



# Complete Genome Sequence of *Buttiauxella agrestis* DSM 9389

Nao Nakamichi,<sup>a</sup> Ryota Moriuchi,<sup>b</sup>  Hideo Dohra,<sup>b,c</sup>  Hiroyuki Futamata,<sup>a,b,d</sup>  Yosuke Tashiro<sup>a,d,e</sup>

<sup>a</sup>Department of Engineering, Graduate School of Integrated Science and Technology, Shizuoka University, Hamamatsu, Japan

<sup>b</sup>Research Institute of Green Science and Technology, Shizuoka University, Suruga-ku, Shizuoka, Japan

<sup>c</sup>Department of Science, Graduate School of Integrated Science and Technology, Shizuoka University, Shizuoka, Japan

<sup>d</sup>Graduate School of Science and Technology, Shizuoka University, Hamamatsu, Japan

<sup>e</sup>JST PRESTO, Kawaguchi, Saitama, Japan

**ABSTRACT** We report here the complete genome sequence of *Buttiauxella agrestis* DSM 9389, which harbors eight 16S rRNA genes classified into three types. The genome sequence of this strain showed a high average nucleotide identity (97.3%) with that of the highly membrane vesicle-producing strain *B. agrestis* ATCC 33320<sup>T</sup>.

**B**uttiauxella agrestis DSM 9389 (S3/6-333), which was isolated from a slug sampled in Braunschweig, Germany (1), is a member of the family Enterobacteriaceae. While some *Buttiauxella* spp. have unique characteristics in membrane vesicle formation (2, 3), genomic information of the species is limited to several reports (4, 5). In this report, we announce the DSM 9389 complete genome sequence and confirm the species with average nucleotide identity (ANI).

*B. agrestis* DSM 9389 was grown in LB medium at 30°C for 16 h, and its genomic DNA was extracted using a Wizard genomic DNA purification kit (Promega). The complete genome sequence of *B. agrestis* DSM 9389 was determined by the combination of PacBio long reads and Illumina short reads. The genomic DNA was sheared using g-TUBE (Covaris) and size selected using the BluePippin system with a High Pass Plus cassette (Sage Science). A PacBio 20-kb library was prepared using the SMRTbell template prep kit and sequenced on the PacBio RS II instrument (Pacific Biosciences) at Macrogen, Inc. (Seoul, South Korea). An Illumina library was constructed using a TruSeq Nano DNA library prep kit and sequenced on the Illumina MiSeq platform (301-bp paired-end sequencing). Information on the PacBio and Illumina reads used in this study is summarized in Table 1. PacBio subreads were filtered (length, ≥6,000 bp; read

**TABLE 1** Summary of reads generated by the MiSeq and PacBio platforms

Parameter	Data for:	
	Illumina MiSeq	PacBio RS II
Raw reads <sup>a</sup>		
No. of reads	2,020,870	103,022
Total bases (bp)	605,620,107	1,143,739,721
$N_{50}$ (bp)		16,358
Filtered reads		
No. of reads	1,743,456	47,499
Total bases (bp)	461,379,893	723,912,023
$N_{50}$ (bp)		17,225
Mean coverage ( $\times$ ) <sup>b</sup>	101.0	158.5
Accession no.	<a href="#">DRR226681</a>	<a href="#">DRR226682</a>

<sup>a</sup> Raw reads from PacBio RS II indicate subreads with a read quality of  $\geq 0.75$ .

<sup>b</sup> Total bases of filtered reads (base pairs)/genome size of DSM 9389 (base pairs).

**Citation** Nakamichi N, Moriuchi R, Dohra H, Futamata H, Tashiro Y. 2021. Complete genome sequence of *Buttiauxella agrestis* DSM 9389. *Microbiol Resour Announc* 10:e00301-21. <https://doi.org/10.1128/MRA.00301-21>.

**Editor** Julie C. Dunning Hotopp, University of Maryland School of Medicine

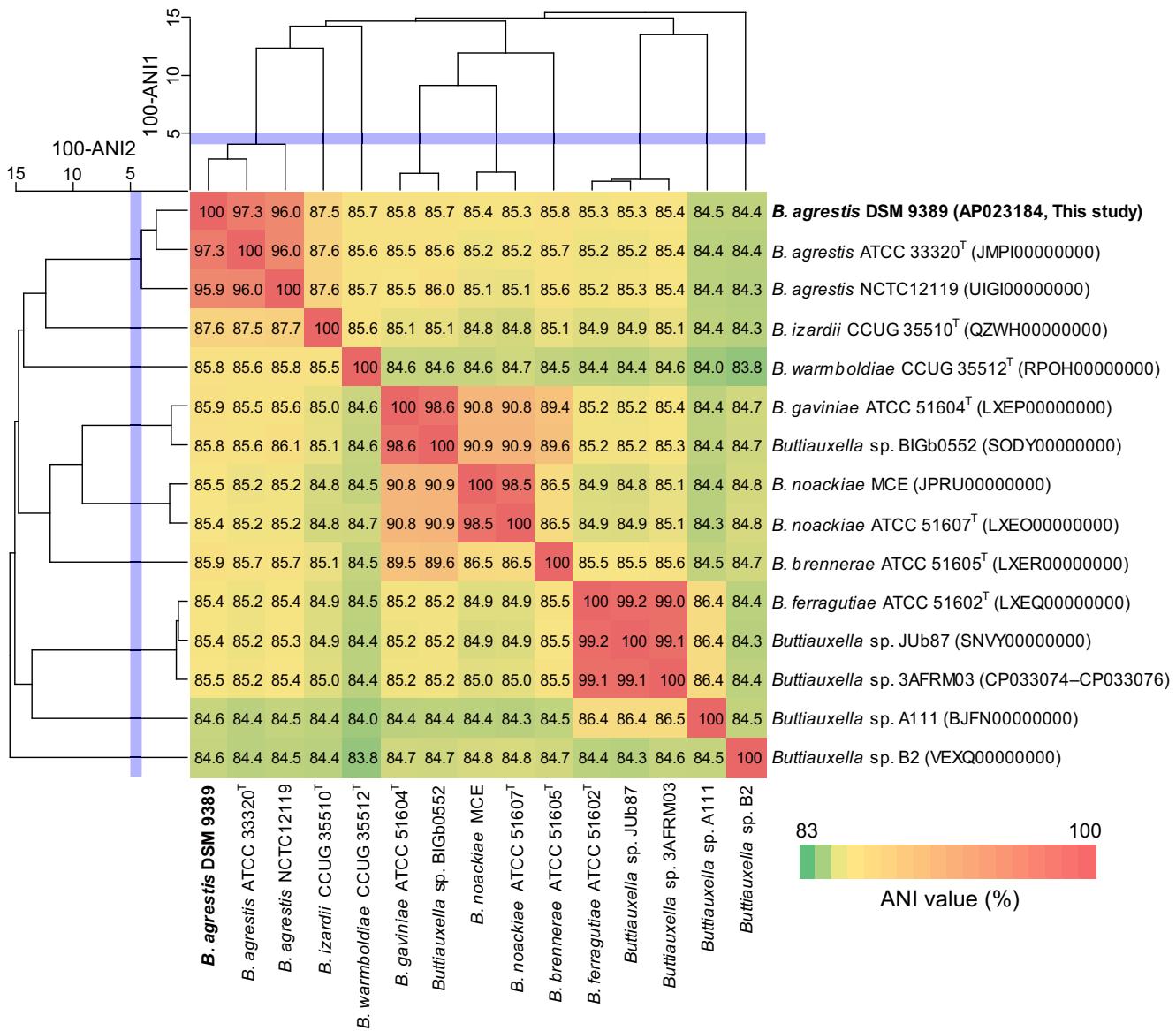
**Copyright** © 2021 Nakamichi et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

Address correspondence to Yosuke Tashiro, tashiro.yosuke@shizuoka.ac.jp.

**Received** 23 March 2021

**Accepted** 14 April 2021

**Published** 13 May 2021



**FIG 1** Average nucleotide identity (ANI) matrix of the genome sequences for *B. agrestis* DSM 9389 and related species. ANI values are visualized by the heatmap, and the relationship of the strains for ANI values are shown by the dendrogram. Blue bands represent the species threshold for ANI values of 95% to 96% (15, 16). GenBank accession numbers for the genome sequences used in this analysis are shown in parentheses.

quality,  $\geq 0.85$ ) using BamTools v. 2.4.1 (6), and the long and high-quality reads were assembled using Canu v. 1.8 (7). The resulting single contig was polished using Arrow v. 2.2.2 (<https://github.com/PacificBiosciences/GenomicConsensus>) and then circularized and rotated to start with the *dnaA* gene using Circlator v. 1.1.1 (8). Illumina reads were cleaned up by trimming adapter sequences and low-quality ends (quality score,  $\geq 15$ ; read length,  $\geq 150$  bp) using Trimmomatic v. 0.38 (9). The high-quality reads were aligned to the polished contig using BWA-MEM v. 0.7.15 (10), and assembly errors were corrected using Pilon v. 1.23 (11). Default parameters were used except where otherwise noted. The complete genome sequence of *B. agrestis* DSM 9389 consisted of a circular chromosome of 4,566,254 bp with a G+C content of 50.7%. The genome was annotated using DFAST v.1.2.3 (12). The genome contains 4,110 protein-coding sequences, 25 rRNA genes, and 83 tRNA genes. Of the eight 16S rRNA genes of strain DSM 9389, five were identical to those of *Buttauxella noackiae* NSW 11 (GenBank accession number NR\_036919.1), and no gene was identical to that of *B. agrestis* ATCC 33320<sup>T</sup> (NR\_041968.1). To confirm the species definition, ANI analysis (13) was

performed using a ruby script (ani.rb) from the enveomics collection (14). The genome sequence of *B. agrestis* DSM 9389 showed a high ANI (97.3%) with that of strain *B. agrestis* ATCC 33320<sup>T</sup> (Fig. 1), resulting in our conclusion that *B. agrestis* DSM 9389 was confirmed to be *B. agrestis*.

**Data availability.** The sequence reads have been deposited in the DDBJ Sequence Read Archive (DRA)/SRA under the accession numbers [DRR226681](#) (Illumina MiSeq) and [DRR226682](#) (PacBio RS II). The complete genome sequence of *B. agrestis* DSM 9389 has been deposited in DDBJ/ENA/GenBank under the accession number [AP023184](#).

## ACKNOWLEDGMENTS

This study is supported by JSPS, KAKENHI (grant number JP19H02920), Japan, and JST, PRESTO (grant number JPMJPR19H8), Japan, to Y.T.

## REFERENCES

- Müller HE, Brenner DJ, Fanning GR, Grimont PAD, Kämpfer P. 1996. Emended description of *Buttiauxella agrestis* with recognition of six new species of *Buttiauxella* and two new species of *Kluyvera*: *Buttiauxella ferruginiae* sp. nov., *Buttiauxella gaviniae* sp. nov., *Buttiauxella brennerae* sp. nov., *Buttiauxella izardii* sp. nov., *Buttiauxella noackiae* sp. nov., *Buttiauxella warmboldiae* sp. nov., *Kluyvera cochleae* sp. nov., and *Kluyvera georgiana* sp. nov. Int J Syst Evol Microbiol 46:50–63. <https://doi.org/10.1099/00207713-46-1-50>.
- Tashiro Y, Hasegawa Y, Shintani M, Takaki K, Ohkuma M, Kimbara K, Futamata H. 2017. Interaction of bacterial membrane vesicles with specific species and their potential for delivery to target cells. Front Microbiol 8:571. <https://doi.org/10.3389/fmicb.2017.00571>.
- Takaki K, Tahara YO, Nakamichi N, Hasegawa Y, Shintani M, Ohkuma M, Miyata M, Futamata H, Tashiro Y. 2020. Multilamellar and multivesicular outer membrane vesicles produced by a *Buttiauxella agrestis tolB* mutant. Appl Environ Microbiol 86:e01131-20. <https://doi.org/10.1128/AEM.01131-20>.
- Jothikumar N, Kahler A, Strockbine N, Gladney L, Hill VR. 2014. Draft genome sequence of *Buttiauxella agrestis*, isolated from surface water. Genome Announc 2:e01060-14. <https://doi.org/10.1128/genomeA.01060-14>.
- Choi J-H, Moriuchi R, Sukprasitchai A, Tokuyama S, Kawagishi H, Dohra H. 2019. Draft genome sequence of *Buttiauxella* sp. strain A111, which converts 2-azahypoxanthine to 2-aza-8-oxohypoxanthine. Microbiol Resour Announc 8:e00664-19. <https://doi.org/10.1128/MRA.00664-19>.
- Barnett DW, Garrison EK, Quinlan AR, Strömberg MP, Marth GT. 2011. BamTools: a C++ API and toolkit for analyzing and managing BAM files. Bioinformatics 27:1691–1692. <https://doi.org/10.1093/bioinformatics/btr174>.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:294. <https://doi.org/10.1186/s13059-015-0849-0>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv 1303.3997 [q-bio.GN]. <https://arxiv.org/abs/1303.3997>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>.
- Konstantinidis KT, Tiedje JM. 2005. Genomic insights that advance the species definition for prokaryotes. Proc Natl Acad Sci U S A 102:2567–2572. <https://doi.org/10.1073/pnas.0409727102>.
- Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for specialized analyses of microbial genomes and metagenomes. PeerJ 4:e1900v1.
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM. 2007. DNA–DNA hybridization values and their relationship to whole-genome sequence similarities. Int J Syst Evol Microbiol 57:81–91. <https://doi.org/10.1099/ij.s.0.64483-0>.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci U S A 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.