCK in 8 mL of deionized water. At a molar ratio of 1:1, DCK solution was added dropwise into the PMX solution with regular vortex mixing. In addition, 80 μL of NaOH (1 M) was added with thorough mixing to maintain the PMX/DCK mixture at pH 7. The mixture was then freeze-dried at -70°C to completely remove the water and obtain PMX/DCK-OP in powder form. Separately, to explore the role of DCK in enhancing the intestinal membrane permeability of PMX, an oral powder formulation of PMX containing Labrasol and P188 (PMX-OP) was prepared. Briefly, 50 mg PMX was dissolved in 10 mL of P188 solution (50 mg/mL) containing 375 μL of Labrasol. The mixture was then well vortexed for 5 min and freeze-dried at -70°C to completely remove water and obtain PMX-OP in powder form. Furthermore, to confirm micelle formation of PMX/DCK-OP in the aqueous phase, the powder formulation was dispersed in deionized water (1:20, w/v) and its droplet size, polydispersity index (PDI), and zeta potential were measured with a dynamic laser light scattering (DLS) analyzer (Zetasizer Nano ZS90; Malvern Instruments, Malvern UK) at 25°C . The morphology and particle size of PMX/DCK-OP micelles were further determined using transmission electron microscopy (TEM). After negative staining of the

PMX/DCK-OP dispersed in deionized water (1:100, w/v) with 2% (w/v) phosphotungstic acid solution, the sample solution was dropped on a copper grid and examined using high-resolution transmission electron microscopy (HR-TEM; JEM-200; JEOL, Tokyo, Japan).

In vitro artificial intestinal membrane permeability

The in vitro artificial membrane permeability of free PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP was determined using the parallel artificial membrane permeability assay. Sample solution (200 μL) at a concentration equivalent to 200 $\mu\text{g}/\text{mL}$ of PMX was added to each well of the donor plates, which were prepared by dilution of free PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP with phosphate-buffered saline (PBS, pH 6.8). We added 300 μL of PBS (pH 6.8) to each well of the receptor plates. After 5 hr incubation at room temperature, the plates were disassembled and samples were withdrawn from both the donor and acceptor plates. The concentration of PMX that permeated through the artificial membrane was measured by HPLC at 254 nm with a C18 column (4.6 \times 250 mm, 5 μm , 100 \AA) at 25°C . The mobile phase consisted of water (pH 3.5 adjusted with phosphoric acid)-acetonitrile (80:20, v/v) and the flow rate was optimized to 1 mL/min.

The effective permeability (P_e) of each sample was calculated using the formula:

$$P_e = -\ln\left[1 - \frac{C_R(t)}{C_{\text{equilibrium}}}\right] / \left[S \times \left(\frac{1}{V_D} + \frac{1}{V_R}\right) \times t\right] \quad (1)$$

where P_e is the permeability (cm/s), S is the effective filter area (0.288 cm^2), V_D is the volume of the donor well (0.2 mL), V_R is the volume of the receptor well (0.3 mL), t is the total time of incubation in seconds, $C_R(t)$ denotes the concentration of the drug in the receptor well at time t , and $C_{\text{equilibrium}}$ represents $[C_D(t) \times V_D + C_R(t) \times V_R] / (V_D + V_R)$, where $C_D(t)$ denotes the concentration of the drug in the donor well at time t .¹⁶

In vitro permeability across a Caco-2 monolayer

To determine the apparent permeability (P_{app}) of PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP, Caco-2 cell monolayers were formed by seeding Caco-2 cells (ATCC[®] HTB-37[™]; American Type Culture Collection, Manassas, VA, USA), at a density of 1×10^5 cells/well, on polystyrene Transwell membranes in Dulbecco's modified

Eagle's medium (DMEM) containing 10% (v/v) fetal bovine serum (FBS) and 1% penicillin/streptomycin, followed by culture for 14 to 16 days. Media in the apical and basolateral compartments were replaced every other day until the transepithelial electrical resistance (TEER) reached $>350 \Omega \cdot \text{cm}^2$. To investigate transcellular and ASBT-mediated transport, the culture medium in both the apical and basolateral compartments was removed and replaced with prewarmed HBSS. HBSS in the apical compartment was then replaced with 0.1 mL of PMX, PMX/DCK, PMX-OP, or PMX/DCK-OP diluted in HBSS (equivalent to 100 $\mu\text{g}/\text{mL}$ PMX), and that in the basolateral compartment was replaced with 0.6 mL of fresh HBSS followed by incubation of the plates at 37 °C. Samples (0.1 mL) from each basolateral compartment were withdrawn at predetermined time points of 0, 0.5, 1, 2, 3, 4, and 5 hrs and replaced with the same volume of fresh HBSS. The collected samples were then filtered through polyvinylidene difluoride (PVDF) membrane filters (0.45 μm), and the concentrations of PMX in the samples were determined by HPLC with a UV detector at 254 nm, as described above.

The P_{app} for PMX in various formulations was calculated as follows:

$$P_{\text{app}} = dQ/dt \times 1/(S \times C_i) \quad (2)$$

where dQ/dt represents the linear appearance rate of mass on the basolateral side ($\mu\text{g}/\text{s}$), S indicates the surface area of the monolayer (cm^2), and C_i is the initial concentration of PMX on the apical side ($\mu\text{g}/\text{mL}$).²⁶

Intestinal transport mechanism of PMX/DCK-OP

Next, we examined the role of ASBT in the permeation of PMX/DCK-OP, as well as the endocytosis pathways of PMX/DCK-OP micelles in the aqueous phase. Before performing the

intestinal transport study, to determine the appropriate Act D concentration, the in vitro cytotoxicity assay results of Act D using Caco-2 cells were evaluated using the Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Rockville, MD, USA). Caco-2 cells were seeded at a density of 1×10^4 cells/well in 200 μL DMEM with 10% FBS in 96-well plates and cultured for 24 h. The cells were then treated with serially diluted sample solutions of 1, 2, 3.2, and 6 μM Act D in HBSS for 5 hrs. To evaluate the cytotoxic effect of Act D on the Caco-2 cells, a 10- μL WST-8 (2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium monosodium salt) solution was added to each well and incubated for 1 hr. The absorbance was then measured using a microplate reader (Multimode Plate Reader; PerkinElmer Inc., Waltham, MA, USA) at 450 nm. The results obtained for the treated cells are expressed as the percentage of viable cells compared to untreated cells. After determining the Act D concentration showing no toxic effect, Caco-2 cells were cultured on Transwell membranes and the medium in the apical and basolateral compartments was replaced every other day until the TEER value of the Caco-2 cell monolayer reached $>350 \Omega \cdot \text{cm}^2$, to establish the in vitro transcellular model. To investigate the possible transcellular pathways of PMX/DCK-OP transport, 0.1 mL of HBSS containing inhibitors of the specific cellular uptake pathway (Table 1) and 0.6 mL of HBSS were then added to the apical and basolateral compartments of each well, respectively. After pre-incubation at 37 °C for 30 mins, the solution in the apical compartment was replaced with 0.1 mL of PMX/DCK-OP diluted in HBSS (equivalent to 100 $\mu\text{g}/\text{mL}$ PMX) with corresponding inhibitors, and that in the basolateral compartment was replaced with 0.6 mL of fresh HBSS followed by incubation at 37 °C. Then, 200 μL of sample solution was withdrawn at 0, 0.5, 1, 2, 3, 4, and 5 hrs from the basolateral compartment of each well and replaced with the same volume of fresh HBSS.^{26,27}

In addition, to evaluate the effects of TJ opening on the transport of PMX/DCK-OP, the extracellular Ca^{+2}

Table 1 Inhibitors with different functions used in the transport study and their concentrations

Inhibitors	Concentrations	Functions
Act D	3.2 μM	Inhibitor of ASBT-mediated transport
Chlorpromazine	32 μM	Inhibitor of clathrin-mediated endocytosis
M β CD	10 mM	Inhibitor of caveola/lipid raft-mediated endocytosis (cholesterol depletion agent)
Genistein	0.1 mM	Inhibitor of caveola/lipid raft-mediated endocytosis (broad inhibitor of protein tyrosine kinase)
Amiloride	0.1 mM	Inhibitor of macropinocytosis
Cyclosporine A	10 mM	Inhibitor of P-gp mediated efflux
Brefeldin A	90 μM	Inhibitor of ER/Golgi pathway

Abbreviations: Act D, actinomycin D; M β CD, methyl- β -cyclodextrin; ASBT, apical sodium-dependent bile acid transporter; P-gp, P-glycoprotein; ER, endoplasmic reticulum.

chelating agent, EGTA, was applied to reversibly open the intracellular TJs in the Caco-2 cell monolayer. Moreover, to examine the effects of EGTA in combination with all inhibitors except cyclosporine A on the transport of PMX/DCK-OP, a Caco-2 cell monolayer with TEER value $>350 \Omega \cdot \text{cm}^2$ was treated with 0.1 mL of HBSS (Ca^{+2} -free medium) containing 2.5 mM EGTA with or without all inhibitors except cyclosporine A. After filling the basolateral side with 0.6 mL of HBSS (Ca^{+2} -free medium), the Transwell membranes were preincubated at 37 °C for 45 mins and the integrity of the cell monolayer was monitored by measuring the TEER value as $\leq 70 \Omega \cdot \text{cm}^2$, to confirm opening of the junctional complex. The medium in the apical compartment was then replaced with 0.1 mL HBSS (with 1.8 mM Ca^{+2}) containing PMX/DCK-OP (equivalent to 100 $\mu\text{g}/\text{mL}$ PMX), alone or with all inhibitors except cyclosporine A, and 0.6 mL of HBSS (with 1.8 mM Ca^{+2}) was added to the basolateral side of each well, followed by incubation at 37 °C for a further 5 hrs. At predetermined time points, 0.2 mL of medium on the basolateral side was withdrawn and replaced with the same volume of fresh HBSS. In addition, the integrity of the monolayer was checked by measuring the TEER value, which returned to $\geq 300 \Omega \cdot \text{cm}^2$ at 2 and 5 hrs after drug loading, respectively, to identify restoration of the transmembrane resistance.²⁸

All collected samples were then filtered through PVDF membrane filters (0.45 μm) and the concentration of PMX that permeated across the Caco-2 cell monolayer was determined by HPLC at 254 nm as described above. Finally, the cumulative permeated amount of PMX from the PMX/DCK-OP was plotted as a function of time and P_{app} was determined from the linear slope of the plot using Equation (2).

In vivo pharmacokinetic study in rats

To evaluate the beneficial effects of ion-pairing complex formation of PMX with DCK on intestinal absorption, 400 μL of PMX in water (25 mg/kg), PMX/DCK in water (25 mg/kg PMX), or PMX/DCK-OP (25 mg/kg PMX) was orally administered to rats. In addition, 150 μL of an aqueous solution of PMX (5 mg/kg) was injected via the tail vein to evaluate the oral bioavailability. Next, blood samples of 150 μL were collected from the retroorbital plexus at predetermined time intervals into heparinized tubes, mixed with 50 μL of 3.8% sodium citrate solution, and immediately centrifuged (2,500 \times g, 15 mins, 4 °C). The plasma samples were separated and

kept frozen at $-70 \text{ }^\circ\text{C}$ until analysis. Liquid chromatography/mass spectrometry (LC/MS) was performed to determine the concentration of PMX in the plasma, as described previously.²⁹ Briefly, 300 μL of 2% NH_4OH in deionized water and 100 μL of standard solution were added to the plasma samples that were defrosted and centrifuged at 2,500 \times g for 5 mins at 4 °C, followed by vortex mixing. Next, 10 μL of DAMPA (5 $\mu\text{g}/\text{mL}$) was added to each sample as an internal standard, vortex-mixed for 2 mins, and subjected to solid-phase extraction using a Plexa Bond Elut PAX cartridge (30 mg, 1 mL; Agilent Technologies, Santa Clara, CA) in accordance with the manufacturer's instructions. The cartridges were conditioned with 500 μL of methanol followed by 500 μL of deionized water, and standards and samples were loaded into the cartridges. After the unadsorbed samples and impurities in the cartridges were washed with 500 μL each of deionized water and methanol, elution of the adsorbed samples was performed with 2 \times 250 μL of 5% formic acid prepared in methanol. The eluate was dried using a centrifugal evaporator (Genevac Ltd., Ipswich, UK), which was then reconstituted with 100 μL of 5% formic acid in methanol. The concentration of PMX in the reconstituted samples was determined using an Agilent 6120 Quadrupole LC/MS system with a Phenomenex Luna C18 column (100 \times 2 mm, 3 μm) as the stationary phase and acetonitrile:0.34% formic acid (15:85, v/v) as the mobile phase, at a flow rate of 0.2 mL/min. PMX and internal standard were ionized using an electrospray ionization (ESI) source in positive ion mode under the following source conditions: capillary voltage of 3.5 kV; drying gas flow rate of 3.1 L/min; and drying gas temperature of 300 °C. Quantitative analysis of protonated molecular ions was performed at ($[\text{M} + \text{H}]^+ = 428$) and ($[\text{M} + \text{H}]^+ = 326.1$) for PMX and DAMPA, respectively.

In vivo tumor growth inhibition efficacy in mice

The antitumor effects of orally administered PMX/DCK-OP, alone or in combination with anti-PD1 administered intraperitoneally, were investigated in BALB/c mice subcutaneously inoculated with B16F10 cells at a density of 1×10^6 cells (in 100 μL of PBS, pH 7.4). Mice with an average tumor volume of 40–60 mm^3 were randomly divided into five groups (10 mice per group): control (untreated), PMX-IV (twice-a-week intravenous administration of 20 mg/kg PMX), Anti-PD1 (once per 3 days intraperitoneal administration of 10 mg/kg

anti-PD1), PMX/DCK-OP (once-a-day oral administration equivalent to 20 mg/kg PMX), and combination of PMX/DCK-OP and Anti-PD1 (once-a-day oral administration equivalent to 20 mg/kg PMX and once per 3 days intraperitoneal administration of 10 mg/kg anti-PD1). The body weight and tumor volumes of mice in all five groups were recorded every 2 days. Tumor volumes in each mouse were calculated based on the following formula: $0.52 \times w^2 \times l$, where w and l represent the width and length of the tumor, respectively. Two weeks after oral administration, mice were sacrificed and tumor masses were measured. Induction of apoptosis in the isolated tumor tissues was detected by terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay. Moreover, microvessel expression and cell proliferation were analyzed using epithelial cell marker CD31 and proliferating cell nuclear antigen (PCNA), respectively.

Pharmacokinetic and statistical analyses

Pharmacokinetic parameters were assessed using a non-compartmental method in WinNonlin[®] software (ver. 5.3; Pharsight Corporation, Mountain View, CA, USA). One-way analysis of variance (ANOVA) followed by Tukey's

multiple-comparison test was used to compare more than two mean values. All data were expressed as means \pm standard deviation. In all analyses, a P -value less than 0.05 was considered statistically significant.

Results and discussion

Preparation and characterization of PMX/DCK-OP

To enhance the lipophilicity and intestinal membrane permeability of PMX, ion-pairing complex and oral powder formulations of PMX and DCK were prepared as shown in Figure 1A. The optimal PMX/DCK-OP diluted in deionized water showed a particle size and PDI of 134 ± 0.098 nm and 0.114 ± 0.005 , respectively, with a zeta potential of 16.3 ± 1.00 mV. The uniformly narrow particle size and high zeta potential supports the stable formation of micelles in the aqueous phase. In addition, TEM images of PMX/DCK-OP dispersed in water confirmed the formation of spherical and homogenous nano-sized micelles with diameters < 100 nm, which was smaller than the size observed using a DLS analyzer (Figure 1B). The

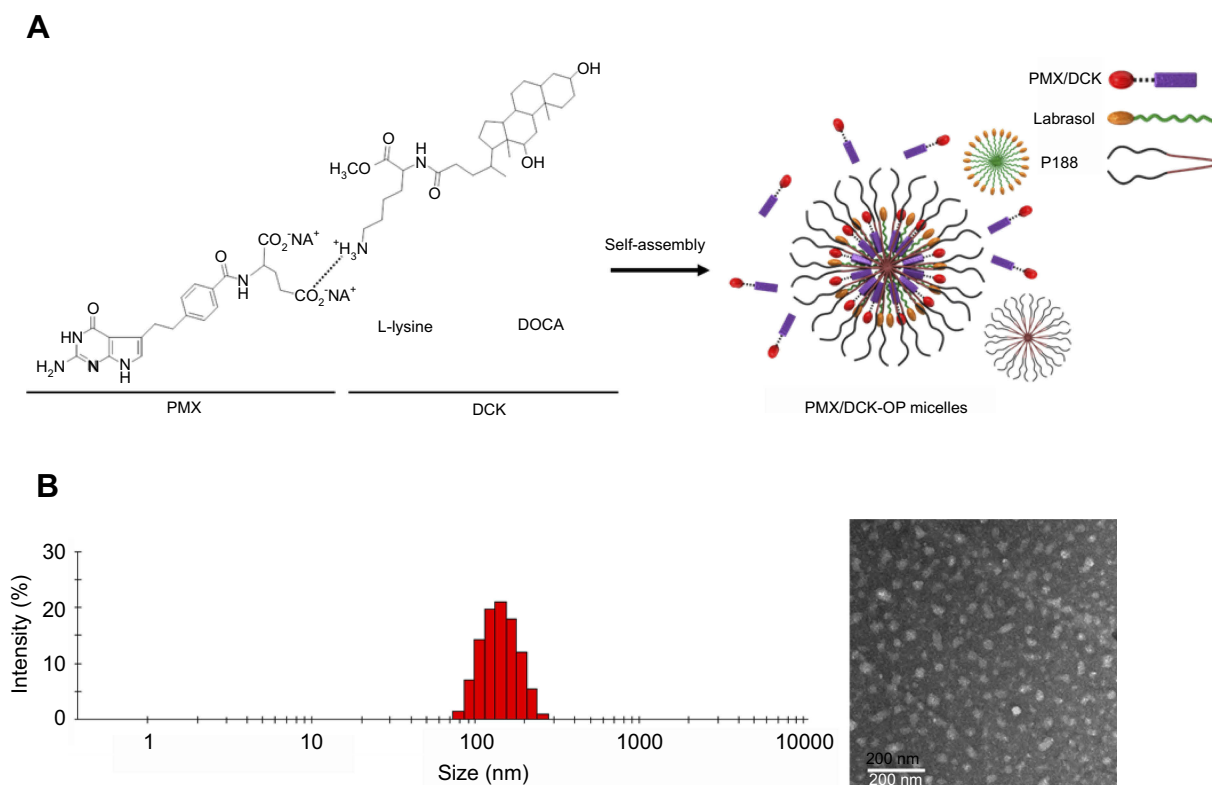


Figure 1 (A) Schematic illustration of ion-pairing complex formation between PMX and DCK, and self-assembled micelle formation of PMX/DCK-OP in the aqueous phase. (B) Transmission electron micrograph of PMX/DCK-OP micelles. Scale bar represents 200 nm.

Abbreviations: PMX, pemetrexed; DCK, N^α-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK.

difference in particle size of the PMX-DCK-OP micelles may be caused by the difference in sample preparation between DLS and TEM measurements. Unlike DLS, a sample for TEM is prepared in a dried state and the loosely aligned micelles can shrink during drying. The presence of Labrasol and P188 as surfactant and dispersing agent, respectively, in the optimum powder formulation were expected to increase the long-term stability and oral bioavailability of PMX. Moreover, Labrasol showed a hydrophilic-lipophilic balance (HLB) value of 14, was less toxic with high tolerance, and enhanced the intestinal absorption of hydrophilic drugs.^{30,31} In addition, use of P188 in the formulation resulted in self-assembly into micelles in the presence of aqueous phase at concentrations above the critical micellar concentration (CMC). Structurally, P188, which is an amphiphilic triblock copolymer of poly(ethylene oxide) and poly(propylene oxide), consists of a hydrophobic central core and hydrophilic outer core that can accommodate lipophilic and water soluble drugs, respectively. Furthermore, P188 has been widely used as a dispersant and solubilizing agent, and is approved for oral drug delivery.^{32,33}

In vitro permeability study of PMX/DCK-OP

The artificial intestinal membrane permeability of PMX was significantly increased after ion-pairing complex formation with DCK and formulation of PMX/DCK-OP. The P_e of PMX after ionic complex formation with DCK was increased by 36.2- and 7.29-fold compared to free PMX and PMX-OP, respectively (Table 2). The enhanced permeability of PMX/DCK may be due to the increased lipophilicity of PMX after complex formation with the oral permeation enhancer, DCK.^{34,35} On the other hand, the permeability of PMX was improved 4.97-fold compared to that of free PMX after formulation with P188 and Labrasol. However, the P_e of PMX/DCK-OP was increased by 3.87-, 28.2-, and 140-fold compared to

PMX/DCK, PMX-OP, and PMX, respectively. Therefore, the enhanced artificial membrane permeability of PMX/DCK-OP could be ascribed to the synergistic action of DCK, Labrasol, and P188 in the membrane permeation of PMX. Moreover, the presence of Labrasol and P188 in the oral powder formulation may have further increased the amphiphilicity of PMX, thus facilitating passive diffusion.

The in vitro permeability data of PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP across Caco-2 cell monolayers are presented in Table 2. The P_{app} of PMX after ion-pairing complex formation with DCK was 3.35- and 1.73-fold higher than those of free PMX and PMX-OP, respectively. The improved permeability of PMX may have been due to the increased membrane flexibility and opening of transient TJs caused by binding of bile acids to the calcium channels, which enhanced paracellular drug transport.³⁶ Moreover, DOCA induces phosphorylation of epidermal growth factor receptor, occludin dephosphorylation, and occludin redistribution, which modulates intestinal membrane permeability.³⁷ In addition, transcellular transport of hydrophilic drugs could be enhanced by the incorporation of bile acids into the cell membrane; this forms reverse micelles and results in the creation of aqueous channels, thereby increasing membrane permeability.³⁸ The permeability of PMX across a Caco-2 cell monolayer was only improved 1.94-fold by adding P188 and Labrasol, while the P_{app} of PMX/DCK-OP was further increased by 2.46-, 4.26-, and 8.26-fold compared to those of PMX/DCK, PMX-OP, and free PMX, respectively. This increase in the P_{app} of PMX/DCK-OP may have been due to the formation of PMX/DCK micelles in combination with Labrasol and P188, which could in turn increase drug absorption through endocytosis and micropinocytosis.³⁹ In addition, nonionic surfactants, such as Labrasol and P188, can modify intestinal membrane fluidity, followed by alteration of the epithelial barrier function.⁴⁰ Labrasol has been

Table 2 Effective and apparent permeability of PMX, PMX/DCK, and PMX/DCK-OP

Test material	Effective permeability (P_e , $\times 10^{-6}$, cm/s) ^a	Apparent permeability (P_{app} , $\times 10^{-6}$, cm/s) ^b
PMX	0.290±0.080	1.78±0.118
PMX/DCK	10.5±1.97***	5.97±1.38*
PMX-OP	1.44±0.180	3.45±0.531
PMX/DCK-OP	40.6±1.48***,####,\$\$\$	14.7±3.98***,####,\$\$\$

Notes: Statistics: one-way ANOVA followed by Tukey's multiple-comparison test. ^aEffective permeability (P_e) of PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP through an artificial intestinal membrane. ^bApparent permeability (P_{app}) of PMX, PMX/DCK, PMX-OP, and PMX/DCK-OP across a Caco-2 cell monolayer. Each value represents the mean \pm standard deviation (n=6). * $P<0.05$, *** $P<0.001$ compared to PMX; #### $P<0.001$ compared to PMX/DCK; \$\$\$ $P<0.001$ compared to PMX-OP.

Abbreviations: PMX, pemetrexed; DCK, N^α-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX-OP, oral powder formulation of PMX; PMX/DCK-OP, oral powder formulation of PMX/DCK.

shown to act as a permeation enhancer by disrupting TJs, causing redistribution of zonula occludens-1 (ZO-1).⁴¹ Therefore, the synergistic activity of DCK, Labrasol, and P188 may have significantly increased the P_{app} of PMX/DCK-OP. However, further permeability studies using gastrointestinal tissues are required to determine the stability and association constant of PMX/DCK and its oral formulation in the gastric and intestinal fluids. In addition, future studies should investigate the effects of ion-pairing complex formation with DCK and formulation with Labrasol and P188 on the intestinal permeability of PMX, and the absorption-enhancing mechanism of PMX/DCK-OP.

Intestinal transport mechanism of PMX/DCK-OP

To determine the transport pathway involved in the active transcellular transport of PMX/DCK-OP, Caco-2 monolayers were pretreated with various biochemical inhibitors of different pathways (Figure 2). In the aqueous phase, PMX/DCK-OP may exist as PMX/DCK, which can be further incorporated into micelles together with P188 and Labrasol, as confirmed in the preparation and characterization section. The main goal of ionic complex formation of PMX with DCK was to increase the lipophilic properties and oral bioavailability of PMX via the bile acid transporter-mediated approach. The transporter specificity, affinity, and capacity are the three major factors taken into consideration in prodrug design. Bile acids are mostly reclaimed in the distal ileum by ASBT.⁴² Moreover, ASBT exhibits high capacity and affinity for native bile acid, suggesting that ASBT may be a useful prodrug target.¹⁵ Coupling of PMX with DCK can increase the uptake of PMX through ASBT-mediated transport. To verify the involvement of ASBT in the uptake of PMX/DCK, we inhibited ASBT using Act D at a concentration that had no cytotoxic effect on Caco-2 cells; the cell viability was greater than 95% and the concentration ranged from 1 to 6 μM Act D. The cell viability at 3.2 μM Act D was $105 \pm 6.81\%$ (Figure S1).⁴³ As a result, the P_{app} of PMX/DCK-OP was markedly decreased, by 75.4%, compared to the control (without inhibitors) (Figure 2A). This observation was suggestive of active involvement of ASBT in the uptake of PMX/DCK-OP. After interacting with ASBT, PMX/DCK from PMX/DCK-OP may have followed a pathway similar to that of bile acid: entry into the enterocytes; binding with intestinal bile acid-binding protein (IBABP); shuttling across the enterocytes to the basolateral membrane; and

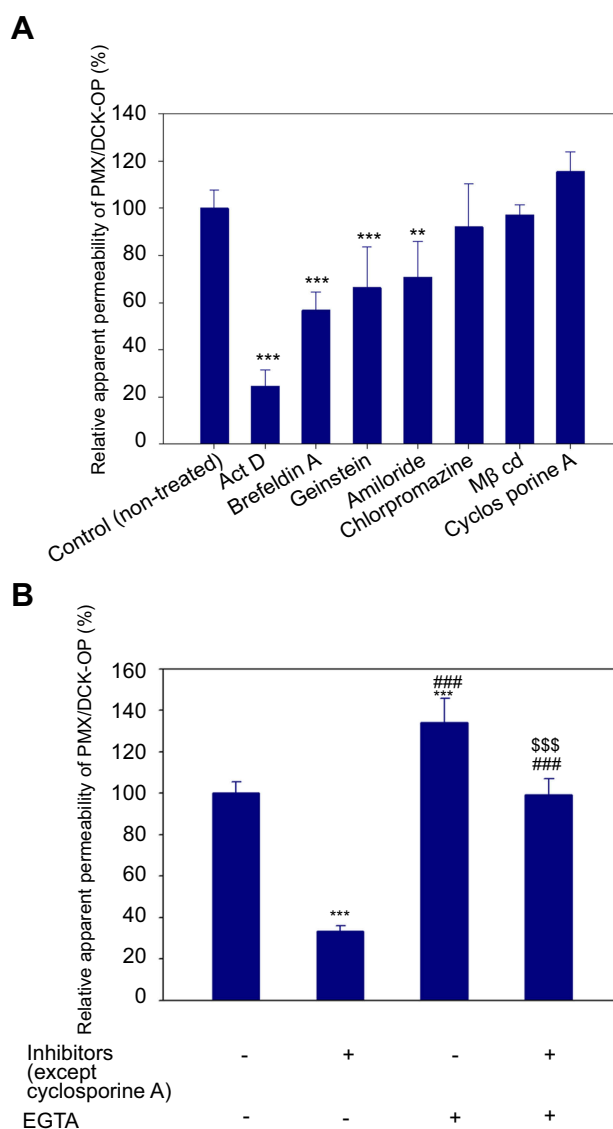


Figure 2 (A) Relative apparent permeability of PMX/DCK-OP across a Caco-2 cell monolayer after incubation with various transport inhibitors and **(B)** with or without EGTA in the presence or absence of all inhibitors except cyclosporine A.

Notes: Each value represents the mean \pm standard deviation ($n=6$). ** $P<0.01$, *** $P<0.001$ compared to the P_{app} of the untreated control group. #### $P<0.001$ compared to the apparent permeability of PMX/DCK-OP after treatment with all inhibitors except cyclosporine A in the absence of EGTA. \$\$\$ $P<0.001$ compared to the P_{app} of PMX/DCK-OP after treatment with EGTA in the presence of all inhibitors except cyclosporine A.

Abbreviations: PMX, pemetrexed; DCK, N⁶-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK; Act D, actinomycin D; M β CD, methyl- β -cyclodextrin; ASBT; EGTA, ethylene glycol-bis-(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid; P_{app} , apparent permeability of PMX/DCK-OP across a Caco-2 cell monolayer.

effusing into the portal circulation via the recently identified heteromeric transporters, organic solute transporter alpha and beta (OST_{α} and OST_{β}) (Figure 3).^{19,44}

In addition to the ASBT-mediated transport, PMX/DCK-OP micelles themselves can be internalized by endocytosis, because they are the major route for nanoparticle transport across the intestinal epithelium by engulfment in membrane invaginations forming intracellular vesicles.

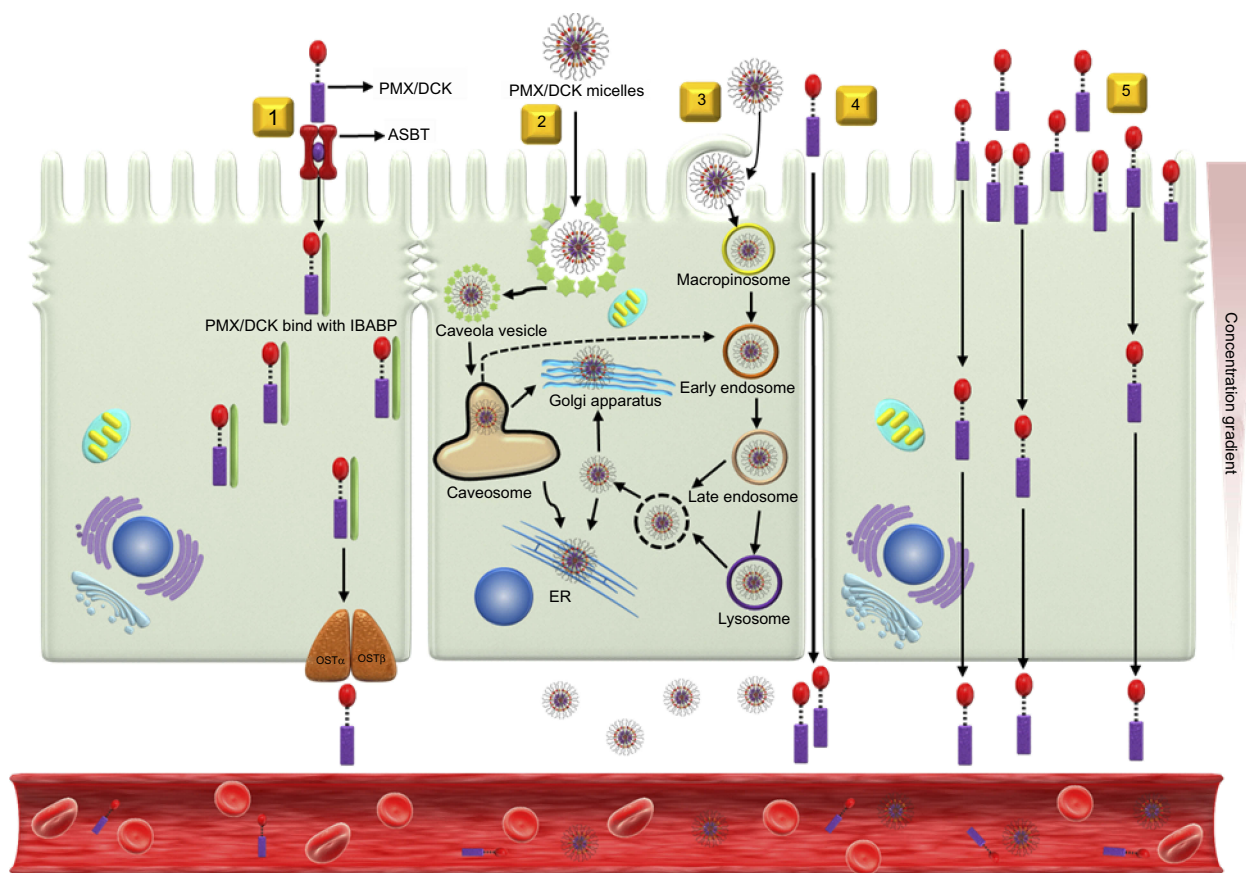


Figure 3 Schematic diagrams of possible specific and nonspecific transport pathways, and predicted intracellular trafficking and transcytosis of PMX/DCK-OP across an intestinal epithelium: (1) ASBT-mediated transport; (2) caveola/lipid raft-mediated endocytosis; (3) macropinosity; (4) paracellular transport; and (5) transcellular passive diffusion.

Abbreviations: PMX, pemetrexed; DCK, N^α-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK; ASBT, apical sodium-dependent bile acid transporter; IBABP, intestinal bile acid-binding protein; OST_α, organic solute transporter alpha; OST_β, organic solute transporter beta; ER, endoplasmic reticulum.

First, we used inhibitors of both clathrin- and caveola/lipid raft-mediated endocytosis to explore their roles in the uptake of PMX/DCK-OP. After treatment with chlorpromazine, an inhibitor of clathrin-mediated endocytosis, the P_{app} of PMX/DCK-OP was only reduced by 7.95% in comparison to the control (without inhibitors), indicating the clathrin-mediated endocytosis plays a trivial role in the uptake of PMX/DCK-OP micelles (Figure 2A). On the other hand, genistein, a caveola/lipid raft-mediated endocytosis inhibitor, significantly decreased the P_{app} by 33.6% compared to the control (without inhibitors) (Figure 2A). Genistein is a commonly used broad-spectrum inhibitor of protein tyrosine kinase (PTK), which catalyzes tyrosine phosphorylation and activates multiple signal transduction pathways, including endocytosis regulation.⁴⁵ Genistein has also been reported to be closely related with the actin network and recruitment of dynamin II, both of which are indispensable for caveola/lipid raft-mediated uptake.⁴⁶ Therefore,

the considerable decline in P_{app} in the presence of genistein confirmed the involvement of the caveola/lipid raft-mediated pathway in the uptake of PMX/DCK-OP. However, there was no significant difference in the P_{app} after treatment of Caco-2 cells with M β CD, a caveola/lipid raft-mediated endocytosis inhibitor (2.37% decrease in P_{app} of PMX/DCK-OP), which may be due to the difference in its mode of action. M β CD generally acts as a cholesterol-depleting agent and effectively inhibits caveola/lipid raft-mediated endocytosis.⁴⁷ However, it was reported previously that the addition of cholesterol together with M β CD may exhaust its function due to its specific interaction with cholesterol, leading to a decrease in the inhibitory efficacy of M β CD.²⁷ Therefore, in the present study, the inhibitory activity of M β CD may have decreased due to the presence of DOCA, a cholesterol derivative, in the DCK molecule. Macropinosity is another well-defined uptake mechanism that may be involved in the

uptake of PMX/DCK-OP micelles. During this process, a large volume of fluid consisting of nanoparticles was engulfed by ruffling the plasma membrane to form cargo-loaded macropinosomes.⁴⁸ Here, the inhibitor of macropinocytosis, amiloride, significantly decreased the uptake of PMX/DCK-OP (by 29.0%) compared to the control (without inhibitors), revealing the active role of macropinocytosis in the absorption of PMX/DCK-OP by the enterocytes (Figure 2A).

After entry into the cell, nanoparticles generally (i) are either captured by endosomes and consequently degraded by the lysosome, (ii) escape into the cytoplasm, or (iii) undergo recycling. In addition, nanoparticles can be localized to various organelles via endocytic vesicles and the complex intracellular trafficking machinery.⁴⁹ Therefore, during the processes of endocytosis and macropinocytosis, PMX/DCK-OP micelles may be engulfed in membrane invaginations forming caveosomes and macropinosomes, and then delivered to various organelles, such as the early endosome, late endosome, and lysosome, after which they escape to the endoplasmic reticulum (ER) and Golgi apparatus, and enter the enterocytes.^{49,50} During this process, endosomal escape may be crucial for the intracellular delivery of PMX/DCK-OP micelles because of the digestive enzymes in the lysosome. The PMX/DCK-OP micelles escaping from the endosomes can enter the cytoplasm or exit the enterocytes via the ER and Golgi apparatus (Figure 3). However, the cytoplasmic route is usually not preferred because most drugs must enter the blood circulation to exert their pharmacological action. In addition, the nanomaterials with the required intracellular delivery density can utilize ER and Golgi routes, as well as the retrograde trafficking pathway.

It has been reported that both the Golgi complex and ER are important regulators of the secretory ER/Golgi pathway, as well as the endocytic recycling pathway.⁵¹ Therefore, to clarify whether the ER-to-Golgi apparatus is involved in the intracellular trafficking of PMX/DCK-OP micelles, we examined the effects of brefeldin A, a specific inhibitor of the secretory pathway from the ER to the Golgi apparatus,⁵² on the permeation of PMX/DCK-OP. The results indicated that the P_{app} of PMX/DCK-OP was significantly decreased (by 43.2%) compared to the control (without inhibitors) after treatment of the cells with brefeldin A, suggesting that the ER and Golgi apparatus are actively involved in the exocytosis of PMX/DCK-OP micelles (Figure 2A). Furthermore, the involvement of the ER-to-Golgi apparatus in the intracellular transport of

PMX/DCK-OP micelles clarified the roles of endocytosis and macropinocytosis; lysosomal escape of PMX/DCK-OP micelles was also observed (Figure 3).

P-gp is responsible for the multidrug resistance phenotype of cancer, by mediating the ATP-dependent efflux of anticancer drugs out of the cell.⁵³ To examine the effects of P-gp-mediated efflux on the transport of PMX/DCK-OP, we used cyclosporine A as an inhibitor of P-gp.^{54,55} The P_{app} of PMX/DCK-OP was increased by 16.0% compared to the control (without inhibitors), but the difference was not statistically significant (Figure 2A). Therefore, P-gp-mediated efflux does not have a significant influence on PMX/DCK-OP uptake.

The oral formulation can also facilitate passive diffusion and paracellular drug transport. To understand the involvement of passive diffusion and paracellular transport in the uptake of PMX/DCK-OP, we examined the effects of EGTA on the P_{app} of PMX/DCK-OP. EGTA is a selective calcium chelator, which is used to bind free extracellular Ca^{+2} ions and can reversibly open the intercellular TJs of the Caco-2 monolayer.²⁸ As shown in Figure 2B, when the Caco-2 monolayer (without EGTA) was treated with all inhibitors except cyclosporine A, the P_{app} of PMX/DCK-OP decreased by 66.7% compared to the control (without inhibitors and EGTA), indicating that 33.3% of PMX/DCK-OP was permeated by passive diffusion and paracellular transport (Figure 2B). As described above, the passive diffusion of PMX through the PAMPA membrane was markedly improved after ion-pairing complex formation with DCK, followed by formulation with Labrasol and P188. Furthermore, the P_{app} of PMX/DCK-OP across the EGTA-treated Caco-2 monolayer (without inhibitors) increased by 34.0% in comparison to the control (without inhibitors and EGTA), suggesting the importance of opening the TJs to increase the uptake of PMX/DCK-OP (Figure 2B). On the other hand, the P_{app} of EGTA-treated Caco-2 monolayer (with all inhibitors except cyclosporine A) was similar to the control (without inhibitors and EGTA) but increased by 66.0% compared to the EGTA-untreated Caco-2 monolayer (with all inhibitors except cyclosporine A). This observation also supported a significant role of EGTA in increasing the transport of PMX/DCK-OP across the Caco-2 monolayer via reversibly opening the intracellular TJs. Therefore, it is clear that reversible opening of intracellular TJs can increase the transport of PMX/DCK-OP, and that the incorporation of components capable of opening TJs in the formulation of PMX/DCK-OP can enhance the intestinal permeation of

PMX (Figure 3). The presence of Labrasol and bile acid, which open TJs, in PMX/DCK-OP may have favored the permeation of PMX/DCK-OP through the paracellular route.^{41,56,57}

In vivo oral absorption in rats

The pharmacokinetic parameters in rats after intravenous administration of PMX, and oral administration of PMX, PMX/DCK, and PMX/DCK-OP, are shown in Figure 4A and B, respectively. The maximum plasma concentration (C_{max}) and area under the plasma concentration-time curve (AUC) achieved after oral administration of PMX/DCK

(equivalent to 25 mg/kg PMX) were $4.58 \pm 4.63 \mu\text{g/mL}$ and $4.13 \pm 2.29 \mu\text{g}\cdot\text{h/mL}$, respectively, which were 3.57- and 1.76-fold higher than those of oral PMX alone, suggesting significant improvement of intestinal absorption of PMX and resulting in 75.3% greater oral bioavailability of PMX/DCK than PMX (Table 3). The improvement of oral bioavailability was supported by the results of the mechanistic study, which demonstrated significant reduction of P_{app} after inhibition of ASBT by Act D, suggesting active involvement of bile acid transporter in the uptake of PMX/DCK. Moreover, the significant increases in in vitro artificial membrane and Caco-2 cell monolayer permeability of PMX/DCK compared to PMX confirmed the major role of paracellular and transcellular transport in the enhancement of oral bioavailability. In addition, DOCA induced disruption of intestinal barrier function by dephosphorylation of occludin, and cytoskeletal rearrangement of epithelial TJs may have enhanced the oral absorption of PMX/DCK.^{37,56–58} Furthermore, the C_{max} and AUC_{last} values for PMX/DCK-OP were 1.29- and 2.25-fold higher than those of PMX/DCK, respectively. Therefore, the oral bioavailability of PMX/DCK-OP was 294% higher than that of PMX solution. The time to reach the maximum concentration (T_{max}) was 0.63 ± 0.25 and 0.50 ± 0.00 hrs after oral administration of PMX and PMX/DCK-OP, respectively, suggesting the immediate dissolution of PMX/DCK from the powder formulation, followed by rapid absorption of PMX/DCK or PMX/DCK-OP micelles across the intestinal membrane. The enhanced oral absorption of PMX in PMX/DCK-OP proceeds as follows: (i) uptake of free PMX/DCK by the bile acid transporter, which was confirmed by inhibition of bile acid transporter; and (ii) significant decrease in P_{app} after inhibition of caveola/lipid raft-mediated endocytosis by genistein, suggesting internalization of PMX/DCK-OP micelles by endocytosis; 3) formation of PMX/DCK-OP macropinosomes that entered the enterocytes (which also revealed the prominent role of macropinocytosis in enhanced oral absorption); and 4) PMX/DCK-OP uptake via paracellular transport and passive diffusion, which was confirmed by reversible opening of TJs by EGTA and in vitro artificial membrane permeability.

In vivo tumor growth inhibitory effect of orally administered PMX/DCK-OP

Based on the above findings, we used the B16F10 cell-bearing mouse tumor model to investigate the efficacy of combined oral administration of PMX/DCK-OP with intraperitoneal anti-PD1 in terms of inhibiting tumor progression. Compared to the untreated control group,

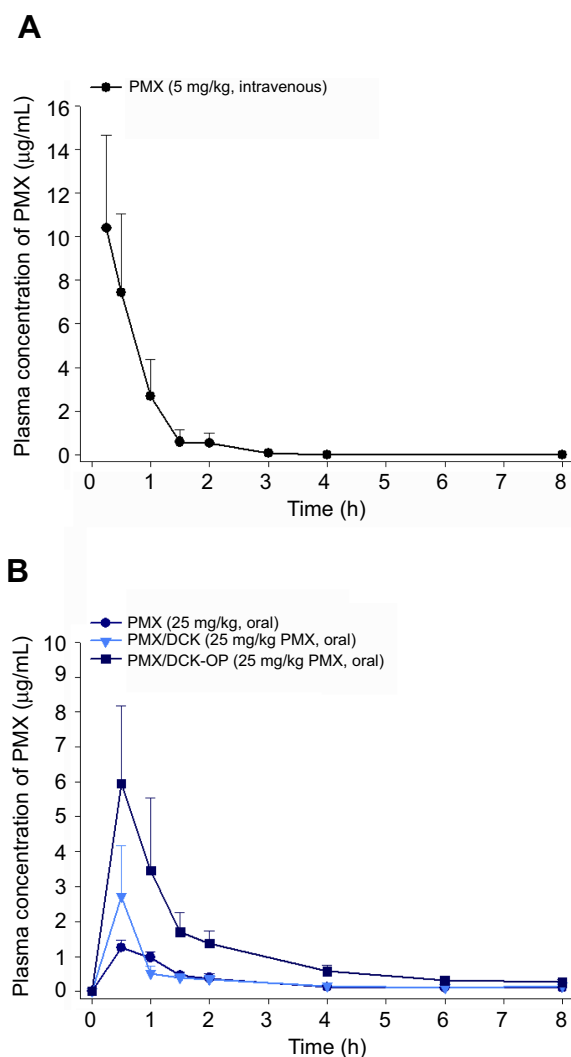


Figure 4 Venous plasma concentration-time profiles of PMX after (A) single intravenous administration of PMX (5 mg/kg) and (B) oral administration of PMX (25 mg/kg), PMX/DCK (25 mg/kg PMX, oral), or PMX/DCK-OP (equivalent to 25 mg/kg of PMX) in aqueous solution to rats.

Notes: Each value represents the mean \pm standard deviation ($n=4$ for each group).

Abbreviations: PMX, pemetrexed; DCK, N^{α} -deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK.

Table 3 Pharmacokinetic parameters of PMX in rats after intravenous injection of PMX and oral administration of PMX, PMX/DCK, or PMX/DCK-OP

Test material	PMX	PMX	PMX/DCK	PMX/DCK-OP
Administration	Intravenous	Oral	Oral	Oral
Dose of PMX (mg/kg)	5	25	25	25
T _{max} (h)	-	0.63±0.25	0.50±0.00	0.50±0.00
T _{1/2} (h)	0.35±0.08	2.50±0.45	4.05±2.02	2.47±0.37
C _{max} (µg/mL)	15.3±6.10	1.28±0.17	4.58±2.63	5.95±3.23
AUC _{last} (µg/mL)	9.38±4.17	2.35±0.27	4.13±1.29	9.27±3.25 ^{***#}
AUC _{inf} (µg/mL)	9.41±4.22	2.77±0.21	4.90±1.73	10.2±2.94 ^{***#}
Bioavailability (%)		5.02±0.57	8.80±2.89	19.8±6.93 ^{***#}

Notes: Statistics: one-way ANOVA followed by Tukey's multiple-comparison test. Each value represents the mean ± standard deviation (n=4). Bioavailability (%), (AUC_{last, oral}/Dose_{PMX, oral})/(AUC_{last, intravenous}/Dose_{PMX, intravenous})×100. ^{**}P<0.01, ^{***}P<0.001 compared to PMX; [#]P<0.05 compared to PMX/DCK.

Abbreviations: PMX, pemetrexed; DCK, N⁶-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK; T_{max}, time to reach maximum plasma concentration; T_{1/2}, half-life of plasma concentration; C_{max}, maximum plasma concentration; AUC_{last}, area under the plasma concentration-time curve from zero to the time of the last measurable plasma concentration; AUC_{inf}, area under the plasma concentration-time curve from zero to infinity.

the tumor growth rates were significantly delayed in mice treated with PMX/DCK-OP or PMX-IV alone (maximally suppressed by 35.5% and 38.4%, respectively; [Figure 5A](#)). The improved tumor growth inhibition efficacy may have been due to the metronomic scheduling of PMX/DCK-OP and PMX-IV, which can activate both innate and adaptive immunity, induce tumor dormancy, suppress tumor angiogenesis, and render tumor cells more sensitive to T cell-mediated lysis.¹ However, PMX/DCK-OP showed similar antitumor efficacy as PMX-IV, although its bioavailability was approximately 20% that of PMX-IV. This may be due to the dosing schedule of PMX/DCK-OP, which allows the drug concentration in plasma to remain constant, resulting in continuous exposure of tumor endothelial cells to the chemotherapeutic agent and allowing them to proliferate slowly, and preventing them from repairing and recovering. PMX-IV dosing (twice-weekly intravenous administration of 20 mg/kg PMX) may not be sufficient to maintain the drug concentration around the tumor tissue, resulting in a drug-free period and allowing the tumor cells to regrow and mobilize circulating endothelial progenitor cells, thus eliciting tumor neovascularization. However, further mechanistic investigations accompanied by dosing studies for metronomic PMX/DCK-OP are required to explore antitumor activity according to anti-angiogenic and immunomodulation effects. On the other hand, after intraperitoneal administration of anti-PD1, the tumor volume was significantly suppressed, by 1.35- and 2.08-fold, compared to the PMX/DCK-OP and control groups, respectively. In addition, the tumor volume showed a more significant

reduction after combined PMX/DCK-OP and intraperitoneal anti-PD1 treatment, which resulted in maximal reduction of tumor volume by 104% and 51.5% relative to PMX/DCK-OP and Anti-PD1, respectively. This synergistic tumor growth-inhibiting activity of chemotherapy with immunotherapy may have been due to enhancement of protective T cell responses and the antigen-presenting function of dendritic cells.⁵⁹ However, oral administration of PMX/DCK-OP or combined treatment with PMX/DCK-OP and anti-PD1 did not affect the body weight of tumor-bearing mice ([Figure 5B](#)), suggesting that the administered drugs do not have systemic toxicity. The tumors isolated on day 14 after combined treatment with oral PMX/DCK-OP and intraperitoneal anti-PD1 also showed significant decreases in tumor mass, by 69.4% and 44.2% compared to the control and oral PMX/DCK-OP groups, respectively ([Figure 5C and D](#)). Langer et al performed an open-label phase 2 cohort study using anti-PD1 (pembrolizumab) along with intravenous carboplatin and PMX, and the results showed significantly improved anticancer efficacy compared to chemotherapy alone.⁶⁰ Therefore, the metronomic combination of PMX/DCK-OP and intraperitoneal anti-PD1 was expected to play a synergistic role in cancer treatment. However, anticancer efficacy of intravenously administered PMX alone, or in combination with anti-PD1, must be demonstrated to confirm that the oral metronomic PMX/DCK-OP allows for a more favorable dosing schedule for synergistic anticancer effects with anti-PD1, via continuous exposure to the tumor endothelial cells, thus decreasing neoangiogenesis and boosting immune activity.

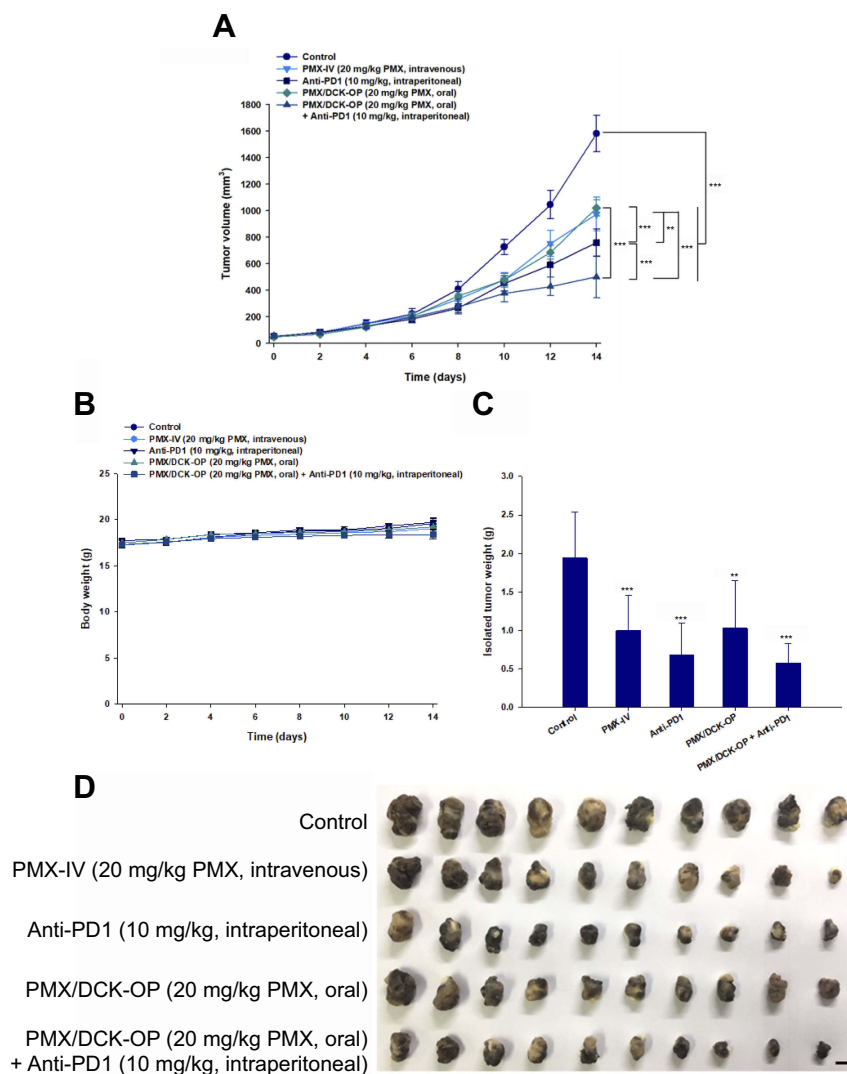


Figure 5 Efficacy of in vivo tumor growth inhibitory effects in B16F10 tumor-bearing mice after biweekly intravenous administration of 20 mg/kg PMX (PMX-IV), intraperitoneal administration of 10 mg/kg anti-PD1 (Anti-PD1) once every 3 days, once-daily oral administration of PMX/DCK-OP as 20 mg/kg PMX (PMX/DCK-OP), and combined treatment with once-daily oral administration of PMX/DCK-OP as 20 mg/kg PMX and intraperitoneal administration of 10 mg/kg anti-PD1 (PMX/DCK-OP+Anti-PD1) once every 3 days for 14 days. **(A)** Tumor volume in mice (** $P < 0.01$, *** $P < 0.001$). **(B)** Variation of body weight in mice during treatment. **(C)** Tumor weight in B16F10 tumor-bearing mice (** $P < 0.01$, *** $P < 0.001$ compared to the control group). **(D)** Photographs of tumors isolated from each group on day 15. Scale bar represents 10 mm. **Notes:** Statistics: one-way ANOVA followed by Tukey's multiple-comparison test. Each value represents the mean \pm standard deviation ($n=10$ for each group). **Abbreviations:** PMX, pemetrexed; DCK, N⁶-deoxycholylyl-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK.

Histological evaluation of the tumor tissues from mice treated with PMX/DCK-OP, alone or in combination with anti-PD1 stained with PCNA, anti-CD31 antibody, and TUNEL (Figure 6) also showed similar trends. Tumors from mice treated with the combination of PMX/DCK-OP and anti-PD1 showed reduced numbers of PCNA-positive proliferative cells compared to the control and oral PMX/DCK-OP groups. In addition, the number of CD31-positive microvessels was markedly decreased in the combined treatment group compared to the control and intravenous PMX groups. In addition, TUNEL assay showed a similar

tendency, with a significantly greater number of apoptotic cells in the combined treatment group compared to the oral PMX/DCK-OP and control groups.

Based on the findings of the present study, PMX/DCK-OP enhanced oral absorption, confirming the improvement in oral bioavailability of PMX with significant tumor growth suppression in combination with intraperitoneal anti-PD1 compared to PMX/DCK and anti-PD1. Therefore, PMX/DCK-OP may be suitable as a drug delivery carrier for PMX and exhibit synergistic activity with immunotherapy.

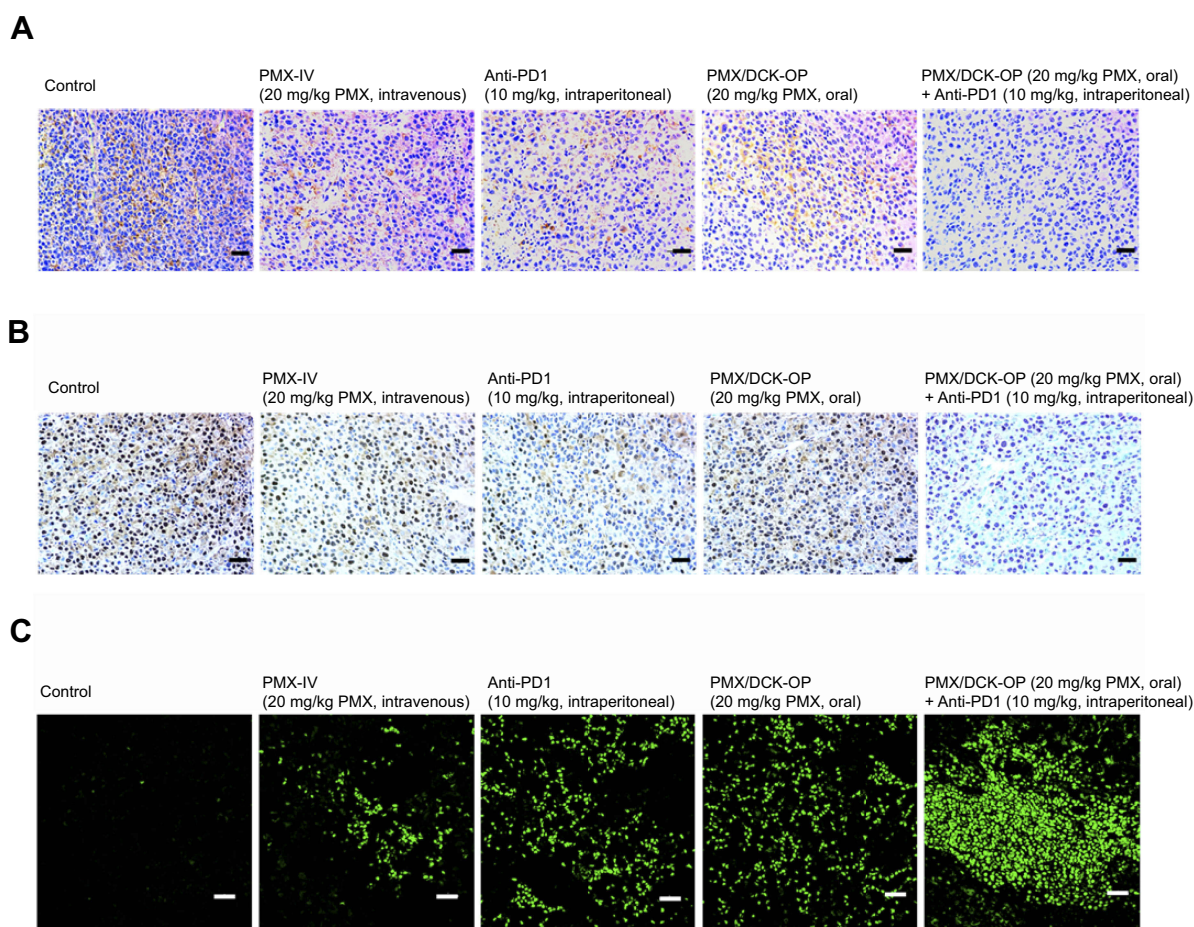


Figure 6 Representative cross-sectional images of isolated tumor tissues stained with **(A)** anti-CD31 antibody for microvessels (brown), **(B)** proliferating cell nuclear antigen (PCNA) for proliferating cells (brown), and **(C)** fluorescent terminal deoxynucleotidyl transferase-mediated dUPT nick end labeling (TUNEL) for apoptosis (green fluorescence) in the tumor tissues taken 14 days after treatment with various modes.

Notes: The treatment modes were biweekly intravenous administration of 20 mg/kg PMX (PMX-IV), intraperitoneal administration of 10 mg/kg anti-PD1 (Anti-PD1) once every 3 days, once-daily oral administration of PMX/DCK-OP as 20 mg/kg of PMX (PMX/DCK-OP), and combined once-daily oral administration of PMX/DCK-OP as 20 mg/kg of PMX with once every 3 days intraperitoneal administration of 10 mg/kg anti-PD1 (PMX/DCK-OP + Anti-PD1). Scale bars represent 50 μ m for anti-CD31 antibody and PCNA staining, and 20 μ m for TUNEL staining, respectively.

Abbreviations: PMX, pemetrexed; DCK, N^o-deoxycholy-L-lysyl-methylester; PMX/DCK, ion-pairing complex between PMX and DCK; PMX/DCK-OP, oral powder formulation of PMX/DCK.

Conclusion

This study confirmed that PMX/DCK-OP enhances the intestinal membrane permeability and oral bioavailability of PMX. PMX/DCK-OP was prepared by ionic complex formation of PMX with DCK at a 1:1 molar ratio, along with Labrasol and P188 as dispersants. The optimum oral powder formulation improved the P_{app} of PMX by 8.26-fold compared to free PMX. The results also indicated that inhibition of bile acid transporters on a Caco-2 cell monolayer significantly curtailed the P_{app} of PMX/DCK-OP, suggesting that an ASBT-mediated pathway is the predominant mechanism underlying intestinal absorption of PMX/DCK-OP, followed by caveola/lipid raft-mediated endocytosis, macropinocytosis, passive diffusion, and paracellular transport. For these reasons, oral PMX/DCK-OP

showed a 294% increase in oral bioavailability in comparison to free PMX, further confirming the role of bile acid derivative and dispersants used in the formulation. Moreover, the tumor growth was maximally suppressed after combined treatment with PMX/DCK-OP and intraperitoneal anti-PD1. These observations suggested the therapeutic potential of combined oral metronomic chemotherapy and immunotherapy for synergistic anticancer efficacy.

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Disclosure

The authors report no conflicts of interest in relation to this work.

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Supplementary material

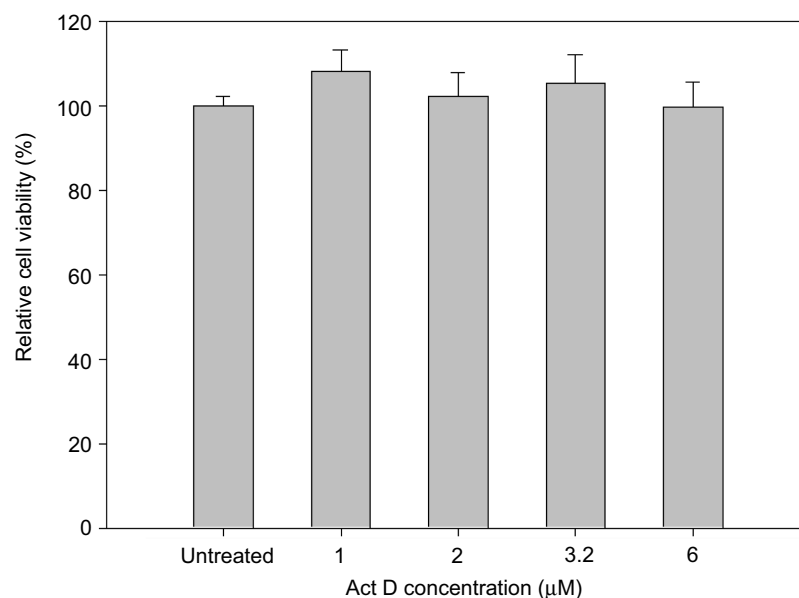


Figure S1 In vitro cytotoxic effect of Act D on Caco-2 cells.

Notes: Cell viability was measured using WST-8 and the growth of Caco-2 cells compared to the untreated control group. All data are expressed as means \pm standard deviation (n=6).

Abbreviation: Act D, actinomycin D.

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