

# BMJ Open Non-typhoidal *Salmonella* in Calabria, Italy: a laboratory and patient-based survey

Valentina Mascaro,<sup>1</sup> Claudia Pileggi,<sup>1</sup> Maria Crinò,<sup>1</sup>  
Yolande Therese Rose Proroga,<sup>2</sup> Maria Rosaria Carullo,<sup>2</sup> Caterina Graziani,<sup>3</sup>  
Fabio Arigoni,<sup>4</sup> Pasquale Turno,<sup>4</sup> Maria Pavia<sup>1</sup>

**To cite:** Mascaro V, Pileggi C, Crinò M, *et al.* Non-typhoidal *Salmonella* in Calabria, Italy: a laboratory and patient-based survey. *BMJ Open* 2017;7:e017037. doi:10.1136/bmjopen-2017-017037

► Prepublication history and additional material for this paper are available online. To view these files please visit the journal online (<http://dx.doi.org/10.1136/bmjopen-2017-017037>).

Received 27 March 2017  
Revised 25 May 2017  
Accepted 28 June 2017



CrossMark

<sup>1</sup>Department of Health Sciences, University of Catanzaro 'Magna Græcia', Catanzaro, Italy

<sup>2</sup>Department of Food Microbiology-Centro Pilota Tipizzazione Salmonelle, Istituto Zooprofilattico Sperimentale del Mezzogiorno, Portici, Naples, Italy

<sup>3</sup>Dipartimento di Sanità Pubblica Veterinaria e Sicurezza Alimentare, Reparto di Epidemiologia Veterinaria e analisi del rischio, Istituto Superiore di Sanità, Rome, Italy

<sup>4</sup>Dipartimento Tutela della Salute, Task Force per le Attività Veterinarie, Regione Calabria, Catanzaro, Italy

## Correspondence to

Maria Pavia; [pavia@unicz.it](mailto:pavia@unicz.it)

## ABSTRACT

**Introduction** Although there has been a decrease in the number of cases of salmonellosis in the European Union, it still represents the primary cause of foodborne outbreaks. In Calabria region, data are lacking for the incidence of human non-typhoid salmonellosis as active surveillance has never been carried out.

**Objective** To report the results of a laboratory and patient-based morbidity survey in Calabria to describe the incidence and distribution of *Salmonella* serovars isolated from humans, with a focus on antimicrobial resistance patterns.

**Methods** Positive cultures from human samples were collected from every laboratory participating in the surveillance, with a minimum set of information about each isolate. A questionnaire was then administered to the patients by telephone interview to assess the potential risk exposures. *Salmonella* isolates underwent biochemical identification, molecular analysis by PCR and antimicrobial susceptibility testing by the disk-diffusion method.

**Results** During a 2-year period, 105 strains of *Salmonella* spp were isolated from samples of patients with diarrhoea, with the highest isolation rate for children aged 1–5 years. The standardised rate was 2.7 cases per 1 00 000 population. The most common *Salmonella* isolates belonged to monophasic variant of *S. Typhimurium* (*S.* 4,[5],12:i:-) (33.3%), followed by *S. Typhimurium* (21.9%). 30.5% of the isolates were susceptible to all microbial agents tested and the most common pan-susceptible serotype was *S. Napoli* (100%). *S.* 4,[5],12:i:- was resistant to ampicillin, streptomycin, sulfonamides and tetracyclines in 42.9% cases, while resistance to quinolones was seen in 14.3% of the isolates.

**Conclusions** The results provide evidence that an active surveillance system effectively enhances *Salmonella* notifications. The high prevalence of antimicrobial resistance, including resistance to quinolones and multiresistance, enforces the need to strengthen strategies of surveillance and monitoring of antimicrobial use.

## INTRODUCTION

*Salmonella* infections are among the most common foodborne diseases: they cause approximately 93.8 million illnesses and 155 000 deaths annually around the world.<sup>1</sup> Although there has been a steady decrease in

## Strengths and limitations of this study

- This study reports for the first time the results of a laboratory and patient-based survey surveillance of human cases of salmonellosis in Calabria, a region with one of the lowest notification rates.
- All hospital and outpatient laboratories (public or private) were contacted and those who performed microbiological analyses for the detection of *Salmonella* spp were considered eligible in a surveillance network.
- As a surveillance system, the study inevitably underestimates diseases occurring in the community, analysing only a fraction of the total number of cases of illness in the population.

the number of cases in the European Union (EU), salmonellosis represents the primary cause of foodborne outbreaks and the second most frequently reported zoonosis. Recently, it has been estimated that the overall incidence of human salmonellosis in the EU general population is approximately 20.4 cases per 1 00 000 population each year, while in Italy the incidence is 7.5 cases per 1 00 000 population each year.

In Italy, surveillance of acute infectious gastroenteritis and outbreaks of food-borne diseases is part of the activities of the Italian National Surveillance System of diseases (SIMI) whereas Enter-net (IT-ENTER-NET) is a laboratory-based surveillance system for enteropathogens based on a network of clinical microbiology diagnostic laboratories.<sup>2,3</sup> It is complementary to SIMI and collects microbiological information on *Salmonella* isolates from human cases each year.

The level of under-reporting of infectious diseases and laboratory surveillance, is expected to vary between countries, depending on differences in organisation and effectiveness of local systems.<sup>4</sup>

These surveillance systems vary greatly in their performance, representativeness and data quality in different Italian regions, since laboratories that participate in the surveillance are not homogeneously distributed nationally. There are differences in sensitivity among Italian regions; in particular, it has been shown that northern regions of Italy are generally more sensitive in detecting cases, leading to significantly higher notification rates in comparison with the national average.<sup>5</sup> In Calabria region, data for the incidence of human non-typhoid salmonellosis (NTS) are lacking.<sup>5</sup> Surveillance has never been carried out, but the information which could be obtained would help to guide clinicians in the treatment of those groups at risk, such as infants, the elderly and immunocompromised patients, who would not benefit from simple rehydration therapy, and would help local health authorities in the identification of appropriate control measures.<sup>6</sup>

Within the framework of a research project supported by the Ministry of Health, the study reports the results of a laboratory and patient-based morbidity survey in Calabria to describe the incidence and distribution of *Salmonella* serovars isolated from humans, with a focus on antimicrobial resistance patterns.

## MATERIALS AND METHODS

A laboratory and patient-based survey for *Salmonella* spp infections has been carried out in Calabria (southern Italy), a region with 1 980 533 inhabitants (population estimates for 2014) living in three major cities and several smaller centres.

The five local health units provided a list of all clinical microbiology diagnostic laboratories of Calabria region. All hospital and outpatient laboratories (public or private) were contacted and those who carried out microbiological analyses for the detection of *Salmonella* spp were considered eligible and invited to participate. A healthcare worker was trained to carry out the survey for each laboratory agreeing to participate. The project took place between February 2013 and March 2015.

### Surveillance system

Positive cultures from human samples were obtained from the different laboratories. Every laboratory sent the isolates to the coordinating unit with accompanying documentation. A minimum set of information about each isolate was collected: name of the laboratory that reached the microbiological diagnosis, isolation date, sample type (stools or blood), patient sex, age, residence and telephone number. The unit provided biochemical identification and collected the strains for dispatch to the reference laboratories (Veterinary Public Health Institute of Southern Italy and the National Institute of Health) for typing.

A questionnaire was then administered to patients by telephone interview by a trained physician after appropriate informed consent had been obtained. If a patient

was aged <18 years, an adult member of the household responded to the questionnaire.

### Review instrument

The questionnaire used in the survey (online supplementary data) was a validated Italian version<sup>7</sup> review instrument currently used in the surveillance of acute gastroenteritis, and was adapted from the Center for Disease Control and Prevention standardised foodborne disease outbreak questionnaire.<sup>8</sup>

Validation of the survey instrument was performed through assessment of internal and test-retest (external) reliability, in addition to face and content validity. Test-retest reliability was checked in a pilot study of a sample of patients to ensure clarity and ease of completion and to improve the validity of responses and the information included. Modifications were then made as necessary.

The questionnaire included 43 questions divided into five sections. Responses were obtained in a variety of formats: closed-ended questions with multiple answers possible, yes or no questions and open-ended questions.

The sections of the questionnaire were as follows: (1) sociodemographic characteristics of the patient; (2) details of whether the patients or their cohabitants had or had not any sign of illness, information about prescribed culture tests, health service use and eventual hospitalisation, drugs taken and travel in the previous 2 weeks; (3) information about food exposures and the origin of foodstuff; (4) other exposures (ie, contact with animals, outdoor activities, water sports); (5) information about the source of commonly used water and purpose of use.

### Microbiological methods

*Salmonella* isolates were biochemically identified using conventional microbiological methods and the automated Vitek system (BioMérieux). Serotyping was performed by the slide agglutination method using commercial O and H antisera (the antisera were purchased from Statens Serum Institut, Denmark) according to Kauffmann-White-Le Minor scheme.<sup>9</sup>

Strains were definitively assigned to serovar Typhimurium or *S.* 4,[5],12:i:- on the basis of the presence or the absence of the *fljB* gene tested by PCR.<sup>10</sup>

Antimicrobial susceptibility was performed by the disk-diffusion method (Kirby-Bauer). The following antimicrobial agents and concentration (µg) were used: amikacin (30), amoxicillin/clavulanic acid (20/10), ampicillin (10), cefoxitin (30), ceftriaxone (30), cephalothin (30), chloramphenicol (30), ciprofloxacin (5), gentamicin (10), kanamycin (30), nalidixic acid (30), neomycin (30), nitrofurantoin (100), streptomycin (10), sulfamethoxazole/trimethoprim (23.75/1.25), trimethoprim (5) and tetracycline (30). Classification of the categories as susceptible, intermediate or resistant was based on the Clinical and Laboratory Standards Institute guidelines<sup>11</sup> and for the purpose of analysis, all readings classified as intermediate were considered as resistant where necessary.

The study protocol was ratified by the institutional ethics committee ('Mater Domini' Hospital of Catanzaro, Italy) (7/05/2013).

### Data analysis

Data were stored and analysed using an appropriate database. Statistical analysis was performed using STATA software programme, version 11 (Stata Corporation, College Station, Texas, USA).

Age-specific and standardised incidence rates were calculated. Direct standardisation using the Italian population as standard was performed. Demographic data were obtained from the National Institute of Statistics (ISTAT).<sup>12</sup>

## RESULTS

The health authorities listed a total of 245 regional microbiology diagnostic laboratories. After exclusion of those which did not perform microbiological analyses for the detection of *Salmonella* spp, the network comprised 114 laboratories. Of these, 110 agreed to participate, with a response rate of 96.5%. Among the participating laboratories, 25 were public laboratories (hospital or outpatient based) and 85 were private.

During the 2-year survey, 108 cultures from stool samples of patients with a clinical picture compatible with salmonellosis were collected from the participating laboratories; among these samples, 105 were confirmed as strains of *S. enterica*. Demographic data were available for 102 of the patients, and complete telephone interviews for 70/105 (66.7%) patients. Four patients (3.8%) were not directly interviewed owing to a lack of contact, and thus data were obtained from medical records.

Distribution of cases and age-specific rates are shown in table 1. The highest isolation rate was for children aged 1–5 years, followed by children aged 6–14 years and in those aged ≥65 years (table 1). The standardised rate was 2.7 cases per 100 000 population. Data from the phone interviews showed that, as expected, the three most frequently reported clinical symptoms were diarrhoea

(100% of responders), fever (89.7%) and abdominal pain (65.4%).

At least one family member of the included patients had similar symptoms in 19 cases and, of these, only four were recommended by their general practitioner or community-based paediatrician to have a culture test for the diagnosis of salmonellosis: one of these was positive for *Salmonella* spp and was included in the study.

All 78 patients who answered the questionnaire sought medical assistance: 54 (69.2%) of them consulted a general practitioner or community-based paediatrician, while 24 (30.8%) went to the emergency department. Antimicrobial agents were used for 78% of the responders, for an average of 9.1 days (±3.9), and 82.3% of the patients from whom we could gather this information were hospitalised, with an average stay of 5.7 days (±3.4). Most frequently used antimicrobial agents were ceftriaxone (41%), sulfamethoxazole/trimethoprim (13.1%), amoxicillin/clavulanic acid (13.1%) and clavulanic acid (8.2%).

All responders referred to the consumption of food deemed to be potentially unsafe. In most cases food eaten within 24–48 hours before the onset of symptoms included milk and milk products (90% of cases); cooked meat (90.5% of cases); sausages (71.4% of cases); cooked eggs (64.3% of cases). Less common was the consumption of cooked (38.6%) and raw (34.3%) vegetables and raw eggs (20%).

*Salmonella* isolates belonged to 19 serotypes with monophasic variant of *S. Typhimurium* (*S.* 4,[5],12:i:-) being the most common (33.3% of the total isolates), followed by *S. Typhimurium* (21.9% of the total isolates), *S. Enteritidis* (13.3% of the total isolates), *S. Napoli* (9.5% of the total isolates) and *S. Infantis* (2.9% of the total isolates). Other serovars represented 19% of the total isolates (table 2).

Of all 105 NTS isolates, 30.5% were susceptible to all microbial agents tested. The most common serotypes of pan-susceptible isolates were *S. Napoli* (100%), *S. Enteritidis* (50%) and *S. Infantis* (33.3%). *S. Typhimurium* serovar 4,[5],12:i:- showed resistance to ampicillin,

**Table 1** Number of *Salmonella* isolates for age group, year and average annual incidence in Calabria, Italy 2014–2015

Age groups	2014 total cases (N. of isolates)	2015 total cases (N. of isolates)	Total cases	Annual average incidence Calabria/100 000	Annual average incidence Italy/100 000
0–11 Months	6	3	9	27.4	13.5*
1–5 Years	36	14	50	28.4	32.5*
6–14 Years	13	7	20	5.9	8*
15–64 Years	6	3	9	0.5	1.6*
≥65 Years	9	5	14	1.8	2.8*
Total	70	32	105†	2.7‡	7.5§

\*Distribution of the annual isolation rates of *Salmonella* spp serovars in Italy during the period between 2000 and 2011.<sup>5</sup>

†Number of total cases is different from the total cases number by age groups because of lack of information about age in three patients.

‡Standardised incidence rate of *Salmonella* spp isolation.

§Notification rate for confirmed cases in 2011.<sup>14</sup>

**Table 2** Antimicrobial resistance patterns of the main *Salmonella* serovars isolated from human cases in Calabria, 2014–2015

Serovars	No of isolates	Antibiotic resistance patterns (%)						Other patterns
		Sensitive	ASSuT	ASSuT +other	ACSSuT	ACSSuT+other	Quinolones	
S. 4,[5],12:i:-	35	0	42.9	17.1	5.7	0	5.7	31.4
S. Typhimurium	23	30.4	13	21.7	0	4.4	17.4	21.7
S. Enteritidis	14	50	0	0	0	0	28.6	21.4
S. Napoli	10	100	0	0	0	0	0	0
S. Infantis	3	33.3	0	0	0	0	66.7	0
Other serovars	20	35	0	0	0	10	15	45
Total*	105	30.5	17.1	10.5	1.9	2.9	14.3	26.7

\*Total rate exceeds 100% because *Salmonella* spp serotypes resistant to quinolones are also present in the categories ASSuT+other or ACSSuT+other.

ACSSuT, ASSuT + chloramphenicol; ASSuT, ampicillin, streptomycin, sulfamethoxazole, tetracycline; Quinolones, this category includes all patterns in which nalidixic acid or ciprofloxacin are present; S. 4,[5],12:i:- monophasic variant of *Salmonella* Typhimurium.

streptomycin, sulfonamides and tetracyclines (R-type ASSuT) with or without additional resistances in 60% of cases. Over the study period resistance to quinolones tested (nalidixic acid and ciprofloxacin) was seen in 15 (14.3%) of the total isolates and among these the most common serovar was *S. Infantis* (66.7% of cases) and *S. Enteritidis* (28.6% of cases) (table 2).

## DISCUSSION

This study reports for the first time the results of surveillance for salmonellosis in the population in Calabria, a region with one of the lowest notification rates.<sup>5 13</sup>

The incidence rate (2.7 cases per 100 000 population) differs significantly from that published by the European Food Safety Authority, which collects data on salmonellosis cases reported in the EU; they reported in 2011 a higher Italian rate of 7.5 cases per 100 000 population.<sup>14</sup> Recently, there has been a significant reduction in reported cases rates across the EU.<sup>14 15</sup> We know that isolation rates are usually considerably lower in the southern part of Italy,<sup>5</sup> but in northern Italy surveillance systems are more sensitive in detecting cases of infectious gastroenteritis,<sup>16</sup> leading to relatively higher national notification rates of salmonellosis.

The total number of cases between the 2 years of our survey differs, showing a reduction of isolates that is only partly explained by the trend for reduction across the EU.<sup>14 15</sup> This might be due to under-reporting of cases.

In addition, any surveillance system inevitably underestimates diseases occurring in the community for different reasons. First of all many subjects do not seek medical attention and for salmonellosis, only a proportion of symptomatic patients submit stool specimens for investigation; moreover, the sensitivity of laboratory identification varies according to the pathogen and not all identified pathogens are reported to the surveillance centre.<sup>15 16</sup>

Isolation rates were highest in children aged 1–5 years. This finding is consistent with that of other studies that

have shown that younger children are at greater risk of infection.<sup>5 17</sup> This might also be due to an overestimation of cases in certain age group. There is a greater proportion of symptomatic infections among children, who also are at risk of dehydration, and they are more likely to see a doctor and therefore to have a stool examination (ie, detection bias).<sup>18</sup>

Almost all the data in our report were collected from hospitalised patients. However, we know that hospitalisations represent only a fraction of the total number of cases of illness in the population: for a pathogen of moderate virulence, a relatively small proportion of those who seek medical attention will be hospitalised. Since our surveillance was extended also to outpatient laboratories, it seems that in non-hospitalised cases, probably with milder disease than in those admitted to hospital, microbiological analyses are less frequently performed. Moreover, asymptomatic and mild cases of disease are difficult to enumerate because of under-reporting by physicians and differences of diagnostic capabilities and protocols among laboratories.<sup>15</sup>

Seventy-eight per cent of patients received antibiotic therapy, even though there is no evidence for its benefit for NTS diarrhoea in otherwise healthy people, and routine antimicrobial therapy is not recommended for mild and moderate cases of NTS.<sup>19</sup> Therefore, such a high percentage might be because in the 2-year survey most of the antibiotic-treated patients were hospitalised and were 0–5-year-old children in more than 50% of cases, justifying the treatment for a severe disease that could not be treated by only electrolyte replacement and rehydration.

The most frequently isolated serovars in the 2 years survey were *S. 4,[5],12:i:-* (33.3% of all isolated) and *S. Typhimurium* (21.9% of all isolated). The monophasic variant of serotype Typhimurium, *S. 4,[5],12:i:-*, has only rarely been reported among *Salmonella* serovars isolated before 1993, whereas since 2000 it has been found in human clinical cases, different animal species and foods in different continents, including Europe, Asia



and South and North America.<sup>5 20</sup> In Europe, although *S. Enteritidis* still remains the most reported serovar, *S. 4,[5],12:i:-* showed a marked increase in foodborne infections associated with pig meat.<sup>21</sup> *S. 4,[5],12:i:-* has well-established reservoirs of livestock that are likely to have favoured its entrance into the food chain, and consequent rapid emergence and dissemination.<sup>22</sup> Our findings are consistent with the results of other Italian studies, with *S. Typhimurium* being the most reported in the period 2000–2011.<sup>5 23</sup> Moreover, a recent study in northeastern Italy reported that the most commonly isolated serotypes were *S. Typhimurium* and its monophasic variant, with *S. Enteritidis* being the third most isolated serovar.<sup>24</sup>

Evidence for the decrease in human salmonellosis in Italy since the late 1990s relates to specific serovars, namely, *S. Enteritidis* and *S. Infantis*, whereas there is evidence that other serovars have emerged (*S. 4,[5],12:i:-*, *S. Derby* and *S. Napoli*) or remained fairly stable (*S. Typhimurium*).<sup>2 13</sup> The decrease of *S. Enteritidis* is probably due to the implementation of control measures against *Salmonella*, especially within the poultry industry, such as the vaccination of laying hens, improved hygiene and education of food workers.<sup>25 26</sup>

In line with other studies in Italy, a substantial proportion of *S. Napoli* was found (9.5% of all isolated) during our 2-year survey. In Italy, there is a slight but constant increase of cases of *S. Napoli*<sup>27</sup> and a low association with foodborne exposure.<sup>28</sup> The number of cases of *S. Napoli* in Italy has been rising since 2000, but no factors accounting for this trend have been identified.<sup>28</sup> There is a possibility of the presence of *S. Napoli* reservoirs in the environment; results obtained applying different molecular typing techniques have shown the presence of *S. Napoli* in different settings, such as wildlife and surface waters.<sup>29</sup>

Rates of antimicrobial resistance varied according to serotype. All isolates belonging to serotype *S. 4,[5],12:i:-* were resistant to at least one antibiotic, while the other serovars were resistant to at least one antimicrobial agent in between 5.7% and 66.7% of cases. The only exception was *S. Napoli* isolates, which were susceptible to all the tested antibiotics. In Europe, *S. Typhimurium* has consistently exhibited the highest percentage of strains with multiple resistances, mainly of the ACSSuT type,<sup>30</sup> but the frequency of dissemination of this multiresistant clone has reduced since 2002.<sup>31</sup> On the other hand, in Italy strains with the ASSuT pattern have become increasingly common, both in *S. Typhimurium* and *S. 4,[5],12:i:-* since 2000,<sup>14 29</sup> and more recent data reported that this resistance pattern is present in 36.1% of *Salmonella* isolates, mainly in *S. 4,[5],12:i:-*. Moreover, *S. 4,[5],12:i:-* only rarely showed the ACSSuT pattern.<sup>13</sup>

The *Salmonella* serovar *4,[5],12:i:-* resistant to ampicillin, streptomycin, sulfonamide and tetracycline (pattern ASSuT) is extensively circulating in Denmark, Italy, the United Kingdom and in Greece,<sup>28 32 33</sup> and it has been frequently isolated from animal sources, in particular, swine.<sup>34</sup> In agreement with previous studies, we found high rates of *S. 4,[5],12:i:-* that presented the

ASSuT pattern with or without other resistances (60% of all *S. 4,[5],12:i:-* isolated).

Multidrug resistant *Salmonella* strains are indeed of worldwide interest and a serious public health concern, because it has been shown that outbreaks caused by multidrug resistant NTS are associated with an increased rate of hospitalisation compared with outbreaks caused by pan-susceptible NTS.<sup>35 36</sup>

Over the study period resistance to quinolones (nalidixic acid or ciprofloxacin) was seen in 14.3% of the isolates. Resistance to quinolones among *Salmonella* around the world was low until the 1990s, when it started to increase<sup>21 37</sup> and the serovar most involved was *S. Enteritidis*.<sup>38</sup> On the contrary, in Italy the resistance rate to quinolones and fluoroquinolones tends to be relatively low.<sup>24</sup>

Emergence of antibiotic resistance has been related to the massive use of antimicrobial agents both in human and in veterinary medicine. Especially in livestock animals, antimicrobial agents are used for treatment and prophylaxis of diseases and in the past have been used as growth promoters.<sup>39</sup> This particularly applied to the poultry industry, until European bans led to a substantial reduction in the amount of antibiotics used in animal production.<sup>40</sup> Indeed, the resistance to fluoroquinolones in *Salmonella* spp is a major concern, since fluoroquinolone is one of the preferred drugs for early empirical treatment of severe gastroenteritis in adults<sup>41</sup> and drug-resistant NTS is associated with increased severity.<sup>42</sup>

## CONCLUSIONS

In conclusion, the results of the survey provide evidence of the effectiveness of an active surveillance system in enhancing the detection of cases of *Salmonella*, and identification of circulating strains associated with human infections. The high prevalence of antimicrobial resistance, including resistance to quinolones and multiresistance (R-type ASSuT), increases the need to strengthen strategies of surveillance and monitoring of antimicrobial use. It might be useful for the future to consider the sustainability of such a survey in order to enforce a true surveillance system for enteric diseases and related antimicrobial resistance problems.

**Acknowledgements** We are very grateful to the clinical microbiology diagnostic laboratories who agreed to participate in this study: A.O. Mater Domini (Professor Alfredo Focà, Dott.ssa Rossana Puccio), A.O. Pugliese Ciaccio (Dott.ssa Rosanna Masciari), P.O. Soverato (Dott. Mimmo Donato, Dott.ssa Donatella Scuteri), P.O. Soveria Mannelli (Dott. Ivan Potente), P.O. Lamezia Terme (Dott.ssa Rosa Anna Leone, Dott. Salvatore Nisticò, Dott.ssa Enza Caruso); Laboratorio Armogida (Dott. Giuseppe Armogida), Poliambulatorio Lametino (Dott.ssa Maria Teresa Germinara), Laboratorio Biomedica (Dott.ssa Anna Bressi), Laboratorio Barillaro (Dott. Rosario Crea), Laboratorio Matozzo (Dott. Francesco Matozzo), A.O. Annunziata Cosenza (Dott.ssa Cristina Giraldi), P.O. Rogliano (Dott.ssa Teresa Spataro), P.O. Praia a Mare (Dott.ssa Francesca Massara, Dott. Raffaele Diana), P.O. San Giovanni in Fiore (Dott. Amalfitano, Dott.ssa Francesca Orrico), P.O. Acri (Dott.ssa Anna Ceraldi, Dott. Michele Fusaro), P.O. Cetraro (Dott.ssa Adele Taranto), P.O. Castrovillari (Dott. Giovanni Gigliotti, Dott.ssa Rossella Maltese), P.O. San Marco Argentano (Dott. Antonio Damis), P.O. Lungro (Dott.ssa Titta Bonifati), P.O. San Francesco di Paola (Dott.ssa Marinella Zacchi, Dott.ssa Maria Maddalena Grossi), P.O. Rossano (Dott. Pasquale Paletta, Dott.ssa Maria Lucia Petrelli, Dott. Luigi Mariano), P.O. Trebisacce

(Dott.ssa Elisabetta Grillo), Laboratorio F.Ili Luca (Dott.ssa Teresa), LABMONACO (Dott. Giovanni Monaco), Laboratorio Lusal (Dott.ssa Angela Argirò), Laboratorio Salus (Dott. Sergio Gravina), BIODIAGNOSTICA (Dott.ssa Carmela Milano), Laboratorio Salimbeni (Dott. Francesco Salimbeni), Laboratorio Cosentino (Dott. Natalino Cosentino), Laboratorio De Pietro (Dott. Francesco De Pietro), Centro Diagnostico S. Nilo (Dott.ssa Annamaria Parise), Laboratorio Ricci (Dott.ssa Elisabetta Pulerà), Laboratorio Bilotta (Dott.ssa Maria Carmela Bilotta), Diagnostica Medica Coscarella (Dott. Fabio Coscarella), Laboratorio Nitti (Dott.ssa Giulia Rossi), Laboratorio Leporace (Dott.ssa Ivana Benvenuti), Laboratorio Biocenter (Dott. Salvatore Di Nardo), Laboratorio Ippolito (Dott.ssa Gabriella Capparelli), Laboratorio Biocenter Check-up (Dott.ssa Emanuela De Marco), Laboratorio Medical (Dott.ssa Laurie-Lynn Carelli), Laboratorio Bios (Dott.ssa Patrizia Marrelli), Laboratorio Perugini (Dott.ssa Anna Perugini), Laboratorio Miceli (Dott. Francesco Miceli), Laboratorio Gamma (Dott.ssa Beatrice Valente), Laboratorio Politano/Loizzo (Dott. Maria Grazia Dielsi), Laboratorio Borzi (Dott. Domenico Borzi), Laboratorio Biolab (Dott.ssa Maria Clorinda Aronna), Laboratorio Biomedical (Dott.ssa Maria Concetta Rummolo), Laboratorio Serio (Dott.ssa Anna Maria Serio), Laboratorio Analysis Centre (Dott. Francesco Sarubbi), Centro diagnostico Sybaris (Dott.ssa Rosina Tirone), Laboratorio Centro Jonico (Dott. Vincenzo Ippolito), Laboratorio Di Donna (Dott. Paolo Di Donna), Laboratorio Vercillo (Dott.ssa Marina Vercillo), Laboratorio Nicastro (Dott. Giancarlo Nicastro), Casa di cura Cascini (Dott. Giuseppe Avolicino), Istituto Tricarico (Dott.ssa Maria Francesca Bianco), P.O. Crotone (Dott.ssa Rita Cizza), Laboratorio Bios (Dott.ssa Fortunata Salvemini), Laboratorio Altomari (Dott.ssa Maria Bonpignano), Laboratorio Fleming (Dott. Oreste Antonio Setti), Laboratorio Dott. Rodio (Dott. Gaetano Papparino), Laboratorio Volante (Dott.ssa Antonietta Rizzo), Laboratorio Biolav (Dott. Salvatore Valente), Laboratorio Via (Dott. Salvatore Cimieri), Laboratorio Riillo (Dott. Pasquale Riillo), Laboratorio Morrone (Dott.ssa Teresa Primerano), Laboratorio Pasteur (Dott. Giovanni Ierardi), A.O. Melacrino-Morelli (Dott. Angelo Barbaro), P.O. Locri (Dott.ssa Antonella Rodinò), P.O. Scilla (Dott.ssa Serafina Rullo), Dott.ssa Francesca Aromato), P.O. Melito Porto Salvo (Dott. Ferdinando Familiari), P.O. Polistena (Dott. Girolamo Cangemi, Dott. Alfredo Rosselli), Laboratorio Saladino (Dott. Rocco Tassone), Laboratorio Tassone (Dott. Rocco Tassone), Laboratorio Antico (Dott. Alfredo Antico), Laboratorio Alfa Bianco (Dott.ssa Maria Giuseppina Strati), Poliambulatorio Specialistico (Dott.ssa Rosita Murdocca), Poliambulatorio Ventra (Dott. Giuseppe Ventra), Laboratorio San Rocco (Dott. Giuseppe Santoro), Laboratorio Cavaliere (Dott. Francesco Cavaliere), Laboratorio L.A.C (Dott. Domenico Fotia, Dott. Ferdinando Maringola), Centro Diagnostico Gamma (Dott. Vincenzo Macino), Laboratorio Caruso (Dott. Antonio Caruso), Laboratorio Salus (Dott. Francesco Mobicci), Laboratorio Minerva (Dott.ssa Maria Campise), Laboratorio Sant'Anna (Dott. Massimo Saiaci), Laboratorio S. Antonio (Dott. Antonino Laurendi), Laboratorio Libri (Dott. Fortunato Libri), Laboratorio Serranò (Dott. Francesco Serranò), Laboratorio Richichi (Dott. Roberto Richichi), Istituto GMM (Dott. Antonio Messina), Laboratorio Zoccali (Dott.ssa De Salvo), Laboratorio Simef (Dott. Stefano Votanol), Laboratorio Pasteur (Dott.ssa Daniela Ricioppo), Laboratorio Siracusa (Dott. Francesco Siracusa), Laboratorio Europa (Dott. Alfio Palazzolo), Laboratorio Barreca (Dott. Giorgio Barreca), Laboratorio I.D.E.A. (Dott. Giuseppe De Angelis), Laboratorio De Blasi (Dott.ssa Antonia Malara), Polidiagnostica Meridionale (Dott. Gregorio Greco), Laboratorio Clinical Control (Dott.ssa Cribari), P.O. Vibo Valentia (Dott. Enzo Majolo, Dott. Tito Rodà), P.O. Tropea (Dott. Michele Cutellè), Laboratorio Salus (Dott. Vincenzo Mangialavori), Laboratorio Nusdeo (Dott. Sergio Pacetti), Laboratorio Biomedical (Dott. Pino Lo Iacono).

**Contributors** VM contributed to the data analysis and its interpretation, and wrote the manuscript. MC made substantial contributions to the acquisition of the data, contributed to the data analysis and its interpretation. YTRP, MRC, FA, and PT participated in the acquisition of the data and contributed to the data analysis. CP and CG made substantial contributions to conception and design of the study, coordinated data collection, contributed to the data analysis and its interpretation, and revised the manuscript critically for important intellectual content. MP conceptualised and designed the study, supervised data collection, was responsible for the data analysis and interpretation and revised the manuscript. All authors had full access to all the data (including statistical reports and tables) in the study and can take responsibility for the integrity of the data and the accuracy of the data analysis.

**Funding** This work was supported by Italian Ministry of Health-Centro di Controllo delle Malattie 'Risk assessment of zoonotic infections due to consumption of fresh produce grown in areas with high livestock density'-grant number CCM 2012/604.

**Competing interests** All authors have completed the ICMJE uniform disclosure form at [www.icmje.org/coi\\_disclosure.pdf](http://www.icmje.org/coi_disclosure.pdf) and declare: no financial relationships with any organisations that might have an interest in the submitted work in the previous three years; no other relationships or activities that could appear to have influenced the submitted work.

**Patient consent** The article does not contain personal medical information about an identifiable living individual.

**Ethics approval** Institutional ethical committee ('Mater Domini' Hospital of Catanzaro, Italy).

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Data sharing statement** Survey data were not included in the present article and are available from the authors.

**Open Access** This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

© Article author(s) (or their employer(s) unless otherwise stated in the text of the article) 2017. All rights reserved. No commercial use is permitted unless otherwise expressly granted.

## REFERENCES

1. Majowicz SE, Musto J, Scallan E, *et al.* International collaboration on enteric disease 'burden of illness' studies. The global burden of nontyphoidal *Salmonella* gastroenteritis. *Clin Infect Dis* 2010;50:882–9.
2. Hardnett FP, Hoekstra RM, Kennedy M, *et al.* Emerging infections program FoodNet Working Group. Epidemiologic issues in study design and data analysis related to FoodNet activities. *Clin Infect Dis* 2004;38(Suppl 3):S121–6.
3. Haagsma JA, Geenen PL, Ethelberg S, *et al.* Med-Vet-Net Working Group. Community incidence of pathogen-specific gastroenteritis: reconstructing the surveillance pyramid for seven pathogens in seven European Union member states. *Epidemiol Infect* 2013;141:1625–39.
4. Gibbons CL, Mangan MJ, Plass D, *et al.* Burden of communicable diseases in Europe (BCoDE) consortium. Measuring underreporting and under-ascertainment in infectious disease datasets: a comparison of methods. *BMC Public Health* 2014;14:147.
5. Graziani C, Mughini-Gras L, Owczarek S, *et al.* Distribution of *Salmonella enterica* isolates from human cases in Italy, 1980 to 2011. *Euro Surveill* 2013;18:20519.
6. Maraki S, Papadakis IS. Serotypes and antimicrobial resistance of human nontyphoidal isolates of *Salmonella enterica* from Crete, Greece. *Interdiscip Perspect Infect Dis* 2014;2014:1–5.
7. Servizio Sanitario della Toscana. Linee Guida per la corretta gestione degli episodi di malattie veicolate da alimenti. <http://www.epicentro.iss.it/problemi/tossinfezioni/pdf/LineeGuidaToscanaTossinfezioni.pdf> (accessed 5 Aug 2016).
8. Center for Disease Control and Prevention. Foodborne disease outbreak investigation and surveillance tools. Minnesota Questionnaire. [https://www.cdc.gov/foodsafety/pdfs/minnesota-standard\\_ques.pdf](https://www.cdc.gov/foodsafety/pdfs/minnesota-standard_ques.pdf) (accessed 5 Aug 2016).
9. Grimont PAD, Weill FX. Antigenic formulae of the *Salmonella* serovars. 9th ed. Paris: World Health Organization collaborating center for reference and research on *Salmonella*, Institut Pasteur, 2007.
10. Echeita MA, Herrera S, Garaizar J, *et al.* Multiplex PCR-based detection and identification of the most common *Salmonella* second-phase flagellar antigens. *Res Microbiol* 2002;153:107–13.
11. Clinical and Laboratory Standards Institute (CLSI). *Performance standards for antimicrobial susceptibility testing; twenty-second informational supplement*. Wayne: PA: clinical and laboratory standards institute, 2012. CLSI document M100S22.
12. Istituto Nazionale di Statistica (ISTAT). Demografia in cifre. Statistiche demografiche ISTAT. <http://demo.istat.it/pop2014/index.htm> (accessed 22 Oct 2015).
13. Dionisi AM, Filetici E, Owczarek S, *et al.* Enter-net: sorveglianza delle infezioni trasmesse da alimenti e acqua. Rapporto dell'attività 2007–2009. *Not Ist Super Sanità* 2011;24:3–10.
14. European Food Safety Authority and European Centre for Disease Prevention and Control (EFSA and ECDC). The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2013. *EFSA Journal* 2015;13:3991.
15. Wheeler JG, Sethi D, Cowden JM, *et al.* Study of infectious intestinal disease in England: rates in the community, presenting to general practice, and reported to national surveillance. *BMJ* 1999;318:1046–50.
16. Mughini-Gras L, Graziani C, Biorci F, *et al.* Surveillance of acute infectious gastroenteritis (1992–2009) and food-borne disease

- outbreaks (1996-2009) in Italy, with a focus on the Piedmont and Lombardy regions. *Euro Surveill* 2012;17:20098.
17. Olsen SJ, Bishop R, Brenner FW, et al. The changing epidemiology of *Salmonella*: trends in serotypes isolated from humans in the United States, 1987-1997. *J Infect Dis* 2001;183:753-61.
  18. Mor SM, DeMaria A, Naumova EN. Hospitalization records as a tool for evaluating performance of food- and water-borne disease surveillance systems: a Massachusetts case study. *PLoS One* 2014;9:e93744.
  19. Onwuezobe IA, Oshun PO, Odigwe CC. Antimicrobials for treating symptomatic non-typhoidal *Salmonella* infection. *Cochrane Database Syst Rev* 2012;11:CD001167.
  20. Switt AI, Soyer Y, Warnick LD, et al. Emergence, distribution, and molecular and phenotypic characteristics of *Salmonella enterica* serotype 4,[5],12:i:-. *Foodborne Pathog Dis* 2009;6:407-15.
  21. Hopkins KL, Kirchner M, Guerra B, et al. Multiresistant *Salmonella enterica* serovar 4,[5],12:i:- in Europe: a new pandemic strain? *Euro Surveill* 2010;15:19580.
  22. EFSA panel on biological hazards (BIOHAZ). Scientific opinion on monitoring and assessment of the public health risk of "Salmonella Typhimurium-like" strains. *EFSA Journal* 2010;8:48.
  23. Busani L, Graziani C, Battisti A, et al. Antibiotic resistance in *Salmonella enterica* serotypes Typhimurium, Enteritidis and Infantis from human infections, foodstuffs and farm animals in Italy. *Epidemiol Infect* 2004;132:245-51.
  24. Frasson I, Bettanello S, De Canale E, et al. Serotype epidemiology and multidrug resistance patterns of *Salmonella enterica* infecting humans in Italy. *Gut Pathog* 2016;8:26.
  25. Thompson CN, Phan VT, Le TP, et al. Epidemiological features and risk factors of *Salmonella* gastroenteritis in children resident in Ho Chi Minh City, Vietnam. *Epidemiol Infect* 2013;141:1604-13.
  26. O'Brien SJ. The "decline and fall" of nontyphoidal *Salmonella* in the United Kingdom. *Clin Infect Dis* 2013;56:705-10.
  27. Graziani C, Luzzi I, Owczarek S, et al. *Salmonella enterica* serovar Napoli infection in Italy from 2000 to 2013: spatial and spatio-temporal analysis of cases distribution and the effect of human and animal density on the risk of infection. *PLoS One* 2015;10:e0142419.
  28. Fisher IS, Jourdan-Da Silva N, Hächler H, et al. Human infections due to *Salmonella* Napoli: a multicountry, emerging enigma recognized by the Enter-net international surveillance network. *Foodborne Pathog Dis* 2009;6:613-9.
  29. Graziani C, Busani L, Dionisi AM, et al. Antimicrobial resistance in *Salmonella enterica* serovar Typhimurium from human and animal sources in Italy. *Vet Microbiol* 2008;128:414-8.
  30. Threlfall EJ. Antimicrobial drug resistance in *Salmonella*: problems and perspectives in food- and water-borne infections. *FEMS Microbiol Rev* 2002;26:141-8.
  31. Meakins S, Fisher IS, Berghold C, et al. Antimicrobial drug resistance in human nontyphoidal *Salmonella* isolates in Europe 2000-2004: a report from the Enter-net International Surveillance Network. *Microb Drug Resist* 2008;14:31-5.
  32. Lucarelli C, Dionisi AM, Torpdahl M, et al. Evidence for a second genomic island conferring multidrug resistance in a clonal group of strains of *Salmonella enterica* serovar Typhimurium and its monophasic variant circulating in Italy, Denmark, and the United Kingdom. *J Clin Microbiol* 2010;48:2103-9.
  33. Mandilara G, Lambiri M, Polemis M, et al. Phenotypic and molecular characterisation of multiresistant monophasic *Salmonella* Typhimurium (1,4,[5],12:i:-) in Greece, 2006 to 2011. *Euro Surveill* 2013;30:18.
  34. Romani C, Aleo A, Pellissier N, et al. Characterization of multi-drug resistance in *Salmonella* strains isolated from animals. *Ann Ist Super Sanita* 2008;44:292-300.
  35. Varma JK, Greene KD, Ovitt J, et al. Hospitalization and antimicrobial resistance in *Salmonella* outbreaks, 1984-2002. *Emerg Infect Dis* 2005;11:943-6.
  36. Majtánová L, Majtán J, Majtán V. Antimicrobial resistance of human *Salmonella enterica* serovar Typhimurium U302 strains: prevalence of R-type ASSuT in Slovakia, 2006-2011. *Jpn J Infect Dis* 2013;66:337-40.
  37. Biedenbach DJ, Toleman M, Walsh TR, et al. Analysis of *Salmonella* spp with resistance to extended-spectrum cephalosporins and fluoroquinolones isolated in North America and Latin America: report from the SENTRY Antimicrobial Surveillance Program (1997-2004). *Diagn Microbiol Infect Dis* 2006;54:13-21.
  38. Mølbak K, Baggesen DL, Aarestrup FM, et al. An outbreak of multidrug-resistant, quinolone-resistant *Salmonella enterica* serotype Typhimurium DT104. *N Engl J Med* 1999;341:1420-5.
  39. van den Bogaard AE, Stobberingh EE. Epidemiology of resistance to antibiotics. Links between animals and humans. *Int J Antimicrob Agents* 2000;14:327-35.
  40. Castanon JI. History of the use of antibiotic as growth promoters in European poultry feeds. *Poult Sci* 2007;86:2466-71.
  41. Hohmann EL. Nontyphoidal salmonellosis. *Clin Infect Dis* 2001;32:263-9.
  42. Mølbak K. Human health consequences of antimicrobial drug-resistant *Salmonella* and other foodborne pathogens. *Clin Infect Dis* 2005;41:1613-20.