Short Paper

Molecular detection of *Brucella abortus* and *Brucella melitensis* in domestic ruminants and their ticks in selected areas of western Iran

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Abstract

Background: Brucellosis is one of the most important zoonotic diseases caused by Gram-negative bacteria belonging to the genus *Brucella*. Detection of *Brucella* species in different countries is of utmost importance. Aims: This study aimed to detect *Brucella abortus* and *Brucella melitensis* in domestic ruminant blood samples and their ticks in western Iran. Methods: Sampling was conducted on ruminants from August to September 2020 in four different counties of Kurdistan Province, including Divandareh, Marivan, Baneh, and Sanandaj. Totally, 250 blood samples were collected from 250 small ruminants. There were no ticks on the skin of six (2.4%) ruminants, and 244 ticks were isolated from 244 animals. After genomic DNA extraction from all the collected samples, quantitative polymerase chain reaction (qPCR) was performed to detect IS711 gene. Results: Based on qPCR results, *Brucella* genus was detected in two blood samples (0.8%) from female sheep and four ticks (1.6%) from male sheep, including three Dermacentor marginatus (1.22%) and one *Rhipicephalus turanicus* (0.4%). Although *B. melitensis* was not detected in any tick or blood sample, one tick sample (*D. marginatus*) was positive for *B. abortus*. Conclusion: Considering the positivity of ticks for brucellosis in this study, there is a possibility of *Brucella* transmission from infected ticks to humans and animals through tick bites, nevertheless, in order to identify the *Brucella* transmission relationship between ticks and animals, serological tests should be used in future studies.

Key words: B. abortus, B. melitensis, Dermacentor marginatus, Rhipicephalus turanicus, Ruminant

Introduction

Brucellosis is one of the most important zoonotic diseases with high economic and health impacts (Rossetti

et al., 2017). The genus *Brucella* is classified into 12 known species, 9 terrestrials, and three marine species (Whatmore *et al.*, 2016). *Brucella melitensis* and *Brucella suis* (except for Biovar 2), followed by *Brucella*

abortus and Brucella canis, are the most dangerous Brucella species (Wakjira and Dilba, 2010). Humans can be infected by contaminated aerosol particles, undercooked meat, unpasteurized milk, and direct contact with infected animals (Ahmed and Ibrahim, 2020; Islam et al., 2023). Additionally, human-to-human brucellosis transmission can occur in infected mothers who breastfeed their infants, sexual transmission, and unusual transmission via tissue transplantation or blood transfusions (Xu et al., 2023). In susceptible animals, brucellosis is most commonly transmitted through direct contact with infected animals or through an environment contaminated by discharge from infected animals (Pal et al., 2020). Cattle, dogs, sheep, goats, and pigs are major reservoirs of Brucella species for human infection. Brucellosis is prevalent in Mediterranean countries, Western Asia, and some parts of Africa and Latin America (Islam et al., 2023). Brucellosis is more prevalent in countries without effective disease control programs (Dadar et al., 2022). It is an endemic disease in many parts of Iran, especially in areas where people are in close contact with animals. According to the Iranian Ministry of Health, there are 22 cases of brucellosis per 100,000 people in Iran (Delam et al., 2022; Zeinali et al., 2022). Arthropods, particularly ticks, spread diseases between humans and animals (Cull, 2021). Despite the research on the prevalence of brucellosis in humans and animals, no study is conducted on Brucella species in ruminant ticks in Iran. There are many tick genera and species, including Dermacentor, Haemaphysalis, and Rhipicephalus, which carry microbes, including Brucella, that transmit tick-borne brucellosis (Wang et al., 2018, 2019; Huang et al., 2020; Jia et al., 2022). Detecting Brucella in reservoirs and nonspecific hosts (such as ticks) provides valuable information regarding the pathogenesis of brucellosis and facilitates effective control measures (Wang et al., 2019). Due to the higher sensitivity and specificity of molecular-based methods, such as quantitative polymerase chain reaction (qPCR), compared to culture and serological tests, these methods have a critical role in the detection and typing of *Brucella* species particularly *B. abortus* and *B. melitensis* for the control and eradication of brucellosis in ruminants. Despite serological evidence in humans and livestock, no molecular data is available on *Brucella* species in Kurdistan, an endemic region of the disease in western Iran. The aim of this study was to detect *B. abortus* and *B. melitensis* in ruminants and ticks of this region by molecular method.

Materials and Methods

Sample collection

Sampling was conducted from August to September 2020 in four different counties of Kurdistan province, including Divandareh, Marivan, Baneh, and Sanandaj. This study was approved by the Ethics Committee of the Pasteur Institute of Iran (IR.PII.REC.1398.051) and all methods were performed in accordance with relevant guidance and regulations.

Blood and tick samples were collected from two hundred and fifty ruminants including sheep and goats. All samples were collected with previous consent. All animals were clinically healthy, without a history of acaricide or antibiotic therapy. Flocks and animals were randomly selected. The mean age (\pm SD) of sheep and goats was 3.31 (\pm 1.4) and 4.78 (\pm 1.8), respectively (Table 1). Five ml of blood was collected in Venoject tubes containing an anticoagulant (EDTA). Blood samples were collected in EDTA-containing tubes and stored at 4-8°C until they were transferred to the laboratory and then stored at 4°C until DNA extraction. The skin surfaces of the animals were examined for tick

Table 1: Prevalence of *Brucella* in ticks based on tick species, tick gender, livestock species, and the geographical location of the sample collection

Category and subcategory	No. of ticks (%)	No. of ticks positive for <i>Brucella</i> genus (%)	No. of ticks positive for <i>Brucella abortus</i> (%)	No. of ticks positive for <i>Brucella melitensis</i> (%)		
Ticks species						
Dermacentor marginatus	164 (67.21)	3 (1.22)	1 (0.4)	0		
Rhipicephalus turanicus	30 (12.30)	1 (0.4)	0	0		
Rhipicephalus sanguineus	26 (10.66)	0	0	0		
Haemaphysalis concinna	24 (9.83)	0	0	0		
Total	244	4 (1.6)	1 (0.4)	0		
Ticks gender						
Female	119 (48.8)	0	0	0		
Male	125 (51.23)	4 (1.6)	1 (0.4)	0		
Total	244 (100)	4 (1.6)	1 (0.4)	0		
Livestock's species						
Sheep	229 (93.85)	4 (1.6)	1 (0.4)	0		
Goat	15 (6.15)	0				
Total	244 (100)	4 (1.6)	1 (0.4)	0		
Geographical location (cities in Kurdistan province)						
Baneh	11 (4.51)	0 (0)	0	0		
Divandareh	185 (75.82)	3 (1.22)	1 (0.4)	0		
Marivan	46 (18.85)	0	0	0		
Sanandaj	2 (0.82)	1 (0.4)	0	0		
Total	244 (100)	4 (1.6)	1 (0.4)	0		

infestation after blood sampling and animals were included in the study if they were infested. One tick per animal was randomly removed using fine-pointed stainless-steel tweezers. Each tick was placed in a separate 2 ml tube, and 70% ethanol was added. Tick samples were transferred to the parasitology laboratory of the Veterinary Faculty of Islamic Azad University of Sanandaj. The collected ticks were examined separately in this study. A parasitologist determined the genus and species of the collected ticks according to the classification and morphological keys previously described for identifying the ticks (Lah et al., 2016; Balinandi et al., 2020). All blood and tick samples collected from the animals were transferred to the Pasteur Institute of Iran (National Reference Laboratory for plague, tularemia, and Q fever) for further experiments under cold conditions (Bounaadja et al., 2009).

DNA extraction

Before DNA was extracted from the ticks, the salivary glands and sexual organs of the ticks were isolated and transferred to sterile microtubes. The samples were dried and crushed in liquid nitrogen in a clean Chinese pounder. Genomic DNA was extracted using G-spin[™] Total DNA Extraction Mini kit (iNtRON Biotechnology, Korea) according to the manufacturer's instructions. Total DNA was extracted from each tick sample. Genomic DNA was extracted from blood samples using a commercial kit (FavorPrep Blood/Culture Cell Genomic DNA Extraction Mini Kit, Favragen Biotech, Taiwan) according to the manufacturer's recommendations. For quality control, the extracted DNA samples were confirmed by gel electrophoresis and stored at -20°C for further experiments (Bounaadja et al., 2009).

Detection of *Brucella* genus using qPCR

The qPCR was performed for the detection of the IS711 gene in blood and tick samples using the previously described specific probe (FAM-AAG CCA ACA CCC GGC CAT TAT GGT-TAMRA) and primers (F: GCT TGA AGC TTG CGG ACA GT; R: GGC CTA CCG CTG CGA AT) on the Corbett 6000 Rotor-Gene system (Corbett, Victoria, Australia) (Hinić *et al.*, 2008). The PCR mixture (final volume, 20 μ L contained 10 μ L of 2X Real Q Plus Master Mix for Probe (Ampliqon, Denmark), 1 μ L of a mixture of probe (200 nM), forward and reverse primers (900 nM), 4 μ L of extracted DNA, and 5 μ L of double-distilled water (DDW). DDW was

used as a negative control, and the *Brucella*-positive sample from the Biobank of the Pasteur Institute of Iran was used as a positive control. The amplification protocol was 10 min at 95°C, followed by 45 cycles of 15 s at 95°C, and 60 s at 60°C. The qPCR results were analyzed using the Rotor-Gene[®] Q 2.3.5 software (QIAGEN).

Identification of *B. abortus* and *B. melitensis* using qPCR

B. abortus and *B. melitensis* were detected using qPCR (Dadar *et al.*, 2019). Primer sequences for the identification of this species are shown in Table 2.

Statistical analysis

Descriptive statistical analysis of the collected data was performed using the statistical SPSS software (version 26).

Results

Sample characteristics

A total of 250 blood samples were collected from small ruminants in four different counties of Kurdistan province: Divandareh (185/250, 74%), Marivan (46/250, 18.4%), Baneh (15/250, 6%), and Sanandaj (4/250, 1.6%). Among 250 ruminants, (233/250, 93.2%) and (17/250, 6.8%) were female and male, respectively. The ruminants selected in this study included 232 sheep (92.8%) and 18 goats (7.2%). There were no ticks on the skin of six ruminants (2.4%), and 244 ticks were isolated from 244 animals. Based on morphological studies, collected ticks included Dermacentor marginatus (67.21%), followed by Rhipicephalus turanicus (12.30%), Rhipicephalus sanguineus (10.66%), and Haemaphysalis concinna (9.83%). All the collected ticks were adults, and 51.2% were male (Table 1 and Fig. 1).

The prevalence of *Brucella* genus in ticks and blood samples of small ruminants

The *Brucella* genus was detected in two blood samples (0.8%) of female sheep, and four (1.6%) tick samples of male sheep, including three *D. marginatus* (1.22%), and one *R. turanicus* (0.4%) (Fig. 2).

The prevalence of *B. abortus* and *B. melitensis* in tick and blood samples of small ruminants

B. abortus and B. melitensis were not detected in blood samples. Additionally, all tick samples were

Table 2: The probe and primers used for the identification of B. abortus and B. melitensis using qPCR in the present study

Bacteria	Target sequence		The sequence of primers $(5^{\prime} \rightarrow 3^{\prime})$	Size fragments (bp)
B. melitensis	BMEII0466	Forward Reverse Probe	TCGCATCGGCAGTTTCAA CCAGCTTTTGGCCTTTTCC TEX-CCTCGGCATGGCCCGCAA-BHQ-2	67
B. abortus	BruAb2_0168	Forward Reverse Probe	GCACACTCACCTTCCACAACAA CCCCGTTCTGCACCAGACT HEX-TGGAACGACCTTTGCAGGCGAGATC-BHQ-1	81



Fig. 1: The collected ticks from small ruminants in Kurdistan province in 2020. A: Dermacentor marginatus, B: Rhipicephalus turanicus, C: Rhipicephalus sanguineus, and D: Haemaphysalis concinna



Fig. 2: Detection of *Brucella* genus using qPCR. The amplifiaction curve of positive and negative samples and positive and negative controlos are showed by arrows

negative for *B. melitensis*, and only one *D. marginatus* (0.4%) of male sheep was positive for *B. abortus*. The relationships between positive brucellosis and tick species, tick gender, livestock species, and the geographical location of the sample collection are presented in Table 1.

Discussion

In the present study, Brucella genus was detected in 1.6% (4/244) of ticks and 0.8% (2/250) of blood samples from western Iran (Kurdistan province) using qPCR. Although B. melitensis was not detected in any tick or blood sample, one tick sample (D. marginatus) was positive for B. abortus. In animals, after a short period of bacteremia, Brucella is localized in specific organs, such as the mammary glands and genital organs of the animal, and in this way, it finds shedding in the environment. Therefore, it is possible to find positive ticks for Brucella but negative in host blood. Brucellosis is commonly caused by B. melitensis and B. abortus in Iran (Golshani and Buozari, 2017). The prevalence of B. abortus and B. melitensis in ticks from domestic ruminants in Iran has not been studied despite warnings of a potential outbreak (Golshani and Buozari, 2017; Zeinali et al., 2022). B. abortus is widespread in Latin America, the Middle East, Africa, and Asia which results in significant economic losses and health problems in humans and animals (Franc et al., 2018). Lack of strict biosafety measures can result in failure to control the disease.

Recently researchers have investigated the role of ectoparasites in the spreading of Brucella species (Saeed et al., 2019). Identification and isolation of Brucella species are important for brucellosis management and control measures (Mu et al., 2022). In addition to Brucella, ticks can also be host pathogenic, nonpathogenic, or endosymbiotic organisms (Zeng et al., 2022). Changes in weather and land use have contributed to the growth of hard ticks and ectoparasites, which transmit microorganisms. Brucellosis may be transmitted naturally by Boophilus, which can be the host of Brucella species (Hosseini-Chegeni et al., 2017). Dermacentor and Rhipicephalus ticks have gained attention because of their geographic distribution, vectorial role, and identification as carriers of *B. abortus* (Zhao et al., 2021). To assess the significance of Dermacentor and Rhipicephalus in Brucella transmission, qPCR has been conducted in various studies (Zhao et al., 2021).

This is the first molecular study to detect of B. abortus and B. melitensis in domestic ruminant blood samples and their ticks in western Iran. Ticks are thought to transmit Brucella pathogenic species such as B. abortus and B. melitensis, but few studies have explored this issue. In China, a significant Brucella infection has been identified in ticks, which can be considered as a potential vector of this pathogen. Among Dermacentor nuttalli ticks in different regions of China, infection rates range from 0-87% (Huang et al., 2020). Other studies on ticks in China reported B. abortus and B. melitensis infection rates of 4.6% in D. marginatus ticks, 24.26% in Haemaphysalis longicornis ticks, and 26.3% in Haemaphysalis anatolicum and D. nuttalli ticks (Li et al., 2020; Zhang et al., 2021). In the present study, the Brucella genus was detected in two blood samples (0.8%) from female sheep, and four ticks (1.6%) from male sheep, including three D. marginatus (1.22%) and one R. turanicus (0.4%). All Brucella genus positive blood and tick samples were negative for B. melitensis, and B. abortus was detected in D. marginatus. The results of the present study are in agreement with those of Khan et al. (2017) who isolated B. abortus from ticks in Pakistan. The detection of Brucella in livestock blood samples and their ticks suggests the possibility of transmission of brucellosis to livestock and humans through tick bites. Based on these results, ticks may have

a role in spreading B. abortus and B. melitensis throughout Iran. Additionally, ticks may transmit Brucella species horizontally and vertically (Omitola and Taylor-Robinson, 2020). According to a previous research (Egyed et al., 2012) tick proximity to infected animals is correlated with the prevalence of tick-borne pathogen infections. Tick control in ruminants may reduce the incidence of the disease, indicating that ticks are brucellosis vectors in domestic animal populations (Miller et al., 2016). In a study performed in Iran, it was found that 38.42% of the examined samples were infected with Brucella, and the species found in this study belonged to B. abortus (Dadar et al., 2019). The Brucella can be transmitted to humans by tick biting, however, in vitro studies are needed to fully understand its mechanism. To prevent Brucella from spreading through tick bites, animals should be immersed in acaricide (Namgyal et al., 2021). Further research is needed to clarify how tick bites spread bacteria. The consistency of transmission of infection from D. marginatus to domesticated animals and humans should also be assessed.

Due to the existence of the genus Brucella in ticks of Kurdistan province, there is a possibility that infection can be transmitted through tick bites to livestock and humans. In addition, ticks are vectors for numerous pathogens, including Brucella species. Thus, tick control is important to prevent the spread of brucellosis. This study shows that D. marginatus and R. turanicus can carry the genus Brucella. In addition, blood samples from animals in which Brucella-infected ticks were isolated were negative. Considering the positivity of ticks for brucellosis in this study, there is a possibility of Brucella transmission from infected ticks to humans and animals through tick bites, nevertheless, in order to identify the Brucella transmission relationship between ticks and animals, serological tests should be used in future studies.

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Conflict of interest

The authors declare no conflict of interest.

References

- Ahmed, ME; Ibrahim, A and Abd El-Rahman, MM (2020). Microbiological and immunological studies on brucellosis in a hospital in Al-Madinah Al-Munawarah. JSRMBS., 1: 24-44.
- Balinandi, S; Chitimia-Dobler, L; Grandi, G; Nakayiki, T; Kabasa, W; Bbira, J; Lutwama, J; Bakkes, D; Malmberg, M and Mugisha, L (2020). Morphological and

molecular identification of ixodid tick species (Acari: Ixodidae) infesting cattle in Uganda. Parasitol. Res., 119: 2411-2420.

- Bounaadja, L; Albert, D; Chénais, B; Hénault, S; Zygmunt, MS; Poliak, S and Garin-Bastuji, B (2009). Real-time PCR for identification of *Brucella* spp: a comparative study of IS711, bcsp31 and per target genes. Vet. Microbiol., 137: 156-164.
- **Cull, B** (2021). Potential for online crowdsourced biological recording data to complement surveillance for arthropod vectors. PLoS One. 16: e0250382.
- Dadar, M; Alamian, S; Behrozikhah, A; Yazdani, F; Kalantari, A; Etemadi, A and Whatmore, AM (2019). Molecular identification of *Brucella* species and biovars associated with animal and human infection in Iran, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran. Ret. Ves. Forum., 10: 315-321.
- Dadar, M; Tabibi, R; Alamian, S; Caraballo-Arias, Y; Mrema, EJ; Mlimbila, J; Chandrasekar, S; Dzhusupov, K; Sulaimanova, C and Alekesheva, LZ (2022). Safety concerns and potential hazards of occupational brucellosis in developing countries: A review. J. Public Health. 31: 1681-1690.
- Delam, H; Keshtkaran, Z; Rezaei, B; Soufi, O and Bazrafshan, MR (2022). Changing patterns in epidemiology of brucellosis in the South of Iran (2015-2020): Based on Cochrane-Armitage Trend test. Ann. Glob. Health. 88: 11, 1-8.
- Egyed, L; Elo, P; Sreter-Lancz, Z; Szell, Z; Balogh, Z and Sreter, T (2012). Seasonal activity and tick-borne pathogen infection rates of Ixodes ricinus ticks in Hungary. Ticks Tick Borne Dis., 3: 90-94.
- Franc, K; Krecek, R; Häsler, B and Arenas-Gamboa, A (2018). Brucellosis remains a neglected disease in the developing world: a call for interdisciplinary action. BMC Public Health. 18: 1-9.
- **Golshani, M and Buozari, S** (2017). A review of brucellosis in Iran: epidemiology, risk factors, diagnosis, control, and prevention. Iran Biomed. J., 21: 349-359.
- Hinić, V; Brodard, I; Thomann, A; Cvetnić, Ž; Makaya, P; Frey, J and Abril, C (2008). Novel identification and differentiation of *Brucella* melitensis, *B. abortus*, *B. suis*, *B. ovis*, *B. canis*, and *B. neotomae* suitable for both conventional and real-time PCR systems. J. Microbiol. Methods. 75: 375-378.
- Hosseini-Chegeni, A; Tavakoli, M; Telmadarraiy, Z; Sedaghat, MM and Faghihi, F (2017). Detection of a *Brucella*-like (Alphaproteobacteria) bacterium in boophilus spp. (Acari: Ixodidae) from Iran. JoMMID., 5: 66-68.
- Huang, T; Zhang, J; Sun, C; Liu, Z; He, H and Wu, J (2020). A novel arthropod host of brucellosis in the arid steppe ecosystem. Front. Vet. Sci., 7: 566253.
- Islam, MS; Islam, MA; Rahman, MM; Islam, K; Islam, MM; Kamal, MM and Islam, MN (2023). Presence of *Brucella* spp. in milk and dairy products: A comprehensive review and its perspectives. J. Food Qual., 2932883: 1-19.
- Jia, W; Chen, S; Chi, S; He, Y; Ren, L and Wang, X (2022). Recent progress on tick-borne animal diseases of veterinary and public health significance in China. Viruses. 14: 355.
- Khan, MZ; Usman, T; Sadique, U; Qureshi, MS; Hassan, MF; Shahid, M and Khan, A (2017). Molecular characterization of *Brucella abortus* and *Brucella melitensis* in cattle and humans at the North West of Pakistan. Pak. Vet. J., 37: 360-363.
- Lah, EFC; Yaakop, S; Ahamad, M; George, E and Nor, SM (2016). Precise identification of different stages of a tick, Ixodes granulatus Supino, 1897 (Acari: Ixodidae). Asian

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Pac. J. Trop. Biomed., 6: 597-604.

- Li, Y; Wen, X; Li, M; Moumouni, PFA; Galon, EM; Guo, Q; Rizk, MA; Liu, M; Li, J and Ji, S (2020). Molecular detection of tick-borne pathogens harbored by ticks collected from livestock in the Xinjiang Uygur Autonomous Region, China. Ticks Tick Borne Dis., 11: 101478.
- Miller, R; Nakavuma, J; Ssajjakambwe, P; Vudriko, P; Musisi, N and Kaneene, J (2016). The prevalence of brucellosis in cattle, goats and humans in rural Uganda: a comparative study. Transbound Emerg. Dis., 63: e197e210.
- Mu, J; Li, Q; Yan, X; Mao, X; Shi, Y; Qin, Y; Liu, C and Wang, W (2022). Detection of *Brucella* S2 vaccine strain by a loop-mediated isothermal amplification (LAMP) method. Front. Cell. Infect., 12: 1023243.
- Namgyal, J; Tenzin, T; Checkley, S; Lysyk, TJ; Rinchen, S; Gurung, RB; Dorjee, S; Couloigner, I and Cork, SC (2021). A knowledge, attitudes, and practices study on ticks and tick-borne diseases in cattle among farmers in a selected area of eastern Bhutan. PloS One. 16: e0247302.
- **Omitola, OO and Taylor-Robinson, AW** (2020). Emerging and re-emerging bacterial zoonoses in Nigeria: current preventive measures and future approaches to intervention. Heliyon. 6: e04095.
- Pal, M; Kerorsa, GB; Desalegn, C and Kandi, V (2020). Human and animal brucellosis: A comprehensive review of biology, pathogenesis, epidemiology, risk factors, clinical signs, laboratory diagnosis, public health significance, economic importance, prevention and control. Am. J. Infect. Dis., 8: 118-126.
- Rossetti, CA; Arenas-Gamboa, AM and Maurizio, E (2017). Caprine brucellosis: A historically neglected disease with significant impact on public health. PloS Negl. Trop. Dis., 11: e0005692.
- Saeed, U; Ali, S; Khan, TM; El-Adawy, H; Melzer, F; Khan, AU; Iftikhar, A and Neubauer, H (2019). Seroepidemiology and the molecular detection of animal brucellosis in Punjab, Pakistan. Microorganisms. 7: 449.

- Wakjira, BS and Dilba, GM (2010). Review on serological and molecular diagnostics techniques of brucellosis. Croat. Med. J., 51: 320-326.
- Wang, Q; Zhao, S; Lu, J and Wang, Y (2019). Transmission of tick-borne *Brucella*. J. Pathog. Biol., 14: 238-243.
- Wang, Q; Zhao, S; Wureli, H; Xie, S; Chen, C; Wei, Q; Cui, B; Tu, C and Wang, Y (2018). Brucella melitensis and B. abortus in eggs, larvae and engorged females of Dermacentor marginatus. Ticks Tick Borne Dis., 9: 1045-1048.
- Whatmore, AM; Koylass, MS; Muchowski, J; Edwards-Smallbone, J; Gopaul, KK and Perrett, LL (2016). Extended multilocus sequence analysis to describe the global population structure of the genus *Brucella*: phylogeography and relationship to biovars. Front Microbiol., 7: 2049.
- Xu, N; Qu, C; Sai, L; Wen, S; Yang, L; Wang, S; Yang, H; Liu, H and Wang, G (2023). Evaluating the efficacy of serological testing of clinical specimens collected from patients with suspected brucellosis. PLoS Negl. Trop. Dis., 17: e0011131.
- Zeinali, M; Doosti, S; Amiri, B; Gouya, MM and Godwin, GN (2022). Trends in the epidemiology of brucellosis cases in Iran during the last decade. Iran. J. Public Health. 51: 2791-2798.
- Zeng, W; Li, Z; Jiang, T; Cheng, D; Yang, L; Hang, T; Duan, L; Zhu, D; Fang, Y and Zhang, Y (2022). Identification of bacterial communities and tick-borne pathogens in *Haemaphysalis* spp. collected from Shanghai, China. Infect. Dis. Trop. Med., 7: 413.
- Zhang, K; Li, A; Wang, Y; Zhang, J; Chen, Y; Wang, H; Shi, R and Qiu, Y (2021). Investigation of the presence of Ochrobactrum spp. and Brucella spp. in Haemaphysalis longicornis. Ticks Tick Borne Dis., 12: 101588.
- Zhao, GP; Wang, YX; Fan, ZW; Ji, Y; Liu, MJ; Zhang, WH; Li, XL; Zhou, SX; Li, H and Liang, S (2021). Mapping ticks and tick-borne pathogens in China. Nat. Commun., 12: 1075.