

Research article

Open Access

Linking mechanistic and behavioral responses to sublethal esfenvalerate exposure in the endangered delta smelt; *Hypomesus transpacificus* (Fam. Osmeridae)

Richard E Connon*^{†1}, Juergen Geist^{†1,4}, Janice Pfeiff^{†2},
Alexander V Loguinov^{†3}, Leandro S D'Abronzio^{†1}, Henri Wintz^{3,5},
Christopher D Vulpe³ and Inge Werner¹

Address: ¹School of Veterinary Medicine, Department of Anatomy, Physiology and Cell Biology, University of California, Davis, California 95616, USA, ²School of Veterinary Medicine, Molecular Biosciences, University of California, Davis, California 95616, USA, ³School of Nutritional Sciences and Toxicology, University of California, Berkeley, California 94720, USA, ⁴Unit of Functional Aquatic Ecology and Fish Biology, Department of Animal Science, Technische Universität München, D-85350 Freising, Germany and ⁵Biorad Laboratories, Life Science Research, Hercules, California, USA

Email: Richard E Connon* - reconnon@ucdavis.edu; Juergen Geist - juergen.geist@wzw.tum.de; Janice Pfeiff - jmpfeiff@ucdavis.edu; Alexander V Loguinov - Avl53@aol.com; Leandro S D'Abronzio - lsdabronzo@ucdavis.edu; Henri Wintz - henri.wintz@gmail.com; Christopher D Vulpe - vulpe@berkeley.edu; Inge Werner - iwerner@ucdavis.edu

* Corresponding author †Equal contributors

Published: 15 December 2009

Received: 25 February 2009

BMC Genomics 2009, 10:608 doi:10.1186/1471-2164-10-608

Accepted: 15 December 2009

This article is available from: <http://www.biomedcentral.com/1471-2164/10/608>

© 2009 Connon et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: The delta smelt (*Hypomesus transpacificus*) is a pelagic fish species listed as endangered under both the USA Federal and Californian State Endangered Species Acts and considered an indicator of ecosystem health in its habitat range, which is limited to the Sacramento-San Joaquin estuary in California, USA. Anthropogenic contaminants are one of multiple stressors affecting this system, and among them, current-use insecticides are of major concern. Interrogative tools are required to successfully monitor effects of contaminants on the delta smelt, and to research potential causes of population decline in this species. We have created a microarray to investigate genome-wide effects of potentially causative stressors, and applied this tool to assess effects of the pyrethroid insecticide esfenvalerate on larval delta smelt. Selected genes were further investigated as molecular biomarkers using quantitative PCR analyses.

Results: Exposure to esfenvalerate affected swimming behavior of larval delta smelt at concentrations as low as 0.0625 µg.L⁻¹, and significant differences in expression were measured in genes involved in neuromuscular activity. Alterations in the expression of genes associated with immune responses, along with apoptosis, redox, osmotic stress, detoxification, and growth and development appear to have been invoked by esfenvalerate exposure. Swimming impairment correlated significantly with expression of aspartoacylase (ASPA), an enzyme involved in brain cell function and associated with numerous human diseases. Selected genes were investigated for their use as molecular biomarkers, and strong links were determined between measured downregulation in ASPA and observed behavioral responses in fish exposed to environmentally relevant pyrethroid concentrations.

Conclusions: The results of this study show that microarray technology is a useful approach in screening for, and generation of molecular biomarkers in endangered, non-model organisms, identifying specific genes that can be directly linked with sublethal toxicological endpoints; such as changes in expression levels of neuromuscular genes resulting in measurable swimming impairments. The developed microarrays were successfully applied on larval fish exposed to esfenvalerate, a known contaminant of the Sacramento-San Joaquin estuary, and has permitted the identification of specific biomarkers which could provide insight into the factors contributing to delta smelt population decline.

Background

The delta smelt (*Hypomesus transpacificus*) is a pelagic fish species endemic to the Sacramento-San Joaquin estuary, whose abundance has dramatically declined since the 1980s, and more precipitously in recent years [1-4]. It was listed as endangered in 2009, under both the Federal Endangered Species Act (ESA) and California Endangered Species Act (CESA). Considerable efforts are presently being made to understand the causes of this recent population decline [4,5], especially because several other pelagic species have shown similar population trends. Delta habitats have been compromised by a number of complex factors, both known and unknown, potentially affecting aquatic species throughout the Sacramento-San Joaquin watersheds and estuary [4]. Pollution, in the form of chemicals contained in runoff from agricultural and urban areas, and old mining sites, treated wastewater effluent, along with the effects of water exports, invasive species and habitat destruction are amongst potential causes for the population decline of several pelagic species [5].

Identifying the sublethal impacts of environmental stressors and their mechanistic effects on resident individuals and populations is a major challenge in ecotoxicology. Contaminants may not only affect organism survival, but can compromise ecological fitness of individual species via sublethal physiological, behavioral or immunological effects (e.g. [6-10]), consequently altering food web and ecosystem dynamics. However, such physiological endpoints are often difficult to determine in field studies, because they either require behavioral observation and measurements, or because affected individuals will not survive in the wild. Similarly, widely used ecotoxicological tools such as standard toxicity tests [11,12] cannot easily be adapted to resident species of concern, and, conversely, it is problematic to extrapolate test results obtained with surrogate species to resident species of concern [13]. Recent comparative studies have demonstrated a need for identifying effects directly in the species of concern, as traditional model organisms may differ in sensitivity and physiological response to environmental contaminants and other stressors [14,15].

Carefully selected molecular biomarkers can provide species-specific and sensitive, mechanistic information on the overall health of an organism, as toxic responses are often preceded by alterations in gene expression [16,17]. In particular microarray gene profiling is a powerful tool for defining genome-wide effects of environmental change on biological function [16,18,19]. The predictive value of microarrays as screening tools, as well as our understanding of these responses and their application in the field of ecotoxicology is rapidly growing. This technology can be applied in vertebrates and invertebrates, plants, algae, cell lines and unicellular organisms [20]. In addition, links are being established between specific molecular biomarkers identified by microarray technology, and higher-level life history parameters, such as metabolism, growth and reproduction [16,18,21,22]. Gene expression studies carried out over short-term exposures have allowed for the prediction of chronic stressor effects, such as reduced fecundity and embryonic arrest, somatic growth, and population dynamics [16,18,21,23]. Thus, specific gene responses in studied organisms would not only be indicative of health status, but when used in conservation studies, could highlight potential causes for population decline. However, few biomarkers are currently understood well enough to provide conclusive evidence of contaminant impacts on aquatic species in field monitoring, and extrapolating effects seen at the biomarker level to individual or population-level toxicity continues to be a challenge. For molecular biomarkers to be used as successful monitoring tools of individual, population and ecosystem damage, strong links need to continue to be established between gene expression and health status.

To better understand the sublethal effects of contaminants upon *H. transpacificus*, and to identify biomarkers for future field investigations, we have constructed a microarray with 8,448 Expressed Sequence Tags (ESTs). No genomic information was available on any database at the time this project began, other than a few mitochondrial sequences used in taxonomic studies [24]. We describe here, the construction and first application of this tool to identify genes in the delta smelt, specifically responding to exposure to esfenvalerate, a pyrethroid insecticide, and

present gene expression quantitation of selected biomarkers, utilizing these to explain observed swimming abnormalities. We used esfenvalerate in our study because biochemical responses and adverse effects on the whole organisms are relatively well understood [25] and therefore would aid interpretation of results in this "proof of principle" test. Esfenvalerate [(S)-a-cyano-3-phenoxybenzyl-(S)-2-(4-chlorophenyl)-3-methylbutyrate] is a synthetic pyrethroid insecticide, widely used in agriculture, with a high risk to aquatic organisms [26]. It causes neurological damage by blocking sodium and potassium channels, resulting in repetitive neurological discharge [25]. In addition, pyrethroid insecticides are highly soluble in myelin sheaths of nerves, causing demyelination, resulting in conduction deficiencies through nerve lesions [27], directly affecting swimming ability, and impinging on foraging and migration. Fish are highly sensitive to this insecticide, with for example effects on bluegill behavior at measured concentrations as low as 0.025 $\mu\text{g/L}$ [28]. Pyrethroids have also been reported to affect growth, induce immune responses, reduce hepatic glycogen levels and delay spawning [9,29].

The main focus of this study was not the development of the microarray, rather the identification of molecular biomarkers specific to the delta smelt and stressors found in the San Joaquin-Sacramento delta. We present here results from annotated genes identified through microarray analyses and specifically quantitative PCR analyses of selected molecular biomarkers.

Results and Discussion

Effects of esfenvalerate exposure: mortality and swimming behavior

Fish larvae are known to be highly sensitive to esfenvalerate, with effects on swimming performance and enhanced susceptibility to predation resulting from concentrations as low as 0.0625 $\mu\text{g/L}$ [10]. Behavior alterations are construed as being consequential to the reported neurological mode of action of this pesticide, further affecting foraging, migration and reproduction [30]. Toxicity of pyrethroids in the Sacramento-San Joaquin estuary is likely alleviated by the presence of particles and organic matter, and to date concentrations of esfenvalerate detected in the water column were low, however, concentrations in winter storm runoff from agricultural lands have been reported up to 0.093 $\mu\text{g/L}$ [31], influencing our decision to investigate dose response exposures to both high and environmentally relevant concentrations in confirmatory studies.

In terms of mortality, 10-d old delta smelt were only slightly more sensitive in this study ($\text{LC}_{50,24\text{ h}} = 0.19 \mu\text{g.L}^{-1}$) than 52-d old ($\text{LC}_{50,24\text{ h}} = 0.24 \mu\text{g.L}^{-1}$), however swimming performance of the younger larvae was affected at a concentration approximating one third of that observed

affecting older fish (figure 1). Swimming abnormality in 10-d old larvae, intensified with increasing esfenvalerate concentration at 4 h, escalating significantly after 24 h exposure (figure 1a). This swimming abnormality was also concentration dependent in 52-d old fish, however swimming effects resulting from different time point measurements differed only at the highest exposure concentration of 0.250 $\mu\text{g.L}^{-1}$ (figure 1b), where effects on motion increased from 22.5% anomaly at 4 h to 45% at 24 h. Behavioral abnormalities, reduced food intake and growth, as well as increased susceptibility to predation were reported in fathead minnow larvae exposed to esfenvalerate for 4 h to concentrations above 0.455 $\mu\text{g.L}^{-1}$ [10]. Significant swimming impairments were determined in this study at 0.250 $\mu\text{g.L}^{-1}$, thus delta smelt are highly sensitive to sublethal esfenvalerate exposure. Furthermore, bioaccumulation in rainbow trout have resulted in concentrations 400 times higher than background ambient levels <http://extoxnet.orst.edu>.

Microarray application and q-PCR

Through the application of the delta smelt microarrays, and combined analytical methods, we have identified 288 ESTs, from which a number of genes of interest could be used to measure the effect of esfenvalerate and potentially other pyrethroid insecticides; further investigating their use as biomarkers in this species. Of the sequenced ESTs that responded significantly, 118 genes were successfully annotated; 170 matched unnamed or hypothetical proteins, or did not match the described annotation selection criteria; i.e. BLASTx searches resulted in expect-values greater than 1×10^{-5} and scores below 50. Based on gene ontology; molecular function, biological processes and cellular components, 94 unique genes were functionally classified (figure 2 and table 1) and described below. Based on the proportions of ESTs responding to successfully annotated unique genes (33%), it is estimated that of the 8,448 ESTs printed on the microarray, there could be above 2,500 unique genes identified in the delta smelt. These numbers however represent responses to a single contaminant, and should not be construed as final as there will have an intrinsic bias exerted upon them, however, the proportion of repeated sequences in the analyses was very low, with a maximum of nine repetitions for CHK1 checkpoint homologue and not more than one or two duplication for a few others. It is also important to note that the microarray was manufactured with incomplete genome data, thus information presented in figure 2 represents proportions of a limited number of available genes.

Differences between methods used allow for greater mining of possible biomarkers. The method by Loguinov *et al* [32] identified a single differentially expressed gene at 0.125 $\mu\text{g/L}$ esfenvalerate (with no significant homol-

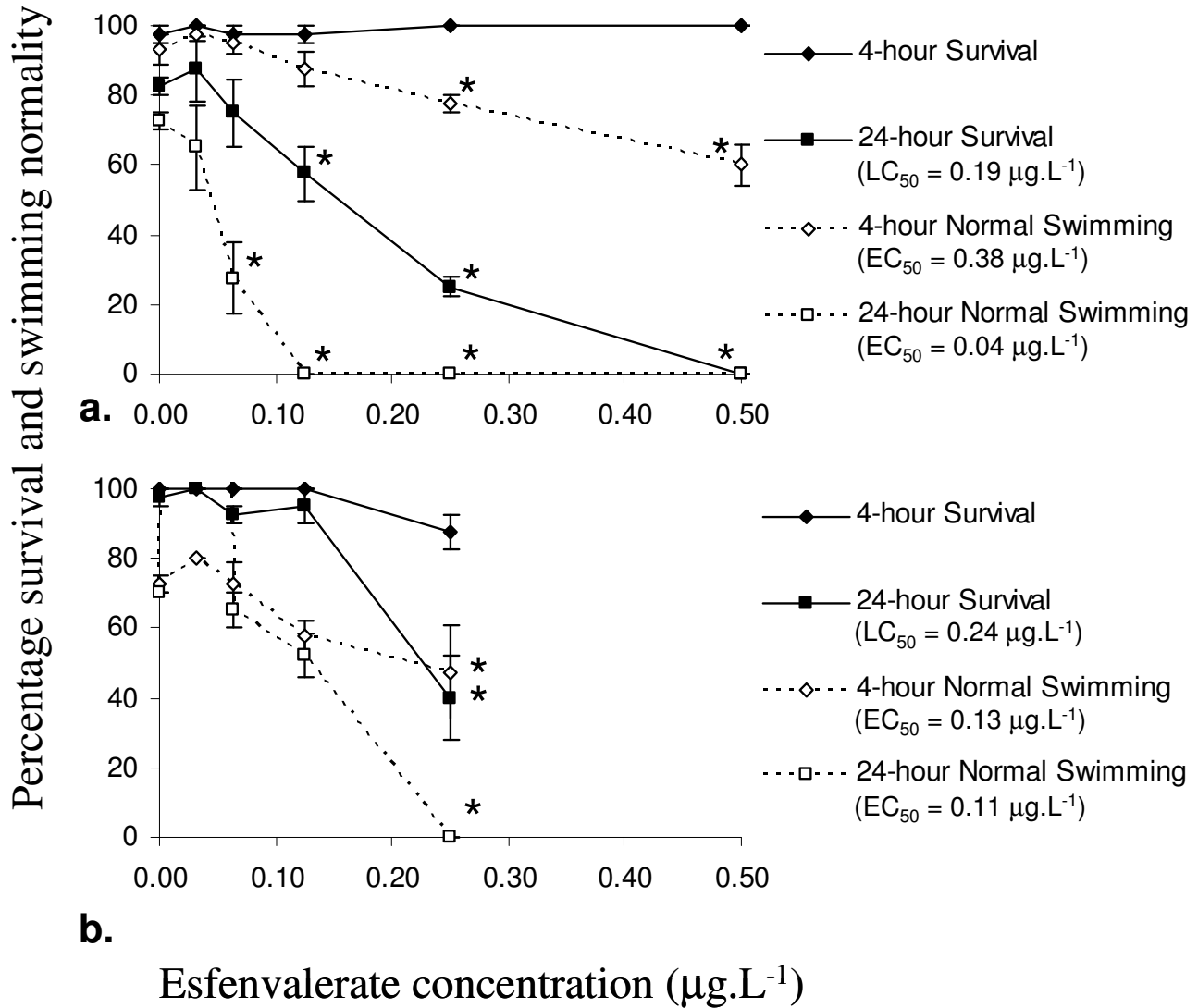


Figure 1
Swimming behavior and mortality. Percentage swimming normality and survival in (a.) 10-d old and (b.) 52-d old *H. transpacificus* exposed to esfenvalerate, ± standard errors (n = 10). * Indicates significant reduction in survival or swimming performance compared to solvent control.

ogy). The addition of LIMMA GUI analysis, also identifying this single gene, generated a broader list of genes for biomarker selection.

A large percentage of uniquely annotated genes, responding to esfenvalerate exposure; 49%, were classified as involved in various biological processes. These included genes encoding for ribosomal proteins, t-RNA synthases,

telomerases, uncoupling proteins and genes involved in chromosome maintenance. Of greater interest was the identification of genes involved in neuromuscular activity; representing 19% of identified sequences, a further 12% eliciting immune responses, along with 6% related to oxidative stress, respiration and iron storage and 5% relevant to apoptosis. Digestion appears to have also been affected, along with growth and development, repre-

Table 1: Classification of differential gene expression of esfenvalerate responding genes in 52-d old *H. transpacificus* following 24 h exposure as identified by microarray analyses.

BLASTX top hit	Species match	Accession No.	Score	E-value	Concentration	Fold Change (+/-)	P-VALUE (or cut-off)	
Neuromuscular								
titin a	<i>Danio rerio</i>	ABG48500	110	4.00E-23	0.0625	1.3277	0.0625	**
smooth muscle cell-specific protein SM22 alpha	<i>Epinephelus coioides</i>	ABW04145	349	1.00E-94	0.1250	1.2252	0.0029	
toxin-I	<i>Oncorhynchus mykiss</i>	AAM21198	116	5.00E-25	0.1250	1.1884	0.0044	
thymosin beta-I2	<i>Lateolabrax japonicus</i>	P33248	80	2.00E-13	0.1250	1.1876	0.0003	
similar to 19.9 kD myosin light chain isoform I	<i>Danio rerio</i>	XP_685183	332	1.00E-89	0.1250	1.1720	0.0046	
ictacalcin	<i>Ictalurus punctatus</i>	AAV86967	145	1.00E-33	0.1250	1.1607	0.0021	
tropomyosin	<i>Theragra chalcogramma</i>	BAC44994	281	2.00E-74	0.1250	1.1368	0.0090	
N-acylsphingosine amidohydrolase	<i>Takifugu rubripes</i>	AAM43813	367	e-100	0.1250	1.1196	0.0082	
alanine-glyoxylate aminotransferase	<i>Platichthys flesus</i>	CAH59400	345	2.00E-93	0.1250	1.0860	0.0059	
titin b	<i>Danio rerio</i>	ABG48499	65	5.00E-09	0.1250	-1.0841	0.0045	
alpha-2,8-polysialyltransferase IV	<i>Oncorhynchus mykiss</i>	BAC77411	70	1.00E-10	0.1250	-1.0878	0.0030	
hedgehog acyltransferase-like, a	<i>Danio rerio</i>	NP_957181	295	1.00E-78	0.1250	-1.1419	0.0074	
parvalbumin	<i>Cyprinus carpio</i>	CAC83659	173	7.00E-42	0.1250	-1.1675	0.0033	
BTEB transcription factor	<i>Pimephales promelas</i>	ABO28528	107	1.00E-21	0.0625	-1.2019	0.0625	*
myosin regulatory light chain 2	<i>Salmo salar</i>	CAD89610	330	7.00E-89	0.1250	-1.2121	0.0020	
similar to Clca1 protein	<i>Danio rerio</i>	XP_694323	198	2.00E-49	0.0625	-1.2547	0.0625	*
ependymin	<i>Perca flavescens</i>	ABU49423	168	2.00E-40	0.0625	-1.2602	0.0625	*
aspartoacylase	<i>Danio rerio</i>	NP_001103573	384	e-105	0.0625	-1.3905	0.0625	*
Immune								
carboxypeptidase B	<i>Paralichthys olivaceus</i>	BAC53789	365	2.00E-99	0.1250	1.2897	0.0041	
fish-egg lectin (FEL)	<i>Cyprinus carpio</i>	P68512	192	2.00E-47	0.1250	1.1769	0.0003	
procathepsin B	<i>Oncorhynchus mykiss</i>	AAK69705	346	1.00E-93	0.0625	-1.1727	0.0625	*
gamma-glutamyl hydrolase	<i>Danio rerio</i>	NP_998487	223	6.00E-57	0.0625	-1.1939	0.0625	*
membrane glycoprotein	<i>Human coronavirus</i>	ABD75532	53	1.00E-05	0.0625	-1.1945	0.0625	*
beta-2 microglobulin	<i>Salmo salar</i>	AAG17525	176	8.00E-43	0.1250	-1.1995	0.0034	
microtubule-associated protein I light chain 3 alpha	<i>Danio rerio</i>	NP_999904	238	3.00E-61	0.0625	-1.2245	0.0625	*
microtubule-associated protein, RP/EB family, member	<i>Danio rerio</i>	NP_998805	272	1.00E-71	0.0625	-1.2298	0.0625	*
T-cell receptor beta chain ANA 11, putative	<i>Brugia malayi</i>	EDP38115	63	2.00E-08	0.1250	-1.2600	0.0052	
glycerophosphodiester phosphodiesterase domain containing I	<i>Danio rerio</i>	NP_001004118	322	2E-86	0.0625	-1.2652	0.0625	*
CHK1 checkpoint homolog	<i>Xenopus tropicalis</i>	CAJ83813	92	2.00E-17	0.1250	-1.3981	0.0013	
Apoptosis								
tissue inhibitor of metalloproteinase 2	<i>Oncorhynchus mykiss</i>	AAU14867	265	3.00E-69	0.1250	1.1235	0.0082	
cathepsin H	<i>Danio rerio</i>	NP_997853	300	5.00E-80	0.1250	1.0898	0.0069	
caspase-3	<i>Dicentrarchus labrax</i>	ABC70996	223	6.00E-68	0.1250	-1.0647	0.0022	

Table 1: Classification of differential gene expression of esfenvalerate responding genes in 52-d old *H. transpacificus* following 24 h exposure as identified by microarray analyses. (Continued)

caspase-1	<i>Dicentrarchus labrax</i>	<u>ABB05054</u>	79	3.00E-13	0.1250	-1.1270	0.0061	
cathepsin S-like	<i>Oncorhynchus mykiss</i>	<u>AAV32964</u>	291	1.00E-77	0.1250	-1.1507	0.0047	
Redox and metal ion binding								
hydroxymethylbilane synthase	<i>Danio rerio</i>	<u>CAM15096</u>	369	e-101	0.1250	1.3333	0.0042	
hemopexin	<i>Danio rerio</i>	<u>NP_001104617</u>	313	2.00E-83	0.1250	1.2968	0.0052	
transferrin	<i>Salvelinus fontinalis</i>	<u>BAA84100</u>	326	1.00E-87	0.1250	1.1769	0.0080	
similar to leprecan-like I protein	<i>Danio rerio</i>	<u>XP_695073</u>	183	1.00E-44	0.1250	1.1543	0.0036	
similar to synaptic glycoprotein SC2	<i>Danio rerio</i>	<u>XP_693420</u>	430	e-119	0.1250	-1.1340	0.0073	
similar to LOC407663 protein	<i>Danio rerio</i>	<u>XP_698537</u>	124	6.00E-27	0.0625	-1.2429	0.0625	*
Growth and development								
yghII (Putative growth hormone like protein-I)	<i>Seriola quinqueradiata</i>	<u>BAB62526</u>	153	1.00E-35	0.1250	-1.1388	0.0072	
ZPA domain containing protein	<i>Oryzias latipes</i>	<u>NP_001098216</u>	168	3.00E-40	0.0625	-1.1568	0.0625	*
Detoxification								
pregnane x receptor	<i>Oncorhynchus mykiss</i>	<u>ABP38412</u>	206	2.00E-51	0.0625	-1.2321	0.0625	*
Osmotic stress								
hyperosmotic glycine rich protein	<i>Salmo salar</i>	<u>AAO32675</u>	134	8.00E-30	0.1250	-1.1053	0.0021	
Digestion								
similar to ApoA4 protein isoform 2 (Apolipoprotein)	<i>Danio rerio</i>	<u>XP_698920</u>	296	7.00E-79	0.1250	1.4959	0.0056	
chymotrypsinogen 2-like protein	<i>Sparus aurata</i>	<u>AAT45254</u>	460	e-128	0.1250	1.4211	0.0095	
pancreatic carboxypeptidase A1 precursor copy 2	<i>Tetraodon nigroviridis</i>	<u>AAR16321</u>	242	9.00E-63	0.1250	1.3204	0.0025	
pancreatic protein with two somatomedin B domains	<i>Paralichthys olivaceus</i>	<u>BAA88246</u>	214	e-100	0.1250	1.1297	0.0021	
chitinase (Zgc:55941)	<i>Danio rerio</i>	<u>AAH44549</u>	369	e-100	0.1250	-1.0618	0.0097	

Genes selected for quantitative PCR analyses are shown in bold. * indicates genes responding at 0.0625 $\mu\text{g.L}^{-1}$ esfenvalerate and ** indicates gene responding to both 0.0625 and 0.125 $\mu\text{g.L}^{-1}$ esfenvalerate. Remaining genes responded only at 0.125 $\mu\text{g.L}^{-1}$ esfenvalerate exposure. Information represents proportions of a limited number of available genes on the applied microarray.

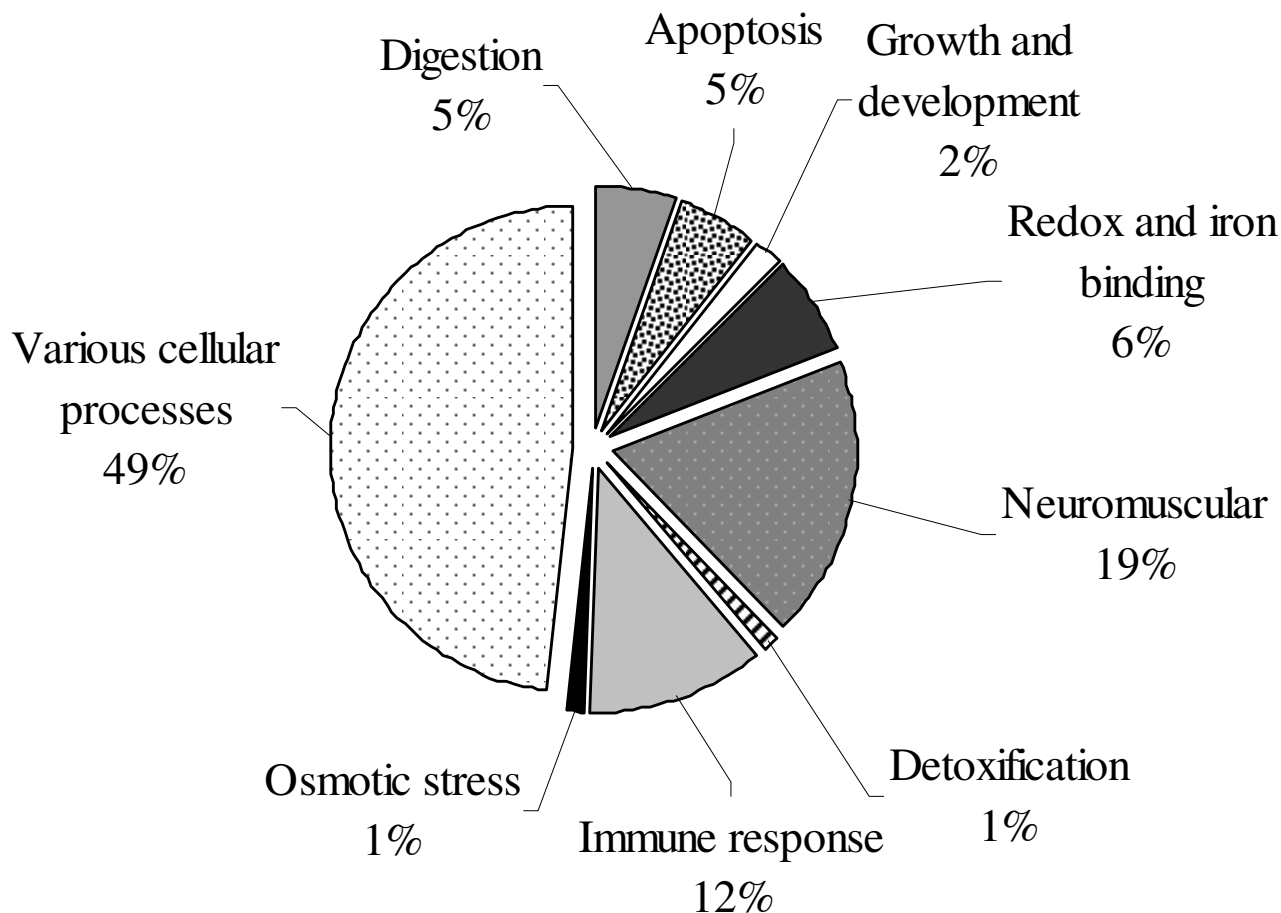


Figure 2

Functional classification of gene expression. Functional classification of genes responding to esfenvalerate exposure (0.0625 and 0.125 $\mu\text{g}\cdot\text{L}^{-1}$) in 52-d old delta smelt. Percentages were calculated based on function and biological processes of 94 unique differentially expressed genes.

sented by genes coding for pancreatic enzymes, a zona pellucida protein; inferred to be choriogenin, and a growth hormone.

A selected group of genes, highlighted in bold in the gene list in table 1, were validated using q-PCR and further investigated for use as molecular biomarkers. These results are described below, in conjunction with differentially expressed genes identified through microarray application.

Specific effects of esfenvalerate exposure on gene expression

Pyrethroid insecticides are known sodium and potassium channel modulators [25], with axon demyelinating effects [27]. This the proof-of-principle microarray assessment of esfenvalerate exposure of 52-d old delta smelt, was suc-

cessfully used to screen for, and further understand its mode of action, identifying neuromuscular responses, that were confirmed as highly significant through qPCR, corroborating known effects of this pesticide, but also pointing at other significant effects upon growth and development, digestion and the immune system. Gene expression assessed by qPCR on 10-d old delta smelt that verified these genomic responses are presented below. Correlations in expression between q-PCR investigated genes are shown in table 2 and fold changes in gene expression are summarized in figure 3 and table 3.

i. Neuromuscular responses

Parvalbumin expression in 10 d old larvae was induced 1.8-fold (t-test, $p = 0.008$) at 0.0313 $\mu\text{g}\cdot\text{L}^{-1}$ esfenvalerate, reducing in expression at higher concentrations. Localized in fast-contracting muscles, some endocrine tissues, in the

Table 2: Pairwise correlations of gene expression in esfenvalerate exposed 10-d old delta smelt.

	Aspartoacylase	Titin a	Microglobulin	Caspase-3	Parvalbumin	Hemopexin	ZPA	Myozenin	Creatine Kinase	PXR
Aspartoacylase	1 (0)									
Titin a	0.490 (0.402)	1 (0)								
Microglobulin	0.576 (0.310)	0.954 (0.012)	1 (0)							
Caspase-3	0.344 (0.571)	0.919 (0.027)	0.921 (0.026)	1 (0)						
Parvalbumin	0.284 (0.644)	0.953 (0.012)	0.928 (0.023)	0.981 (0.003)	1 (0)					
Hemopexin	0.629 (0.256)	0.819 (0.090)	0.951 (0.013)	0.860 (0.061)	0.822 (0.088)	1 (0)				
ZPA	-0.017 (0.979)	0.788 (0.113)	0.646 (0.239)	0.853 (0.066)	0.871 (0.054)	0.472 (0.422)	1 (0)			
Myozenin	0.309 (0.613)	0.898 (0.038)	0.949 (0.014)	0.909 (0.033)	0.943 (0.016)	0.892 (0.042)	0.677 (0.210)	1 (0)		
Creatine Kinase	0.255 (0.679)	0.916 (0.029)	0.865 (0.058)	0.984 (0.003)	0.978 (0.004)	0.759 (0.137)	0.932 (0.021)	0.859 (0.062)	1 (0)	
PXR	0.880 (0.049)	0.087 (0.890)	0.208 (0.737)	-0.113 (0.857)	-0.153 (0.805)	0.307 (0.616)	-0.475 (0.418)	-0.038 (0.952)	-0.218 (0.725)	1 (0)

Numbers represent correlation coefficients; r, and significance probabilities; (p), between ten selected biomarkers assessed with quantitative PCR. Bold p-values highlight significant correlations.

nervous system in GABAergic interneurons and in the brain [33], parvalbumin removes calcium from myofibrils, protecting neurons from hyper-excitability and facilitating muscle relaxation [34]. Accumulation of calcium in muscular tissue contributes to muscle degradation, muscular dystrophy and muscle fiber necrosis [35]. Estrogen is required for parvalbumin expression, thus estrogen receptor-β co-expresses with parvalbumin [36]. Estrogen is also required in brain development and has a protective neurological role, by regulating the activity of GABAergic systems within the hippocampus, basal forebrain and

hypothalamus [37]. Differential expression of parvalbumin on exposure to esfenvalerate may be resultant of estrogenic effects. Pyrethroid pesticides have steroid receptor-binding activity [38] linked with endocrine disruption [39], thus exposure is likely to affect the population dynamics of wildlife not only through neuromuscular impairments, but also by affecting reproductive output [29,40]. Parvalbumin, could therefore, be a good indicator of possible endocrine-disruption as well as neuromuscular impairments.

Table 3: Mean and standard deviations in fold changes in gene expression of ten selected biomarkers in esfenvalerate exposed, 10-d old, delta smelt, assessed by quantitative PCR.

Gene\Concentration		0.000	0.031	0.063	0.125	0.250
Aspartoacylase	Mean	1.000	0.629	0.346	0.299	0.244
	SE	0.240	0.100	0.078	0.035	0.088
Titin	Mean	1.000	1.515	0.909	0.933	0.475
	SE	0.529	0.198	0.095	0.192	0.096
Microglobulin	Mean	1.000	1.420	0.760	0.828	0.628
	SE	0.404	0.127	0.104	0.154	0.221
Caspase	Mean	1.000	2.024	1.136	0.818	0.670
	SE	0.336	0.432	0.043	0.117	0.094
Parvalbumin	Mean	1.000	1.718	1.097	1.037	0.771
	SE	0.241	0.151	0.125	0.062	0.107
Hemopexin	Mean	1.000	1.501	0.521	0.548	0.612
	SE	0.089	0.296	0.051	0.147	0.295
ZPA	Mean	1.000	1.612	1.455	1.165	0.912
	SE	0.415	0.321	0.224	0.210	0.574
Myozenin	Mean	1.000	1.730	0.857	1.069	0.835
	SE	0.212	0.205	0.121	0.093	0.188
Creatine Kinase	Mean	1.000	1.799	1.265	0.968	0.750
	SE	0.270	0.249	0.272	0.162	0.145
PXR	Mean	1.000	0.737	0.668	0.737	0.729
	SE	0.126	0.083	0.124	0.080	0.178

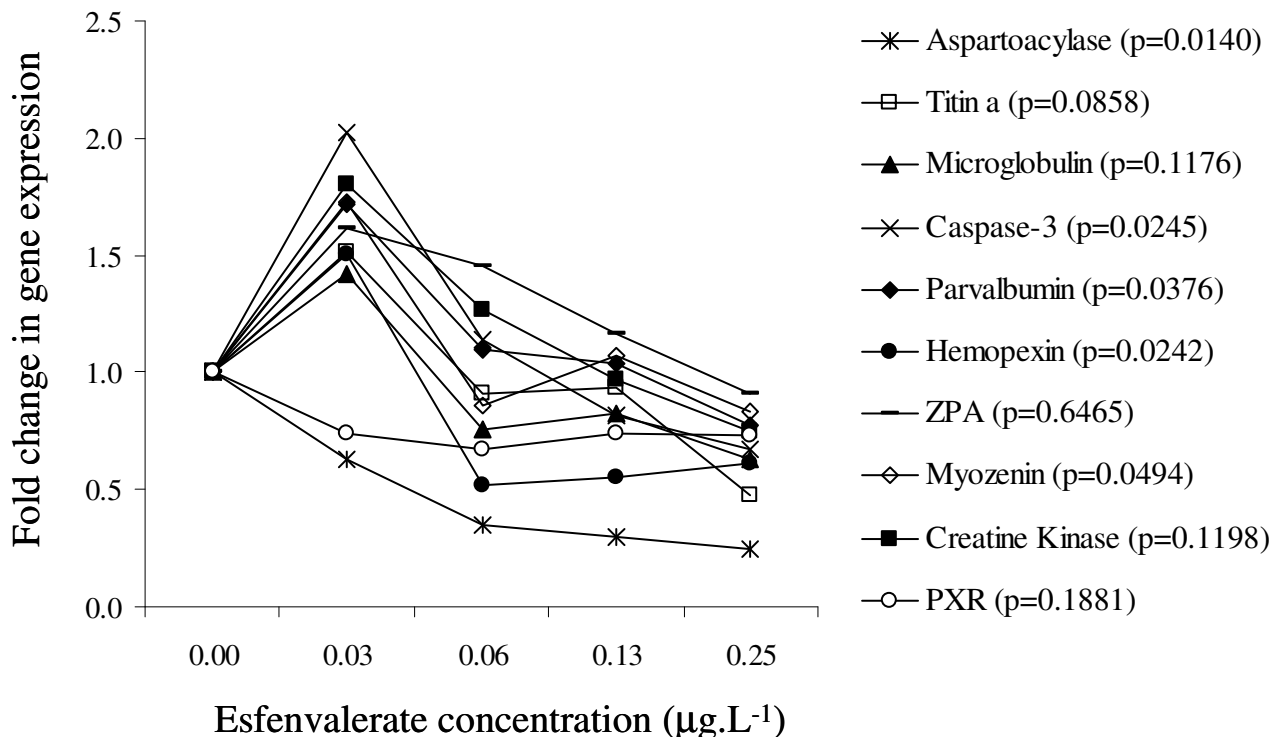


Figure 3

Molecular biomarkers responses. Fold changes in gene expression of ten selected biomarkers in esfenvalerate exposed, 10-d old, delta smelt, assessed by quantitative PCR. Significance in expression differences, as determined by One-way ANOVA, is shown in brackets in legend.

Interestingly, expression of aspartoacylase (ASPA) in exposed 10-d old delta smelt larvae was significantly affected at all concentrations, downregulating with increasing esfenvalerate concentration in a dose response manner, and correlating significantly with swimming anomaly at 24 h ($r = 0.913$, $p = 0.029$). Aspartoacylase catalyzes hydrolysis of *N*-acetyl-L-aspartate (NAA) to aspartate and acetate in the vertebrate brain [41]. Variations in NAA measured in urine, blood and brain, have been used as diagnosis of nervous system diseases such as Alzheimer's and multiple sclerosis [42,43]. Measurements of NAA, along with ADP levels determined by creatine kinase activity, are used to evaluate the energetic state of the brain, a positive linear correlation existing between NAA and ADP synthesis [44]. Deficiency in ASPA activity leads to degeneration of the myelin; an ensheathment that isolates and controls axonal activity, it is associated with schizophrenia [45], and is the established cause of leukodystrophy in Canavan's disease [43]. Abnormal myelination is known to result from acyltransferase deficiency [46]. Ependymin, a myelin associated glycoprotein

related to memory formation and involved in neuronal regeneration [47], was also negatively affected by esfenvalerate. Myelin has been postulated as a probable modulator of ASPA activity [48] further affecting this critical pathway of neurological function. ASPA protein activity is a strong biomarker of brain damage and neurological impairment investigation, used regularly in human and veterinary disease diagnostics [49].

Creatine kinase was significantly up-regulated at 0.0313 $\mu\text{g.L}^{-1}$ esfenvalerate (t-test, $p < 0.05$). Creatine kinase protein is used not only as a diagnosis of brain energetic value as mentioned above, but also of diseases like cardiac infarction and skeletal muscle necrosis [50]. In muscle, creatine kinase is specifically bound to sarcoendoplasmic reticulum, and regulates calcium uptake and ATP/ADP ratios [51], thus is directly involved in muscle contraction. Of interest here, are the pathway links and correlating responses ($r = 0.98$) between parvalbumin; facilitating muscle relaxation by binding calcium, and creatine kinase, which regulates calcium uptake. These two param-

eters on their own indicate muscular activity impairments, creating strong links with observed larval swimming behavior.

Titin expression also correlated significantly with parvalbumin ($r = 0.95$) and creatine kinase ($r = 0.92$), though not statistically significant in q-PCR assessments of 10-d old smelt larvae. Titin is an important protein also involved in muscle contraction, responsible for muscle elasticity and is the molecular scaffold for thick actin filament formation, forming a connection between filaments and the muscle Z-line [52]. Myozenin, another protein involved in muscle contraction, was significantly influenced by esfenvalerate exposure (t-test, $p < 0.05$) in 10-d old larvae. Co-regulating with Titin ($r = 0.90$), myozenin is a Z-line, α -actinin- and γ -filamin-binding protein expressed predominantly in skeletal muscle, and has been suggested as a biomarker for muscular dystrophy and other neuromuscular disorders [53].

ii Immune responses

In this study we have identified a significant alteration in expression of several genes involved in immune responses, most of them with links to neurological damage. β -microglobulin, a small protein normally found on the surface of many cells, including lymphocytes, is known to be involved in cell protection [54]. β -microglobulin is almost exclusively catabolized in the kidney and its excretion is an indication of long term nephrotoxicity [55]. High concentrations of β -microglobulin are reported to inhibit generation of functional dendritic cells [56], thus an increased amount in the blood or urine may be a sign of neural degeneration and of certain diseases, including some types of cancer, such as multiple myeloma or lymphoma. β -microglobulin levels are also reported to rise following viral infection and its reduced expression can compromise the immune system [57]. Interestingly, exposure to esfenvalerate resulted in a significant increase of pathogen susceptibility in chinook salmon [9]. β -microglobulin assessment with q-PCR did not show any significance in expression in 10 d old larvae (t-test, $p = 0.131$), however, an overall increase was observed at low concentrations of esfenvalerate, correlating significantly with expression of other genes investigated, further discussed below.

Multiple sclerosis is caused by an immunological attack on myelin [58], decreasing NAA and resulting in neurological instability. Furthermore, oxidative stress is inductive of apoptosis of myelin-reactive T cells [59]. A putative T cell receptor gene was identified through microarray screening, probably reacting to pyrethroid exposure, acting upon the myelin sheath and causing further neurological damage and cell death.

iii Apoptotic responses

Caspases (*cysteine-aspartic acid protease*) belong to a family of cysteine proteases that cleave other proteins, such as the precursor forms of the inflammatory cytokines interleukin 1- β and interleukin 18, into active mature peptides and are also involved in programmed cell death; or apoptosis [60]. Enzymatic activity requires an aspartic acid residue, and plays a critical role in the regulation of proinflammatory cytokines [61] that are associated with septic shock and autoimmune syndromes [62]. Upregulation of proinflammatory cytokines were reported in viral infected salmon, which further increased in expression following esfenvalerate exposure [63]. Caspases contribute to the pathogenesis of neurodegenerative disorders such as ischemia, Krabbes and Huntington's diseases, Alzheimer's, and other leukodystrophic diseases resulting in neural degeneration [64,65]. Moreover, caspase inhibitors have been suggested as therapeutic treatments for neurodegenerative diseases [66]. Low concentrations of esfenvalerate; $0.0313 \mu\text{g.L}^{-1}$, significantly induced caspase-3 expression 2-fold (t-test, $p = 0.002$) in 10-d old delta smelt. As caspases are activated by aspartic acid, induction may be suggestive of increases in substrate residues, along with inflammatory cytokines, probable effects upon the immune system, and subsequent neurodegeneration. Furthermore, a decrease in ASPA expression could be suggestive of reduced breakdown of NNA to aspartate and acetate, required as substrate for caspase activity, and synthesis of proteins required in repair mechanisms.

iv. Redox and metal ion binding

Upregulation of hemopexin was confirmed by quantitative PCR in 10-d old delta smelt exposed to $0.0313 \mu\text{g.L}^{-1}$ esfenvalerate. Hemopexin-like protein, a gene sequence displaying vast similarities to warm-temperature-acclimation-related-65 protein (WAP65) on BLAST homologies with Japanese medaka (*Oryzias latipes*), was identified as significantly upregulated through microarray screening. Hemopexin is synthesized by Schwann cells following nerve injury [67], accumulation has been reported in the peripheral nervous system following axonal lesions, and is specifically regulated during repair, returning to normal levels on axonal regeneration [68]. Wallerian degeneration occurs after axonal injury and is critical for repair, it is characterized by axonal and myelin degeneration [69], is accompanied by macrophage invasion and subsequent synthesis of hemopexin [67]. Hemopexin appears to play a significant role in neural regeneration, but may be resultant of oxidative stress mediated T cell activity on myelin sheaths (described above, under immune responses). Though upregulation follows nerve injury and there are strong connections with apoptosis, we classify hemopexin under oxidative stress as it has a high affinity with heme and reportedly plays a strong role in both heme transport and preventing heme-catalyzed oxidative

damage [70]. Moreover, pyrethroids have been shown to generate free radicals and induce oxidative stress [71]. Heme is known to respond to nerve injury, and has been suggested to play a role in neurodegenerative disorders [72], and hemopexin-mediated heme transport was reported to significantly decrease levels of transferrin receptor mRNA in HeLa cells [70]. Transferrin was also identified by microarray screening as significantly upregulated by esfenvalerate exposure. The primary role of transferrin is the delivery of iron across the blood brain barrier, and its expression in brain is not only related to myelin production, but may be a permissive agent in the process of myelination [73]. Furthermore, hemopexin and transferrin reportedly act by similar receptor-mediated mechanisms [74].

v. Growth and development

Microarray analyses identified a gene with high homology to egg envelope glycoproteins within the zona pellucida (ZPA) referred to as choriogenins, in fish [75]. This was significantly expressed in 52-d old larvae, however, no statistical differences in expression of ZPA were measured with qPCR in 10-d old larvae exposed to esfenvalerate. Choriogenin is reportedly more sensitive to endocrine disrupting chemicals (EDCs) than estrogen receptors and vitellogenin [76]. Composed primarily of glycoproteins with various functions during fertilization and development, choriogenin has been suggested as a biomarker of exposure to endocrine disrupting chemicals, as it is induced in late stage embryos, larvae and adult male fish exposed to estrogens [76,77]. Choriogenin is synthesized in liver of adult females, in response to estrogen, transported in blood and incorporated into the fish egg envelope; chorion or zona pellucida (ZPA), an extracellular matrix that surrounds the oocyte and early embryo [78]. Expression was notably elevated at low pyrethroid concentration, and it responded in a similar fashion to creatine kinase ($r = 0.93$), though no significant links were identified between these two biomarkers.

vi. Detoxification

Pregnane \times receptor (PXR), involved in the detection of toxic substances and a key regulator of xenobiotic metabolism, was identified through microarray assessments, as downregulated in 52-d old larvae. PXR is a steroid receptor and transcriptional regulator of detoxification mechanisms such as cytochrome oxidases, and phase II conjugating enzymes such as glutathione-S-transferases [79,80]. Downregulation of PXR expression has been linked with growth inhibition and cell death in rats and human cell lines following exposure to medroxyprogesterone and estradiol, known PXR ligands [80], further supporting identified apoptotic responses, steroid receptor-binding and endocrine disruption activity of esfenvalerate [38,39]. PXR expression was not significantly

different in q-PCR assessments of 10-d old larvae exposed to esfenvalerate, however, overall expression declined in a dose response manner, correlating with ASPA ($r = 0.880$; $p = 0.049$), making it a notable candidate of xenobiotic detection for future biomarker investigations in the delta smelt.

vii Osmotic Stress

A hyperosmotic glycine rich protein was identified with the microarrays as significantly downregulated by exposure to $0.0625 \mu\text{g.L}^{-1}$ esfenvalerate in 52-d old larvae. Osmoregulation is physiologically controlled by chemical messages from the endocrine system, along with cell signalling and nerve transmission [81]. Pyrethroids have been suggested to induce osmotic imbalances in common carp larvae [82] which are linked to effects on ATPase activity responsible for maintaining the Sodium transmembrane electrochemical gradient [83]. Larval fish are under direct exposure to osmotic stress as their endocrine system is not fully developed [84]. Parvalbumin and choriogenin expression have indicated possible effects on the endocrine system, thus expression of this hyperosmotic glycine rich protein may be directly caused by conditions affecting endocrine regulation.

viii Digestion

Chitinase was identified through microarray analysis as being downregulated by esfenvalerate exposure. Chitinase is the principal enzyme involved in digesting chitin, a major component of insect and crustacean exoskeleton [85]. Larval smelt were fed on artemia during the pre-exposure acclimation period. Not only chitinase but, other digestive enzymes; apolipoproteins, pancreatic enzymes, carboxypeptidase precursors and chemotrypsinogen, were also significantly upregulated following exposure in 52-d old larvae. Effects on digestion alone will undoubtedly have significant effects on growth, which when combined with hypothesized feeding reduction resulting from impaired swimming would lead to significant malnutrition. Contaminants affect a whole ecosystem, at all levels, and dramatic reductions in copepods, cladoceran and amphipod populations; organisms predated upon by the delta smelt, have been reported in the Sacramento delta [86]. Scarcity in food and reduced ingestion ability, besides digestion will significantly affect population dynamics of any specie.

Conclusions

Microarray technology was used as an initial screening of probable genes responding to esfenvalerate exposure therefore no multiple testing correction was applied. We have, however, examined and confirmed effect of esfenvalerate upon some of the genes in a different age group of larval delta smelt, identifying significant responses that are primarily linked with swimming behavior. Some

responding genes can be classified within different functional groups. Due to the measured behavioral responses, the classification approach contains a certain bias towards understanding neuromuscular effects. It is interesting that qPCR measurements have identified a greater response at the lower concentrations, implying homeostatic alterations, at environmentally relevant concentrations. Most genes did not display a desired dose response correlation associated with usable biomarkers, but did support responses within the suite of genes investigated, somewhat validating their use within a broader biomarker approach. Hemopexin for example is known to be involved in axon repair, and the myelin sheath surrounding the axon needs to be degraded for this repair to be processed, hypothesizing therefore that ASPA downregulation is resultant of neurological damage. The subsequent decrease of hemopexin expression at higher exposure concentrations, and further decrease in ASPA, may be indicative of repair impairments.

What becomes apparent from this study is that exposure to sublethal concentrations of esfenvalerate results in neurological damage and a series of compensatory molecular responses that attempt to repair nerve damage. We would hypothesize that induction of transcription of the genes encoding ASPA, hemopexin, parvalbumin and creatine kinase are part of a pathway of damage triggered repair mechanisms, responding to esfenvalerate insult. Reduction in expression of ASPA indicates that myelin sheaths may be degraded, resulting in a number of detrimental effects on the lesion sites, and similarly, muscular structure and function is being compromised as measured by alterations in titin and myozenin expression. The expression of β -microglobulins could be a compensatory reaction to toxic damage, protecting cells from infections in a susceptible immune system caused by exposure to esfenvalerate. Previous studies, carried out in esfenvalerate exposed chinook salmon have reported a compromised immunity and significantly higher susceptibility to infection [9,63]. This is particularly important in younger organisms that are generally more susceptible than adults. Furthermore, polluted waters not only contain mixtures of contaminants, but also harbor multiple pathogens that will further affect health parameters.

Behavioral endpoints, such as swimming behavior, are amongst the most sensitive and ecologically relevant parameters to assess sublethal toxicity of neurotoxic chemicals [29]. The high susceptibility of delta smelt to esfenvalerate, mediated neurological damage resulting in impaired swimming ability, also raises questions on the likely effects upon their chemosensory system; olfactory system, important in sensing reproductive pheromones, mediating reproduction. Females synthesize sex hormones stimulating male reproductive behavior [87]. Neu-

rological damage affecting the olfactory nerves, the brain and or entire nervous system, could lead to further impairments in reproductive success following exposure to pyrethroids. Damage to the olfactory system has been used as a sublethal toxicological endpoint in fish, in studies investigating behavior following pesticide exposures [88]. Pyrethroids are known to affect the olfactory system [89]. A chemosensory gene, ictalcalcin, responding to esfenvalerate exposure was also differentially expressed on the microarray. Ictalcalcin is a gene originally identified in catfish (*Ictalurus punctatus*), involved in chemosensory tissues, and highly expressed in barbell, olfactory mucosa and gill [90]. Differential expression of this gene may indicate that further behavioral parameters, not investigated in this study, such as recognition, alarm response, feeding, imprinting and homing, gamete release and synchronization, contaminant avoidance [88], and other behavioral parameters that are governed by chemosensory system, could be compromised. We could speculate that outside laboratory conditions, neuromuscular and chemosensory impairments would probably result in higher ecological parameters being affected through inability to swim against water currents, making them more susceptible to predation and reducing their ability to obtain food. Furthermore, effects on chemosensory parameters would lead to migratory, reproductive, predator and contaminant avoidance impairments.

Inhibition of repair mechanisms, leading to neuromuscular damage and eventual death, was behaviorally observed throughout exposure, as impairment in swimming ability. The ability to use molecular biomarkers of neuromuscular effect further strengthens links between mechanistic effects with parameters of ecological relevance. Our study supports the use of gene expression as a productive way of understanding modes of actions of individual chemicals in endangered species. Furthermore, this screening and interrogative approach permits the identification and development of biomarkers for species of concern in which prior information is limited and allows for investigations into problems specific to the organism in question; assessing possible causes of detrimental effects and resulting influences on individual performance and hypothesizing effects upon population dynamics. A suite of biomarkers developed in this manner, though additions and subtractions are required from the presented list, could be used to aid monitor impacts of stressors upon organisms within a specific environment and could be an essential tool in determining causative factors of population decline in the delta smelt and other threatened species. The selected biomarkers clearly need to be further investigated and validated against other known contaminants, and suitability in field applications.

Methods

Microarray construction

We constructed a delta smelt microarray using 8448 PCR amplified fragments from a normalized cDNA library. A cDNA library was created using expressed sequence tags (ESTs) ligated into *p-BS* plasmid vectors and cloned into chemically competent *Escherichia coli* cells (BioS&T Inc, Montreal, Quebec, Canada). RNA for library construction was obtained from a number of larval, juvenile and adult delta smelt, ranging from unexposed, control conditions, to fish from exposures to high temperature (25°C), and sublethal concentrations of copper, esfenvalerate, and a six field water samples from throughout the Sacramento-San Joaquin estuary. Products were PCR amplified from 1 µl bacterial suspension, and visualized on agarose gels. Purified PCR fragments ranging in size from 1-4 kb, along with control spots, were pin-printed in duplicate onto epoxysilane coated glass slides (Schott-Nexterion, USA) in a 20 × 19 block format, with 48 blocks per microarray. Microarrays were printed using a Lucidea Array Spotter (Amersham) at the Array Core facility at UC Davis (since closed down). Microarray control spots included a number of hybridization tags comprised of a pooled PCR product from all spots on the array, *H. transpacificus* DNA, and four Spot Report System of alien PCR products from *Arabidopsis thaliana*; CAB, RCA, RBCL and LPT4 (Stratagene, USA). Blank control spots consisted of 1 × Nexterion buffer solution.

Esfenvalerate exposures

Delta smelt larvae aged 10 d and 52 d were exposed for 24 h, in two separate experiments, to a range of esfenvalerate concentrations; 0.0313, 0.0625, 0.125, 0.250 and 0.500 µg.L⁻¹ (nominal) in laboratory control water, with corresponding laboratory and solvent controls. Concentrations were measured at the start of the experiment by the Water Pollution Control Laboratory at the Department of Fish and Game (Rancho Cordova, California, USA), only single measurements were taken per treatment (results not shown), therefore we present the data in terms of nominal concentrations. Laboratory control water consisted of deionized water amended to US EPA moderately hard standards (80-100 mg.L⁻¹ CaCO₃) and 200 µl/L methanol was used as solvent carrier. Salinity was adjusted with Instant Ocean salt to match hatchery rearing conditions (range 650 µS.cm⁻¹ to 900 µS.cm⁻¹).

Average wet weights of 10-d to 52-d old larval delta smelt ranged from 0.5 to 2.5 mg respectively. Larvae were obtained from the Fish Conservation and Culture Laboratory (FCCL) UC Davis, Byron, CA, transported in cool, oxygenated 2-gallon black buckets, and held overnight in the laboratory at 17°C and a 8 h:16 h D:L light cycle. The following day, ten larvae were transferred to each 2-L beaker containing 1 L of aerated control water or esfenvalerate treatment.

Each treatment consisted of 4 replicates and tests were performed at 8 h:16 h D:L cycle, at a water temperature of 17°C ± 1.2°C. The pH during the tests was 7.1 - 7.5. Dissolved oxygen levels were within the acceptable range for delta smelt (above 6.5 mg.L⁻¹)[91]. Larvae were fed rotifers the day before the test start, but not during the 24 h exposure. Rotifer cultures were obtained from FCCL. During exposure, larvae were observed for aberrant swimming behavior, and surviving fish were scored after 4 h and 24 h. Swimming behavior was assessed by observing each tank for 5 min as described in Geist *et. al.* [17]. Any pronounced deviation (> 1 min) from normal (control) swimming patterns were recorded as abnormal. Effects on swimming performance (EC₅₀) and mortality (LC₅₀) were assessed using linear regression analysis with Environmental Toxicity Information System (CETIS) by Tidepool Scientific Software (McKinleyville, CA, USA).

Four surviving 52 d old larvae from solvent controls, and exposures to 0.0625 and 0.125 µg.L⁻¹ esfenvalerate, were used for microarray analyses, hybridized in a reference design against a pool of RNA from all treatments. Four replicate 10 d old larvae from each treatment (controls and 0.0312, 0.0625, 0.125 µg.L⁻¹ esfenvalerate) were used for biomarker analyses and gene expression verification using quantitative PCR (q-PCR).

All experiments and use of test organisms were approved by the UC Davis Institutional Animal Care and Use Committee (Animal Use Protocol for Animal Care and Use #13361). This institution is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care, International (AAALAC) and has an Animal Welfare Assurance on file with the Office of Laboratory Animal Welfare (OLAW). The Assurance Number is A3433-01. The IACUC is constituted in accordance with U.S. Public Health Service (PHS) Animal Welfare Policy and includes a member of the public and a non-scientist.

RNA isolation, cDNA synthesis and fluorescence labeling

RNA was extracted from whole, individual organisms using a standard phenol:chloroform protocol with Trizol Reagent (Invitrogen). Fifteen micrograms of total RNA were used for cDNA synthesis, spiked with control RNA (CAB, RCA, RBCL and LTP4 (SpotReport, Stratagene) and labeled with Alexa fluor dyes, using SuperScript™ Plus Indirect cDNA labeling System (Invitrogen). Each experimental sample and control was combined with a reference pool cDNA prior to hybridization using an automated Tecan HS4800 hybridization station. Slides were scanned using a GenePix 4000B scanner (Axon Instruments).

Microarray images and data from esfenvalerate exposed delta smelt can be accessed at <http://www.vet>

med.ucdavis.edu/apc/WernerLab/subpapelagic_organism_decline.html; POD archive data.

Microarray Analyses

Normalization and analytical methods are described in Loguinov *et. al.* [32] and Smyth [92]. In brief, print tip normalization was carried out within slides and sequential single slide data analysis was carried out as an alternative to between-slide normalization. An α -outlier-generating model was used to identify differentially expressed genes by applying the following decision rule for multiple-slide data analysis: a given gene was selected as a candidate if it was detected as significantly up- or downregulated in 4 of 4 replicates (raw p-value = 0.0625 using exact binomial test and considering outcomes as Bernoulli trials). The approach did not use scale estimator for statistical inference and, therefore, it did not require between-slide normalization. This method however, detected only one significant differentially expressed candidate gene at the highest exposure concentration (0.125 $\mu\text{g}/\text{L}^{-1}$), (with no significant annotation identity - see results and discussion). As a result, a second analytical method was applied to increase the number of probable genes for consideration in biomarker development. Thus we further analyzed the data using LIMMA GUI (Linear model for microarray analysis graphical user interface) [92], written in the R-programming language available through Bioconductor <http://www.Bioconductor.org>. Data was normalized within arrays using print-tip Lowess and between arrays applying aquantile normalization methods [92]. A linear model fit was computed using the

duplicates on the arrays and the least-squares method, no multiple assessment methods were applied to eliminate false positives as our aim was to increase the number of genes available for biomarker assessment, and qualify these through quantitative PCR.

Sequencing and Annotation

Sequencing was carried out at the CA&ES Genomic Facility, UC Davis. Basic Local Alignment Search Tool; translated nucleotide (BLASTx) searches were performed on specific fragments that responded significantly to the exposure treatments. Only genes that were differentially expressed following esfenvalerate exposure were sequenced. Sequences were annotated according to homologies to protein database searches using translated nucleotide sequences and direct nucleotide queries <http://blast.ncbi.nlm.nih.gov/Blast.cgi>. Sequences were only annotated if they were found to have a BLASTx match with the expect value smaller than 1×10^{-5} and a score above 50.

Functional Classifications

Differentially expressed genes were classified according to gene ontology <http://www.uniprot.org/uniprot>, and information gathered from literature, into functional groups. Classification was carried out based on gene expression changes in respect of control subjects, regardless of whether these were up or downregulated, or exposure concentrations. Specific genes of interest were selected for further investigation using quantitative PCR.

Table 4: Primers and probes used for molecular biomarkers: Primer pairs and TaqMan probes used in q-PCR assessments.

Accession no.	Gene	Primer Sequences	Probe No.
EJ711577	Aspartoacylase	Left: ggaggcacacatgggaatg Right: cttcctctgaatctctgtccattatc	109
EJ711576	Parvalbumin	Left: gaccaagacaagagtggcttca Right: tctggcaccagcagagaagt	101
EJ711580	β -2 microglobulin	Left: tcttcgcggtcatctttctc Right: gggtgtggccatacacctgaa	22
EJ711579	Hemopexin	Left: catgcactacgaggacgacaag Right: tggtagtagtgaaacacctgctg	143
EJ711578	Caspase-3	Left: gagaaccggtatgaaccaacg Right: tccaagctcccaaacactttc	159
EJ711575	Titin a	Left: tgatcactggcgtgaaagagg Right: caagctcattggacagtttgagg	159
EJ711581	ZPA	Left: catgcgctgagtttgataa Right: tgccattgatagcatcaacttca	106
EJ711583	Myozenin*	Left: ccaatgtcgtgctggtacacc Right: ctgccagacattgatgtagcca	106
EJ711584	Creatine kinase*	Left: cgatcggcgttgagatg Right: gccaaagttcaacgagattctgg	163
EJ711582	PXR	Left: tgaggcggaggagaagag Right: gaggcggaggagaagag	144

* indicates biomarkers obtained from preliminary studies carried out during microarray development.

Biomarker development

Genes were selected according to level of expression significance, knowledge base from literature, and functional classification. Primer and probes for qPCR analyses were designed using Roche Universal Probe Library Assay Design Center <https://www.roche-applied-science.com>. Designed primers were obtained from Eurofins MWG Operon <http://www.eurofinsdna.com>, and TaqMan probes were supplied by Roche. Sequences for all genes assessed by qPCR analyses have been submitted to genbank <http://www.ncbi.nlm.nih.gov>. Primers and probes, and genbank accession numbers, for investigated biomarkers are detailed in table 4. Myozenin and creatine kinase, though not resulting from the current study, were genes identified in investigations carried out during microarray development, and added to the selected biomarkers due to our interest in neuromuscular activity.

Quantitative PCR

A total of 1.5 µg RNA was cDNA synthesized using random primers, and diluted to a total of 50 µl with nuclease free water to generate sufficient template for qPCR analysis. TaqMan Universal PCR Mastermix (Applied Biosystems) was used in q-PCR amplifications in a reaction containing 10mM Tris-HCl (pH 8.3), 50 mM KCl, 5 mM MgCl₂, 2.5 mM deoxynucleotide triphosphates, 0.625 U AmpliTaq Gold DNA polymerase per reaction, 0.25 U AmpErase UNG per reaction and 5 µL of cDNA sample in a final volume of 12 µL. The samples were placed in 384 well plates and cDNA was amplified in an automated fluorometer (ABI PRISM 7900 Sequence Detection System, Applied Biosystems). Amplification conditions were 2 min at 50°C, 10 min at 95°C, 40 cycles of 15 s at 95°C and 60 s at 60°C. Fluorescence of samples was measured every 7 s and signals were considered positive if fluorescence intensity exceeded 10 times the standard deviation of the baseline fluorescence (threshold cycle, CT). SDS 2.2.1 software (Applied Biosystems) was used to quantify transcription.

Quantitative PCR data was analyzed using the relative quantification 2^(-Delta Delta CT) method ([93]). In the absence of house keeping genes, expression was calculated relative to the mean of controls in respective exposures. Surviving larvae from each replicate of the 10-d old exposed delta smelt were used for q-PCR analyses. One-way ANOVA was used to assess differences in gene expression through out the exposure concentrations, and data were further assessed using Student's T-test at individual concentrations in respect to solvent controls.

Abbreviations

EST: expressed sequence tag; q-PCR: real-time quantitative polymerase chain reaction; ASPA: aspartoacylase; ESA: endangered species act; CESA: California endangered spe-

cies act; LIMMA GUI: linear model for microarray analysis, graphical user interface; BLAST: basic local alignment search tool; GABA: gamma-aminobutyric acid; NAA: N-acetyl-L-aspartate; ADP: adenosine diphosphate; WAP65: warm-temperature-acclimation-related-65 protein; EDC: endocrine disrupting chemical; ZPA: zona pellucida; PXR: pregnane × receptor; p-BS: p-Bluescript; IPTG: isopropyl β-galactosidase; LB: Luria Bertani; FCCL: Fish Conservation and Culture Laboratory; UNG: Uracil N-glycosylase; CT: cycle threshold.

Authors' contributions

All authors were involved in designing the experiments. JG carried out esfenvalerate exposures recording swimming abnormality and mortality. REC designed, developed and applied the microarrays with assistance from JP and HW. REC and JP prepared, hybridized and scanned microarrays. REC and AVL performed microarray analyses. REC and LSD designed qPCR assays and performed respective assessments and analysis. REC drafted the manuscript and all authors read, contributed intellectually and approved the final manuscript.

Acknowledgements

We thank E. Campbell, UC Davis, for assistance with the microarray development, J. Lindberg and B. Baskerville-Bridges (FCCL) for supplying delta smelt along with invaluable knowledge on culture and handling. UC Davis ATL was used for all exposure tests; we thank all staff for assistance. Funding provided by the Interagency Ecological Program, Sacramento, California (Contract No. 4600004445 to I. Werner).

References

- Bryant M, Souza K: **Summer ternet and fall midwater trawl survey status and trends.** *Interagency Ecological Program Newsletter* 2004, **17**:14-17.
- Hieb K, Bryant M, Souza K, Greiner T, Slater S: **Place holder for bay and estuary species 2004 status and trends report.** *Interagency Ecological Program Newsletter* 2005, **18**:6-10.
- Feyrer F, Nobriga ML, Sommer TR: **Multidecadal trends for three declining fish species: habitat patterns and mechanisms in the San Francisco Estuary, California, USA.** *Journal of Fisheries and Aquatic Sciences* 2007, **64**:723-734.
- Brown LR, Kimmerer W, Brown R: **Managing water to protect fish: a review of California's Environmental Water Account, 2001-2005.** *Environmental management* 2009, **43**(2):357-368.
- Sommer T, Armor C, Baxter R, Breuer R, Brown L, Chotkowski M, Culbertson S, Feyerer F, Gingras M, Herbold B, et al.: **The collapse of pelagic fishes in the upper san francisco estuary.** *Fisheries* 2007, **32**(6):270-277.
- Scholz NL, Truelove NK, French BL, Berejikian BA, Quinn TP, Casillas E, Collier TK: **Diazinon disrupts antipredator and homing behaviors in chinook salmon (*Oncorhynchus tshawytscha*).** *Canadian Journal of Fisheries and Aquatic Sciences* 2000, **57**:1911-1918.
- de Vlaming V, Connor V, DiGiorgio C, Bailey HC, Deanovic LA, Hinton DE: **Application of whole effluent toxicity test procedures to ambient water quality assessment.** *Environmental Toxicology and Chemistry* 2000, **19**(1):42-62.
- Sandahl JF, Baldwin DH, Jenkins JJ, Scholz NL: **Comparative thresholds for acetylcholinesterase inhibition and behavioral impairment in coho salmon exposed to chlorpyrifos.** *Environmental toxicology and chemistry/SETAC* 2005, **24**(1):136-145.
- Clifford MA, Eder KJ, Werner I, Hedrick RP: **Synergistic effects of esfenvalerate and infectious hematopoietic necrosis virus on juvenile chinook salmon mortality.** *Environmental toxicology and chemistry/SETAC* 2005, **24**(7):1766-1772.

10. Floyd EY, Geist JP, Werner I: **Acute, sublethal exposure to a pyrethroid insecticide alters behavior, growth, and predation risk in larvae of the fathead minnow (*Pimephales promelas*).** *Environmental toxicology and chemistry/SETAC* 2008, **27(8)**:1780-1787.
11. Rand GM: *Fundamentals of Aquatic Toxicology* Taylor and Francis, Bristol, PA; 1995.
12. U.S. EPA: **Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms.** 4th edition. U.S. EPA OoW, Washington, DC, EPA-821-R-02-013; 2002.
13. Denslow ND, Colbourne JK, Dix D, Freedman J, Helbing C, Kennedy S, Williams P: **Selection of surrogate animal species for comparative toxicogenomics.** In *Genomic Approaches for Cross-Species Extrapolation in Toxicology* Edited by: Benson WH, Di Giulio RT. Portland, Oregon, USA: CRC Press; 2006:33-75.
14. Little EE, Calfee RD: **Acute toxicity of three herbicides, DMA*41VM, Renovate 3, and Sonar A.S., to two life stages of white sturgeon from the Kootenai River and to rainbow trout.** USGS Report: Columbia Environmental Research Center, Columbia, MO; 2008.
15. Little EE, Calfee RD: **Toxicity of Chlorine and Copper to Rainbow Trout and to White Sturgeon from the Kootenai River and Columbia River.** USGS Report: Columbia Environmental Research Center, Columbia, MO; 2008.
16. Connon R, Hooper HL, Sibly RM, Lim FL, Heckmann LH, Moore DJ, Watanabe H, Soetaert A, Cook K, Maund SJ, et al.: **Linking molecular and population stress responses in *Daphnia magna* exposed to cadmium.** *Environmental science & technology* 2008, **42(6)**:2181-2188.
17. Geist J, Werner I, Eder KJ, Leutenegger CM: **Comparisons of tissue-specific transcription of stress response genes with whole animal endpoints of adverse effect in striped bass (*Morone saxatilis*) following treatment with copper and esfenvalerate.** *Aquatic toxicology (Amsterdam, Netherlands)* 2007, **85(1)**:28-39.
18. Heckmann LH, Sibly RM, Connon R, Hooper HL, Hutchinson TH, Maund SJ, Hill CJ, Bouetard A, Callaghan A: **Systems biology meets stress ecology: linking molecular and organismal stress responses in *Daphnia magna*.** *Genome biology* 2008, **9(2)**:R40.
19. Denslow ND, Garcia-Reyer N, Barber DS: **Fish 'n' chips: the use of microarrays for aquatic toxicology.** *Molecular bioSystems* 2007, **3(3)**:172-177.
20. Steinberg CE, Sturzenbaum SR, Menzel R: **Genes and environment - Striking the fine balance between sophisticated biomonitoring and true functional environmental genomics.** *The Science of the total environment* 2008, **400(1-3)**:142-161.
21. Soetaert A, Vandenbrouck T, Ven K van der, Maras M, van Remortel P, Blust R, De Coen WM: **Molecular responses during cadmium-induced stress in *Daphnia magna*: integration of differential gene expression with higher-level effects.** *Aquatic toxicology (Amsterdam, Netherlands)* 2007, **83(3)**:212-222.
22. Poynton HC, Varshavsky JR, Chang B, Cavigliolo G, Chan S, Holman PS, Loguinov AV, Bauer DJ, Komachi K, Theil EC, et al.: ***Daphnia magna* ecotoxicogenomics provides mechanistic insights into metal toxicity.** *Environmental science & technology* 2007, **41(3)**:1044-1050.
23. Ahluwalia A, Clodfelter KH, Waxman DJ: **Sexual dimorphism of rat liver gene expression: regulatory role of growth hormone revealed by deoxyribonucleic Acid microarray analysis.** *Molecular endocrinology (Baltimore, Md)* 2004, **18(3)**:747-760.
24. Ilves KL, Taylor EB: **Molecular resolution of the systematics of a problematic group of fishes (Teleostei: Osmeridae) and evidence for morphological homoplasy.** *Molecular phylogenetics and evolution* 2009, **50(1)**:163-178.
25. Soderlund DM, Clark JM, Sheets LP, Mullin LS, Piccirillo VJ, Sargent D, Stevens JT, Weiner ML: **Mechanisms of pyrethroid neurotoxicity: implications for cumulative risk assessment.** *Toxicology* 2002, **171(1)**:3-59.
26. Werner I, Geist J, Okihiro M, Rosenkranz P, Hinton DE: **Effects of dietary exposure to the pyrethroid pesticide esfenvalerate on medaka (*Oryzias latipes*).** *Marine environmental research* 2002, **54(3-5)**:609-614.
27. Lees G: **Effects of pyrethroid molecules on rat nerves in vitro: potential to reverse temperature-sensitive conduction block of demyelinated peripheral axons.** *British journal of pharmacology* 1998, **123(3)**:487-496.
28. Little EE, Dwyer FJ, Fairchild JF, DeLonay AJ, Zajicek JL: **Survival of bluegill and their behavioral responses during continuous and pulsed exposures to esfenvalerate, a pyrethroid insecticide.** *Environmental Toxicology and Chemistry* 1993, **12(5)**:871-878.
29. Werner I, Moran K: **Effects of pyrethroid insecticides on aquatic organisms.** *Synthetic pyrethroids, occurrence and behaviour in aquatic environments* 2008:310-334 [<http://pubs.acs.org/doi/abs/10.1021/bk-2008-0991.ch014>]. Washington DC: American Chemical Society
30. Swanson C, Young P: **Swimming performance of delta smelt: maximum performance, and behavioral and kinematic limitations on swimming at submaximal velocities.** *J Exp Biol* 1998, **201(3)**:333-345.
31. Bacey J, Spurlock F, Starner K, Feng H, Hsu J, White J, Tran DM: **Residues and toxicity of esfenvalerate and permethrin in water and sediment, in tributaries of the Sacramento and San Joaquin rivers, California, USA.** *Bulletin of environmental contamination and toxicology* 2005, **74(5)**:864-871.
32. Loguinov AV, Mian IS, Vulpe CD: **Exploratory differential gene expression analysis in microarray experiments with no or limited replication.** *Genome biology* 2004, **5(3)**:R18.
33. Freund TF: **GABAergic septohippocampal neurons contain parvalbumin.** *Brain Res* 1989, **478(2)**:375-381.
34. Rewal M, Wren Y, Wilson A, Simpkins JW, Jung ME: **Role of parvalbumin in estrogen protection from ethanol withdrawal syndrome.** *Alcoholism, clinical and experimental research* 2005, **29(10)**:1837-1844.
35. Olive M, Rivera R, Ferrer I: **Parvalbumin immunocytochemistry and calcium deposition in muscle fiber necrosis and subsequent regeneration following intramuscular injection of metoclopramide.** *Muscle & nerve* 1994, **17(5)**:494-499.
36. Blurton-Jones M, Tuszyński MH: **Estrogen receptor-beta colocalizes extensively with parvalbumin-labeled inhibitory neurons in the cortex, amygdala, basal forebrain, and hippocampal formation of intact and ovariectomized adult rats.** *The Journal of comparative neurology* 2002, **452(3)**:276-287.
37. Ross NR, Porter LL: **Effects of dopamine and estrogen upon cortical neurons that express parvalbumin in vitro.** *Brain Res Dev Brain Res* 2002, **137(1)**:23-34.
38. Tyler C, Beresford N, Woning M van der, Sumpter J, Thorpe K: **Metabolism and environmental degradation of pyrethroid insecticides produce compounds with endocrine activities.** *Environmental Toxicology and Chemistry* 2002, **19**:801-809.
39. Moniz AC, Cruz-Casallas PE, Salzgeber SA, Varoli FM, Spinosa HS, Bernardi MM: **Behavioral and endocrine changes induced by perinatal fenvalerate exposure in female rats.** *Neurotoxicology and teratology* 2005, **27(4)**:609-614.
40. Gutjahr-Gobell RE, Zaroogian GE, Horowitz DJ, Gleason TR, Mills LJ: **Individual effects of estrogens on a marine fish, Cunner (*Tautoglabrus adspersus*), extrapolated to the population level.** *Ecotoxicology and environmental safety* 2006, **63(2)**:244-252.
41. Baslow MH: **Functions of N-acetyl-L-aspartate and N-acetyl-L-aspartylglutamate in the vertebrate brain: role in glial cell-specific signaling.** *Journal of neurochemistry* 2000, **75(2)**:453-459.
42. Bitto E, Bingman CA, Wesenberg GE, McCoy JG, Phillips GN Jr: **Structure of aspartoacylase, the brain enzyme impaired in Canavan disease.** *Proceedings of the National Academy of Sciences of the United States of America* 2007, **104(2)**:456-461.
43. Matalon R, Kaul R, Casanova J, Michals K, Johnson A, Rapin I, Gashkoff P, Deanching M: **SSIEM Award. Aspartoacylase deficiency: the enzyme defect in Canavan disease.** *Journal of inherited metabolic disease* 1989, **12(Suppl 2)**:329-331.
44. Pan JW, Takahashi K: **Interdependence of N-acetyl aspartate and high-energy phosphates in healthy human brain.** *Annals of neurology* 2005, **57(1)**:92-97.
45. Holmes E, Tsang TM, Huang JT, Leweke FM, Koethe D, Gerth CW, Nolden BM, Gross S, Schreiber D, Nicholson JK, et al.: **Metabolic profiling of CSF: evidence that early intervention may impact on disease progression and outcome in schizophrenia.** *PLoS medicine* 2006, **3(8)**:e327.
46. Sztriha LS, Nork MP, Abdulrazzaq YM, al-Gazali LI, Bakalnova DB: **Abnormal myelination in peroxisomal isolated dihydroxyacetonephosphate acyltransferase deficiency.** *Pediatric neurology* 1997, **16(3)**:232-236.

47. Adams DS, Hasson B, Boyer-Boiteau A, El-Khishin A, Shashoua VE: **A peptide fragment of ependymin neurotrophic factor uses protein kinase C and the mitogen-activated protein kinase pathway to activate c-Jun N-terminal kinase and a functional AP-1 containing c-Jun and c-Fos proteins in mouse NB2a cells.** *Journal of neuroscience research* 2003, **72(3)**:405-416.
48. Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mork S, Bo L: **Axonal transection in the lesions of multiple sclerosis.** *The New England journal of medicine* 1998, **338(5)**:278-285.
49. Matalon R, Michals-Matalon K: **Biochemistry and molecular biology of Canavan disease.** *Neurochemical research* 1999, **24(4)**:507-513.
50. Jockusch H, Friedrich G, Zippel M: **Serum parvalbumin, an indicator of muscle disease in murine dystrophy and myotonia.** *Muscle & nerve* 1990, **13(6)**:551-555.
51. Rossi AM, Eppenberger HM, Volpe P, Cotrufo R, Wallimann T: **Muscle-type MM creatine kinase is specifically bound to sarcoplasmic reticulum and can support Ca²⁺ uptake and regulate local ATP/ADP ratios.** *The Journal of biological chemistry* 1990, **265(9)**:5258-5266.
52. Tskhovrebova L, Trinick J, Sleep JA, Simmons RM: **Elasticity and unfolding of single molecules of the giant muscle protein titin.** *Nature* 1997, **387(6630)**:308-312.
53. Takada F, Woude DL, Vander, Tong HQ, Thompson TG, Watkins SC, Kunkel LM, Beggs AH: **Myozenin: an alpha-actinin- and gamma-filamin-binding protein of skeletal muscle Z lines.** *Proceedings of the National Academy of Sciences of the United States of America* 2001, **98(4)**:1595-1600.
54. Tanaka T, Ebata T, Tajima A, Kinoshita K, Okumura K, Yagita H: **Beta2-microglobulin required for cell surface expression of blastocyst MHC.** *Biochemical and biophysical research communications* 2005, **332(1)**:311-317.
55. Sorensen PG, Nissen MH, Groth S, Rorth M: **Beta-2-microglobulin excretion: an indicator of long term nephrotoxicity during cis-platinum treatment?** *Cancer chemotherapy and pharmacology* 1985, **14(3)**:247-249.
56. Xie J, Wang Y, Freeman ME, Barlogie B, Yi Q: **Beta 2-microglobulin as a negative regulator of the immune system: high concentrations of the protein inhibit in vitro generation of functional dendritic cells.** *Blood* 2003, **101(10)**:4005-4012.
57. Tay CH, Welsh RM, Bruckiewicz RR: **NK cell response to viral infections in beta 2-microglobulin-deficient mice.** *J Immunol* 1995, **154(2)**:780-789.
58. Ota K, Matsui M, Milford EL, Mackin GA, Weiner HL, Hafler DA: **T-cell recognition of an immunodominant myelin basic protein epitope in multiple sclerosis.** *Nature* 1990, **346(6280)**:183-187.
59. Zettl UK, Mix E, Zielasek J, Stangel M, Hartung HP, Gold R: **Apoptosis of myelin-reactive T cells induced by reactive oxygen and nitrogen intermediates in vitro.** *Cellular immunology* 1997, **178(1)**:1-8.
60. Jarry A, Vallette G, Cassagnau E, Moreau A, Bou-Hanna C, Lemarre P, Letessier E, Le Neel JC, Galmiche JP, Laboisie CL: **Interleukin 1 and interleukin 1beta converting enzyme (caspase 1) expression in the human colonic epithelial barrier.** *Gut* 1999, **45(2)**:246-251.
61. Ghayur T, Banerjee S, Hugunin M, Butler D, Herzog L, Carter A, Quintal L, Sekut L, Talanian R, Paskind M, et al.: **Caspase-1 processes IFN-gamma-inducing factor and regulates LPS-induced IFN-gamma production.** *Nature* 1997, **386(6625)**:619-623.
62. Mariathasan S, Newton K, Monack DM, Vucic D, French DM, Lee WP, Roose-Girma M, Erickson S, Dixit VM: **Differential activation of the inflammasome by caspase-1 adaptors ASC and Ipaf.** *Nature* 2004, **430(6996)**:213-218.
63. Eder KJ, Clifford MA, Hedrick RP, Kohler HR, Werner I: **Expression of immune-regulatory genes in juvenile Chinook salmon following exposure to pesticides and infectious hematopoietic necrosis virus (IHNV).** *Fish & shellfish immunology* 2008, **25(5)**:508-516.
64. Kruidering M, Schouten T, Evan GI, Vreugdenhil E: **Caspase-mediated cleavage of the Ca²⁺/calmodulin-dependent protein kinase-like kinase facilitates neuronal apoptosis.** *The Journal of biological chemistry* 2001, **276(42)**:38417-38425.
65. Zaka M, Wenger DA: **Psychosine-induced apoptosis in a mouse oligodendrocyte progenitor cell line is mediated by caspase activation.** *Neuroscience letters* 2004, **358(3)**:205-209.
66. Wellington CL, Hayden MR: **Caspases and neurodegeneration: on the cutting edge of new therapeutic approaches.** *Clinical genetics* 2000, **57(1)**:1-10.
67. Camborieux L, Bertrand N, Swerts JP: **Changes in expression and localization of hemopexin and its transcripts in injured nervous system: a comparison of central and peripheral tissues.** *Neuroscience* 1998, **82(4)**:1039-1052.
68. Swerts JP, Soula C, Sagot Y, Guinaudy MJ, Guillemot JC, Ferrara P, Duprat AM, Cochard P: **Hemopexin is synthesized in peripheral nerves but not in central nervous system and accumulates after axotomy.** *The Journal of biological chemistry* 1992, **267(15)**:10596-10600.
69. Camborieux L, Julia V, Pipy B, Swerts JP: **Respective roles of inflammation and axonal breakdown in the regulation of peripheral nerve hemopexin: an analysis in rats and in C57BL/6J mice.** *Journal of neuroimmunology* 2000, **107(1)**:29-41.
70. Tolosano E, Altruda F: **Hemopexin: structure, function, and regulation.** *DNA and cell biology* 2002, **21(4)**:297-306.
71. Kale M, Rathore N, John S, Bhatnagar D: **Lipid peroxidative damage on pyrethroid exposure and alterations in antioxidant status in rat erythrocytes: a possible involvement of reactive oxygen species.** *Toxicology letters* 1999, **105(3)**:197-205.
72. Ferreira G, Moura J, Franco R: **Iron metabolism: inorganic biochemistry and regulatory mechanisms.** Germany: Wiley-VCH; 1999:65-93.
73. Connor JR: **Iron acquisition and expression of iron regulatory proteins in the developing brain: manipulation by ethanol exposure, iron deprivation and cellular dysfunction.** *Developmental neuroscience* 1994, **16(5-6)**:233-247.
74. Smith A, Hunt RC: **Hemopexin joins transferrin as representative members of a distinct class of receptor-mediated endocytic transport systems.** *European journal of cell biology* 1990, **53(2)**:234-245.
75. Yu RM, Wong MM, Kong RY, Wu RS, Cheng SH: **Induction of hepatic choriogenin mRNA expression in male marine medaka: a highly sensitive biomarker for environmental estrogens.** *Aquatic toxicology (Amsterdam, Netherlands)* 2006, **77(4)**:348-358.
76. Chen X, Li VW, Yu RM, Cheng SH: **Choriogenin mRNA as a sensitive molecular biomarker for estrogenic chemicals in developing brackish medaka (Oryzias melastigma).** *Ecotoxicology and environmental safety* 2008, **71(1)**:200-208.
77. Lee C, Na JG, Lee KC, Park K: **Choriogenin mRNA induction in male medaka, Oryzias latipes as a biomarker of endocrine disruption.** *Aquatic toxicology (Amsterdam, Netherlands)* 2002, **61(3-4)**:233-241.
78. Ueno T, Yasumasu S, Hayashi S, Iuchi I: **Identification of choriogenin cis-regulatory elements and production of estrogen-inducible, liver-specific transgenic Medaka.** *Mechanisms of development* 2004, **121(7-8)**:803-815.
79. Kliewer SA, Goodwin B, Willson TM: **The nuclear pregnane X receptor: a key regulator of xenobiotic metabolism.** *Endocrine reviews* 2002, **23(5)**:687-702.
80. Masuyama H, Nakatsukasa H, Takamoto N, Hiramatsu Y: **Down-regulation of pregnane X receptor contributes to cell growth inhibition and apoptosis by anticancer agents in endometrial cancer cells.** *Molecular pharmacology* 2007, **72(4)**:1045-1053.
81. Bentley PJ: **Endocrines and Osmoregulation: A Comparative Account in Vertebrates.** Volume 1. 2nd edition. New York, NY.: Springer; 2002.
82. Aydın R, Köprüçü K, Dörücü M, Köprüçü SŞ, Pala M: **Acute Toxicity of Synthetic Pyrethroid Cypermethrin on the Common Carp (Cyprinus carpio L.) Embryos and Larvae.** *Aquaculture International* 2005, **13(5)**:451-458.
83. Tipsmark CK, Madsen SS: **Rapid modulation of Na⁺/K⁺-ATPase activity in osmoregulatory tissues of a salmonid fish.** *J Exp Biol* 2001, **204(Pt 4)**:701-709.
84. Takeuchi K, Toyohara H, Sakaguchi M: **A hyperosmotic stress-induced mRNA of carp cell encodes Na⁽⁺⁾- and Cl⁽⁻⁾-dependent high affinity taurine transporter.** *Biochimica et biophysica acta* 2000, **1464(2)**:219-230.
85. Krogdahl Å, Hemre G-I, Mommsen T: **Carbohydrates in fish nutrition: digestion and absorption in postlarval stages.** *Aquaculture Nutrition* 2005, **11(2)**:103-122.
86. Baxter R, Breuer R, Brown L, Chotkowski M, Feyrer F, Gingras M, Herbold B, Mueller-Solger A, Nobriga M, Sommer T, et al.: **Pelagic**

Organism Decline Progress Report: 2007 Synthesis of Results. *Interagency Ecological Program for the San Francisco Estuary* 2008:1-86.

87. Fujita I, Sorensen PW, Stacey NE, Hara TJ: **The olfactory system, not the terminal nerve, functions as the primary chemosensory pathway mediating responses to sex pheromones in male goldfish.** *Brain, behavior and evolution* 1991, **38(6)**:313-321.
88. Tierney KB, Singh CR, Ross PS, Kennedy CJ: **Relating olfactory neurotoxicity to altered olfactory-mediated behaviors in rainbow trout exposed to three currently-used pesticides.** *Aquatic toxicology (Amsterdam, Netherlands)* 2007, **81(1)**:55-64.
89. Moore A, Waring CP: **The effects of a synthetic pyrethroid pesticide on some aspects of reproduction in Atlantic salmon (*Salmo salar* L.** *Aquatic toxicology (Amsterdam, Netherlands)* 2001, **52(1)**:1-12.
90. Bettini E, Porta AR, Dahmen N, Wang H, Margolis FL: **Expressed sequence tags (EST) identify genes preferentially expressed in catfish chemosensory tissues.** *Brain research* 1994, **23(4)**:285-291.
91. Baskerville-Bridges B, Lindberg JC, Doroshov SI: **Manual for the Intensive Culture of Delta Smelt (*Hypomesus transpacificus*).** Second edition. University of California, Department of Animal Science Davis, CA; 2005:1-63.
92. Smyth GK: **Limma: linear models for microarray data.** *Bioinformatics and computational biology solutions using R and Bioconductor* 2005:397-420
[<http://www.springerlink.com/content/g26110k024423738/>]. New York, NY.: Springer
93. Livak KJ, Schmittgen TD: **Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method.** *Methods (San Diego, Calif)* 2001, **25(4)**:402-408.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

