

Citation: Marques AM, Ananina G, Costa VP, Vasconcellos JPCd, de Melo MB (2018) Estimating the age of the p.Cys433Arg variant in the *MYOC* gene in patients with primary open-angle glaucoma. PLoS ONE 13(11): e0207409. <u>https://</u> doi.org/10.1371/journal.pone.0207409

Editor: Amanda Ewart Toland, Ohio State University Wexner Medical Center, UNITED STATES

Received: August 13, 2018

Accepted: October 30, 2018

Published: November 16, 2018

Copyright: © 2018 Marques et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: This work was supported by José Paulo Cabral de Vasconcellos (JPCV), São Paulo Research Foundation (FAPESP) grant 2002/11575-0, URL: http://www.fapesp.br/; Mônica Barbosa de Melo (MBM), São Paulo Research Foundation (FAPESP) grant 2010/18353-9, URL: http://www. fapesp.br/; Ana Maria Marques (AMM), **RESEARCH ARTICLE**

Estimating the age of the p.Cys433Arg variant in the *MYOC* gene in patients with primary open-angle glaucoma

Ana Maria Marques¹[#], Galina Ananina¹, Vital Paulino Costa², José Paulo Cabral de Vasconcellos², Mônica Barbosa de Melo¹*

1 Center for Molecular Biology and Genetic Engineering (CBMEG), University of Campinas (UNICAMP), Campinas, São Paulo, Brazil, 2 Department of Ophthalmology and Otorhinolaryngology, School of Medical Sciences, University of Campinas (UNICAMP), Campinas, São Paulo, Brazil

 Current address: Department of Genetics, Evolution, Microbiology and Immunology, Biology Institute, University of Campinas (UNICAMP), Campinas, São Paulo, Brazil
* melomb@uol.com.br

Abstract

The aim of this study was to estimate the age of the Cys433Arg (c.1297T>C, p.Cys433Arg) variant by comparing the genotypes of individuals affected and not affected by primary open angle glaucoma juvenile onset (JOAG). Our sample consisted of 35 JOAG-affected individuals from three families, 16 unrelated patients with the *MYOC* p.Cys433Arg variant and 16 unaffected individuals. Genomic DNA was amplified by PCR; nine short tandem repeats were genotyped through automated electrophoresis and three single nucleotide polymorphisms through Sanger sequencing. The determination of haplotypes was performed using Arlequin software and age estimation was performed using DMLE+ 2.3 and BDMC21 softwares. Four markers constituted the haplotypes associated with the p.Cys433Arg variant. The software DMLE+2.3 predicted an age of 43 generations for this variant with a 95% confidence interval ranging from 28 to 76 generations (560–1520 years) and BDMC21 predicted an age of 59 generations (1180 years) (95% CI: 40 to 100).

Introduction

Glaucoma is the leading cause of irreversible blindness [1], affecting more than 60 million people worldwide and projected to reach 76 million by 2020 [2]. It is characterized by degeneration of retinal ganglion cells, loss of neural tissue from the optic nerve head and correspondent visual field defect [3].

Primary open-angle glaucoma [POAG] is the most common form of the disease with a complex inheritance pattern [4,5]. The main risk factor for POAG is elevated intraocular pressure (IOP), in addition to positive family history, advanced age and black ethnicity [6,7]. Primary open angle glaucoma juvenile onset (JOAG) is a subtype of POAG characterized by an early onset (3 to 40 years of age), usually following an autosomal dominant inheritance



Coordination for the Improvement of Higher Education Personnel (CAPES)-Program of Academic Excellence (PROEX) grant 0230081, URL: http://www.capes.gov.br/. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

pattern and more aggressive clinical presentation, which frequently requires surgical intervention [8,9].

Several *loci* have already been associated with POAG, and the most reproducible and representative is the *GLC1A locus*, where the *MYOC* gene was subsequently identified [10,11]. The *MYOC* gene is composed by three exons with 606, 126 and 718 bp, respectively, and encodes the myocillin protein which is expressed in a variety of ocular tissues including the trabecular meshwork, cornea, retina and ciliary body [12–14]. It has been reported that 8% to 30% of patients with JOAG and 2 to 4% of patients with POAG have disease causing variants in the *MYOC* gene [15,16], evidencing the importance of this gene to glaucoma's etiology.

Vasconcellos et al carried out an investigation in 25 unrelated JOAG patients from Brazil and first reported the *MYOC* p.Cys433Arg (c.1297T>C) variant, which is the replacement of a cysteine for an arginine at codon 433 in the exon 3 of the *MYOC* gene [17]. This variant is involved in the intracellular sequestration (in the endoplasmic reticulum of trabecular meshwork cells) of insoluble aggregates of mutant myocilin, which represents the same pattern of other disease-causing variants. This condition may lead to an increase in oxidative stress that may facilitate the death of these cells and thus impair the output of aqueous humor, ultimately leading to increased IOP [18–20].

This variant was found only in Brazilian patients and has never been reported in other countries [17,21]. Searches performed in the Brazilian databases BIPMed [http://bipmed.iqm. unicamp.br/] and ABraOM (http://abraom.ib.usp.br/) and in international databases ExAC Browser (http://exac.broadinstitute.org/) and 1000 genomes (http://www.internationalgenome. org/) found no registration of this variant. The few studies about this variant reveal high pene-trance in patients with JOAG, ranging from 28 to 100% depending on the age of the patient [17,21,22]. Case-control studies report a high prevalence of the p.Cys433Arg variant in the state of São Paulo; approximately 30% of unrelated JOAG patients that harbor variants in the *MYOC* gene have this variant [23].

In the first study, which reported the p.Cys433Arg variant, Vasconcellos and colleagues analyzed four microsatellite markers in unrelated patients with JOAG who carried this variant and showed that six patients exhibited the same haplotype, indicating that these patients inherited the variant from a common ancestor [17]. Based on this fact, the aim of this study was to estimate the age of the p.Cys433Arg variant, contributing to the understanding of the demographic history of this variant in the Brazilian population, and to estimate the population at risk to harbor it.

Methods

Patients

This study consisted of 51 individuals, of whom 35 came from three different families and 16 were unrelated individuals, all affected by JOAG, with age at diagnosis less than 40 years-old and harboring the p.Cys433Arg variant in the *MYOC* gene. The control group consisted of 32 unrelated individuals unaffected by the disease and above 50 years of age. Both patients and controls are from the State of São Paulo, Southeast region of Brazil (Fig 1). In terms of ethnicity most of the patients and controls analyzed in this study were classified by the observers as "pardos", which is a term used in the Brazilian census by the Brazilian Institute of Geography and Statistics (Instituto Brasileiro de Geografia e Estatística—IBGE) to refer to individuals of mixed race origin, as a result of a miscegenation among European, African and Amerindian descendants. The study was approved by the Research Ethics Committee of the Faculty of Medical Sciences, University of Campinas (CAAE: 49120615.4.0000.5404). Written informed consent was obtained from all participants.



Fig 1. Map of Brazil emphasizing the Southeast region. Southeast region of Brazil formed by the states of São Paulo (SP), Minas Gerais (MG), Rio de Janeiro (RJ) and Espírito Santo (ES). The red points represent the hometowns of the families (Botucatu, Tanabi and Mogi das Cruzes). Yellow points represent the hometowns of the unrelated patients (Campinas, Limeira, Americana, Mogi Guaçu, Jundiaí, Pirassununga, Itapira and Paraguaçu Paulista).

https://doi.org/10.1371/journal.pone.0207409.g001

A comprehensive ophthalmological examination, including visual acuity measurement, slit-lamp biomicroscopy, Goldmann applanation tonometry, gonioscopy and optic disc evaluation (with the pupils dilated and using a 78-diopter lens) were performed. The definition of JOAG was based on structural and functional changes observed in the optic disc and visual field evaluations, respectively, as well as an open angle at gonioscopy. The optic disc was considered to have glaucoma if it exhibited at least two of the following signs: thinning of the neuroretinal rim, haemorrhage, notch, cup/disc ratio >0.7, or asymmetry of the cup/disc ratio >0.2. Glaucomatous visual field defect was defined as a pattern standard deviation outside of 95% of the normal limits or a glaucoma hemifield test result outside of the 99% limit and a correspondent cluster of points with p < 0.05% in the pattern deviation graphic.

The participants had their biological materials collected at the Glaucoma Service, Clinical Hospital of the University of Campinas and were identified as having the *MYOC* p.Cys433Arg variant in previous studies [17,20,22,23].

Genotyping

Nine short tandem repeats (STRs) and three single nucleotide polymorphisms (SNPs) have been selected in a 5Mb region around the p.Cys433Arg variant in the *MYOC* gene (see S1 Table). The PRIMER3 (http://primer3.ut.ee/) software was employed to predict the primers. The forward primers of the STRs were labeled with HEX or FAM fluorophores at the 5' end in order to assemble multiplex reactions with three markers on each. The fragments of interest were amplified by PCR and confirmed in 1.5% agarose gel electrophoresis. The descriptions of the primers are shown in Tables 1 and 2.

Molecular Marker	Primer sequences (5'-3')	M/m	Annealing temperatures (°C)	Product sizes (bp)
rs6133 (F)	ACTGCTGTCCATTGTCCTGA	C/A	59	197
rs6133 (R)	ACAGGCATAGCATCACTTCCT			
rs2266782 (F)	TGCTGTAATGGTTTGTTCCGG	G/A	59	238
rs2266782 (R)	GCGACCTTGTGAATAGATGCA			
rs2266780 (F)	TGGCATTGTGTCCGTAAAGC	A/G	59	237
rs2266780 (R)	TGGACAAAGCCAATCACTGC			

Table 1. Primers used in the polymerase chain reaction (PCR) for single nucleotide polymorphisms (SNPs) genotyping.

M/m denotes major allele/minor allele

https://doi.org/10.1371/journal.pone.0207409.t001

Repetition unit

(CA)

(TG)

(TG)

(AC)

(CA)

(TG)

(CA)

(TG)

(GGAA)

117-210

196-224

215-249

	Μ	MYOC Cys433Arg va			
Table 2. Primers used in th	e polymerase chain reaction (PCR) for sl	nort tandem repea	ts (STRs) genotyping.		
Molecular marker		Fluorophores	Annealing temperatures (C)	Product sizes (bp)	
rs3219828 (R)	ACTCAATGTTCAAGTTCTCCCTG	FAM	57	133-177	
rs3221612 (F)	GCCCAGCAAAGAGACCTTCA	HEX	57	179-250	
rs3221612 (R)	CTTCGAACACGTCTTCCCAC	_			
rs3219958 (D1S218) (F)	TGTAAAAGCAAACTGTAGACGAT	FAM	57	266-286	
rs3219958 (D1S218) (R)	TTTATGTTATCACCAAGGCTTCT				
rs2234708 (F)	ACATCCCAATTTATGCACCCA	FAM	59	190-207	
rs2234708 (R)	CACAGTGCAGGTTCTCAATGA				
D1S2815 (F)	CTGACATGGAATACCTCTATGATGC	HEX	59	210-237	
D1S2815 (R)	CTCCAAATCTAGTCACACTGGAAG				
rs3223566 (D1S2790) (F)	GAAGCCTTTCTGTGCTAGCC	FAM	59	243-253	

FAM

HEX

FAM

58

58

62

AGGCAACAGAGTGAGACTC

CTTCCATAGCTGATACTGCT

GCGCTTGTAATCCCAGCTAC

AGTTGATGGGTAGGGGTGTG

GAGCCACCACTCCAGTTTGA

CTCTCTACCACTTGAATTCCTGT

GCACAATGGCCAATGTTTTCA

https://doi.org/10.1371/journal.pone.0207409.t002

rs3223566 (D1S2790) (R)

D1S1165 (F)

D1S1165 (R)

rs3220994 (F)

rs3220994 (R)

rs3220452 (F)

rs3220452 (R)

SNPs were sequenced by Sanger method in the ABI PRISM 3500 DNA Analyzer (Applied Biosystems, Foster City, CA, USA) and the sequences obtained were analyzed by the FinchTV software (Geopiza, Seattle, WA, USA). The STRs PCR products were genotyped by automated electrophoresis in the same equipment and analyzed with Geneious version 10.0 software (http://www.geneious.com) [24] (see S2 Table for genotypes).

To determine the haplotypes of unrelated individuals we used the Arlequin v3.5 software [25] and for the families' individuals the haplotypes were determined by pedigree analysis.

Age estimation

The age of the variant was estimated by the DMLE+ 2.3 [26] and BDMC21 [27] softwares. DMLE+ 2.3 allows Bayesian inference to calculate the age of the variant (in generations) based on the linkage disequilibrium observed in multiple genetic markers. This software uses the Markov Chain Monte Carlo algorithm (MCMC) which is based on massive random simulations, to calculate the probability of the occurrence of each event. For this study, four million repetitions have been used to obtain the results.

To estimate the age of the variant using DMLE+ 2.3, it was necessary to determine the proportion of mutant chromosomes (0.0294). This parameter was calculated based on the frequency of JOAG in the Brazilian population (150,000 affected people-http://www.who.int), the frequency of the variant in the affected individuals (30%) [17,21–23] and the size of the population in the Southeast region of Brazil, the region where the variant has been reported so far (80,350,000 inhabitants—http://www.ibge.gov.br/). Using this data, we were also able to calculate the number of mutant chromosomes in the population of the Southeast region of Brazil. These calculations were performed using the world population (7.465.000.000 inhabitants—http://www.ibge.gov.br/), the population affected by JOAG and the frequency of the variant in people affected by JOAG. The genetic distances between the markers and between the markers and the variant were extrapolated from physical distances using the Map Interpolator software from The Rutgers Combined Linkage—Physical Map [28]. The average

Brazilian population growth rate for the last 300 years was taken as 0.095 (http://www.ibge. gov.br/home/). In addition to these data, haplotypes containing and not containing the variant were added to the software input file. BDMC21 is a maximum likelihood method based on the extent of genetic variation within an allelic class and the allele frequency [27, 29]. To calculate the age of the variant using BDMC21, we applied the same Brazilian population growth rate and the proportion of mutant chromosomes of DMLE+ 2.3. In this software we used ten thousand repetitions of MCMC.

Labuda et al. (1996, 1997) [30–32] suggested that the age of the variant can be underestimated when the study population is growing because the genetic clock becomes slower than expected. Due to recombination events, LD has been decreasing over the time from the founding event until now and the estimation method takes into account the LD measure and recombination rates. However, in growing populations, in the first generations, immediately after the founding event, recombination occurs less than expected. Therefore, a possible approach to this problem is to define the genetic clock through the Luria-Delbrüch correction [33,34]. For the parameters of the calculation we used the population growth rate (0.095) and the recombination rate (equivalent to genetic distance—0.0076 cM) between markers flanking the haplotype (among markers rs3219828 and rs2234708).

Results

Population analysis of SNPs

Population analysis of the markers showed that, in the sample of unrelated individuals, the rs6133 SNP was monomorphic, presenting only the C allele. The marker rs2266782 exhibited the G allele more frequently than the A allele, whereas for marker rs2266780, the A allele was identified as the more frequent.

Haplotypes and age of variant

The haplotypes associated with the p.Cys433Arg variant were formed by the markers rs3219828 (STR), rs2266782 (SNP), rs2266780 (SNP) and rs2234708 (STR) (Fig 2). The haplotypes determined for individuals in the control group are in S1 Fig. In this group, we found a greater variation of haplotypes when compared to the group that harbors the variant, with the presence of alleles not observed in the haplotypes of the case group.

Analysis performed by the DMLE+ 2.3 and BDMC21 softwares estimated that the age of the *MYOC* p.Cys433Arg variant might be 43 generations (95% CI: 28–76) (Fig 3) and 59 generations (95% CI: 40–100) (Fig 4), respectively. If the time interval between one generation and the next is considered to be 20 years [27,29, 35–37], the age of the variant lies in a range of approximately 560 to 1520 years (mode = 860 years) by DMLE+2.3 and approximately 800 to 2000 years (mode = 1180 years) by BDMC21. The application of the Luria-Delbrüch correction, resulted in $g_0 = 4.25$, which means approximately four more generations than the numbers reached using the softwares for the age prediction of the variant. This correction lies within the confidence interval determined in the calculation of the age of the variant.

We also calculated the number of mutant chromosomes in the population of the Southeastern region of Brazil, resulting in approximately 484 chromosomes with the variant in this region of the country.

Discussion

Founder variants are variants "observed with high frequency in a group that is or was geographically or culturally isolated, in which one or more of the ancestors was a carrier of the

1p 36.32	1p35.3 1p34.2 1p32.2	1p31.1 1p21.3 1p12	lq21.2 lq23.2	1q24.3 1q31.1 1q32.2	1942.12 1943		
36.23	036.12 035.1 033 033	022.2 013.2	21 ZZ	q24.1 q25.2 q31.3	42.2		
					MYOC	gene	
	(%)			~	8	~	
se de de	ype iy (%	328	782	780	233	708	
and elate tient	plot	198	.99	566	131	34	
e aunre pat	Hal	s32	s22	s22	s74	s22	
	fr	<u> </u>	-	<u> </u>	-	<u> </u>	
5 (family 1, patients 1, 2, 3, 4)	26.32	2	G	A	С	1	
2 (families 2 and 3)	10.53	1	G	A	С	1	
1 (patient 9)	5.26	2	G	A	С	2	
1 (patient 10)	5.26	1	G	A	С	3	
1 (patient 13)	5.26	2	G	А	С	5	
2 (patients 11 and 12)	10.53	3	G	А	С	5	
3 (patients 5, 6, 7)	15.79	2	А	А	С	1	
1 (patient 14)	5.26	2	А	G	С	1	
1 (patient 15)	5.26	4	А	G	С	2	
1 (patient 16)	5.26	1	А	G	С	5	
1 (patient 8)	5.26	1	А	А	С	1	

Fig 2. Representation of the haplotypes determined for individuals with the p.Cys433Arg variant at chromosome region 1q24.3. Haplotypes are formed by the markers rs3219828 (STR), rs2266782 (SNP), rs2266780 (SNP) and rs2234708 (STR). Each number represents a different allele in each microsatellite marker, as the letters represent different alleles in each SNP. The rs74315338, highlighted in bold, corresponds to the variant of interest. The first column indicates the number of each haplotype constructed and corresponding proband identity for each haplotype (family and unrelated patients). The position of the *MYOC* gene in relation to the markers is represented.

https://doi.org/10.1371/journal.pone.0207409.g002



Age of variant (generations)

Fig 3. Age estimation for the p.Cys433Arg variant as assessed by DMLE+2.3. Histogram representing the age of the p.Cys433Arg variant in generations according to the probability of each age to occur, resulting from the iterations performed by the DMLE+ 2.3 software. Green bars indicate 95% CIs.

https://doi.org/10.1371/journal.pone.0207409.g003



Fig 4. Age estimation for the p.Cys433Arg variant as assessed by BDMC21. Distribution of likelihoods of the p. Cys433Arg variant age (in generations).

https://doi.org/10.1371/journal.pone.0207409.g004

altered gene" (https://www.cancer.gov/publications/dictionaries/genetics-dictionary?cdrid= 570712). Founder variants are important for studies in human genetics. Whenever these variants are discovered, they may help the molecular diagnosis of diseases, as part of an initial screening, which is cheaper than other genomic screening techniques [38]. Genetic tests for one or more founder variants are considered to be more efficient than for other rare variants [39–42]. Founder variants exhibit linkage disequilibrium (LD) with nearby genetic markers and with the use of LD mapping they can be helpful in the identification of the related genes. The size of the LD interval correlates inversely with the time since the variant occurred in the population and the identification of the same founder variant in different populations supports studies of the relationship among them [41,43].

A number of founder variants was detected in the *MYOC* gene, as in patients from Spain [44], where the heterozygous A>C transversion in codon 380 was detected in all affected members with JOAG from five families and was not detected in any of the 100 individuals without the disease. To determine whether a founder effect occurred, four microsatellite markers comprising the *MYOC* gene were analyzed. The results showed that a common haplotype associated with the disease was found in three families, indicating that the members of these families inherited the variant from a common ancestor. There are reports about founder variants in the *MYOC* gene in populations from France [45], Quebec [46] and Australia [47, 48]. These variants may have arisen within the population itself or may have been brought to the population immigrants [41]. In this case, immediately after the founder event, the allele could be found at higher frequency than it was before and could reach even higher frequency because of the strong genetic drift that might have occurred while the population was still small [49]. In our study we suggest a founder effect of the p.Cys433Arg variant, supporting the study of Vasconcellos et al [17].

As mentioned before, the p.Cys433Arg variant was reported exclusively in Brazil. We have suggested that this variant is associated with one common haplotype in individuals carrying the variant, and therefore it would have been inherited from a common ancestor [17].

Estimating the age of the p.Cys433Arg variant could help to understand the demographic history of this variant in our population and the projection of its prevalence and distribution in the Brazilian population. We estimate that about 484 people in the Southeast region of Brazil carry this variant. However, this estimate cannot be expanded to the whole country, because the presence of this variant has not been investigated in individuals from other Brazilian states.

The Brazilian population is the fifth largest population in the world with a high degree of miscegenation among native indigenous and immigrants. It is known that more than 2 million natives of several ethnic groups inhabited the territory before the arrival of the immigrants [50]. Colonization of the Brazilian territory by Europeans began about 500 years ago. During the early colonization history, approximately 500 thousand Portuguese individuals arrived in Brazil, followed by more than 4 million European immigrants in subsequent immigration waves, including more Portugueses, Germans, Italians and Spaniards. Meanwhile, during the slavery period (from the 16th to 19th century), approximately 4 million Africans from different regions of Africa were brought to the country [50–52].

Our study estimated the age of the p.Cys433Arg variant between 43 (860 years) and 59 (1180 years) generations. The estimates were obtained with two computational approaches, using DMLE+ 2.3 (43 generations) and BDMC21 (59 generations). We know that age-based variant analysis methods coalescent-based (DMLE+ 2.3) and likelihood-based (BDMC21) are quite efficient for young variants with fast population growth rate, which are the cases of the p. Cys433Arg variant and the Brazilian population, respectively [53]. The study of Clendenning et al (2008) [54], showed that the coalescent intra-allelic model to assess LD used by DMLE+ 2.3 software was more efficient to date the origin of the study variant than the method that gives a separate age estimate for each marker. The age of the variant was already known and the result obtained by DMLE was very accurate. Therefore, we believe that DMLE+ 2.3 software was the best choice to identify the age of the variant in this case. Besides, we defined the haplotype structure from the segregation analysis in the three families and also used statistically inferred haplotypes, analyzing a relatively small group of non-related individuals. Naturally sampling variation in allele frequencies can occur in the studies of rare variants and this inevitably will produce variance in LD estimations. Different marker types also tend to influence the estimates.

These results bring us to the beginning of the European colonization or even before that. Naturally, the question is whether the variant has arisen in the native population that inhabited Brazil or if it was brought by immigrants. The alternative about the appearance of this variant in native Brazilians is uncertain, since there are no studies directly addressing the incidence of glaucoma in the Brazilian indigenous population. A review of the ocular health of Brazilian natives has identified studies on indigenous peoples from the Amazon basin [55]. However, most of the reported cases were primary angle-closure glaucoma, and we know that the p. Cys433Arg variant is associated with primary open-angle glaucoma. Notwithstanding, it is important to notice that there are no studies investigating indigenous groups from the Southeast Brazil, where the p.Cys433Arg variant was detected. This fact, in addition to the absence of this variant in other studies around the world, does not exclude its origin among Brazilian native indigenous from the Southeast region.

The alternative hypothesis includes the arrival of the variant with the settlers. It could have arrived in Brazil with the slave trade among Africans but, if it was the case, it should have resulted in a younger variant with a lower number of generations compared to what we observed. In the present study, we analyzed a relatively small number of markers. The rs6133 SNP was the only marker that can be considered as an "ancestry marker", because it has prominently different allele frequencies in European and African populations. Information from the Ensembl database (http://www.ensembl.org/index.html) shows that the C allele is more

frequent in populations from South and East Asia (98% and 100%, respectively), Europe (88%) and America (87%), while in the African population the A allele is the most frequent (66%). We found the C allele as the most prevalent in our study sample. Since this, is the least frequent allele in the African population, it suggests that the variant may not have arrived in Brazil with the Africans.

Another possibility is the arrival of the variant with European colonizers. This is supported by the confidence interval of the age of the variant, which is close to the time of the arrival of Europeans in Brazil, when a founder effect could have occurred, as suggested by Vasconcellos et al [17]. The p.Cys433Arg variant may have come with European colonizers to Brazil, being extremely rare in Europe or even unique among settlers. Upon arrival in Brazil, its frequency could have decreased until nearly disappearing or it could have increased, as is the case in the Brazilian population. Noteworthy is the fact that this variant has never been described in European populations, which have been extensively studied.

The Brazilian population is highly heterogeneous, so the most accurate way to determine the ethnicity of the studied individuals would be with the use of ancestry markers or high density marker panels, an analysis that was not in the scope of this study. However, we know that Brazilians are the result of a number of migration events that were accompanied by intensive mixing of three main ancestral roots: Amerindians, Europeans and African [56]. In the Southeast region of Brazil, the European origin prevails, but with continuous migration events, the intermarriage between individuals of different ancestry is frequent [57]. A study conducted in our laboratory evaluated a group of 145 individuals from the same region of Brazil (São Paulo State), using high density array markers and found 62.4% European, 27.3% African and 10.2% Amerindian ancestry; however, the analyzed cohort was not preselected by ethnicity or skin color (personal communication).

Technological advances facilitating molecular genetic testing and overall request of societies for the preventing health care draw attention to the population-based screening for adultonset disorders. The screening tests have a different purpose when compared to the diagnostic tests and aim to identify individuals who are more likely to have diseases. This may help to prevent the development of a serious illness or alleviate the severity of the disease. Knowledge of population-specific variants is necessary to compose population screening programs and proposes population-based intensive genetic studies.

The investigation about the demographics and age of the p.Cys433Arg variant, reported only in the Brazilian population, suggests a founder effect and its origin in native indigenous from the Southeast of Brazil or its arrival with European colonizers. The estimated number of about 484 people in Southeast Brazil carrying the variant contributes to our understanding of the distribution of JOAG in this region and reinforces the importance of screening JOAG suspects for this variant in the *MYOC* gene and may help to optimize screening tests for this population at risk.

Supporting information

S1 Table. Physical distance among markers and the *MYOC* gene. Location of *MYOC* gene: 1: 171,635,417–171,652,683.

(PDF)

S2 Table. Table summarizing JOAG families and unrelated patients genotyping data.

Determined genotypes of families and unrelated patients affected by JOAG. Each column represents a marker and each line represents an individual (in the case of unrelated patients) or each family. Each number (in case of microsatellites) represents a different allele from each marker. These numbers were assigned in ascending order according to the alleles found in each sample. The markers are sorted according to the position on the chromosome. x represents an undefined allele.

(PDF)

S1 Fig. Representation of the haplotypes determined for control group. Haplotypes formed by the same markers as in the mutated group. Each number represents a different allele in each microsatellite marker, as the letters represent different alleles in each SNP. The rs74315338 highlighted in bold corresponds to the variant of interest. The first column indicates the amount of each haplotype constructed. The position of the *MYOC* gene in relation to the markers is represented.

(PDF)

Acknowledgments

We would like to thank all the patients who participated in this study and Dr. Dulcinéia Martins Albuquerque for technical assistance. This work was funded by Fapesp grants 2002/11575-0 and 2010/18353-9 and CAPES/PROEX 0230081.

Author Contributions

Conceptualization: José Paulo Cabral de Vasconcellos, Mônica Barbosa de Melo.

Data curation: Ana Maria Marques.

Formal analysis: Ana Maria Marques, Galina Ananina.

Investigation: Ana Maria Marques, José Paulo Cabral de Vasconcellos.

Methodology: Ana Maria Marques, Galina Ananina.

Project administration: Ana Maria Marques, Mônica Barbosa de Melo.

Resources: José Paulo Cabral de Vasconcellos, Mônica Barbosa de Melo.

Software: Ana Maria Marques, Galina Ananina.

Supervision: Galina Ananina, Mônica Barbosa de Melo.

Writing - original draft: Ana Maria Marques.

Writing – review & editing: Ana Maria Marques, Galina Ananina, Vital Paulino Costa, José Paulo Cabral de Vasconcellos, Mônica Barbosa de Melo.

References

- 1. Goldberg I. How common is glaucoma worldwide? Glaucoma 21st Century. 2000;1-8.
- Tham YC, Li X, Wong TY, Quigley HA, Aung T, Cheng CY. Global prevalence of glaucoma and projections of glaucoma burden through 2040: A systematic review and meta-analysis. Ophthalmology [Internet]. 2014; 121(11):2081–90. Available from: http://dx.doi.org/10.1016/j.ophtha.2014.05.013 PMID: 24974815
- Ray K, Mukhopadhyay A, Acharya M. Recent advances in molecular genetics of glaucoma. Mol Cell Biochem. 2003; 253(1–2):223–31. PMID: 14619973
- 4. Quigley HA. Open Angle Glaucoma. N Engl J Med. 1993; 281.
- Weinreb RN, Khaw PT. Primary open-angle glaucoma. Lancet (London, England) [Internet]. 2004; 363 (9422):1711–20. Available from: http://www.sciencedirect.com/science/article/pii/S0140673604162570
- Liu Y, Allingham RR. Molecular genetics in glaucoma. Exp Eye Res [Internet]. 2011; 93(4):331–9. Available from: http://dx.doi.org/10.1016/j.exer.2011.08.007 PMID: 21871452
- 7. Cristina M, Connell AMS. The Barbados Eye Study. 2013;

- Khan AO. Genetics of primary glaucoma. Curr Opin Ophthalmol [Internet]. 2011; 22(5):347–55. Available from: http://content.wkhealth.com/linkback/openurl?sid=WKPTLP:landingpage&an=00055735-201109000-00007 https://doi.org/10.1097/ICU.0b013e32834922d2 PMID: 21730848
- Johnson AT, Richards JE, Boehnke M, Stringham HM, Herman SB, Wong DJ, et al. Clinical phenotype of juvenile-onset primary open-angle glaucoma linked to chromosome 1q. Ophthalmology [Internet]. 1996; 103(5):808–14. Available from: <u>http://dx.doi.org/10.1016/S0161-6420(96)30611-8</u> PMID: 8637692
- Sheffield VC, Stone EM, Alward WLM, Drack A V., Johnson AT, Streb LM, et al. Genetic linkage of familial open angle glaucoma to chromosome 1q21–q31. Nat Genet. 1993; 4(1):47–50. https://doi.org/ 10.1038/ng0593-47 PMID: 8513321
- Stone EM. Identification of a Gene That Causes Primary Open Angle Glaucoma. Science (80-) [Internet]. 1997; 275(5300):668–70. Available from: http://www.sciencemag.org/cgi/doi/10.1126/science. 275.5300.668
- Polansky JR. Cellular Pharmacology and Molecular Biology of the Trabecular Meshwork Inducible Glucocorticoid Response Gene Product. Ophthalmologica. 1997;
- Ortego J, Escribano J, Coca-Prados M. Cloning and characterization of subtracted cDNAs from a human ciliary body library encoding TIGR, a protein involved in juvenile open angle glaucoma with homology to myosin and olfactomedin. FEBS Lett [Internet]. 1997; 413(2):349–53. Available from: http://dx.doi.org/10.1016/S0014-5793(97)00934-4 PMID: 9280311
- Lütjen-Drecoll E, May CA, Polansky JR, Johnson DH, Bloemendal H, Nguyen TD. Localization of the stress proteins alpha B-crystallin and trabecular meshwork inducible glucocorticoid response protein in normal and glaucomatous trabecular meshwork. Investig Ophthalmol Vis Sci [Internet]. 1998; 39 (3):517–25. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9501861%5Cnhttp://iovs. arvojournals.org/data/Journals/IOVS/933428/517.pdf
- 15. Yen Y-C, Yang J-J, Chou M-C, Li S-Y. Identification of mutations in the myocilin (MYOC) gene in Taiwanese patients with juvenile-onset open-angle glaucoma. Mol Vis. 2007; 13(June):1627–34.
- Mimivati Z, Nurliza K, Marini M, Liza-Sharmini A. Identification of MYOC gene mutation and polymorphism in a large Malay family with juvenile-onset open angle glaucoma. Mol Vis [Internet]. 2014; 20 (May):714–23. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid= 4037532&tool=pmcentrez&rendertype=abstract
- 17. Vasconcellos JPC, Melo MB, COSTA FF. Novel mutation in the MYOC gene in primary open angle glaucoma patients. J Med Genet. 2000; 37(4):297–8.
- Fautsch MP, Johnson DH. Characterization of Myocilin—Myocilin Interactions AND Collection of Aqueous Humor. Invest Ophthalmol. 2001;2324–31.
- Aslan M, Dogan S, Kucuksayan E. Oxidative stress and potential applications of free radical scavengers in glaucoma. Redox Rep [Internet]. 2013; 18(2):76–87. Available from: http://ovidsp.ovid.com/ovidweb. cgi?T=JS&PAGE=reference&D=medl&NEWS=N&AN=23485101 https://doi.org/10.1179/ 1351000212Y.0000000033 PMID: 23485101
- Braghini CA, Neshich IAP, Neshich G, Soardi FC, de Mello MP, Costa VP, et al. New mutation in the myocilin gene segregates with juvenile-onset open-angle glaucoma in a Brazilian family. Gene [Internet]. 2013; 523(1):50–7. Available from: http://dx.doi.org/10.1016/j.gene.2013.02.054 PMID: 23566828
- Povoa CA, Santiago Malta RF, de Moraes Rezende M, Santana de Melo KF, Giannella-Neto D. Correlation between genotype and phenotype in primary open angle glaucoma of Brazilian families with mutations in exon 3 of the TIGR/MYOC gene. Arq Bras Oftalmol. 2006; 69(3):289–97. PMID: 16936947
- Vasconcellos JPC, de Melo MB, Schimiti R, Costa FF, Costa VP. Penetrance and phenotype of the Cys433Arg myocilin mutation in a family pedigree with primary open-angle glaucoma. J Glaucoma [Internet]. 2003; 12(2):104–7. Available from: http://www.ncbi.nlm.nih.gov/pubmed/12671463 PMID: 12671463
- **23.** Vasconcellos JPC. New mutations in the MYOC gene in patients with juvenile openangle glaucoma. In: 64° Annual Meeting of the American Society of Human Genetics, 2014. 2014;2014.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, et al. Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 2012; 28(12):1647–9. https://doi.org/10.1093/bioinformatics/bts199 PMID: 22543367
- 25. Excoffier L, Laval G, Schneider S. Arlequin (version 3.0): An integrated software package for population genetics data analysis. Evol Bioinform Online [Internet]. 2005; 1:47–50. Available from: https://www.researchgate.net/publication/24238444_Arlequin_(version_3.0)_an_integrated_software_package_for_population_genetics_data_analysis
- Reeve JP, Rannala B. DMLE+: Bayesian linkage disequilibrium gene mapping. Bioinforma Appl Note. 2002; 18(6):894–5.

- Slatkin M, Rannala BH. Estimating the age of alleles by use of intraallelic variability. Am J Hum Genet [Internet]. 1997; 60(2):447–58. Available from: papers2://publication/uuid/7819B9D5-EC40-45C9-91F9-09B72E18011E PMID: 9012419
- Nato, AQ, Buyske, S, and Matise T. The Rutgers map: A third-generation combined linkage-physical map of the human genome. Hum Genet Inst New Jersey Second Res Day.
- Behura SK, Severson DW. Motif mismatches in microsatellites: Insights from genome-wide investigation among 20 insect species. DNA Res. 2015; 22(1):29–38. https://doi.org/10.1093/dnares/dsu036 PMID: 25378245
- Luria S, Delbrück M. Mutations of Bacteria from Virus Sensitivity to Virus Resistance. Genetics. 1943; 28(6):491–511. PMID: 17247100
- Labuda M, Labuda D, Korab-Laskowska M, Cole DE, Zietkiewicz E, Weissenbach J, et al. Linkage disequilibrium analysis in young populations: pseudo-vitamin D-deficiency rickets and the founder effect in French Canadians. Am J Hum Genet. 1996; 59(3):633–43. PMID: 8751865
- Labuda D, Zietkiewicz E LM. The Genetic Clock and the Age of the Founder Effect in Growing Populations: A Lesson from French Canadians and Ashkenazim. Am J Hum Genet [Internet]. 1997; 61:768– 71. Available from: Kriegsbild PMID: 9326343
- **33.** Colombo R. Age estimate of the N370S mutation causing Gaucher disease in Ashkenazi Jews and European populations: A reappraisal of haplotype data. Am J Hum Genet [Internet]. 2000; 66(2):692–7. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1288120&tool= pmcentrez&rendertype=abstract https://doi.org/10.1086/302757 PMID: 10677327
- Blumen SC, Korczyn AD, Lavoie H, Medynski S, Chapman J, Asherov A, et al. Oculopharyngeal MD among Bukhara Jews is due to a founder (GCG)9 mutation is in the PABP2 gene. Neurology. 2000; 55 (9):1267–70. PMID: 11087766
- 35. Griffiths RC, Tavaré S. The age of a mutation in a general coalescent tree. Commun Stat Stoch Model [Internet]. 1998; 14(1–2):273–95. Available from: http://www.tandfonline.com/doi/abs/10.1080/ 15326349808807471
- 36. Slatkin M, Rannala B. Estimating Allele Age. New York [Internet]. 2000; 1:225–49. Available from: http://www.annualreviews.org/doi/abs/10.1146/annurev.genom.1.1.225
- Laitman Y, Feng B-J, Zamir IM, Weitzel JN, Duncan P, Port D, et al. Haplotype analysis of the 185delAG BRCA1 mutation in ethnically diverse populations. Eur J Hum Genet [Internet]. 2013; 21 (2):212–6. Available from: http://www.nature.com/doifinder/10.1038/ejhg.2012.124 PMID: 22763381
- Ossa CA, Torres D. Founder and Recurrent Mutations in *BRCA1* and *BRCA2* Genes in Latin American Countries: State of the Art and Literature Review. Oncologist [Internet]. 2016; 21(7):832–9. Available from: http://theoncologist.alphamedpress.org/lookup/doi/10.1634/theoncologist.2015-0416 PMID: 27286788
- 39. Kronn D, Oddoux C, Phillips J, Ostrer H. Prevalence of Canavan disease heterozygotes in the New York metropolitan Ashkenazi Jewish population. Am J Hum Genet [Internet]. 1995; 57(5):1250–2. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1801394&tool= pmcentrez&rendertype=abstract PMID: 7485179
- Pastinen T. Dissecting a population genome for targeted screening of disease mutations. Hum Mol Genet [Internet]. 2001; 10(26):2961–72. Available from: https://academic.oup.com/hmg/article-lookup/ doi/10.1093/hmg/10.26.2961 PMID: 11751678
- 41. Zeegers MPA, Van Poppel F, Vlietinck R, Spruijt L, Ostrer H. Founder mutations among the Dutch. Found Mutat Inherit Card Dis Netherlands. 2014;(October 2003):3–12.
- Ponti G, Castellsagué E, Ruini C, Percesepe A, Tomasi A. Mismatch repair genes founder mutations and cancer susceptibility in Lynch syndrome. Clin Genet. 2015; 87(6):507–16. <u>https://doi.org/10.1111/</u> cge.12529 PMID: 25345868
- Ostrer H. A genetic profile of contemporary Jewish populations. Nat Rev Genet. 2001; 2(11):891–8. https://doi.org/10.1038/35098506 PMID: 11715044
- 44. Campos-Mollo E, Sanchez-Sanchez F, Lopez-Garrido MP, Lopez-Sanchez E, Lopez-Martinez F, Escribano J. MYOC gene mutations in Spanish patients with autosomal dominant primary open-angle glaucoma: a founder effect in southeast Spain. Mol Vis. 2007; 13(September):1666–73.
- Brézin AP, Adam MF, Belmouden A, Lureau MA, Chaventré A, Copin B, et al. Founder effect in GLC1A-linked familial open-angle glaucoma in Northern France. Am J Med Genet. 1998; 76(5):438– 45. PMID: 9556305
- Duchesne A, Faucher M, Anctil J, Bergeron D, Blondeau P, Co G, et al. Founder TIGR / myocilin mutations for glaucoma in ' bec population the Que. 2002; 11(18):2077–90.
- **47.** Baird PN, Craig JE, Rochtchina E, Stat M. The Q368STOP Myocilin Mutation in a Population-based Cohort: The Blue Mountains Eye Study.

- Baird PN, Craig JE, Richardson AJ, Ring Ma, Sim P, Stanwix S, et al. Analysis of 15 primary openangle glaucoma families from Australia identifies a founder effect for the Q368STOP mutation of myocilin. Hum Genet [Internet]. 2003; 112(2):110–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/ 12522550 PMID: 12522550
- Slatkin M. A Population-Genetic Test of Founder Effects and Implications for Ashkenazi Jewish Diseases. Am J Hum Genet. 2004; 75:282–93. https://doi.org/10.1086/423146 PMID: 15208782
- Reis JJ. Presença negra: conflitos e encontros. Instituto Brasileiro de Geografia Estatística (2000) Brasil: 500 Anos de Povoamento. 2000. 80–99 p.
- Salzano, Francisco M.; Freire-Maia N. Populações brasileiras: aspestos demogràficos, genéticos e antropológicos [por] FM Salzano [e] N. Freire-Maia. Cia. Ed. Nacional, Ed. da Universidade, 1967. SAL-ZANO, Francisco M.; FREIRE-MAIA, Newton. 1967;1967.
- Levy MS. O papel da migração internacional na evolução da população brasileira (1872 a 1972). Rev Saude Publica. 1974; Suppl:49–90.
- Greenwood CMT, Sun S, Veenstra J, Hamel N, Niell B, Gruber S, et al. How old is this mutation?—A study of three Ashkenazi Jewish founder mutations. BMC Genet. 2010; 11(1990).
- 54. Clendenning M, Baze ME, Sun S, Walsh K, Liyanarachchi S, Fix D, et al. Origins and prevalence of the American Founder Mutation of MSH2. Cancer Res. 2008; 68(7):2145–53. <u>https://doi.org/10.1158/0008-5472.CAN-07-6599</u> PMID: 18381419
- 55. Biberg-Salum TG, Rodrigues MDLV. Ocular health of brazilian indigenous populations. Med. 2016; 49 (3):265–72.
- Kehdy FSG, Gouveia MH, Machado M, Magalhães WCS, Horimoto AR. Origin and dynamics of admixture in Brazilians and its effect on the pattern of deleterious mutations. 2015; 112(28):8696–701.
- 57. Giolo SR, Soler JMP, Greenway SC, Almeida MAA, De Andrade M, Seidman JG, et al. Brazilian urban population genetic structure reveals a high degree of admixture. Eur J Hum Genet. 2012; 20(1):111–6. https://doi.org/10.1038/ejhg.2011.144 PMID: 21863058