



Article Coupling Reactions of Anhydro-Aldose Tosylhydrazones with Boronic Acids

Tímea Kaszás, Balázs Áron Baráth, Bernadett Balázs, Tekla Blága, László Juhász 跑, László Somsák *🗅 and Marietta Tóth *🖻

> Department of Organic Chemistry, University of Debrecen, P.O. Box 400, H-4002 Debrecen, Hungary; kaszas.timea@science.unideb.hu (T.K.); balazs.barath01@gmail.com (B.Á.B.); balazs.bernadetti@gmail.com (B.B.); tekla.tblaga@gmail.com (T.B.); juhasz.laszlo@science.unideb.hu (L.J.)

* Correspondence: somsak.laszlo@science.unideb.hu (L.S.); toth.marietta@science.unideb.hu (M.T.)

Abstract: A catalyst-free coupling reaction between *O*-peracetylated, *O*-perbenzoylated, *O*-permethyl ated, and *O*-permethoxymethylated 2,6-anhydro-aldose tosylhydrazones (*C*-(β -D-glycopyranosyl)for maldehyde tosylhydrazones) and aromatic boronic acids is reported. The base-promoted reaction is operationally simple and exhibits a broad substrate scope. The main products in most of the transformations were open-chain 1-*C*-aryl-hept-1-enitol type compounds while the expected β -D-glycopyranosylmethyl arenes (benzyl *C*-glycosides) were formed in subordinate yields only. A mechanistic rationale is provided to explain how a complex substrate may change the well-established course of the reaction.

Keywords: coupling; anhydro-aldose tosylhydrazones; C-glycosides; heptenitols



Citation: Kaszás, T.; Baráth, B.Á.; Balázs, B.; Blága, T.; Juhász, L.; Somsák, L.; Tóth, M. Coupling Reactions of Anhydro-Aldose Tosylhydrazones with Boronic Acids. *Molecules* **2022**, *27*, 1795. https:// doi.org/10.3390/molecules27061795

Academic Editor: Kenneth Laali

Received: 5 February 2022 Accepted: 6 March 2022 Published: 9 March 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

1. Introduction

N-Tosylhydrazones have extensively been used in organic synthesis for more than half a century. In the past decade *N*-tosylhydrazones were generally applied in a variety of carbon–carbon and carbon–heteroatom bond forming reactions [1–6]. These transition metal catalyzed or catalyst-free cross-coupling reactions proceed through the in situ generated diazo compounds, followed by the formation of metal–carbene or carbene intermediates, which lead to the corresponding coupled products. Carbohydrate tosylhydrazones are also known, but their application in coupling reactions is poorly investigated.

In our research group an easy, one-step method was worked out for the synthesis of anhydro-aldose tosylhydrazones from readily accessible glycosyl cyanides [7–9]. We began a systematic study aimed at the investigation of the applicability of anhydro-aldose-tosylhydrazones 1 [7–9] in coupling reactions. In this project C-O [10], C-S [11], and C-N [12] bonds were successfully formed under metal-free conditions, while C-C bonds [13,14] were obtained in Pd-catalyzed reactions (Scheme 1).

The metal-free reaction between the diazo precursor *N*-tosylhydrazones and alkyl, alkenyl, and arylboronic acids has been established in recent years as a powerful C(sp³)–C bond-forming transformation (Scheme 2a) that avoids the application of precious metal catalysts and highly air/moisture-sensitive or expensive coupling partners [15,16]. However, this reaction was primarily limited to benzylic, α -heterocyclic, and/or aldehyde-derived tosylhydrazones at the substrate level, with lower yields observed for substrates that differed from these [15,17–20]. Dai and coworkers expanded this reductive coupling to acylferrocene tosylhydrazones, producing highly substituted α -arylalkylferrocenes [21]. *N*-Tosylhydrazones derived from 2-, 3-, and 4-substituted cyclohexanones and 4-substituted cyclopentanone were also used in couplings with alkenyl boronic acids [22]. The reductive coupling of *N*-tosylhydrazones under the standard reaction conditions was also examined with diarylborinic acids (Ar₂B(OH)) to give diarylmethanes with good yields [23]. Kirschning developed a flow protocol for the reductive coupling reaction of *N*-tosylhydrazones

with aryl boronic acids. To increase the practical applicability of the reaction, a two-step continuous flow protocol, starting with carbonyl compounds and tosylhydrazide, was also developed [24]. Nakagawa and coworkers expanded the scope of the transformation to a set of challenging heterocycle-containing aldehyde tosylhydrazones, such as those of protected azetidine, imidazole, and azaindole derivatives. These couplings resulted in low to good yields of drug-like molecules, bicyclic products, with a methylene linker between the rings (Scheme 2b) [25]. This type of coupling of indole-3-carbaldehyde tosylhydrazone with boronic acids was used for the synthesis of biologically important 3-benzyl indole derivatives (Scheme 2b) [26]. Ley and coworkers used the procedure for the metal-free coupling of 4-, 5-, and 6-membered saturated heterocyclic p-methoxyphenyl (PMP) sulfonylhydrazones with (het)aryl boronic acids to form sp^2-sp^3 linked bicyclic building blocks, including oxetanes, piperidines, and azetidines, from their parent ketones (Scheme 2c) [27]. The reductive coupling was also applied for the synthesis of 9-arylfluorenes (Scheme 2d) [28]. Thus, a wide range of 9-arylfluorenes was prepared in a one-pot process from 9-fluorenones by treatment with N-tosylhydrazide, followed by the reductive coupling of (het)aryl and alkyl boronic acids in the presence of potassium carbonate. A similar protocol was applied for the synthesis of triarylmethanes from less reactive diaryl ketones (Scheme 2d) [29] and 1(or 2)-(1-phenylethyl)naphthalenes from acetyl naphthalene derivatives [30]. Wang and coworkers developed a three-component transition-metal-free reaction from α -halo-Ntosylhydrazones in the presence of N-alkylindoles and arylboronic acids to form a range of 3-substituted indoles [31]. A new type of cascade cyclization by reaction of alkenylboronic acids with 2-cyanoethyl or 3-cyanopropylcyclohexanone N-tosylhydrazones was developed by Valdés et al. [32,33]. A similar reaction between γ -azido-N-tosylhydrazones and boronic acids led to the formation of 2,2-disubstituted pyrrolidines in a domino process under microwave activation [34]



Scheme 1. Synthetic applications of anhydro-aldose tosylhydrazones in coupling reactions.

As the tosylhydrazone-boronic acid coupling can be of a great potential to avoid the utility of costly and poisonous metals and ligands, metal-free coupling reactions of boronic acids with anhydro-aldose tosylhydrazones were examined as a new type of substrate with higher complexity in comparison to the previous ones (Scheme 2e). This transformation offers a simple possibility for the formation of *C*-glycosylmethyl derivatives whose preparation is rather cumbersome in the literature [13,35–43]. Herein we disclose our experiences with this reaction using various sugar configurations, protecting groups and boronic acids.



Scheme 2. Selected examples of *N*-tosylhydrazone-boronic acid coupling (**a**–**d**) and the reaction studied in this work (**e**).

2. Results and Discussion

We started our study with the reaction between O-perbenzoylated C-(β -D-glucopyran osyl)formaldehyde tosylhydrazone 1a [7-9] and phenylboronic acid (Table 1). First, the literature conditions [15] were applied using 1.5 equivalents of boronic acid and 1.5 equivalents of K_2CO_3 as the base in dry dioxane at reflux temperature (entry 1). The transformation resulted in a complex mixture, containing heptenitols 3a and 4a and exo-glucal 5 [8,44,45] but we did not observe the formation of the expected C-glucoside 2a [13]. However, it can be assumed that the formation of the open chain compounds might occur by a base mediated ring-opening process, whose driving force could be the resonance stabilization of styrene 3a. Similar heptenitols were obtained by the Wittig reaction [46,47]. Migration of a benzoyl protecting group could result in 4a, and intramolecular carbene insertion into the C-2-H bond yielded exo-glucal 5 [8,44,45]. With other bases (Bu₄NF, LiOtBu, and K₃PO₄) the formation of the coupled product **2a** could also not be observed (entries 2–4). Instead, we obtained variable amounts of the heptenitols 3a and 4a, and exo-glucal 5. Increasing the amount of K_3PO_4 raised the yield of heptenitol **3a** to 43% (entry 5). The effects of solvents other than dioxane were also studied, but in each case, complex reaction mixtures were obtained (entries 6-8). On the other hand, performing the reaction in the presence of five equivalents of phenylboronic acid with three or four equivalents of K_3PO_4 gave the

C-glucoside **2a** in a very low yield beside **3a**, while **4a** and **5** were also isolated (entries 9 and 10). Raising the base excess gave *exo*-glucal **5** in moderate yield and heptenitols **3a** and **4a** in traces (entry 11). The best result was achieved with 20-fold excess of phenylboronic acid and 10-fold excess of K_3PO_4 , to give heptenitol **3a** in 70% yield (entry 12). Thus, instead of the expected *C*-glycosylmethylarene derivative **2a**, an open chain compound, **3a**, proved to be the main product of the transformation.

Table 1. Optimization of the coupling reaction of 1 with phenylboronic acid.

BzO Bzı	05	$ \begin{array}{c} $		OH BOH base dry solvent reflux + H	20 320 2 10 320 2 10 320 4	z OBz DBz OBz OBz	+	BzO BzO BzO	OBZ OH OBZ OBZ OBZ 5 OBZ	=	
Е.			Reac	tion Conditions				Yield (%)			
	1	PhB(OH) ₂ (Equiv.)	Base (Equiv.)	Solvent	Т (°С)	t (h)	2a	3a	4a	5	
1	а	1.5	K ₂ CO ₃ (1.5)	1,4-dioxane	101	3	-	-	28	-	
2	a	1.5	Bu ₄ NF (1.5)	1,4-dioxane	101	3	comp	olex read	tion mi	ixture	
3	a	1.5	LiOtBu (1.5)	1,4-dioxane	101	3	-	+ ^a	+ ^a	16	
4	а	1.5	K ₃ PO ₄ (1.5)	1,4-dioxane	101	3	-	+ ^a	-	38	
5	а	1.5	K ₃ PO ₄ (3)	1,4-dioxane	101	3	-	43	-	-	
6	a	1.5	K ₃ PO ₄ (3)	fluorbenzene	85	3.5	comp	olex read	tion mi	ixture	
7	а	1.5	K ₃ PO ₄ (3)	acetonitrile	82	3	comp	olex read	tion mi	ixture	
8	а	1.5	K ₃ PO ₄ (3)	toluene	111	3.5	comp	olex read	tion mi	ixture	
9	а	5	K ₃ PO ₄ (3)	1,4-dioxane	101	3.5	2	36 ^b	11 ^b	2 ^b	
10	a	5	K ₃ PO ₄ (4)	1,4-dioxane	101	3	4	38 ^b	12 ^b	-	
11	a	5	K ₃ PO ₄ (10)	1,4-dioxane	101	3	-	+ ^a	+ ^a	39	
12	а	20	K ₃ PO ₄ (10)	1,4-dioxane	101	2.5	-	70	-	-	
13	b	2	-	1,4-dioxane	101	2	+ ^a	22	-	-	
14	b	5	-	1,4-dioxane	101	2	+ ^a	19	-	- ,	
15	b	10	-	1,4-dioxane	101	2	7 ^b	17 ^b	15 ^b	15 ^b	

^a Compounds were detected in the mixture. ^b Yields were calculated on the basis of the ¹H NMR spectra of the worked-up reaction mixture.

To avoid base mediated side reactions, such as the acyl migration, C-(β -D-glucopyra nosyl)formaldehyde tosylhydrazone Li-salt **1b** [10,12] was used for the couplings, where no added base is needed. Attempted reactions under UV irradiation (λ = 254 nm and 368 nm) carried out in a quartz tube proved to be totally ineffective, resulting in complex reaction mixtures. However, thermic conditions gave, generally, **3a** as the main product, besides *C*-glucoside **2a** and *exo*-glucal **5** (entries 13 and 14). Although the application of 10 equivalents of boronic acid significantly increased the yields (entry 15), the Li-salt

reactions appeared less effective. Thus, tosylhydrazone 1a and 1.5 or 20-fold excess of a boronic acid and 3 or 10-fold excesses of K₃PO₄ were used in further transformations.

The coupling reaction of **1a** was also examined with a variety of aryl boronic acids under the conditions selected above. These reactions resulted in varying yields of compound types **2–5**, among which the heptenitols **3** and **4** were the main products (Table 2). Application of higher excess of boronic acids and K_3PO_4 improved the yields in couplings with 4-(dibenzofuranyl) and 4-methoxyphenyl boronic acids (compare entries 3–4 and 6–7), but in other cases, this had no significant effect on the reaction outcome (compare entries 1–2, 10–11 and 12–13). The coupling was found to be significantly affected by the substituents on the aromatic ring; boronic acids with electron-releasing (entries 1–7) and chloro (entries 8 and 9)-substituents gave better yields. However, with the strong electron-withdrawing nitro group (entries 10–13) *exo*-glucal **5** was the main product, the coupled compound **2h** was observed in only one case. Isolation of the products in pure state often encountered difficulties. Due to very similar mobilities in silica gel column chromatography, *C*-glucosyl compounds **2** were polluted with the *exo*-glucal **5**, and heptenitols **3** and **4** polluted each other, therefore the yields were generally calculated on the basis of the ¹H NMR spectra (Supplementary Materials).

Table 2. Reactions of tosylhydrazone 1a with aryl boronic acids.

$\begin{array}{c} B_{ZO} \longrightarrow OBz \\ B_{ZO} \longrightarrow OBz \\ 1a \end{array} \xrightarrow{OH} Ar - B \\ OH \\ OH \\ OBz \\ 1a \end{array} \xrightarrow{OH} C \\ OBz \\ OBz \\ OBz \\ OBz \\ Ar + B \\ OBz \\ OBz \\ OBz \\ Ar \\ HO \\ OBz \\ OBz \\ OBz \\ OBz \\ Ar \\ Ar - B \\ OBz \\ OBz \\ Ar \\ Ar \\ BzO \\ OBz \\ Ar \\ F \\ BzO \\ OBz \\ F \\ BzO \\ OBz \\ Ar \\ F \\ BzO \\ OBz \\ F \\ $									H Bz Ar Bz
Entr	у	Read	tion Conditi	ons			Yield	d (%)	
	Ar		Boronic Acid (Equiv.)	K ₃ PO ₄ (Equiv.)	t (h)	2	3	4	5
1	b	2-naphthyl	1.5	3	2	7 ^a	39 a	14 ^a	3 a
2		2-naphthyl	20	10	1.5	4	44 ^a	31 ^a	-
3	с	4-(dibenzofuranyl)	1.5	3	2	3 ^a	16	+ ^b	18 ^a
4		4-(dibenzofuranyl)	20	10	2	19 ^a	9 ^a	47	15 ^a
5	d	$4-MeC_6H_4$	1.5	3	2	-	31 ^a	12 ^a	-
6	e	$4-MeOC_6H_4$	1.5	3	3	-	34 ^a	14 ^a	-
7		$4-MeOC_6H_4$	20	10	3	-	+ ^b	42	-
8	f	$3-ClC_6H_4$	1.5	3	1.5	5 a	34 ^a	18 ^a	11 ^a
9	g	$4-ClC_6H_4$	1.5	3	2	-	68	-	4 ^a
10	h	$4-NO_2C_6H_4$	1.5	3	2	-	-	-	63
11		$4-NO_2C_6H_4$	20	10	2.5	10 ^a	-	-	12 ^a
12	i	$3-NO_2C_6H_4$	1.5	3	2	com	plex rea mixture	ction	22
13 3-NO ₂ C ₆ H ₄		20	10	2	complex reaction mixture		62		

^a Yields were calculated on the basis of the ¹H NMR spectra of the worked-up reaction mixture. ^b Compounds were detected in the mixture.

The coupling of *O*-peracetylated *C*-(β -D-galactopyranosyl)formaldehyde tosylhydrazone (**6**, Table 3) with phenylboronic acid was also investigated. With 1.5 equivalents of phenylboronic acid and 3 equivalents of potassium carbonate, only traces of the known compound types **7**, **8**, and **10** [8,44,45] were detected in the complex product mixture (entry 1), but with a 20-fold excess of the boronic acid *C*-(galactosyl)phenylmethane **7** was formed in low yield and heptenitols **8** and **9** proved to be the main products (entry 2). A compound

AcO AcO OAc OAc -OAc OH ÒAc ÒAc -B AcO Ph-7 8 ЮH HO AcO base OAc OAc ÒAc ó ò dry 1,4-dioxane -OAc 6 reflux AcC ÒAc ÒAc 10 9 Entry **Reaction Conditions** Isolated Yield (%) Base t 7 9 PhB(OH)₂ (Equiv.) 8 10 (Equiv.) (h) 3 a 2 ^a 1 1.5 K₂CO₃ (3) 3.5 6 _ 2 20 K₃PO₄ (10) 1.5 4 5 75

 Table 3. Reactions of tosylhydrazone 6 with phenyl boronic acids.

due to a faster acetyl migration to give 8 and 9.

with a free 6-OH (analogue of 3), though might be formed, could not be detected possibly

^a Yields were calculated on the basis of the ¹H NMR spectra of the worked-up reaction mixture.

The NMR analysis provided evidence for the structure of all of the above derivatives and these are illustrated here by the examples of compounds 2, 3, and 4. Anhydro-heptitol **2a**, synthesized in our group earlier [13], showed characteristic ¹H NMR resonances for the C-1 methylene (δ 2.96 ppm (H-1_a), 2.92 ppm (H-1_b), with a great geminal coupling constant (12.3 Hz) between them) and the H-2 ('anomeric') protons (4.00 ppm). The characteristic ¹³C NMR resonances were δ 38.0 ppm (C-1) and 79.2 ppm (C-2). ¹H and ¹³C NMR analysis of C-glycosyl derivatives **2b**,**c**,**f**,**h** showed similar chemical shifts for H-1a (2.92–3.44 ppm), H-1b (2.90-3.29 ppm), H-2 (3.98-4.33 ppm), C-1 (32.1-38.1), and C-2 (77.9-79.2) with geminal coupling constants of $H-1_a-1_b$ in the range of 14.3–15.0 Hz. These data indicated the similar structure of the C-glycosyl derivatives 2. Ring-opened heptenitols 3 and 4 showed quite different spectral data. Signals characteristic for C-1 and C-2 of compounds 2 in the above ranges were missing in the ¹³C NMR spectra of **3** and **4**, instead resonances for -CH= type carbons in the ranges 130.8–136.9 ppm (for C-1) and 119.6–125.9 ppm (for C-2) appeared to prove the presence of a double bond in the molecules. The acyclic form was evidenced by the small vicinal coupling constants (in the range of 0.8–8.9 Hz). The great values (14.9–16.3 Hz) of coupling constant ${}^{3}J_{1,2}$ proved the *E*-configured double bond C-1=C-2 in these structures. The position of the free OH groups of heptenitols 3 and 4 were confirmed by observing cross peaks between OH and H-6 in heptenitols 3 and OH and H-5 in molecules 4 in their ¹H–¹H COSY spectra.

To further prove the formation of heptenitols and acyl group migration, benzoylation/acetylation of the corresponding compounds under standard conditions were carried out. Benzoylation [47] of the mixture of heptenitols **3** and **4** resulted in a single product **11** (Table 4) while acetylation [48] of heptenitol **9** gave *O*-peracetylated product **12** in good to excellent yields (Scheme 3).



Scheme 3. Acetylation of heptenitol 9.

	BZO BZO OBz Ar + 3	HO BZO OBZ Ar 4	7 equiv. BzCl 6.3 equiv. Py dry CHCl ₃ reflux	Bzo Bzo OBz OBz Ar
Entry	7	Reaction	Conditions	Yield of 11 (%)
		Ar	(t h)
1	а	Ph		2 90
2	b	4-(dibenzofurany	l)	2 54

 Table 4. Benzoylation of heptenitols 3 and 4.

To get an insight into the effect of hydrolytically resistant ether type protecting groups on the outcome of the studied coupling reactions, *O*-permethylated (β -D-glucopyranosyl)for maldehyde tosylhydrazone **17** was synthesized. Methyl glucoside **13** was *O*-permethyled to get **14** [49] which was converted to the acetate derivative **15** [50] (Scheme 4). On reacting **15** with trimethylsilyl cyanide in the presence of boron trifluoride etherate, cyanide **16** [51] was obtained. The anomers were separated by column chromatography. Then, β -cyanide **16** β was reduced in the presence of tosylhydrazide to give β -D-glucosyl tosylhydrazone **17** as a mixture of *E* and *Z* isomers.



Scheme 4. Synthesis of *O*-permethylated (β-D-glucopyranosyl)formaldehyde tosylhydrazone 17.

Couplings with **17** gave cleaner product mixtures in better yields, and resulted in *C*-glucosides **18** (Table 5, entries 2, 4, and 8) or open-chain heptenitols **19** and **20** as the main products (entries 1, 5, 6, 7, 9, 10). *Exo*-glucal **21** [52] was always formed as a by-product. Compounds **18** and **21** proved inseparable, similar to open chain isomers **19** and **20**.

The transformation was extended to the acetal protected galactose derivative **24**, which was synthesized from the galactosyl cyanide **22** in two steps. Compound **22** was reacted with methoxymethyl chloride to obtain cyanide **23** [53], then a reduction step in the presence of tosylhydrazide gave a mixture of *E* and *Z* isomers of **24** (Scheme 5).



Scheme 5. Synthesis of *O*-permethoxymethylated (β-D-galactopyranosyl)formaldehyde tosylhydrazone **24**.

	MeO MeO OMe OMe N S OMe 17			OH →B OH → SgPO ₄ 4-dioxane reflux +	MeO MeO 18 MeO OMe Ar MeO OH MeO OMe 20	Ar + +	MeO MeO II 19 MeO MeO 21	Vie Me		
Entry			Reaction Cond	Conditions			Yield (%)			
		Ar	Boronic Acid (Equiv.)	K ₃ PO ₄ (Equiv.)	t (h)	18	19	20	21	
1.	а	Ph	1.5	3	3.5	17 ^a	61 ^a	7 ^a	13 ^a	
2.	b	4-(dibenzofuranyl)	1.5	3	1	8	-	-	+ ^b	
3.		4-COOHC ₆ H ₄	1.5	3	3		complex read	tion mixtu	re	
4.	с	$4-CF_3C_6H_4$	1.5	3	1.5	45	16 ^a	+ ^b	26	
5.	d	$4-FC_6H_4$	1.5	3	5.5	14	55 ^a	18 ^a	+ ^b	
6.	e	$3-ClC_6H_4$	1.5	3	2.5	29	37 ^a	4 ^a	+ ^b	
7.	f	$4-BrC_6H_4$	1.5	3	1.5	22	35 ^a	4 ^a	+ ^b	
8.	g	$4-NO_2C_6H_4$	1.5	3	1.5	46	-	-	13	
9.	h	$4-MeOC_6H_4$	1.5	3	1.5	9	52 ^a	2 ^a	+ ^b	
10.	i	$4-MeC_6H_4$	1.5	3	3.5	20 ^a	50 ^a	6 ^a	7 ^a	

Table 5. Reactions of tosylhydrazone 17 with aryl boronic acids.

^a Yields calculated on the basis of the ¹H NMR spectra of the worked-up reaction mixture. ^b Compounds were detected in the mixture.

The coupling reation of **24** with phenylboronic acid resulted in *E* heptenitol **26** as the main product and an inseparable mixture of *C*-(galactopyranosyl)phenylmethane **25** and *exo*-galactal **28** [53]. The *Z* isomer **27** was also detected in the mixture (Scheme 6).



Scheme 6. Coupling of tosylhydrazone 24 with phenylboronic acid.

For the structure elucidation of Me (**18–20**) and MOM (**25–27**), protected derivatives 1D-NMR (¹H, ¹³C) and 2D-NMR (¹H–¹H COSY, HSQC, and HMBC) spectra were recorded. The characteristic chemical shifts of C-1 (32.0–38.1 ppm vs. 132.3–134 ppm) and C-2 (79.0–80.9 ppm vs. 124.3–130.4 ppm) clearly revealed the structures of the anhydro-heptitols **18**, **25**, and heptenitols **19**, **20**, **26**, **27**, respectively.

In contrast to the transformations of acylated derivatives **2** and **7**, those of tosylhydrazones **17** and **24** possessing ether-type protecting groups (Me, MOM) resulted in no migration of the protecting groups as expected, but the *E* and *Z* isomers of the acyclic derivatives were isolated. The configuration of the double bonds was identified by the vicinal coupling constants being 16.0 Hz for the *E* and 11.4–12.1 Hz for the *Z* isomers. The measured vicinal coupling constants showed high variety for heptenitols **19** and **20**, in contrast to the cyclic ${}^{4}C_{1}$ conformers **18**, where these values were 8.7 and 9.8 Hz for the trans diaxial protons. The position of the free OH groups of heptenitols **19**, **20**, **26**, and **27** were confirmed by observing cross peaks between OH and H-6 in their ${}^{1}H_{-}{}^{1}H$ COSY spectra.

		PG F	GO GO OPG 2a, 18c,g	OH Ar−B OH K₃PO₄ dry 1,4-dioxane reflux	PGO PGO OH 3a, 19	PG Pc,g		
				Reaction Conditi	ons			
Ent	ry	PG	Ar	Boronic Acid (Equiv.)	K ₃ PO ₄ (Equiv.)	t (h)	Experience	
1	2a	Bz	Ph	-	10	22	partial deprotection	
2	18g	Me	$4-NO_2C_6H_4$	-	3	21	no conversion	
3 18g Me		Me	$4-NO_2C_6H_4$	1.5	-	21	no conversion	
4	18c	Me	$4-CF_3C_6H_4$	1.5	3	21	no conversion	

Table 6. Examination of possible ring opening of some anhydro-heptitols.

Table 7. Examination of possible ring closing of heptenitols.



To obtain more information about the formation of the open-chain heptenitols, first we checked the possibility of the ring opening of the anhydro-heptitols under the reaction conditions. Thus, **2a** was reacted with K_3PO_4 but partial deprotection of **2a** was observed only, without the formation of **3a** (Table 6, entry 1). The methyl protected derivatives **18c** or **18g** reacted neither in the presence of K_3PO_4 , nor of a boronic acid or both (entries 2–4).

Next, formation of *C*-(glycosyl)arylmethane derivatives **18c**,**d**,**e** was examined from the corresponding heptenitols **19c**,**d**,**e** and **20c**,**d**,**e**. Attempted reactions in the presence of base and/or boronic acid resulted in no conversion (Table 7).

Based on these observations, it can be concluded that the cyclic *C*-glycosylmethyl derivatives and the open-chain heptenitols are not interconvertible under the applied conditions, they must be formed from the same intermediate during the reaction.

To explain these experiences, the following mechanistic possibilities can be considered (Scheme 7). Loss of a sulfinate ion from tosylhydrazones I upon deprotonation or from Li-salt V may lead to the diazo intermediate VI which can give rise to carbene VII by eliminating a nitrogen molecule. The zwitterionic intermediate VIII, which arises from carbene VII (*path a*) or boronate complex **X**, formed from the diazo compound VI (*path b*), may lead to intermediate IX. Then, protodeboronation of IX under basic conditions can give anhydro-heptitol type products III (*path c*). Nevertheless, in intermediate IX, the ring

oxygen, as a Lewis base, can attack the electron deficient boron atom to form the open chain heptenitol borate **XI** (*path e*) which, upon hydrolysis, can lead to the isolated heptenitols **IV**. The driving force of this rearrangement may be the conjugation of the double bond with the aromatic system, leading to an energetically more stable species. The standard by-product *exo*-glycal **II** can be formed by an intramolecular insertion reaction of carbene **VII** (*path d*).



Scheme 7. Mechanistic possibilities for the coupling reactions.

3. Conclusions

This study on the metal-free coupling reactions of C-(β -D-glycopyranosyl)formaldehyde (2,6-anhydro-aldose) tosylhydrazones with aromatic boronic acids revealed that the main reaction pathway was the formation of ring-opened hept-1-enitol derivatives, while the expected *C*-glycopyranosyl compounds (benzyl *C*-glycosides) were formed only in low to moderate yields. The corresponding *exo*-glycals always appeared as unavoidable by-products. *O*-Acyl protecting groups on the carbohydrate moieties underwent migrations which further increased the number of products in the otherwise rather complex reaction mixtures. Tosylhydrazones with ether type *O*-protections gave cleaner reactions but resulted in the same product types in similar ratios. The suggested mechanistic rationale explained how the complex sugar-derived tosylhydrazone substrates changed the reaction pathway. We think that this study also highlights the importance of transformations of high complexity which, though resulting in several products, may lead to a better understanding of their mechanism and may thus inspire further work.

4. Experimental

4.1. General Methods

Optical rotations were determined with a Perkin–Elmer 241 polarimeter or Jasco P-2000 (Easton, MD, USA) at room temperature. NMR spectra were recorded with a Bruker AM Avance DRX 360 MHz (360/90 MHz for ¹H/¹³C) or Bruker AM Avance I 400 MHz (400/100 MHz for ¹H/¹³C) or Bruker AM Avance II 500 MHz (500/125 MHz for ¹H/¹³C) spectrometers. Chemical shifts are referenced to TMS as the internal reference (¹H), or to the residual solvent signals (¹³C). The assignments of the ¹H and ¹³C NMR signals of compounds 2–4, 7–9, 11, 12, 18–20, and 25–27 were performed by their COSY (2a, 3a,c, 4a,e, 7, 8, 9, 11a,b, 12, 18b,f,i, 19a,c,h,i, 20a,d,i, 25, 26, 27), HSQC (2a, 3a,c, 4a,e, 7, 8, 9, 11a,b, 12, 18b,f,i, 25, 26, 27), or HMBC (3a,c, 4a,e, 7, 8, 9, 11a,b, 12, 18b,e,f,i, 19a,c,h,i, 20a,d,i, 25, 26, 27), were recorded with maXis II UHR ESI-

QTOF MS (Bruker Daltonik, Bremen, Germany) instruments in positive ion mode with the electrospray ionization technique, or Thermo LTQ XL (Thermo Electron Corp., San Jose, CA, USA) mass spectrometers operated in a full scan positive ion ESI and APCI mode. TLC was performed on a DCAlurolle Kieselgel 60 F254 (Merck). TLC plates were visualized under UV light, and by gentle heating (generally no spray reagent was used but, if more intense charring was necessary, the plate was sprayed with the following solution: abs. EtOH (95 mL), cc. H_2SO_4 (5 mL), anisaldehyde (1 mL)). For column chromatography Kieselgel 60 (Merck, particle size (0.063–0.200 mm) was applied. The compound 1,4-dioxane was distilled from sodium benzophenone ketyl and stored over sodium wires.

4.2. General Procedure I: Conditions for the Reaction of Anhydro-Aldose Tosylhydrazones with Boronic Acids

A boronic acid (1.5 or 20 mmol, specified with the particular reactions) and K_3PO_4 (3 or 10 mmol, specified with the particular reactions) were suspended in dry 1,4-dioxane (15 mL). The suspension was stirred and heated to reflux, and then a solution of a tosylhydrazone (1; 17 or 24, 1 mmol) in dry 1,4-dioxane (15 mL) was added dropwise over ~20 min. When TLC (1:2 EtOAc–hexane for 1 and 17, 1:1 EtOAc–hexane for 24) indicated complete consumption of the starting compound (20 min–4 h), the mixture was cooled down and the insoluble material was filtered off and washed thoroughly with dry 1,4-dioxane (3 × 20 mL). The solvent was removed under reduced pressure, and the residue was purified by column chromatography, with eluents indicated for the particular compounds to give anhydro heptitols and hept-1-enitols.

4.3. Characterization of Anhydro-Heptitols 2

4.3.1. 2,6-Anhydro-3,4,5,7-Tetra-O-Benzoyl-1-Deoxy-1-Phenyl-D-glycero-D-gulo-Heptitol (2a)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), phenylboronic acid (1.5 equiv., 0.02 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 3 mg (4%) of **2a** as a white amorphous product. Optical rotation, NMR and MS spectra are identical with those reported [13].

4.3.2. 2,6-Anhydro-3,4,5,7-Tetra-O-Benzoyl-1-Deoxy-1-(Naphth-2-yl)-D-*glycero*-D-*gulo*-Heptitol (**2b**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), naphthalen-2ylboronic acid (20 equiv., 0.44 g, 2.57 mmol), and K₃PO₄ (10 equiv., 0.27 g, 1.29 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 4 mg (4%) of **2b** as a pale brown amorphous solid. R_f: 0.42 (1:2 EtOAc–hexane); $[\alpha]_D$ + 6 (*c* 0.16, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.99–7.19 (27H, m, aromatics), 5.90 (1H, pszeudo t, *J*_{4,5} 9.6 Hz, H-4), 5.62 (1H, pseudo t, *J*_{5,6} 9.7 Hz, H-5), 5.52 (1H, pseudo t, *J*_{3,4} 9.6 Hz, H-3), 4.57 (1H, dd, *J*_{7a,7b} 12.0 Hz, H-7_a), 4.41 (1H, dd, H-7_b), 4.09 (1H, ddd, *J*_{1a,2} 5.1, *J*_{1b,2} 6.6, *J*_{2,3} 9.8 Hz, H-2), 4.04 (1H, ddd, *J*_{6,7a} 2.7, *J*_{6,7b} 6.3 Hz, H-6), 3.12 (1H, dd, *J*_{1a,1b} 14.8 Hz, H-1_a), 3.08 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 166.3, 166.1, 165.6, 165.5 (4 × CO), 136.6–124.7 (aromatics), 79.2 (C-2), 76.3 (C-6), 74.7 (C-4), 72.6 (C-3), 70.1 (C-5), 63.6 (C-7), 38.3 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 743.2252, found: [M + Na]⁺ = 743.2253; C₄₅H₃₆O₉ (720.24).



4.3.3. 2,6-Anhydro-3,4,5,7-Tetra-O-Benzoyl-1-(4-Dibenzo[b,d]furanyl)-1-Deoxy-D-glycero-D-gulo-Heptitol (**2c**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), dibenzo[*b,d*]furan-4-ylboronic acid (20 equiv., 0.55 g, 2.57 mmol), and K₃PO₄ (10 equiv., 0.27 g, 1.29 mmol) according to General procedure I by column chromatography (1:3 EtOAc–hexane) to yield 30 mg pale brown amorphous solid containing **2c** and **5** in 1:1 ratio. R_f: 0.50 (1:2 EtOAc– hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.21–6.93 (27H, m, aromatics), 5.91 (1H, pseudo t, *J*_{4,5} 9.5 Hz, H-4), 5.64 (1H, pseudo t, *J*_{5,6} 9.8 Hz, H-5), 5.52 (1H, pseudo t, *J*_{3,4} 9.8 Hz, H-3), 4.56 (1H, dd, *J*_{7a,7b} 12.0 Hz, H-7a), 4.42 (1H, dd, H-7b), 4.33 (1H, ddd, *J*_{1a,2} 3.2, *J*_{1b,2} 8.0, *J*_{2,3} 9.8 Hz, H-2), 4.07 (1H, ddd, *J*_{6,7a} 2.9, *J*_{6,7b} 5.9 Hz, H-6), 3.44 (1H, dd, *J*_{1a,1b} 14.6 Hz, H-1a), 3.29 (1H, dd, H-1b). ¹³C NMR (100 MHz, CDCl₃) δ 166.3, 166.1, 165.5 (4 × CO), 135.6–110.4 (aromatics), 77.9 (C-2), 76.2 (C-6), 74.8 (C-4), 72.5 (C-3), 70.1 (C-5), 63.5 (C-7), 32.1 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 761.2381, found: [M + H]⁺ = 761.2379; C₄₇H₃₆O₁₀ (760.23).



4.3.4. 2,6-Anhydro-3,4,5,7-Tetra-O-Benzoyl-1-(3-Chlorophenyl)-1-Deoxy-D-glycero-D-gulo-Heptitol (2f)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), 3-chlorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 13 mg white amorphous solid containing **2f** and **5** in 1:2 ratio. R_f: 0.48 (1:2 EtOAc–hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.13–7.76 (12H, m, aromatics), 7.63–6.94 (12H, m, aromatics), 5.89 (1H, pseudo t, *J*_{4,5} 9.7 Hz, H-4), 5.60 (1H, pseudo t, *J*_{5,6} 9.7 Hz, H-5), 5.45 (1H, pseudo t, *J*_{3,4} 9.5 Hz, H-3), 4.57 (1H, dd, *J*_{7a,7b} 12.1 Hz, H-7a), 4.42 (1H, dd, H-7b), 4.05 (1H, ddd, *J*_{6,7a} 2.8, *J*_{6,7b} 6.2 Hz, H-6), 3.98 (1H, ddd, *J*_{1a,2} 5.3, *J*_{1b,2} 6.6, *J*_{2,3} 9.7 Hz, H-2), 2.92 (1H, dd, *J*_{1a,1b} 15.0 Hz, H-1a), 2.90 (1H, dd, H-1b). ¹³C NMR (90 MHz, CDCl₃) δ 166.3, 166.1, 165.7, 165.6 (4 × CO), 156.3–125.7 (aromatics), 78.8 (C-2), 76.4 (C-6), 74.6 (C-4), 72.6 (C-3), 70.1 (C-5), 63.5 (C-7), 37.4 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 727.1705, found: [M + Na]⁺ = 727.1708; C₄₁H₃₃ClO₉ (704.18).



4.3.5. 2,6-Anhydro-3,4,5,7-Tetra-O-Benzoyl-1-Deoxy-1-(4-Nitrophenyl)-D-glycero-D-gulo-Heptitol (**2h**)

Isolated from a reaction of tosylhydrazone **1a** (0.30 g, 0.39 mmol), 4-nitrophenylboronic acid (20 equiv., 1.30 g, 7.72 mmol), and K₃PO₄ (10 equiv., 0.82 g, 3.86 mmol) according to General procedure I by column chromatography (1:3 EtOAc–hexane) to yield 32 mg pale brown amorphous solid containing **2h** and **5** in 4:1 ratio. R_f: 0.44 (1:2 EtOAc–hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.32–7.72 (8H, m, aromatics), 7.69–7.16 (16H, m, aromatics), 5.90 (1H, pseudo t, $J_{4,5}$ 9.5 Hz, H-4), 5.58 (1H, pseudo t, $J_{5,6}$ 9.8 Hz, H-5), 5.45 (1H, pseudo t, $J_{3,4}$ 9.7 Hz, H-3), 4.53 (1H, dd, $J_{7a,7b}$ 12.2 Hz, H-7_a), 4.48 (1H, dd, H-7_b), 4.05 (1H, ddd, $J_{6,7a}$ 3.2, $J_{6,7b}$ 6.6 Hz, H-6), 3.99 (1H, ddd, $J_{1a,2}$ 5.1, $J_{1b,2}$ 7.0, $J_{2,3}$ 9.7 Hz, H-2), 3.03 (1H, dd, $J_{1a,1b}$ 14.3 Hz, H-1_a), 3.02 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 166.4, 166.2, 165.7, 165.6 (4 × CO), 161.7–115.1 (aromatics), 78.3 (C-2), 76.4 (C-6), 74.5 (C-4), 72.5 (C-3), 70.0 (C-5), 63.3 (C-7), 37.8 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 738.1946, found: [M + Na]⁺ = 738.1950; C₄₁H₃₃NO₁₁ (715.21).



4.4. *Characterization of Hept-1-Enitols* **3** *and* **4** 4.4.1. (*E*)-3,4,5,7-Tetra-O-Benzoyl-1,2-Dideoxy-1-Phenyl-D-gluco-Hept-1-Enitol (**3***a*)

Prepared from tosylhydrazone **1a** (0.80 g, 1.03 mmol), phenylboronic acid (20 equiv., 2.51 g, 20.60 mmol), and K₃PO₄ (10 equiv., 2.19 g, 10.30 mmol) according to General procedure I. Purified by column chromatography (1:4 EtOAc–hexane) to yield 484 mg (70%) of **3a** as a white amorphous solid. R_f: 0.36 (1:2 EtOAc–hexane); $[\alpha]_D + 21$ (*c* 0.20, CH₂Cl₂). ¹H NMR (500 MHz, CDCl₃) δ 8.18–7.82 (8H, m, aromatics), 7.64–7.15 (17H, m, aromatics), 6.78 (1H, d, *J*_{1,2} 15.9 Hz, H-1), 6.32 (1H, dd, *J*_{2,3} 6.9 Hz, H-2), 6.14–6.02 (2H, m, H-3, H-4), 5.76 (1H, dd, *J*_{4,5} 0.8, *J*_{5,6} 8.9 Hz, H-5), 4.53 (1H, dd, *J*_{6,7a} 2.6, *J*_{7a,7b} 11.9 Hz, H-7_a), 4.34 (1H, dd, *J*_{6,7b} 5.7 Hz, H-7_b), 4.21–4.11 (1H, m, H-6), 3.58 (1H, d, *J*_{6,OH} 4.3 Hz, OH). ¹³C NMR (125 MHz, CDCl₃) δ 167.3, 166.7, 165.6, 165.4 (4 × CO), 136.7 (C-1), 136.3–125.9 (aromatics), 122.1 (C-2), 73.9 (C-3), 73.3 (C-4), 71.3 (C-5), 68.6 (C-6), 65.5 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 693.2095, found: [M + Na]⁺ = 693.2095; C₄₁H₃₄O₉ (670.22).



4.4.2. (E)-3,4,6,7-Tetra-O-Benzoyl-1,2-Dideoxy-1-Phenyl-D-gluco-Hept-1-Enitol (4a)

Isolated from a reaction of tosylhydrazone **1a** (0.30 g, 0.39 mmol), phenylboronic acid (20 equiv., 0.94 g, 7.72 mmol), and K₃PO₄ (10 equiv., 8.20 g, 3.86 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 24 mg yellow amorphous solid containing **4a** and **3a** in 10:2 ratio. R_f: 0.37 (1:2 EtOAc–hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.22–7.77 (8H, m, aromatics), 7.63–7.06 (17H, m, aromatics), 6.99 (1H, d, *J*_{1,2} 15.6 Hz, H-1), 6.31 (1H, dd, *J*_{2,3} 8.0 Hz, H-2), 6.23 (1H, pseudo t, *J*_{3,4} 8.6 Hz, H-3), 5.82 (1H, dd, *J*_{4,5} 1.3 Hz, H-4), 5.44 (1H, ddd, *J*_{6,7a} 3.3, *J*_{6,7b} 4.4, *J*_{5,6} 8.0 Hz, H-6), 4.81 (1H, dd, *J*_{7a,7b} 12.4 Hz, H-7_a), 4.74 (1H, dd, H-7_b), 4.39 (1H, pseudo t, H-5), 3.25 (1H, d, *J*_{5,OH} 8.4 Hz, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 166.7, 165.7, 165.4 (4 × CO), 136.9 (C-1), 136.3–124.1 (aromatics), 122.7 (C-2), 74.6 (C-3), 72.4 (C-4), 71.7 (C-6), 68.5 (C-5), 63.4 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 693.2095, found: [M + Na]⁺ = 693.2096; C₄₁H₃₄O₉ (670.22).



4.4.3. (*E*)-3,4,5,7-Tetra-O-Benzoyl-1,2-Dideoxy-1-Naphth-2-yl-D-*gluco*-Hept-1-Enitol (**3b**) and (*E*)-3,4,6,7-Tetra-O-Benzoyl-1,2-Dideoxy-1-Naphth-2-yl-D-*gluco*-Hept-1-Enitol (**4b**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), naphthalen-2ylboronic acid (20 equiv., 0.44 g, 2.57 mmol), and K_3PO_4 (10 equiv., 0.27 g, 1.29 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 70 mg pale brown amorphous solid containing **3b** and **4b** in 1.5:1 ratio. R_f : 0.25 (1:2 EtOAc–hexane).



3b: ¹H NMR (400 MHz, CDCl₃) δ 8.20–7.03 (27H, m, aromatics), 6.94 (1H, d, $J_{1,2}$ 15.9 Hz, H-1), 6.45 (1H, dd, $J_{2,3}$ 6.7 Hz, H-2), 6.19–6.09 (2H, m, H-3, H-4), 5.82 (1H, dd, $J_{4,5}$ 1.2, $J_{5,6}$ 8.9 Hz, H-5), 4.54 (1H, dd, $J_{6,7a}$ 3.0, $J_{7a,7b}$ 11.9 Hz, H-7_a), 4.35 (1H, dd, $J_{6,7b}$

5.7 Hz, H-7_b), 4.24–4.13 (1H, m, H-6), 3.66 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.2, 166.7, 165.6, 165.4 (4 × CO), 136.6 (C-1), 136.4–123.3 (aromatics), 122.4 (C-2), 73.9 (C-3), 73.2 (C-4), 71.3 (C-5), 68.5 (C-6), 65.5 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 743.2252, found: [M + Na]⁺ = 743.2250; C₄₅H₃₆O₉ (720.24).

4b: ¹H NMR (400 MHz, CDCl₃) δ 8.20–7.03 (28H, m, aromatics, H-1), 6.44 (1H, dd, $J_{1,2}$ 15.8, $J_{2,3}$ 8.4 Hz, H-2), 6.31 (1H, pseudo t, $J_{3,4}$ 8.9 Hz, H-3), 5.88 (1H, dd, $J_{4,5}$ 1.4 Hz, H-4), 5.48 (1H, ddd, $J_{6,7a}$ 3.3, $J_{6,7b}$ 4.4, $J_{5,6}$ 8.0 Hz, H-6), 4.82 (1H, dd, $J_{7a,7b}$ 12.4 Hz, H-7a), 4.75 (1H, dd, H-7b), 4.44 (1H, d, H-5), 3.57 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 166.9, 166.3, 165.8, 165.4 (4 × CO), 136.9 (C-1), 136.4–123.0 (aromatics), 123.1 (C-2), 74.8 (C-3), 72.5 (C-4), 71.6 (C-6), 68.4 (C-5), 63.4 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 743.2252, found: [M + Na]⁺ = 743.2254; C₄₅H₃₆O₉ (720.24).

4.4.4. (*E*)-3,4,5,7-Tetra-O-Benzoyl-1-(4-Dibenzo[b,d]furanyl)-1,2-Dideoxy-D-*gluco*-Hept-1-Enitol (**3c**)

Prepared from tosylhydrazone **1a** (0.10 g, 0.13 mmol), dibenzo[*b*,*d*]furan-4-ylboronic acid (1.5 equiv., 0.04 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:2 EtOAc–hexane) to yield 16 mg (16%) of **3c** as a pale brown amorphous solid. R_f: 0.32 (1:2 EtOAc–hexane); $[\alpha]_D$ + 5 (*c* 0.11, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.38–7.68 (12H, m, aromatics), 7.64–7.16 (15H, m, aromatics), 7.15–6.92 (2H, m, H-1, H-2), 6.22 (1H, dd, *J*_{2,3} 5.5, *J*_{3,4} 8.0 Hz, H-3), 6.16 (1H, dd, *J*_{4,5} 1.7 Hz, H-4), 5.87 (1H, dd, *J*_{5,6} 8.9 Hz, H-5), 4.54 (1H, dd, *J*_{6,7a} 3.0, *J*_{7a,7b} 11.9 Hz, H-7_a), 4.35 (1H, dd, *J*_{6,7b} 5.7 Hz, H-7_b), 4.23–4.15 (1H, m, H-6), 3.60 (1H, d, *J*_{6,0H} 5.3 Hz, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.2, 166.7, 165.6, 165.4 (4 × CO), 130.8 (C-1), 156.5–111.9 (aromatics), 125.9 (C-2), 74.1 (C-3), 73.3 (C-4), 71.3 (C-5), 68.6 (C-6), 65.5 (C-7). C₄₇H₃₆O₁₀ (760.23). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 783.2201; found: [M + Na]⁺ = 783.2202; C₄₇H₃₆O₁₀ (760.23).



4.4.5. (*E*)-3,4,6,7-Tetra-O-Benzoyl-1-(4-Dibenzo[b,d]furanyl)-1,2-Dideoxy-D-*gluco*-Hept-1-Enitol (**4c**)

Prepared from tosylhydrazone **1a** (0.10 g, 0.13 mmol), dibenzo[*b*,*d*]furan-4-ylboronic acid (20 equiv., 0.55 g, 2.57 mmol), and K₃PO₄ (10 equiv., 0.27 g, 1.29 mmol) according to General procedure I. Purified by column chromatography (1:3 EtOAc–hexane) to yield 29 mg (30%) of **4c** as a yellow amorphous solid. R_f: 0.32 (1:2 EtOAc–hexane); $[\alpha]_D$ + 5 (*c* 0.11, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.23–6.76 (27H, m, aromatics), 7.02 (1H, d, *J*_{1,2} 16.2 Hz, H-1), 6.97 (1H, dd, *J*_{2,3} 8.2 Hz, H-2), 6.29 (1H, pseudo t, *J*_{3,4} 9.0 Hz, H-3), 5.91 (1H, dd, *J*_{4,5} 1.5 Hz, H-4), 5.44 (1H, ddd, *J*_{6,7a} 3.5, *J*_{6,7b} 4.4, *J*_{5,6} 8.0 Hz, H-6), 4.77 (1H, dd, *J*_{7a,7b} 12.4 Hz, H-7_a), 4.68 (1H, dd, H-7_b), 4.50–4.41 (1H, m, H-5), 3.28 (1H, d, *J*_{5,OH} 6.0 Hz, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.1, 166.9, 165.9, 165.8 (4 × CO), 131.7 (C-1), 156.3–111.0 (aromatics), 120.6 (C-2), 75.1 (C-3), 72.5 (C-4), 71.7 (C-6), 68.4 (C-5), 63.3 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 783.2201, found: [M + Na]⁺ = 783.2202; C₄₇H₃₆O₁₀ (760.23).



4.4.6. (*E*)-3,4,5,7-Tetra-*O*-Benzoyl-1,2-Dideoxy-1-(4-Methylphenyl)-D-*gluco*-Hept-1-Enitol (**3d**) and (*E*)-3,4,6,7-Tetra-*O*-Benzoyl-1,2-Dideoxy-1-(4-Methylphenyl)-D-*gluco*-Hept-1-enitol (**4d**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), 4-methylphenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 44 mg pale yellow amorphous solid containing **3d** and **4d** in 2:1 ratio. R_f : 0.38 (1:2 EtOAc–hexane).



3d: ¹H NMR (400 MHz, CDCl₃) δ 8.20–7.81 (8H, m, aromatics), 7.64–7.01 (16H, m, aromatics), 6.74 (1H, d, $J_{1,2}$ 15.9 Hz, H-1), 6.26 (1H, dd, $J_{2,3}$ 6.7 Hz, H-2), 6.12–6.02 (2H, m, H-3, H-4), 5.75 (1H, dd, $J_{4,5}$ 1.1, $J_{5,6}$ 8.9 Hz, H-5), 4.52 (1H, dd, $J_{6,7a}$ 2.9, $J_{7a,7b}$ 11.9 Hz, H-7_a), 4.34 (1H, dd, $J_{6,7b}$ 5.9 Hz, H-7_b), 4.20–4.10 (1H, m, H-6), 3.60 (1H, d, $J_{6,0H}$ 5.2 Hz, OH), 2.33 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 166.7, 165.6, 165.4 (4 × CO), 136.8 (C-1), 139.5–126.1 (aromatics), 120.9 (C-2), 74.0 (C-3), 73.4 (C-4), 71.4 (C-5), 68.6 (C-6), 65.5 (C-7), 21.4 (CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 707.2252, found: [M + Na]⁺ = 707.2251; C₄₂H₃₆O₉ (684.24).

4d: ¹H NMR (400 MHz, CDCl₃) δ 8.21–7.73 (8H, m, aromatics), 7.72–7.01 (16H, m, aromatics), 6.96 (1H, d, $J_{1,2}$ 14.9 Hz, H-1), 6.25 (1H, dd, $J_{2,3}$ 6.6 Hz, H-2), 6.21 (1H, pseudo t, $J_{3,4}$ 8.6 Hz, H-3), 5.81 (1H, dd, $J_{4,5}$ 1.3 Hz, H-4), 5.44 (1H, ddd, $J_{6,7a}$ 3.4, $J_{6,7b}$ 4.5, $J_{5,6}$ 8.5 Hz, H-6), 4.79 (1H, dd, $J_{7a,7b}$ 12.4 Hz, H-7_a), 4.74 (1H, dd, H-7_b), 4.39 (1H, pseudo t, $J_{5,OH}$ 8.8 Hz, H-5), 3.16 (1H, bs, OH), 2.35 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 167.1, 166.9, 165.9, 165.3 (4 × CO), 135.9 (C-1), 139.5–115.0 (aromatics), 121.6 (C-2), 74.7 (C-3), 72.5 (C-4), 71.7 (C-6), 68.5 (C-5), 63.4 (C-7), 21.4 (CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 707.2252, found: [M + Na]⁺ = 707.2254; C₄₂H₃₆O₉ (684.24).

4.4.7. (*E*)-3,4,5,7-Tetra-*O*-Benzoyl-1,2-Dideoxy-1-(4-Methoxyphenyl)-D-*gluco*-Hept-1-Enitol (**3e**) and (*E*)-3,4,6,7-Tetra-*O*-Benzoyl-1,2-Dideoxy-1-(4-Methoxyphenyl)-D-*gluco*-Hept-1-Enitol (**4e**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), 4-methoxyphenylbor onic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 43 mg yellow amorphous solid containing **3e** and **4e** in 2.5:1 ratio. R_f : 0.31 (1:2 EtOAc–hexane).



3e: ¹H NMR (400 MHz, CDCl₃) δ 8.28–7.67 (8H, m, aromatics), 7.65–7.09 (14H, m, aromatics), 6.81 (2H, d, *J* 8.8 Hz, aromatics), 6.71 (1H, d, *J*_{1,2} 15.7 Hz, H-1), 6.25–6.11 (1H, m, H-2), 6.10–6.02 (2H, m, H-3, H-4), 5.75 (1H, dd, *J*_{4,5} 1.0, *J*_{5,6} 8.9 Hz, H-5), 4.52 (1H, dd, *J*_{6,7a} 3.0, *J*_{7a,7b} 11.9 Hz, H-7_a), 4.34 (1H, dd, *J*_{6,7b} 5.9 Hz, H-7_b), 4.21–4.09 (1H, m, H-6), 3.80 (3H, s, OCH₃), 3.63 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 166.7, 165.6, 165.4 (4 × CO), 136.5 (C-1), 160.9–112.7 (aromatics), 119.6 (C-2), 74.2 (C-3), 73.4 (C-4), 71.4 (C-5), 68.5 (C-6), 65.5 (C-7), 55.4 (OCH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 723.2201; found: [M + Na]⁺ = 723.2204; C₄₂H₃₆O₁₀ (700.23).

4e: ¹H NMR (400 MHz, CD₃OD) δ 8.19–7.74 (8H, m, aromatics), 7.65–7.12 (14H, m, aromatics), 6.96 (1H, d, $J_{1,2}$ 15.8 Hz, H-1), 6.90 (2H, d, J 8.7 Hz, aromatics), 6.32 (1H, dd, $J_{2,3}$ 8.2 Hz, H-2), 6.21 (1H, pseudo t, $J_{3,4}$ 9.1 Hz, H-3), 5.82 (1H, dd, $J_{4,5}$ 1.5 Hz, H-4), 5.45 (1H, dd, $J_{6,7a}$ 2.5, $J_{6,7b}$ 5.1 Hz, H-6), 4.93 (1H, dd, $J_{7a,7b}$ 12.2 Hz, H-7_a), 4.57 (1H, dd, H-7_b), 4.52 (1H, dd, $J_{5,6}$ 9.1 Hz, H-5), 3.80 (3H, s, OCH₃), 3.58 (1H, bs, OH). ¹³C NMR (90 MHz, CDCl₃) δ 167.0, 166.9, 165.8, 165.4 (4 × CO), 136.6 (C-1), 160.9–112.7 (aromatics), 120.3 (C-2), 74.5

(C-3), 72.5, 71.7 (C-4, C-6), 68.8 (C-5), 63.4 (C-7), 55.5 (OCH₃). HR-ESI-MS positive mode (m/z): calc. for $[M + H]^+ = 701.2381$, found: $[M + H]^+ = 701.2381$; C₄₂H₃₆O₁₀ (700.23).

4.4.8. (*E*)-3,4,5,7-Tetra-O-Benzoyl-1-(3-Chlorophenyl)-1,2-Dideoxy-D-*gluco*-Hept-1-Enitol (**3f**) and (*E*)-3,4,6,7-Tetra-O-Benzoyl-1-(3-Chlorophenyl)-1,2-Dideoxy-D-*gluco*-Hept-1-Enitol (**4f**)

Isolated from a reaction of tosylhydrazone **1a** (0.10 g, 0.13 mmol), 3-chlorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 60 mg white amorphous solid containing **3f** and **4f** in 2:1 ratio with two unidentified species. R_f : 0.35 (1:2 EtOAc–hexane).



3f: ¹H NMR (400 MHz, CDCl₃) δ 8.18–7.82 (8H, m, aromatics), 7.64–7.02 (16H, m, aromatics), 6.70 (1H, d, $J_{1,2}$ 15.9 Hz, H-1), 6.34 (1H, dd, $J_{2,3}$ 6.9 Hz, H-2), 6.12–6.04 (2H, m, H-3, H-4), 5.76 (1H, dd, $J_{4,5}$ 0.6, $J_{5,6}$ 8.9 Hz, H-5), 4.54 (1H, dd, $J_{6,7a}$ 3.0, $J_{7a,7b}$ 11.9 Hz, H-7a), 4.34 (1H, dd, $J_{6,7b}$ 5.7 Hz, H-7b), 4.23–4.14 (1H, m, H-6), 3.64 (1H, d, $J_{6,OH}$ 3.9 Hz, OH). ¹³C NMR (90 MHz, CDCl₃) δ 167.1, 166.7, 165.7, 165.5 (4 × CO), 134.9 (C-1), 138.4–123.4 (aromatics), 123.7 (C-2), 73.5 (C-3), 73.0 (C-4), 71.2 (C-5), 68.5 (C-6), 65.5 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 727.1705, found: [M + Na]⁺ = 727.1706; C₄₁H₃₃ClO₉ (704.18).

4f: ¹H NMR (400 MHz, CDCl₃) δ 8.18–7.79 (8H, m, aromatics), 7.69–7.06 (16H, m, aromatics), 6.91 (1H, d, $J_{1,2}$ 15.8 Hz, H-1), 6.32 (1H, dd, $J_{2,3}$ 8.0 Hz, H-2), 6.21 (1H, pseudo t, $J_{3,4}$ 8.4 Hz, H-3), 5.81 (1H, dd, $J_{4,5}$ 1.5 Hz, H-4), 5.43 (1H, ddd, $J_{6,7a}$ 3.3, $J_{6,7b}$ 4.3, $J_{5,6}$ 8.0 Hz, H-6), 4.84 (1H, dd, $J_{7a,7b}$ 12.4 Hz, H-7_a), 4.73 (1H, dd, H-7_b), 4.38–4.27 (1H, m, H-5), 3.34 (1H, d, $J_{5,OH}$ 8.4 Hz, OH). ¹³C NMR (90 MHz, CDCl₃) δ 167.1, 166.8, 165.8, 165.4 (4 × CO), 135.2 (C-1), 153.1–123.4 (aromatics), 124.4 (C-2), 74.4 (C-3), 72.4 (C-4), 71.7 (C-6), 68.4 (C-5), 63.4 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 727.1705, found: [M + Na]⁺ = 727.1706; C₄₁H₃₃ClO₉ (704.18).

4.4.9. 3,4,5,7-Tetra-O-Benzoyl-1-(4-Chlorophenyl)-1,2-Dideoxy-D-gluco-Hept-1-Enitol (3g)

Prepared from tosylhydrazone **1a** (0.10 g, 0.13 mmol), 4-chlorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:2 acetone–hexane) to yield 62 mg (68%) of **3g** as a white amorphous solid. R_f: 0.36 (1:2 EtOAc–hexane); $[\alpha]_D + 9$ (*c* 0.57, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.16–7.85 (8H, m, aromatics), 7.64–7.12 (16H, m, aromatics), 6.71 (1H, d, $J_{1,2}$ 16.0 Hz, H-1), 6.34–6.24 (1H, m, H-2), 6.10–6.02 (2H, m, H-3, H-4), 5.74 (1H, dd, $J_{4,5}$ 0.9, $J_{5,6}$ 8.9 Hz, H-5), 4.53 (1H, dd, $J_{6,7a}$ 2.9, $J_{7a,7b}$ 11.9 Hz, H-7_a), 4.34 (1H, dd, $J_{6,7b}$ 5.7 Hz, H-7_b), 4.21–4.10 (1H, m, H-6), 3.60 (1H, d, $J_{6,OH}$ 5.1 Hz, OH). ¹³C NMR (100 MHz, CDCl₃) δ 167.2, 166.8, 165.5, 165.4 (4 × CO), 135.2 (C-1), 134.7–127.2 (aromatics), 122.7 (C-2), 73.7 (C-3), 73.1 (C-4), 71.2 (C-5), 68.5 (C-6), 65.5 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 727.1705, found: [M + Na]⁺ = 727.1703; C₄₁H₃₃ClO₉ (704.18).



4.5. Characterization of Molecules **7–9** Isolated from the Reaction of Tosylhydrazone **6** and Phenylboronic Acid

4.5.1. 3,4,5,7-Tetra-O-Acetyl-2,6-Anhydro-1-Deoxy-1-Phenyl-D-glycero-L-manno-Heptitol (7)

Isolated from a reaction of tosylhydrazone **6** (0.10 g, 0.19 mmol), phenylboronic acid (20 equiv., 0.46 g, 3.78 mmol), and K_3PO_4 (10 equiv., 0.40 g, 1.89 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 6 mg (7%) of **7** as a

white amorphous product. Optical rotation, NMR and MS spectra are identical with those reported [13].



4.5.2. (E)-3,5,6,7-Tetra-O-Acetyl-1,2-Dideoxy-1-Phenyl-D-galacto-Hept-1-Enitol (8)

Isolated from a reaction of tosylhydrazone **6** (0.30 g, 0.56 mmol), phenylboronic acid (20 equiv., 1.38 g, 11.35 mmol), and K₃PO₄ (10 equiv., 1.20 g, 5.68 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 12 mg (5%) of **8** as a white amorphous solid. R_f: 0.15 (1:2 EtOAc–hexane); $[\alpha]_D + 40$ (*c* 0.18, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.43–7.22 (5H, m, aromatics), 6.71 (1H, d, *J*_{1,2} 16.0 Hz, H-1), 6.32 (1H, dd, *J*_{2,3} 7.6 Hz, H-2), 5.54 (1H, ddd, *J*_{6,7a} 4.6, *J*_{6,7b} 7.7 Hz, H-6), 5.48 (1H, dd, *J*_{3,4} 1.1 Hz, H-3), 5.19 (1H, dd, *J*_{5,6} 1.7 Hz, H-5), 4.26 (1H, dd, *J*_{7a,7b} 11.8 Hz, H-7_a), 4.05 (1H, dd, H-7_b), 3.72 (1H, dd, *J*_{4,5} 9.6 Hz, H-4), 3.09 (1H, bs, OH), 2.16, 2.11, 2.10, 2.04 (12H, 4s, 4 × CH₃). ¹³C NMR (90 MHz, CDCl₃) δ 171.8, 170.6, 170.4, 170.0 (4 × CO), 134.9 (C-1), 136.7–123.2 (aromatics), 123.7 (C-2), 72.4 (C-3), 70.6 (C-4), 70.1, 70.0 (C-5, C-6), 62.8 (C-7), 21.3, 21.0, 20.8 (4 × CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 445.1469, found: [M + Na]⁺ = 445.1470; C₂₁H₂₆O₉ (422.16).



4.5.3. (E)-3,4,6,7-Tetra-O-Acetyl-1,2-Dideoxy-1-Phenyl-D-galacto-Hept-1-Enitol (9)

Prepared from tosylhydrazone **6** (0.30 g, 0.56 mmol), phenylboronic acid (20 equiv., 1.38 g, 11.35 mmol), and K₃PO₄ (10 equiv., 1.20 g, 5.68 mmol) according to General procedure I. Purified by column chromatography (1:4 EtOAc–hexane) to yield 180 mg (75%) of **9** as a white amorphous solid. R_f: 0.10 (1:2 EtOAc–hexane); $[\alpha]_D + 37$ (*c* 0.40, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.57–7.09 (5H, m, aromatics), 6.63 (1H, d, *J*_{1,2} 15.8 Hz, H-1), 6.03 (1H, dd, *J*_{2,3} 6.0 Hz, H-2), 5.88 (1H, dd, *J*_{3,4} 1.9 Hz, H-3), 5.22 (1H, ddd, *J*_{6,7a} 4.7, *J*_{6,7b} 7.8 Hz, H-6), 5.17 (1H, dd, *J*_{4,5} 9.7 Hz, H-4), 4.43 (1H, dd, *J*_{7a,7b} 11.7 Hz, H-7_a), 4.17 (1H, dd, H-7_b), 3.85 (1H, dd, *J*_{5,6} 1.5 Hz, H-5), 3.53 (1H, bs, OH), 2.21, 2.07, 2.04, 2.01 (12H, 4s, 4 × CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 171.5, 171.2, 170.7, 170.0 (4 × CO), 133.0 (C-1), 136.0–121.2 (aromatics), 123.4 (C-2), 72.5 (C-3), 71.6 (C-4), 69.0, (C-6), 67.9 (C-5), 63.4 (C-7), 21.1, 20.8, 20.7 (4 × CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 445.1469, found: [M + Na]⁺ = 445.1467; C₂₁H₂₆O₉ (422.16).



4.6. General Procedure II for the Synthesis of 1-Aryl-3,4,5,6,7-Penta-O-Benzoyl-1,2-Dideoxy-D-gluco-Hept-1-Enitols **11** and **12**

A mixture of 1-aryl-tetra-*O*-benzoyl-1,2-dideoxy-D-*gluco*-hept-1-enitol (**3** and **4**, 1 mmol) and dry pyridine (6.3 mmol) were dissolved in dry chloroform (3 mL). Then, benzoyl-chloride (7 mmol) was added dropwise to the solution. The reaction mixture was stirred and heated at 80 °C. When TLC (1:2 EtOAc–hexane) showed complete consumption of the starting compound (~2 h), the mixture was cooled down. The organic layer was washed with 2M aqueous hydrogen chloride solution (1 × 3 mL), cold, saturated sodium hydrogen carbonate solution (1 × 3 mL), water (1 × 3 mL), and then dried on anhydrous magnesium

sulfate. The solvent was removed under reduced pressure, and the residue was purified by column chromatography (1:2 EtOAc-hexane) to give hept-1-enitols.

4.6.1. (E)-3,4,5,6,7-Penta-O-Benzoyl-1,2-Dideoxy-1-Phenyl-D-gluco-Hept-1-Enitol (11a)

Prepared from (*E*)-3,4,5,7-tetra-*O*-benzoyl-1,2-dideoxy-1-phenyl-D-*gluco*-hept-1-enitol **3a** and (*E*)-3,4,6,7-tetra-*O*-benzoyl-1,2-dideoxy-1-phenyl-D-*gluco*-hept-1-enitol **4a** (0.10 g, 0.15 mmol) according to General procedure II. Purified by column chromatography (1:2 EtOAc–hexane) to yield 104 mg (90%) of **11a** as a white amorphous solid. R_f: 0.41 (1:2 EtOAc–hexane); $[\alpha]_D - 2$ (*c* 0.50, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.24–7.84 (8H, m, aromatics), 7.66–7.17 (17H, m, aromatics), 6.80 (1H, d, $J_{1,2}$ 15.9 Hz, H-1), 6.40–6.29 (1H, m, H-2), 6.18 (1H, dd, $J_{4,5}$ 2.0 Hz, H-5), 6.12–6.04 (2H, m, H-3, H-4), 5.91 (1H, ddd, $J_{6,7a}$ 3.6, $J_{6,7b}$ 5.9, $J_{5,6}$ 7.2 Hz, H-6), 4.82 (1H, dd, $J_{7a,7b}$ 12.3 Hz, H-7_a), 4.55 (1H, dd, H-7_b). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 165.7, 165.5, 165.4, 165.3 (5 × CO), 136.7 (C-1), 135.8–127.0 (aromatics), 122.0 (C-2), 73.8 (C-3), 71.8 (C-4), 69.9, 69.7 (C-5, C-6), 62.8 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 797.2357, found: [M + Na]⁺ = 797.2355; C₄₈H₃₈O₁₀ (774.25).



4.6.2. (*E*)-3,4,5,6,7-Penta-O-Benzoyl-1-(4-Dibenzo[b,d]furanyl)-1,2-Dideoxy-D-*gluco*-Hept-1-Enitol (**11b**)

Prepared from (*E*)-3,4,5,7-tetra-*O*-benzoyl-1-(4-dibenzo[*b*,*d*]furanyl)-1,2-dideoxy*gluco*-hept-1-enitol **3c** and (*E*)-3,4,6,7-tetra-*O*-benzoyl-1-(4-dibenzo[*b*,*d*]furanyl)-1,2-dideoxy-D-*gluco*-hept-1-enitol **4c** (0.05 g, 0.06 mmol), according to General procedure II. Purified by column chromatography (1:2 EtOAc–hexane) to yield 28 mg (55%) of **11b** as a yellow amorphous solid. R_f: 0.39 (1:2 EtOAc–hexane); $[\alpha]_D - 1$ (*c* 0.48, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 8.44–6.81 (32H, m, aromatics), 7.08–7.04 (2H, m, H-1, H-2), 6.28 (1H, dd, *J*_{4,5} 2.0 Hz, H-5), 6.20–6.14 (2H, m, H-3, H-4), 5.92 (1H, ddd, *J*_{6,7a} 3.8, *J*_{6,7b} 5.8, *J*_{5,6} 7.1 Hz, H-6), 4.82 (1H, dd, *J*_{7a,7b} 12.2 Hz, H-7_a), 4.53 (1H, dd, H-7_b). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 165.7, 165.5, 165.4, 165.3 (5 × CO), 130.9 (C-1), 162.8–110.9 (aromatics), 125.8 (C-2), 74.0 (C-3), 71.8 (C-4), 69.9 (C-6), 69.6 (C-5), 62.8 (C-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 887.2463, found: [M + Na]⁺ = 887.2460; C₅₄H₄₀O₁₁ (864.26).



4.6.3. (E)-3,4,5,6,7-Penta-O-Acetyl-1,2-Dideoxy-1-Phenyl-D-galacto-Hept-1-Enitol (12)

3,4,6,7-Tetra-O-acetyl-1,2-dideoxy-1-phenyl-D-*galacto*-hept-1-enitol (**9**, 0.12 g, 0.29 mmol) was dissolved in dry pyridine (1 mL) and cooled to 0 °C. Then, acetic anhydride (1.5 equiv., 0.04 mL, 0.04 g, 0.43 mmol) was added dropwise to the solution. The reaction mixture was stirred for a day at room temperature and the pyridine was evaporated. The residue was dissolved in dichloromethane and washed with water (1 × 2 mL), then dried on anhydrous magnesium sulfate. The solution was concentrated under reduced pressure and traces of pyridine were removed by repeated co-evaporations with toluene to yield 122 mg (91%) of **12** as a white amorphous solid. R_f: 0.36 (1:2 EtOAc–hexane); $[\alpha]_D$ + 115 (*c* 0.02, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.57–7.06 (5H, m, aromatics), 6.58 (1H, dd, *J*_{Ar,1} 0.7, *J*_{1,2} 15.9 Hz, H-1), 5.97 (1H, dd, *J*_{2,3} 6.1 Hz, H-2), 5.67–5.59 (1H, m, H-3), 5.45 (1H, dd, *J*_{5,6} 1.8 Hz, H-5), 5.41–5.31 (1H, m, H-6), 5.37 (1H, dd, *J*_{3,4} 2.5, *J*_{4,5} 10.0 Hz, H-4), 4.29 (1H, dd, *J*_{6,7a} 5.0, *J*_{7a,7b} 11.6 Hz, H-7_a), 3.88 (1H, dd, *J*_{6,7a} 7.5 Hz, H-7_b), 2.14, 2.10, 2.08, 2.04, 202 (15H, 5s, 5 × CH₃).

¹³C NMR (90 MHz, CDCl₃) δ 170.5, 170.3, 170.1, 169.8 (5 × CO), 133.5 (C-1), 136.5–122.2 (aromatics), 122.9 (C-2), 71.1 (C-3), 69.5 (C-4), 68.1 (C-5), 68.0 (C-6), 62.3 (C-7), 21.0, 20.8, 20.7 (5 × CH₃). HR-ESI-MS positive mode (m/z): calc. for $[M + Na]^+ = 487.1575$, found: $[M + Na]^+ = 487.11577$; C₂₃H₂₈O₁₀ (464.17).



4.7. General Procedure III for the Synthesis of Anhydro-Aldose Tosylhydrazones (C-(2,3,4,6-Tetra-O-Alkyl-β-D-Glycopyranosyl) Formaldehyde Tosylhydrazones) (**17**, **24**)

Raney-nickel (1.5 g, an aqueous suspension, Merck) was added at room temperature to a vigorously stirred solution of pyridine (6 mL), acetic acid (4 mL), and water (4 mL). Then, sodium hypophosphite (0.75 g, 8.50 mmol), tosylhydrazine (0.37 g, 2.00 mmol), and nitrile (**16** β [52] or **23**) (1.00 mmol) were added to the mixture. When TLC (2:1 EtOAc–hexane) indicated complete consumption of the starting compound, the insoluble material was filtered off through a pad of celite and washed with dichloromethane (10 mL). The organic layer of the filtrate was separated, washed with water (3 mL), 10% aqueous hydrogen chloride solution (2 × 3 mL), cold, saturated sodium hydrogen carbonate solution (2 × 3 mL), water (3 mL), and then dried on anhydrous magnesium sulfate. The solution was concentrated under reduced pressure, and traces of pyridine were removed by repeated co-evaporations with toluene. The residue was purified by silica gel column chromatography with eluents indicated for the particular compounds to give anhydro-aldose tosylhydrazones **17** or **24**.

2,6-Anhydro-3,4,5,7-Tetra-O-Methyl-D-*glycero*-D-*gulo*-Heptose Tosylhydrazone (C-(2,3,4,6-Tetra-O-Methyl- β -D-Glucopyranosyl) Formaldehyde Tosylhydrazone) (17)

Prepared from cyanide 16β [52] (1.00 g, 4.08 mmol) according to General procedure III. Purified by column chromatography (1:2 EtOAc–hexane) to yield 1.02 g (60%) two unidentified isomers 17-1 and 17-2 in 1:3 ratio as a colourless oil.



17-1 R_f: 0.11 (1:1 EtOAc–hexane). ¹H NMR (360 MHz, CDCl₃) δ 7.92 (1H, bs, NH), 7.86–7.76 (2H, m, aromatics), 7.31 (2H, d, *J* 8.2 Hz, aromatics), 7.05 (1H, d, *J*_{1,2} 6.0 Hz, H-1), 3.74 (1H, dd, *J*_{2,3} 9.5 Hz, H-2), 3.61–3.45 and 3.32–3.00 (6H, m, H-3–H-7_b), 3.63, 3.52, 3.35, 3.25 (12H, 4s, 4 × CH₃), 2.41 (3H, s, CH₃-Ts). ¹³C NMR (90 MHz, CDCl₃) δ 146.7 (C-1), 144.3–127.5 (aromatics), 88.5, 82.7, 79.4, 78.6, 74.1 (C-2–C-6), 71.0 (C-7), 60.9, 60.7, 60.6, 59.3 (4 × CH₃), 21.6 (CH₃-Ts). HR-ESI-MS positive mode (m/z): calcd. for [M + H]⁺ = 417.1692, found: [M + H]⁺ = 417.1694; C₁₈H₂₈N₂O₇S (416.16).

17-2 R_f: 0.10 (1:1 EtOAc–hexane). ¹H NMR (360 MHz, CDCl₃) δ 9.31 (1H, bs, NH), 7.83 (2H, d, *J* 8.2 Hz, aromatics), 7.31 (2H, d, *J* 8.2 Hz, aromatics), 6.80 (1H, d, *J*_{1,2} 4.6 Hz, H-1), 4.02 (1H, dd, *J*_{2,3} 10.3 Hz, H-2), 3.61–3.45 and 3.32–3.00 (6H, m, H-3–H-7_b), 3.63, 3.52, 3.41, 3.30 (4s, 12H, 4 × CH₃), 2.41 (s, 3H, CH₃-Ts). ¹³C NMR (90 MHz, CDCl₃) δ 146.2 (C-1), 144.3–127.5 (aromatics), 87.9, 81.2, 79.4, 78.4, 77.9 (C-2–C-6), 71.3 (C-7), 60.8, 60.4, 59.8, 59.2 (4 × CH₃), 21.6 (CH₃-Ts). HR-ESI-MS positive mode (m/z): calcd. for [M + H]⁺ = 417.1692, found: [M + H]⁺ = 417.1694; C₁₈H₂₈N₂O₇S (416.16).

4.8. Characterization of Anhydro-Heptitols 18

4.8.1. 2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methyl-1-Phenyl-D-glycero-D-gulo-Heptitol (18a)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), phenylboronic acid (1.5 equiv., 0.02 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 18 mg amorphous solid containing **18a** and **21** in 1.3:1 ratio. R_f : 0.50 (1:2 EtOAc–hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.34–6.74 (5H, m, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.59 (3H, s, *CH*₃OC-3), 3.53

(3H, s, CH_3OC-5), 3.55–3.50 (2H, m, H-7_a, H-7_b), 3.36 (3H, s, CH_3OC-7), 3.30 (1H, ddd, $J_{1a,2}$ 2.4, $J_{1b,2}$ 8.8, $J_{2,3}$ 8.9 Hz, H-2), 3.23–3.15 (2H, m, H-4, H-5), 3.12 (1H, ddd, $J_{6,7a}$ 2.0, $J_{6,7b}$ 4.0, $J_{5,6}$ 9.8 Hz, H-6), 3.07 (1H, dd, $J_{1a,1b}$ 14.3 Hz, H-1_a), 2.90 (1H, pseudo t, $J_{3,4}$ 9.0 Hz, strongly coupled, H-3), 2.74 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 139.4–126.1 (aromatics), 89.2 (C-4), 83.7 (C-3), 80.3 (C-2), 80.1 (C-5), 78.8 (C-6), 71.5 (C-7), 60.8 (CH₃OC-4) 60.7 (CH₃OC-3), 60.4 (CH₃OC-5), 59.5 (CH₃OC-7), 37.9 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 333.1672; found: [M + Na]⁺ = 333.1672; C₁₇H₂₆O₅ (310.39).



4.8.2. 2,6-Anhydro-1-Deoxy-1-(4-Dibenzo[b,d]furanyl)-3,4,5,7-Tetra-O-Methyl-D-*glycero*-D-*gulo*-Heptitol (**18b**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), dibenzo[*b,d*]furan-4-ylboronic acid (1.5 equiv., 0.04 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 4 mg (8%) of **18b** as a white amorphous solid. R_f: 0.47 (1:2 EtOAc–hexane). ¹H NMR (500 MHz, CDCl₃) δ 7.93 (1H, d, *J* 7.7 Hz, aromatic), 7.80 (1H, dd, *J* 1.1, 7.7 Hz, aromatic), 7.58 (1H, d, *J* 8.2 Hz, aromatic), 7.47–7.39 (2H, m, aromatics), 7.35–7.30 (1H, m, aromatic), 7.29–7.23 (1H, m, aromatics), 3.67 (3H, s, *CH*₃OC-4), 3.62 (3H, s, *CH*₃OC-3), 3.58 (1H, ddd, *J*_{1a,2} 2.9, *J*_{1b,2} 8.9, *J*_{2,3} 9.2 Hz, H-2), 3.54 (1H, dd, H-1_a), 3.53 (3H, s, *CH*₃OC-5), 3.48 (1H, dd, H-7_a), 3.46 (1H, dd, *J*_{7a,7b} 11.2 Hz, H-7_b), 3.28 (3H, s, *CH*₃OC-7), 3.26 (1H, pseudo t, *J*_{3,4} 8.7 Hz, H-4), 3.21 (1H, pseudo t, *J*_{4,5} 8.8 Hz, H-5), 3.14 (1H, ddd, *J*_{6,7a} 2.5, *J*_{6,7b} 3.4, *J*_{5,6} 9.5 Hz, H-6), 3.09 (1H, dd, *J*_{1a,1b} 14.4 Hz, H-1_b) 3.02 (1H, pseudo t, *J*_{3,4} 8.9 Hz, H-3). ¹³C NMR (90 MHz, CDCl₃) δ 129.4–110.9 (aromatics), 89.2 (C-4), 84.2 (C-3), 80.2 (C-5), 79.0, 78.9 (C-2, C-6), 71.5 (C-7), 60.9 (*CH*₃OC-4), 60.8 (*CH*₃OC-3), 60.5 (*CH*₃OC-5), 59.5 (*CH*₃OC-7), 32.0 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 423.1778, found: [M + Na]⁺ = 423.1777; C₂₃H₂₈O₆ (400.19).



4.8.3. 2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methyl-1-(4-Trifluoromethylphenyl)-D-glycero-D-gulo-Heptitol (18c)

Prepared from tosylhydrazone **17** (0.05 g, 0.13 mmol), (4-trifluoromethly)phenylboronic acid (1.5 equiv., 0.04 g, 0.39 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:3 EtOAc–hexane) to yield 22 mg (45%) of **18c** as a white amorphous solid. R_f: 0.50 (1:2 EtOAc–hexane); $[\alpha]_D - 6 (c 0.30, CH_2Cl_2)$. ¹H NMR (400 MHz, CDCl₃) δ 7.52 (2H, d, *J* 8.1 Hz, aromatics), 7.38 (2H, d, *J* 8.1 Hz, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.59 (3H, s, *CH*₃OC-3), 3.54 (1H, dd, H-7_a), 3.53 (3H, s, *CH*₃OC-5), 3.50 (1H, dd, *J*_{7a,7b} 11.1 Hz, H-7_b), 3.36 (3H, s, *CH*₃OC-7), 3.29 (1H, ddd, *J*_{1a,2} 2.3, *J*_{1b,2} 8.9, *J*_{2,3} 9.2 Hz, H-2), 3.24–3.08 (3H, m, H-1_a, H-4, H-5), 3.13 (1H, ddd, *J*_{6,7a} 2.0, *J*_{6,7b} 3.9, *J*_{5,6} 9.8 Hz, H-6), 2.88 (1H, pseudo t, *J*_{3,4} 8.9 Hz, strongly coupled, H-3), 2.80 (1H, dd, *J*_{1a,1b} 14.2 Hz, H-1_b). ¹³C NMR (100 MHz, CDCl₃) δ 143.7–124.1 (aromatics), 89.2 (C-4), 83.6 (C-3), 80.0 (C-2), 79.8 (C-5), 78.8 (C-6), 71.4 (C-7), 60.8 (*CH*₃OC-4), 60.8 (*CH*₃OC-3), 60.5 (*CH*₃OC-5), 59.5 (*CH*₃OC-7), 37.7 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 379.1727, found: [M + H]⁺ = 379.1727; C₁₈H₂₅F₃O₅ (378.17).



4.8.4. 2,6-Anhydro-1-Deoxy-1-(4-Fluorophenyl)-3,4,5,7-Tetra-O-Methyl-D-*glycero*-D-*gulo*-Heptitol (**18d**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-fluorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 6 mg (14%) of **18d** as a white amorphous solid. R_f: 0.41 (1:2 EtOAc–hexane); $[\alpha]_D + 0.5$ (*c* 0.20, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.22 (2H, dd, *J* 5.6, 8.6 Hz, aromatics), 6.94 (2H, t, *J* 8.8 Hz, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.58 (3H, s, *CH*₃OC-3), 3.54 (1H, dd, H-7_a), 3.53 (3H, s, *CH*₃OC-5), 3.50 (1H, dd, *J*_{7a,7b} 10.8 Hz, H-7_b), 3.37 (3H, s, *CH*₃OC-7), 3.24 (1H, ddd, *J*_{1a,2} 2.1, *J*_{1b,2} 8.8, *J*_{2,3} 9.1 Hz, H-2), 3.21–3.13 (2H, m, H-4, H-5), 3.12 (1H, ddd, *J*_{6,7a} 1.9, *J*_{6,7b} 3.6, *J*_{5,6} 8.7 Hz, H-6), 3.04 (1H, dd, *J*_{1a,1b} 14.3 Hz, H-1_a), 2.87 (1H, pseudo t, *J*_{3,4} 9.0 Hz, strongly coupled, H-3), 2.71 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 131.4–109.0 (aromatics), 89.3 (C-4), 83.6 (C-3), 80.2, (C-2), 80.1 (C-5), 78.8 (C-6), 71.5 (C-7), 60.8 (*CH*₃OC-3, *CH*₃OC-4), 60.5 (*CH*₃OC-5), 59.5 (*CH*₃OC-7), 37.1 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 329.1759, found: [M + Na]⁺ = 329.1759; C₁₇H₂₅FO₅ (328.17).



4.8.5. 2,6-Anhydro-1-(3-Chlorophenyl)-1-Deoxy-3,4,5,7-Tetra-O-Methyl-D-*glycero*-D-*gulo*-Heptitol (**18e**)

Prepared from tosylhydrazone **17** (0.05 g, 0.13 mmol), 3-chorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:2 EtOAc–hexane) to yield 13 mg (29%) of **18e** as a pale-yellow amorphous solid. R_f: 0.48 (1:2 EtOAc–hexane); $[\alpha]_D - 3$ (*c* 0.24, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.32–7.24 (1H, m, aromatic), 7.23–7.08 (3H, m, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.59 (3H, s, *CH*₃OC-3), 3.56 (1H, dd, *J*_{7a,7b} 11.0 Hz, H-7a), 3.53 (3H, s, *CH*₃OC-5), 3.51 (1H, dd, H-7_b), 3.37 (3H, s, *CH*₃OC-7), 3.27 (1H, ddd, *J*_{1a,2} 2.3, *J*_{1b,2} 8.8, *J*_{2,3} 9.1 Hz, H-2), 3.23–3.14 (2H, m, H-4, H-5), 3.13 (1H, ddd, *J*_{6,7a} 1.6, *J*_{6,7b} 3.4, *J*_{5,6} 8.6 Hz, H-6), 3.04 (1H, dd, *J*_{1a,1b} 14.3 Hz, H-1a), 2.87 (1H, pseudo t, *J*_{3,4} 8.8 Hz, H-3), 2.71 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 141.3–126.0 (aromatics), 89.2 (C-4), 83.5 (C-3), 80.1 (C-2), 79.9 (C-5), 78.8 (C-6), 71.5 (C-7), 60.9 (*CH*₃OC-4), 60.8 (*CH*₃OC-3), 60.5 (*CH*₃OC-5), 59.6 (*CH*₃OC-7), 37.6 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 345.1463, found: [M + H]⁺ = 345.1460; C₁₇H₂₅ClO₅ (344.14).



4.8.6. 2,6-Anhydro-1-(4-Bromophenyl)-1-Deoxy-3,4,5,7-Tetra-O-Methyl-D-*glycero*-D-*gulo*-Heptitol (**18f**)

Prepared from tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-bromophenylboronic acid (1.5 equiv., 0.04 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:4 EtOAc–hexane) to yield 11 mg (22%) of **18f** as a white amorphous solid. R_f: 0.53 (1:2 EtOAc–hexane); $[\alpha]_D - 6 (c 0.21, CH_2Cl_2)$. ¹H NMR (400 MHz, CDCl₃) δ 7.38 (2H, d, *J* 8.4 Hz, aromatics), 7.14 (2H, d, *J* 8.4 Hz, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.58 (3H, s, *CH*₃OC-3), 3.53 (3H, s, *CH*₃OC-5), 3.53 (1H, dd, H-7_a), 3.49 (1H, dd, *J*_{7a,7b} 10.8 Hz, H-7_b), 3.37 (3H, s, *CH*₃OC-7), 3.24 (1H, ddd, *J*_{1a,2} 2.2, *J*_{1b,2} 8.9, *J*_{2,3} 9.1 Hz, H-2), 3.21–3.13 (2H, m, H-4, H-5), 3.11 (1H, ddd, *J*_{6,7a} 2.1, *J*_{6,7b} 3.5,

 $J_{5,6}$ 9.8 Hz, H-6), 3.02 (1H, dd, $J_{1a,1b}$ 14.3 Hz, H-1_a), 2.87 (1H, pseudo t, $J_{3,4}$ 9.0 Hz, strongly coupled, H-3), 2.69 (1H, dd, H-1_b). ¹³C NMR (100 MHz, CDCl₃) δ 138.4–119.3 (aromatics), 89.2 (C-4), 83.5 (C-3), 80.0 (C-2, C-5), 78.8 (C-6), 71.4 (C-7), 60.9 (CH₃OC-4), 60.8 (CH₃OC-3), 60.5 (CH₃OC-5), 59.5 (CH₃OC-7), 37.3 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 389.0958, found: [M + H]⁺ = 389.0959; C₁₇H₂₅BrO₅ (389.29).

4.8.7.

2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methyl-1-(4-Nitrophenyl)-D-*glycero*-D-*gulo*-Heptitol (18g)

(18g) Prepared from tosylhydrazone 17 (0.05 g, 0.13 mmol), 4-nitrophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I. Purified by column chromatography (1:2 EtOAc–hexane) to yield 21 mg (46%) of 18g as a yellow amorphous solid. R_f: 0.26 (1:2 EtOAc–hexane); $[\alpha]_D + 3 (c 0.22, CH_2Cl_2)$. ¹H NMR (400 MHz, CDCl₃) δ 8.13 (2H, d, *J* 8.7 Hz, aromatics), 7.43 (2H, d, *J* 8.7 Hz, aromatics), 3.66 (3H, s, *CH*₃OC-4), 3.60 (3H, s, *CH*₃OC-3), 3.53 (3H, s, *CH*₃OC-5), 3.53 (1H, dd, H-7_a), 3.49 (1H, dd, *J*_{7a,7b} 10.8 Hz, H-7_b), 3.37 (3H, s, *CH*₃OC-7), 3.29 (1H, ddd, *J*_{1a,2} 2.4, *J*_{1b,2} 9.0, *J*_{2,3} 9.2 Hz, H-2), 3.23–3.14 (3H, m, H-1_a, H-4, H-5), 3.12 (1H, ddd, *J*_{6,7a} 1.3, *J*_{6,7b} 3.1, *J*_{5,6} 9.6 Hz, H-6), 2.89 (1H, pseudo t, *J*_{3,4} 8.8 Hz, strongly coupled, H-3), 2.85 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 147.6–121.1 (aromatics), 89.2 (C-4), 83.5 (C-3), 80.0 (C-2), 79.6 (C-5), 78.8 (C-6), 71.4 (C-7), 60.9 (*CH*₃OC-3, *CH*₃OC-4), 60.5 (*CH*₃OC-5), 59.5 (*CH*₃OC-7), 37.8 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 356.1704, found: [M + H]⁺ = 356.1704; C₁₇H₂₅NO₇ (355.16).



4.8.8. 2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methyl-1-(4-Methoxyphenyl)-D-glycero-D-gulo-Heptitol (**18h**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-methoxyphenylbor onic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 4 mg (9%) of **18h** as a pale-yellow amorphous solid. R_f: 0.41 (1:2 EtOAc–hexane); $[\alpha]_D + 5$ (*c* 0.57, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.19 (2H, d, *J* 8.6 Hz, aromatics), 6.81 (2H, d, *J* 8.7 Hz, aromatics), 3.79 (3H, s, OCH₃), 3.65 (3H, s, *CH*₃OC-4), 3.59 (3H, s, *CH*₃OC-3), 3.55 (1H, dd, *J*_{7a,7b} 10.9 Hz, H-7_a), 3.53 (3H, s, *CH*₃OC-5), 3.50 (1H, dd, H-7_b), 3.38 (3H, s, *CH*₃OC-7), 3.24 (1H, ddd, *J*_{1a,2} 2.3, *J*_{1b,2} 8.8, *J*_{2,3} 9.1 Hz, H-2), 3.21–3.14 (2H, m, H-4, H-5), 3.12 (1H, ddd, *J*_{6,7a} 2.0, *J*_{6,7b} 3.9, *J*_{5,6} 9.8 Hz, H-6), 3.01 (1H, dd, *J*_{1a,1b} 14.3 Hz, H-1_a), 2.88 (1H, pseudo t, *J*_{3,4} 9.0 Hz, strongly coupled, H-3), 2.69 (1H, dd, H-1_b). ¹³C NMR (90 MHz, CDCl₃) δ 162.4–109.6 (aromatics), 89.3 (C-4), 83.7 (C-3), 80.5 (C-2), 80.1 (C-5), 78.8 (C-6), 71.6 (C-7), 60.8 (*CH*₃OC-3, *CH*₃OC-4), 60.5 (*CH*₃OC-5), 59.6 (*CH*₃OC-7), 55.4 (OCH₃), 37.0 (C-1). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 341.1959, found: [M + H]⁺ = 341.1957; C₁₈H₂₈O₅ (340.42).



4.8.9. 2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methyl-1-(4-Methylphenyl)-D-glycero-D-gulo-Heptitol (18i)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-methylphenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 11 mg white amorphous solid containing **18i** and **21** in 3:1 ratio. R_f: 0.48 (1:2 EtOAc–hexane); $[\alpha]_D + 0.5$ (*c* 0.08, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.16 (2H, d, *J* 7.9 Hz, aromatics), 7.07 (2H, d, *J* 7.9 Hz, aromatics), 3.65 (3H, s, *CH*₃OC-4), 3.59 (3H, s, *CH*₃OC-3), 3.53 (3H, s, *CH*₃OC-5), 3.55–3.50 (2H, m, H-7_a, H-7_b), 3.37 (3H, s, *CH*₃OC-7), 3.26 (1H, ddd, *J*_{1a,2} 2.1, *J*_{1b,2} 8.8, *J*_{2,3} 9.1 Hz, H-2), 3.23–3.14 (2H, m, H-4, H-5), 3.11 (1H, ddd, *J*_{6,7a} 1.9, *J*_{6,7b} 3.6, *J*_{5,6} 9.7 Hz, H-6), 3.03 (1H, dd, *J*_{1a,1b} 14.3 Hz, H-1_a), 2.88 (1H, pseudo t, *J*_{3,4} 9.0 Hz, strongly coupled, H-3), 2.70 (1H, dd, H-1_b), 2.31 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 136.2–128.6 (aromatics), 89.3 (C-4), 83.7 (C-3), 80.4 (C-2), 80.1 (C-5), 78.8 (C-6), 71.5 (C-7), 60.8 (*CH*₃OC-3, *CH*₃OC-4), 60.5 (*CH*₃OC-5), 59.6 (*CH*₃OC-7), 37.5 (C-1), 21.2 (CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + H]⁺ = 325.2010, found: [M + H]⁺ = 325.2008; C₁₈H₂₈O₅ (324.42).



4.9. Characterization of Heptenitols 19 and 20

4.9.1. (*E*)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methyl-1-Phenyl-D-*gluco*-Hept-1-Enitol (**19a**) and (*Z*)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methyl-1-Phenyl-D-*gluco*-Hept-1-Enitol (**20a**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), phenylboronic acid (1.5 equiv., 0.02 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 29 mg white amorphous solid containing **19a** and **20a** in 9:1 ratio. R_f : 0.16 (1:2 EtOAc–hexane), $[\alpha]_D$ + 28 (*c* 0.16, CH₂Cl₂).



19a: ¹H NMR (500 MHz, CDCl₃) δ 7.42 (2H, d, *J* 7.6 Hz, aromatics), 7.38–7.30 (2H, m, aromatics), 7.29–7.23 (1H, m, aromatic), 6.63 (1H, d, *J*_{1,2} 16.0 Hz, H-1), 6.16 (1H, dd, *J*_{2,3} 8.2 Hz, H-2), 4.05 (1H, dd, *J*_{3,4} 6.0 Hz, H-3), 3.96 (1H, ddd, *J*_{6,7a} 3.9, *J*_{6,7b} 5.5, *J*_{5,6} 6.7 Hz, H-6), 3.60 (3H, s, *CH*₃OC-4), 3.59–3.50 (3H, m, H-4, H-7_a, H-7_b), 3.40 (6H, 2s, *CH*₃OC-5, *CH*₃OC-7), 3.40–3.37 (1H, m, H-5), 3.37 (3H, s, *CH*₃OC-3), 3.32 (1H, bs, OH). ¹³C NMR (125 MHz, CDCl₃) δ 134.0 (C-1), 137.0–126.3 (aromatics), 126.7 (C-2), 83.8 (C-4), 83.4 (C-3), 79.8 (C-5), 73.8 (C-7), 70.2 (C-6), 60.8 (*CH*₃OC-4), 59.4 (*CH*₃OC-5), 59.2 (*CH*₃OC-7), 56.8 (*CH*₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 333.1672, found: [M + Na]⁺ = 333.1679; C₁₇H₂₆O₅ (310.39).

20a: ¹H NMR (500 MHz, CDCl₃) δ 7.45–7.38 (2H, m, aromatics), 7.38–7.30 (2H, m, aromatics), 7.29–7.23 (1H, m, aromatic), 6.77 (1H, d, $J_{1,2}$ 12.0 Hz, H-1), 5.58 (1H, dd, $J_{2,3}$ 10.0 Hz, H-2), 4.59 (1H, dd, $J_{3,4}$ 4.6 Hz, H-3), 3.99–3.91 (1H, m, H-6), 3.57 (3H, s, CH_3OC -4), 3.57–3.49 (3H, m, H-4, H-7_a, H-7_b), 3.45 (1H, dd, $J_{4,5}$ 3.6, $J_{5,6}$ 6.4 Hz, H-5), 3.40 (3H, s, CH_3OC -7), 3.32 (3H, s, CH_3OC -5), 3.23 (3H, s, CH_3OC -3), 3.02 (1H, bs, OH). ¹³C NMR (125 MHz, CDCl₃) δ 133.9 (C-1), 137.0–126.3 (aromatics), 129.5 (C-2), 84.1 (C-4), 79.6 (C-5), 76.8 (C-3), 73.9 (C-7), 70.4 (C-6), 60.7 (CH_3OC -4), 59.2 (CH_3OC -7), 59.1 (CH_3OC -5), 56.4 (CH_3OC -3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 333.1672, found: [M + Na]⁺ = 333.1669; C₁₇H₂₆O₅ (310.39).

4.9.2. (*E*)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-1-(4-Trifluoromethylphenyl)-D-*gluco*-Hept-1-Enitol (**19c**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), (4-trifluoromethly)ph enylboronic acid (1.5 equiv., 0.04 g, 0.39 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:3 EtOAc–hexane) to yield 11 mg white amorphous solid containing **19c** and an unidentified impurity in 3:1 ratio. R_f: 0.41 (1:2 EtOAc–hexane). ¹H NMR (500 MHz, CDCl₃) δ 8.40–7.40 (4H, m, aromatics), 6.68 (1H, d, $J_{1,2}$ 16.0 Hz, H-1), 6.29 (1H, dd, $J_{2,3}$ 7.7 Hz, H-2), 4.09 (1H, dd, $J_{3,4}$ 5.9 Hz, H-3), 4.00–3.91 (1H, m, H-6), 3.60 (3H, s, *CH*₃OC-4), 3.60–3.53 (3H, m, H-4, H-7_a, H-7_b), 3.41 (3H, s, *CH*₃OC-5), 3.40 (3H, s, *CH*₃OC-7), 3.39 (3H, s, *CH*₃OC-3), 3.40–3.36 (1H, m, H-5), 3.07 (1H, bs, OH). ¹³C NMR (90 MHz, CDCl₃) δ 132.1 (C-1), 140.0–120.5 (aromatics), (2H₃OC-7), 59.2 (*CH*₃OC-5), 57.1 (*CH*₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 401.1546, found: [M + Na]⁺ = 401.1542; C₁₈H₂₅F₃O₅ (378.17).



4.9.3. (*E*)-1,2-Dideoxy-1-(4-Fluorophenyl)-3,4,5,7-Tetra-*O*-Methyl-D-*gluco*-Hept-1-Enitol (**19d**) and (*Z*)-1,2-Dideoxy-(4-Fluorophenyl)-3,4,5,7-Tetra-*O*-Methyl-1-D-*gluco*-Hept-1-Enitol (**20d**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-fluorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 31 mg white amorphous solid containing **19d** and **20d** in 3:1 ratio. R_f : 0.11 (1:2 EtOAc–hexane).



19d: ¹H NMR (400 MHz, CDCl₃) δ 7.39 (2H, dd, *J* 5.4, 8.7 Hz, aromatics), 7.03 (2H, t, *J* 8.7 Hz, aromatics), 6.60 (1H, d, *J*_{1,2} 16.0 Hz, H-1), 6.09 (1H, dd, *J*_{2,3} 8.1 Hz, H-2), 4.04 (1H, dd, *J*_{3,4} 5.9 Hz, H-3), 4.00–3.91 (1H, m, H-6), 3.59 (3H, s, *CH*₃OC-4), 3.59–3.49 (3H, m, H-4, H-7_a, H-7_b), 3.40 (6H, 2s, *CH*₃OC-5, *CH*₃OC-7), 3.38 (1H, dd, *J*_{4,5} 3.1, *J*_{5,6} 7.3 Hz, H-5), 3.36 (3H, s, *CH*₃OC-3), 3.03 (1H, bs, OH). ¹³C NMR (90 MHz, CDCl₃) δ 132.7 (C-1), 129.7–110.2 (aromatics), 126.4 (C-2), 83.7 (C-4), 83.2 (C-3), 79.8 (C-5), 73.7 (C-7), 70.2 (C-6), 60.8 (*CH*₃OC-4), 59.4 (*CH*₃OC-5), 59.2 (*CH*₃OC-7), 56.9 (*CH*₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 351.1578, found: [M + Na]⁺ = 351.1579; C₁₇H₂₅FO₅ (328.17).

20d: ¹H NMR (500 MHz, CDCl₃) δ 7.34 (2H, dd, *J* 5.5, 8.5 Hz, aromatics), 7.03 (2H, t, *J* 8.7 Hz, aromatics), 6.72 (1H, d, *J*_{1,2} 11.9 Hz, H-1), 5.66 (1H, dd, *J*_{2,3} 10.1 Hz, H-2), 4.54 (1H, dd, *J*_{3,4} 4.8 Hz, H-3), 3.98–3.91 (1H, m, H-6), 3.57 (3H, s, *CH*₃OC-4), 3.57–3.50 (3H, m, H-4, H-7_a, H-7_b), 3.45 (1H, dd, *J*_{4,5} 3.3, *J*_{5,6} 6.6 Hz, H-5), 3.40 (3H, s, *CH*₃OC-7), 3.33 (3H, s, *CH*₃OC-5), 3.22 (3H, s, *CH*₃OC-3), 3.17 (1H, bs, OH). ¹³C NMR (125 MHz, CDCl₃) δ 132.7 (C-1), 131.2–114.5 (aromatics), 129.4 (C-2), 84.1 (C-4), 79.6 (C-5), 76.8 (C-3), 73.9 (C-7), 70.4 (C-6), 60.7 (*CH*₃OC-4), 59.3 (*CH*₃OC-7), 59.0 (*CH*₃OC-5), 56.4 (*CH*₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 351.1578, found: [M + Na]⁺ = 351.1579; C₁₇H₂₅FO₅ (328.17).

4.9.4. (*E*)-1-(3-Chlorophenyl)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-D-*gluco*-Hept-1-Enitol (**19e**) and (*Z*)-1-(3-Chlorophenyl)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-D-*gluco*-Hept-1-Enitol (**20e**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 3-chorophenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 18 mg pale yellow amorphous solid containing **19e** and **20e** in 9:1 ratio. R_f : 0.13 (1:2 EtOAc–hexane).



19e: ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.37 (1H, m, aromatic), 7.36–7.19 (3H, m, aromatics), 6.58 (1H, d, $J_{1,2}$ 16.0 Hz, H-1), 6.19 (1H, dd, $J_{2,3}$ 7.9 Hz, H-2), 4.06 (1H, dd, $J_{3,4}$ 6.2 Hz, H-3), 4.01–3.90 (1H, m, H-6), 3.60 (3H, s, CH_3OC -4), 3.59–3.44 (3H, m, H-4, H-7_a, H-7_b), 3.40 (6H, 2s, CH_3OC -5, CH_3OC -7), 3.38 (1H, dd, $J_{4,5}$ 2.6, $J_{5,6}$ 7.4 Hz, H-5), 3.37 (3H, s, CH_3OC -3), 3.11 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 132.5 (C-1), 138.8–120.5 (aromatics), 124.9 (C-2), 83.5 (C-4), 82.9 (C-3), 79.7 (C-5), 73.6 (C-7), 70.2 (C-6), 60.8 (CH_3OC -4), 59.4 (CH_3OC -5), 59.2 (CH_3OC -7), 57.0 (CH_3OC -3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 367.1283, found: [M + Na]⁺ = 367.1282; C₁₇H₂₅ClO₅ (344.14).

20e: ¹H NMR (400 MHz, CDCl₃) δ 7.84–6.06 (5H, m, H-1, aromatics), 5.75 (1H, dd, $J_{1,2}$ 11.9, $J_{2,3}$ 10.01 Hz, H-2), 4.52 (1H, dd, $J_{3,4}$ 4.3 Hz, H-3), 4.12–3.71 (1H, m, H-6), 3.57 (3H, s, CH_3OC -4), 3.58–3.42 (4H, m, H-4, H-5, H-7_a, H-7_b), 3.40 (3H, s, CH_3OC -7), 3.40–3.23 (3H, m, CH_3OC -5), 3.23 (3H, s, CH_3OC -3), 3.11 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 132.4 (C-1), 138.8–120.5 (aromatics), 129.3 (C-2), 83.9 (C-4), 83.0 (C-5), 81.4 (C-3), 74.5 (C-7), 73.9 (C-6), 60.4 (CH_3OC -4), 59.3 (CH_3OC -7), 59.1 (CH_3OC -5), 57.0 (CH_3OC -3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 367.1283, found: [M + Na]⁺ = 367.1282; C₁₇H₂₅ClO₅ (344.14).

4.9.5. (*E*)-1-(4-Bromophenyl)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methyl-D-*gluco*-Hept-1-Enitol (**19f**) and (*Z*)-1-(4-Bromophenyl)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methyl-D-*gluco*-Hept-1-Enitol (**20f**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-bromophenylboronic acid (1.5 equiv., 0.04 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:4 EtOAc–hexane) to yield 20 mg white amorphous solid containing **19f** and **20f** in 9:1 ratio. R_f : 0.10 (1:2 EtOAc–hexane).



19f: ¹H NMR (400 MHz, CDCl₃) δ 7.46 (2H, d, *J* 8.5 Hz, aromatics), 7.28 (2H, d, *J* 8.5 Hz, aromatics), 6.57 (1H, d, *J*_{1,2} 16.0 Hz, H-1), 6.18 (1H, dd, *J*_{2,3} 7.9 Hz, H-2), 4.04 (1H, dd, *J*_{3,4} 5.9 Hz, H-3), 3.99–3.90 (1H, m, H-6), 3.59 (3H, s, *CH*₃OC-4), 3.58–3.50 (3H, m, H-4, H-7_a, H-7_b), 3.40 (3H, s, *CH*₃OC-5), 3.39 (3H, s, *CH*₃OC-7), 3.37 (3H, s, *CH*₃OC-3), 3.37 (1H, dd, *J*_{4,5} 2.8, *J*_{5,6} 6.7 Hz, H-5), 3.00 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 132.4 (C-1), 136.1–117.1 (aromatics), 127.6 (C-2), 83.6 (C-4), 83.0 (C-3), 79.7 (C-5), 73.7 (C-7), 70.2 (C-6), 60.7 (*CH*₃OC-4), 59.4 (*CH*₃OC-5), 59.2 (*CH*₃OC-7), 57.0 (*CH*₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 411.0778, found: [M + Na]⁺ = 411.0777; C₁₇H₂₅BrO₅ (389.29).

20f: ¹H NMR (400 MHz, CDCl₃) δ 7.47 (2H, d, *J* 8.4 Hz, aromatics), 7.25 (2H, d, *J* 8.4 Hz, aromatics), 6.69 (1H, d, *J*_{1,2} 11.9 Hz, H-1), 5.71 (1H, dd, *J*_{2,3} 10.1 Hz, H-2), 4.53 (1H, dd, *J*_{3,4}

4.7 Hz, H-3), 3.99–3.90 (1H, m, H-6), 3.57 (3H, s, CH_3OC-4), 3.56–3.47 (3H, m, H-4, H-7_a, H-7_b), 3.45 (1H, dd, $J_{4,5}$ 3.3, $J_{5,6}$ 6.5 Hz, H-5), 3.40 (3H, s, CH_3OC-7), 3.34 (3H, s, CH_3OC-5), 3.21 (3H, s, CH_3OC-3), 3.00 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 132.6 (C-1), 136.1–117.1 (aromatics), 130.2 (C-2), 84.0 (C-4), 79.5 (C-5), 76.8 (C-3), 73.8 (C-7), 70.4 (C-6), 60.7 (CH₃OC-4), 59.3 (CH₃OC-7), 59.0 (CH₃OC-5), 56.4 (CH₃OC-3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 411.0778, found: [M + Na]⁺ = 411.0777; C₁₇H₂₅BrO₅ (389.29).

4.9.6. (*E*)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-1-(4-Methoxyphenyl)-D-*gluco*-Hept-1-Enitol (**19h**) and (*Z*)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-1-(4-Methoxyphenyl)-D-*gluco*-Hept-1-Enitol (**20h**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-methoxyphenylbor onic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K_3PO_4 (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 24 mg pale yellow amorphous solid containing **19h** and **20h** in 23:1 ratio. R_f : 0.13 (1:2 EtOAc–hexane).



19h: ¹H NMR (400 MHz, CDCl₃) δ 7.36 (2H, d, *J* 8.7 Hz, aromatics), 6.88 (2H, d, *J* 8.7 Hz, aromatics), 6.57 (1H, d, $J_{1,2}$ 16.0 Hz, H-1), 6.01 (1H, dd, $J_{2,3}$ 8.3 Hz, H-2), 4.02 (1H, dd, $J_{3,4}$ 6.0 Hz, H-3), 3.98–3.91 (1H, m, H-6), 3.82 (3H, s, OCH₃), 3.60 (3H, s, *CH*₃OC-4), 3.59–3.53 (2H, m, H-7_a, H-7_b), 3.54–3.49 (1H, m, H-4), 3.40 (3H, s, *CH*₃OC-5), 3.39 (3H, s, *CH*₃OC-7), 3.38 (1H, dd, $J_{4,5}$ 2.9, $J_{5,6}$ 6.9 Hz, H-5), 3.35 (3H, s, *CH*₃OC-3), 3.02 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 133.6 (C-1), 159.9–112.7 (aromatics), 124.3 (C-2), 83.9 (C-4), 83.6 (C-3), 79.8 (C-5), 73.8 (C-7), 70.3 (C-6), 60.8 (*CH*₃OC-4), 59.4 (*CH*₃OC-5), 59.2 (*CH*₃OC-7), 56.7 (*CH*₃OC-3), 55.5 (OCH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 363.1778, found: [M + Na]⁺ = 363.1779; C₁₈H₂₈O₅ (340.42).

20h: ¹H NMR (400 MHz, CDCl₃) δ 7.32 (2H, d, *J* 8.7 Hz, aromatics), 6.88 (2H, d, *J* 8.7 Hz, aromatics), 6.69 (1H, d, *J*_{1,2} 12.0 Hz, H-1), 5.56 (1H, dd, *J*_{2,3} 10.0 Hz, H-2), 4.63 (1H, dd, *J*_{3,4} 4.7 Hz, H-3), 3.99–3.90 (1H, m, H-6), 3.82 (3H, s, OCH₃), 3.58 (3H, s, *CH*₃OC-4), 3.57–3.49 (3H, m, H-4, H-7_a, H-7_b), 3.46 (1H, dd, *J*_{4,5} 3.3, *J*_{5,6} 6.6 Hz, H-5), 3.40 (3H, s, *CH*₃OC-7), 3.33 (3H, s, *CH*₃OC-5), 3.23 (3H, s, *CH*₃OC-3), 3.02 (1H, bs, OH). ¹³C NMR (100 MHz, CDCl₃) δ 133.3 (C-1), 161.0–112.7 (aromatics), 130.4 (C-2), 84.2 (C-4), 79.6 (C-5), 77.0 (C-3), 73.9 (C-7), 70.4 (C-6), 60.7 (*CH*₃OC-4), 59.3 (*CH*₃OC-7), 59.0 (*CH*₃OC-5), 56.3 (*CH*₃OC-3), 55.4 (OCH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 363.1778, found: [M + Na]⁺ = 363.1776; C₁₈H₂₈O₅ (340.42).

4.9.7. (*E*)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-1-(4-Methylphenyl)-D-*gluco*-Hept-1-Enitol (**19i**) and (*Z*)-1,2-Dideoxy-3,4,5,7-Tetra-*O*-Methyl-1-(4-Methylphenyl)-D-*gluco*-Hept-1-Enitol (**20i**)

Isolated from a reaction of tosylhydrazone **17** (0.05 g, 0.13 mmol), 4-methylphenylboronic acid (1.5 equiv., 0.03 g, 0.19 mmol), and K₃PO₄ (3 equiv., 0.08 g, 0.39 mmol) according to General procedure I by column chromatography (1:2 EtOAc–hexane) to yield 24 mg pale white amorphous solid containing **19i** and **20i** in 8:1 ratio. R_f: 0.13 (1:2 EtOAc–hexane), $[\alpha]_D + 28$ (*c* 0.36, CH₂Cl₂).



19i: ¹H NMR (400 MHz, CDCl₃) δ 7.32 (2H, d, J 8.1 Hz, aromatics), 7.15 (2H, d, J 7.9 Hz, aromatics), 6.60 (1H, d, $J_{1,2}$ 16.0 Hz, H-1), 6.10 (1H, dd, $J_{2,3}$ 8.3 Hz, H-2), 4.03 (1H, dd, $J_{3,4}$ 6.0 Hz, H-3), 3.98–3.90 (1H, m, H-6), 3.60 (3H, s, *CH*₃OC-4), 3.58–3.49 (3H, m, H-4, H-7_a, H-7_b), 3.39 (6H, 2s, *CH*₃OC-5, *CH*₃OC-7), 3.39–3.36 (1H, m, H-5), 3.35 (3H, s, *CH*₃OC-3), 3.03 (1H, bs, OH), 2.35 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 134.0 (C-1), 138.3–125.3 (aromatics), 125.5 (C-2), 83.8 (C-4), 83.6 (C-3), 79.8 (C-5), 73.8 (C-7), 70.2 (C-6), 60.8 (*CH*₃OC-4), 59.4 (*CH*₃OC-5), 59.2 (*CH*₃OC-7), 56.7 (*CH*₃OC-3), 21.3 (CH₃). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 347.1829, found: [M + Na]⁺ = 347.1828; C₁₈H₂₈O₅ (324.42).

20i: ¹H NMR (400 MHz, CDCl₃) δ 7.28–7.23 (4H, m, aromatics), 6.73 (1H, d, $J_{1,2}$ 12.1 Hz, H-1), 5.62 (1H, dd, $J_{2,3}$ 10.0 Hz, H-2), 4.61 (1H, dd, $J_{3,4}$ 4.6 Hz, H-3), 3.98–3.90 (1H, m, H-6), 3.57 (3H, s, CH_3OC -4), 3.58–3.49 (3H, m, H-4, H-7_a, H-7_b), 3.46 (1H, dd, $J_{4,5}$ 3.4, $J_{5,6}$ 6.5 Hz, H-5), 3.40 (3H, s, CH_3OC -7), 3.33 (3H, s, CH_3OC -5), 3.23 (3H, s, CH_3OC -3), 3.03 (1H, bs, OH), 2.36 (3H, s, CH₃). ¹³C NMR δ 133.7 (C-1), 138.2–125.3 (aromatics), 128.7 (C-2), 84.1 (C-4), 79.6 (C-5), 76.7 (C-3), 73.9 (C-7), 70.5 (C-6), 60.7 (CH_3OC -4), 59.2 (CH_3OC -7), 59.1 (CH_3OC -5), 56.4 (CH_3OC -3), 21.3 (CH_3). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 347.1829, found: [M + Na]⁺ = 347.1828; C₁₈H₂₈O₅ (324.42).

4.10. 2,6-Anhydro-3,4,5,7-Tetra-O-Methoxymethyl-D-glycero-L-manno-Heptononitrile (2,3,4,6-Tetra-O-Methoxymethyl- β -D-Galactopyranosyl Cyanide) (**23**)

β-D-Galactopyranosyl cyanide 22 (0.10 g, 0.53 mmol) was suspended in dichloromethane (7 mL). The suspension was stirred under nitrogen atmosphere and cooled to 0 $^{\circ}$ C, and then N-diisopropylethylamine (6.4 equiv. / OH, 2.3 mL, 1.75 g, 13.55 mmol) was added, followed by careful addition of chloromethyl methyl ether (10 equiv. / OH, 1.6 mL, 1.70 g, 21.13 mmol), dropwise. The reaction mixture was stirred in the dark at room temperature. When TLC (1:1 EtOAc-hexane) indicated complete consumption of the starting compound (3 day), the mixture was cooled to 0 °C. Saturated aqueous NH₄Cl solution (1 mL) was added to the reaction mixture. The organic layer was separated, washed with water (1 mL), then the aquous phase was washed with dichloromethane $(3 \times 3 \text{ mL})$. The combined organic phase was washed with water (1 mL) and dried on anhydrous magnesium sulfate. The solution was concentrated under reduced pressure and purified by column chromatography (1:1 EtOAc-hexane) to yield 163 mg (84%) of 23 as a colourless oil. R_f: 0.45 (1:1 EtOAc–hexane); [α]_D – 40 (*c* 0.29, CHCl₃). ¹H NMR (400 MHz, DMSO-*d*₆) § 4.86 (1H, d, J 6.7 Hz, CH₂), 4.77 (1H, d, J 6.7 Hz, CH₂), 4.72 (1H, d, J 6.6 Hz, CH₂), 4.71 (1H, d, J 6.6 Hz, CH₂), 4.65–4.58 (3H, m, H-2, CH₂), 4.57 (2H, s, 2 × CH₂), 4.00 (1H, dd, J_{5.6} 0.6 Hz, H-5), 3.91 (1H, pseudo t, J_{2.3} 9.8, J_{3.4} 9.6 Hz, H-3), 3.85 (1H, ddd, J_{6.7a} 5.9, J_{6,7b} 5.9 Hz, H-6), 3.76 (1H, dd, J_{4,5} 2.7 Hz, H-4), 3.58 (1H, dd, J_{7a,7b} 11.0 Hz, H-7a), 3.56 (1H, dd, H-7_b), 3.37, 3.32, 3.31, 3.26 (12H, 4s, 4 × CH₃). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 117.5 (C-1 = CN), 97.0, 95.9, 94.6 (4 × CH₂), 77.7, 77.2, 72.3, 72.2, 66.6 (C-2–C-6), 66.2 (C-7), 56.1, 55.4, 55.3, 54.8 ($4 \times CH_3$). HR-ESI-MS positive mode (m/z): calcd. for [M + H]⁺ = 366.1759, found: $[M + H]^+ = 366.1761; C_{15}H_{27}NO_9$ (365.17).

4.11. 2,6-Anhydro-3,4,5,7-Tetra-O-Methoxymethyl-D-glycero-L-manno-Heptose Tosylhydrazone (C-(2,3,4,6-Tetra-O-Methoxymethyl-β-D-Galactopyranosyl) Formaldehyde Tosylhydrazone) (**24**)

Prepared from cyanide **23** (0.10 g, 0.27 mmol) according to General procedure III. Purified by column chromatography (2:1 EtOAc–hexane) to get two unidentified isomers **24-1** and **24-2**.

24-1 yellow oil, 19 mg (13%); R_f : 0.33 (2:1 EtOAc–hexane). ¹H NMR (360 MHz, CDCl₃) δ 9.50 (1H, s, NH), 7.84–7.75 (2H, m, aromatics), 7.33–7.22 (2H, m, aromatics), 4.89 (1H, d, *J* 6.8 Hz, CH₂), 4.85 (1H, d, *J* 6.5 Hz, CH₂), 4.79 (1H, d, *J* 6.8 Hz, CH₂), 4.73–4.59 (4H, m, CH₂), 4.57 (1H, d, *J* 6.5 Hz, CH₂), 4.03 (1H, dd, *J*_{4,5} 2.4, *J*_{5,6} 0.6 Hz, H-5), 4.03–3.99 (1H, m, H-2 or H-4), 3.98 (1H, pseudo t, *J*_{2,3} 9.9, *J*_{3,4} 9.9 Hz, H-3), 3.78–3.65 (4H, m, H-2 or H-4, H-6, H-7_a, H-7_b), 3.41, 3.39, 3.21 (12H, 4s, 4 × CH₃), 2.42 (3H, s, CH₃-Ts). HR-ESI-MS positive mode (m/z): calcd. for [M + H]⁺ = 537.2113, found: [M + H]⁺ = 537.2111; C₂₂H₃₆N₂O₁₁S (536.20).

24-2 yellow oil, 96 mg (65%); R_f : 0.19 (2:1 EtOAc–hexane).¹H NMR (360 MHz, CDCl₃) δ 8.25 (1H, s, NH), 7.86–7.73 (2H, m, aromatics), 7.35–7.23 (2H, m, aromatics), 7.05 (1H, d, $J_{1,2}$ 4.4 Hz, H-1), 4.87 (1H, d, J 6.7 Hz, CH₂), 4.77 (1H, d, J 6.6 Hz, CH₂), 4.72–4.67 (2H, m, CH₂), 4.65 (1H, d, J 6.7 Hz, CH₂), 4.60 (2H, s, CH₂), 4.42 (1H, d, J 6.7 Hz, CH₂), 4.02 (1H, dd, $J_{4,5}$ 2.6, $J_{5,6}$ 0.6 Hz, H-5), 3.88–3.78 (2H, m) and 3.75–3.55 (4H, m) and 3.46–3.19 (1H, m): (H-2, H-3, H-4, H-6, H-7a, H-7b), 3.39, 3.32, 3.05 (12H, 4s, 4 × CH₃), 2.42 (3H, s, CH₃-Ts). ¹³C NMR (90 MHz, CDCl₃) δ 146.6 (C-1), 144.8–127.4 (aromatics), 98.2, 97.6, 96.9, 95.7 (4 × CH₂), 79.1, 78.8, 77.3, 74.6, 72.9 (C-2–C-6), 66.9 (C-7), 56.2, 55.9, 55.6 (4 × CH₃-Ts). HR-ESI-MS positive mode (m/z): calcd. for [M + H]⁺ = 537.2113, found: [M + H]⁺ = 537.2111; C₂₂H₃₆N₂O₁₁S (536.20).

4.12. Characterization of Anhydro-Heptitol **25** and Heptenitols **26** and **27** 4.12.1.

2,6-Anhydro-1-Deoxy-3,4,5,7-Tetra-O-Methoxymethyl-1-Phenyl-D-*glycero*-D-*gulo*-Heptitol (25)

Isolated from a reaction of tosylhydrazone 24 (0.10 g, 0.19 mmol), phenylboronic acid (1.5 equiv., 0.03 g, 0.28 mmol), and K_3PO_4 (3 equiv., 0.12 g, 0.56 mmol) according to General procedure I by column chromatography (1:6 EtOAc-hexane) to yield 7 mg white amorphous solid containing 25 and 28 in 2.6:1 ratio. R_f: 0.35 (1:2 EtOAc-hexane). ¹H NMR (400 MHz, CDCl₃) δ 7.32–7.16 (5H, m, aromatics), 4.98 (1H, d, J 6.5 Hz, CH₃OCH₂OC-3), 4.92 (1H, d, J 6.8 Hz, CH₃OCH₂OC-5), 4.83 (1H, d, J 6.8 Hz, CH₃OCH₂OC-4), 4.76 (H, d, J 6.5 Hz, CH₃OCH₂OC-3), 4.70 (1H, d, J 6.8 Hz, CH₃OCH₂OC-4), 4.69 (1H, d, J 7.0 Hz, CH₃OCH₂OC-5), 4.55 (1H, d, J 6.5 Hz, CH₃OCH₂OC-7), 4.50 (1H, d, J 6.5 Hz, CH₃OCH₂OC-7), 4.05 (1H, dd, J_{4,5} 2.0, J_{5,6} 0.6 Hz, H-5), 3.76–3.67 (2H, m, H-3, H-4), 3.67 (1H, dd, J_{6,7a} 6.3, J_{7a,7b} 10.2 Hz, H-7a), 3.58 (1H, dd, J_{6.7b} 6.5 Hz, H-7b), 3.48 (3H, s, CH₃OCH₂OC-3), 3.50–3.44 (1H, m, H-6), 3.43 (3H, s CH₃OCH₂OC-4), 3.42 (3H, s CH₃OCH₂OC-5), 3.42–3.39 (1H, m, H-2), 3.27 (3H, s, CH₃OCH₂OC-7), 3.23 (1H, dd, J_{1a.1b} 14.2, J_{1a.2} 1.5 Hz, H-1_a), 2.77 (1H, dd, J_{1b.2} 10.0 Hz, H-1_b). ¹³C NMR (100 MHz, CDCl₃) δ 139.7–125.1 (aromatics), 98.9 (CH₃OCH₂OC-3), 97.5 (CH₃OCH₂OC-5), 96.9 (CH₃OCH₂OC-7), 95.4 (CH₃OCH₂OC-4), 80.9 (C-2), 80.2 (C-4), 77.5 (C-3), 77.2 (C-6), 72.9 (C-5), 66.7 (C-7), 56.7 (CH₃OCH₂OC-3), 56.1 (CH₃OCH₂OC-5), 56.0 (CH₃OCH₂OC-4), 55.5 (CH₃OCH₂OC-7), 38.1 (C-1). HR-ESI-MS positive mode (m/z): calc. for $[M + Na]^+ = 453.2095$, found: $[M + Na]^+ = 453.2093$; $C_{21}H_{34}O_9$ (430.49).



4.12.2. (*E*)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methoxymethyl-1-Phenyl-D-*gluco*-Hept-1-Enitol (26) and (*Z*)-1,2-Dideoxy-3,4,5,7-Tetra-O-Methoxymethyl-1-Phenyl-D-*gluco*-Hept-1-Enitol (27)

Isolated from a reaction of tosylhydrazone **24** (0.10 g, 0.19 mmol), phenylboronic acid (1.5 equiv., 0.03 g, 0.28 mmol), and K_3PO_4 (3 equiv., 0.12 g, 0.56 mmol) according to General procedure I by column chromatography (1:6 EtOAc–hexane) to yield 19 mg white amorphous solid containing **26** and **27** in 100:1 ratio. R_f : 0.29 (1:2 EtOAc–hexane), $[\alpha]_D + 1$ (*c* 0.30, CH₂Cl₂).

26: ¹H NMR (500 MHz, CDCl₃) δ 7.39 (2H, d, *J* 8.7 Hz, aromatics), 7.35–7.29 (2H, m, aromatics), 7.29–7.23 (1H, m, aromatic), 6.65 (1H, d, *J*_{1,2} 16.0 Hz, H-1), 6.15 (1H, dd, *J*_{2,3} 8.1 Hz, H-2), 4.86 (1H, d, *J* 6.6 Hz, CH₃OCH₂OC-4), 4.84 (2H, d, *J* 6.7 Hz, CH₃OCH₂OC-4, CH₃OCH₂OC-5), 4.78 (1H, d, *J* 6.7 Hz, CH₃OCH₂OC-3), 4.71 (1H, d, *J* 6.8 Hz, CH₃OCH₂OC-5), 4.64 (1H, d, *J* 6.7 Hz, CH₃OCH₂OC-3), 4.62 (1H, d, *J* 6.5 Hz, CH₃OCH₂OC-7), 4.60 (1H, d, *J* 6.5 Hz, CH₃OCH₂OC-7), 4.47 (1H, dd, *J*_{3,4} 5.4 Hz, H-3), 4.20–4.12 (1H, m, H-6), 4.00 (1H, pseudo t, *J*_{4,5} 4.6 Hz, H-4), 3.89 (1H, dd, *J*_{5,6} 2.1 Hz, H-5), 3.66 (1H, dd, *J*_{6,7a} 6.4, *J*_{7a,7b} 10.3 Hz, H-7a), 3.64 (1H, dd, *J*_{6,7b} 6.1 Hz, H-7b), 3.49 (1H, dd, *J*_{6,OH} 3.9 Hz, OH), 3.46 (3H, s CH₃OCH₂OC-7). ¹³C NMR (125 MHz, CDCl₃) δ 134.6 (C-1), 136.4–125.5 (aromatics), 125.9 (C-2), 98.5 (CH₃OCH₂OC-4), 97.5 (CH₃OCH₂OC-5), 96.9 (CH₃OCH₂OC-7), 94.3 (CH₃OCH₂OC-3), 81.3 (C-4), 77.2 (C-3), 76.9 (C-5), 69.8 (C-6), 69.1 (C-7), 56.4 (CH₃OCH₂OC-4, CH₃OCH₂OC-5), 56.0 (CH₃OCH₂OC-3), 55.4 (CH₃OCH₂OC-7). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 453.2095, found: [M + Na]⁺ = 453.2099; C₂₁H₃₄O₉ (430.49).

27: ¹H NMR (500 MHz, CDCl₃) δ 7.43–7.36 (2H, m, aromatics), 7.35–7.29 (2H, m, aromatics), 7.29–7.23 (1H, m, aromatic), 6.75 (1H, d, $J_{1,2}$ 11.4 Hz, H-1), 5.70 (1H, dd, $J_{2,3}$ 9.9 Hz, H-2), 4.93–4.22 (11H, m, H-3, H-4, H-5, 4 × CH₃OCH₂), 4.20–4.12 (1H, m, H-6), 3.96–3.83 (2H, m, H-7_a, H-7_b), 3.44, 3.35, 3.34 (12H, 4s, 4 × CH₃OCH₂). ¹³C NMR (125 MHz, CDCl₃) δ 133.9 (C-1), 136.4–125.5 (aromatics), 129.2 (C-2), 98.9, 97.6, 97.0, 94.6 (4 × CH₃OCH₂), 81.6 (C-4), 76.9 (C-5), 71.7 (C-3), 69.5 (C-6), 65.7 (C-7), 56.6, 56.5, 55.9, 55.7 (4 × CH₃OCH₂). HR-ESI-MS positive mode (m/z): calc. for [M + Na]⁺ = 453.2095, found: [M + Na]⁺ = 453.2099; C₂₁H₃₄O₉ (430.49).

Supplementary Materials: The following are available online at https://www.mdpi.com/article/ 10.3390/molecules27061795/s1, The NMR spectral analysis.

Author Contributions: M.T., T.K. designed the experiments; M.T., T.K., B.B., B.Á.B., T.B. performed the synthetic work, M.T., T.K., L.J. carried out the structure elucidation, L.S., M.T. conceived the research, M.T., T.K., L.S., L.J. wrote the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Research, Development and Innovation Office, grant number FK128766, and by the EU co-financed by the European Regional Development Fund under the project GINOP-2.3.2-15-2016-00008.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples are not available.

References

- 1. Barluenga, J.; Valdes, C. Tosylhydrazones: New uses for classic reagents in palladium-catalyzed cross-coupling and metal-free reactions. *Angew. Chem. Int. Ed.* **2011**, *50*, 7486–7500. [CrossRef]
- Shao, Z.; Zhang, H. N-Tosylhydrazones: Versatile reagents for metal-catalyzed and metal-free cross-coupling reactions. *Chem. Soc. Rev.* 2012, 41, 560–572. [CrossRef]
- 3. Xiao, Q.; Zhang, Y.; Wang, J.B. Diazo Compounds and *N*-Tosylhydrazones: Novel cross-coupling partners in transition-metalcatalyzed reactions. *Acc. Chem. Res.* 2013, *46*, 236–247. [CrossRef]

- Qiu, D.; Mo, F.; Zhang, Y.; Wang, J. Chapter Two—Recent advances in transition-metal-catalyzed cross-coupling reactions with N-tosylhydrazones. In Advances in Organometallic Chemistry; Pérez, P.J., Ed.; Academic Press: Cambridge, MA, USA, 2017; Volume 67, pp. 151–219.
- 5. Xia, Y.; Qiu, D.; Wang, J. Transition-metal-catalyzed cross-couplings through carbene migratory insertion. *Chem. Rev.* 2017, 117, 13810–13889. [CrossRef]
- 6. Wang, H.; Deng, Y.-H.; Shao, Z. An update of *N*-tosylhydrazones: Versatile reagents for metal-catalyzed and metal-free coupling reactions. *Synthesis* **2018**, *50*, 2281–2306. [CrossRef]
- Tóth, M.; Somsák, L. One-pot transformation of nitriles into aldehyde tosylhydrazones. *Tetrahedron Lett.* 2001, 42, 2723–2725. [CrossRef]
- 8. Tóth, M.; Kövér, K.E.; Bényei, A.; Somsák, L. C-Glycosylmethylene carbenes: Synthesis of anhydro-aldose tosylhydrazones as precursors; generation and a new synthetic route to *exo*-glycals. *Org. Biomol. Chem.* **2003**, *1*, 4039–4046. [CrossRef]
- Tóth, M.; Somsák, L.; Goyard, D. Preparation of 2,6-anhydro-aldose-tosylhydrazones. In *Carbohydrate Chemistry: Proven Synthetic Methods*; Kováč, P., Ed.; CRC Press: Boca Raton, FL, USA, 2012; Volume 1, pp. 355–365.
- Kaszás, T.; Tóth, M.; Kun, S.; Somsák, L. Coupling of anhydro-aldose tosylhydrazones with hydroxy compounds and carboxylic acids: A new route for the synthesis of C-[β]-D-glycopyranosylmethyl ethers and esters. RSC Adv. 2017, 7, 10454–10462. [CrossRef]
- 11. Kaszás, T.; Tóth, M.; Somsák, L. A new synthesis of C-[β]-D-glycopyranosylmethyl sulfides by metal-free coupling of anhydroaldose tosylhydrazones with thiols. *New J. Chem.* **2017**, *41*, 13871–13880. [CrossRef]
- Kaszás, T.; Cservenyák, I.; Juhász-Tóth, É.; Kulcsár, A.E.; Granatino, P.; Nilsson, U.J.; Somsák, L.; Tóth, M. Coupling of *N*-tosylhydrazones with tetrazoles: Synthesis of 2-β-D-glycopyranosylmethyl-5-substituted-2*H*-tetrazole type glycomimetics. *Org. Biomol. Chem.* 2021, *19*, 605–618. [CrossRef]
- 13. Kaszás, T.; Ivanov, A.; Tóth, M.; Ehlers, P.; Langer, P.; Somsák, L. Pd-catalyzed coupling reactions of anhydro-aldose tosylhydrazones with aryl bromides to produce substituted *exo*-glycals. *Carbohydr. Res.* **2018**, *466*, 30–38. [CrossRef] [PubMed]
- Kaszás, T.; Tóth, M.; Langer, P.; Somsák, L. C-Glycosyl styrene type compounds by Pd-catalyzed cross-coupling reactions of anhydro-aldose tosylhydrazones with benzyl bromides. *Adv. Synth. Catal.* 2019, 361, 105–117. [CrossRef]
- 15. Barluenga, J.; Tomas-Gamasa, M.; Aznar, F.; Valdes, C. Metal-free carbon-carbon bond-forming reductive coupling between boronic acids and tosylhydrazones. *Nat. Chem.* **2009**, *1*, 494–499. [CrossRef] [PubMed]
- 16. Paraja, M.; Plaza, M.; Valdés, C. Transition-metal-free reactions between boronic acids and *N*-sulfonylhydrazones or diazo compounds: Reductive coupling processes and beyond. *Synlett* **2017**, *28*, 2373–2389. [CrossRef]
- 17. Pérez-Aguilar, M.C.; Valdés, C. Olefination of carbonyl compounds through reductive coupling of alkenylboronic acids and tosylhydrazones. *Angew. Chem. Int. Ed.* **2012**, *51*, 5953–5957. [CrossRef] [PubMed]
- Merchant, R.R.; Lopez, J.A. A general C(sp³)–C(sp³) cross-coupling of benzyl sulfonylhydrazones with alkyl boronic acids. *Org. Lett.* 2020, 22, 2271–2275. [CrossRef] [PubMed]
- 19. He, J.; Zhang, J.; Dai, B.; Liu, P. Sequentially Formations of Csp³-Csp² and Csp²-Csp² bonds by a one-pot reaction involving *N*-tosylhydrazone and *p*-bromobenzeneboronic acid. *ChemistrySelect* **2019**, *4*, 4496–4498. [CrossRef]
- 20. Gu, N.; Wei, Y.; Liu, P.; Liu, Y.; Dai, B. Multi-component one-pot reaction of aromatic carbonyl compounds, tosylhydrazide, and arylboronic acids. *Molecules* **2017**, *22*, 2168. [CrossRef]
- Liu, Y.; Ma, X.; Liu, Y.; Liu, P.; Dai, B. Synthesis of α-arylalkylferrocenes through cesium fluoride-promoted coupling of arylboronic acids with N-tosylhydrazones. Synth. Comm. 2018, 48, 921–928. [CrossRef]
- 22. Plaza, M.; Pérez-Aguilar, M.C.; Valdés, C. Stereoselective Csp³–Csp² bond-forming reactions by transition-metal-free reductive coupling of cyclic tosylhydrazones with boronic acids. *Chem. Eur. J.* **2016**, *22*, 6253–6257. [CrossRef]
- 23. Li, X.; Feng, Y.; Lin, L.; Zou, G. Synthesis of diarylmethanes via metal-free reductive cross-coupling of diarylborinic acids with tosyl hydrazones. *J. Org. Chem.* **2012**, *77*, 10991–10995. [CrossRef] [PubMed]
- Kupracz, L.; Kirschning, A. Two-step flow synthesis of biarylmethanes by reductive arylation of tosylhydrazones. J. Flow Chem. 2013, 3, 11–16. [CrossRef]
- 25. Nakagawa, S.; Bainbridge, K.A.; Butcher, K.; Ellis, D.; Klute, W.; Ryckmans, T. Application of Barluenga boronic coupling (BBC) to the parallel synthesis of drug-like and drug fragment-like molecules. *ChemMedChem* **2012**, *7*, 233–236. [CrossRef] [PubMed]
- Khan, I.; Sharma, A.; Kamboj, P.; Maity, B.; Tyagi, V. Base-mediated reductive coupling of indole-3-tosylhydrazone with thiols/boronic acids: Facile synthesis of 3-(phenylthio)methyl/benzyl indole derivatives. *ChemistrySelect* 2020, 5, 591–600. [CrossRef]
- Allwood, D.M.; Blakemore, D.C.; Brown, A.D.; Ley, S.V. Metal-free coupling of saturated heterocyclic sulfonylhydrazones with boronic acids. J. Org. Chem. 2014, 79, 328–338. [CrossRef]
- 28. Shen, X.; Gu, N.; Liu, P.; Ma, X.; Xie, J.; Liu, Y.; He, L.; Dai, B. A simple and efficient synthesis of 9-arylfluorenes via metal-free reductive coupling of arylboronic acids and *N*-tosylhydrazones in situ. *RSC Adv.* **2015**, *5*, 63726–63731. [CrossRef]
- 29. Shen, X.; Gu, N.; Liu, P.; Ma, X.; Xie, J.; Liu, Y.; Dai, B. One-pot synthesis of triarylmethanes via metal-free reductive coupling of diaryl ketones, tosylhydrazide, and arylboronic acids. *Chin. J. Chem.* **2016**, *34*, 1033–1038. [CrossRef]
- Shen, X.; Liu, P.; Liu, Y.; Liu, Y.; Dai, B. One-pot reductive coupling reactions of acetyl naphthalene derivatives, tosylhydrazide, with arylboronic acids. *Tetrahedron* 2017, 73, 785–793. [CrossRef]
- Wu, G.; Deng, Y.; Luo, H.; Zhou, J.; Li, T.; Zhang, Y.; Wang, J. Transition-metal-free cascade reaction of α-halo-N-tosylhydrazones, indoles and arylboronic acids. *Chem. Commun.* 2016, 52, 5266–5268. [CrossRef]

- 32. Plaza, M.; Valdés, C. Stereoselective domino carbocyclizations of γ and δ -cyano-*N*-tosylhydrazones with alkenylboronic acids with formation of two different C(sp³)–C(sp²) bonds on a quaternary stereocenter. *J. Am. Chem. Soc.* **2016**, *138*, 12061–12064. [CrossRef]
- Plaza, M.; Parisotto, S.; Valdés, C. Heterocyclization and spirocyclization processes based on domino reactions of *N*-tosylhydrazones and boronic acids involving intramolecular allylborylations of nitriles. *Chem. Eur. J.* 2018, 24, 14836–14843. [CrossRef] [PubMed]
- 34. Florentino, L.; López, L.; Barroso, R.; Cabal, M.-P.; Valdés, C. Synthesis of pyrrolidines by a Csp³-Csp³/Csp³-N transition-metalfree domino reaction of boronic acids with γ-azido-*N*-tosylhydrazones. *Angew. Chem. Int. Ed.* **2021**, *60*, 1273–1280. [CrossRef] [PubMed]
- Panigot, J.; Curley, W. Reaction of glycosyl halides with benzyl Grignard reagents: Unexpected *o*-tolyl alkylation of tetra-Oacetylglucopyranosyl bromide and direct synthesis of (β-glycosyl)phenylmethanes. *J. Carbohydr. Chem.* 1994, 13, 293–302. [CrossRef]
- Sanhueza, C.A.; Mayato, C.; García-Chicano, M.; Díaz-Peñate, R.; Dorta, R.L.; Vázquez, J.T. Antiproliferation and apoptosis induced by C-glycosides in human leukemia cancer cells. *Bioorg. Med. Chem. Lett.* 2006, 16, 4223–4227. [CrossRef]
- Kolympadi, M.; Fontanella, M.; Venturi, C.; André, S.; Gabius, H.-J.; Jiménez-Barbero, J.; Vogel, P. Synthesis and conformational analysis of (α-D-galactosyl)phenylmethane and α-,β-difluoromethane analogues: Interactions with the plant lectin viscumin. *Chem. Eur. J.* 2009, 15, 2861–2873. [CrossRef]
- Pasetto, P.; Chen, X.; Drain, C.M.; Franck, R.W. Synthesis of hydrolytically stable porphyrin C- and S-glycoconjugates in high yields. *Chem. Commun.* 2001, 81–82. [CrossRef]
- Ichikawa, S.; Tatebayashi, N.; Matsuda, A. Synthesis of C-glycosyl pyrrolo[3,4-C]carbazole-1,3(2H,6H)-diones as a scaffold for check point kinase 1 inhibitors. J. Org. Chem. 2013, 78, 12065–12075. [CrossRef]
- 40. Brenna, E.; Fuganti, C.; Grasselli, P.; Serra, S.; Zamboti, S. A novel general route for the synthesis of C-glycosyl tyrosine analogues. *Chemistry* **2002**, *8*, 1872–1878. [CrossRef]
- Fatima, S.; Pandey, V.P.; Bisht, S.S.; Tripathi, R.P. Application of butenonyl-C-glucosides in the synthesis of pyrazolinyl-, aminopyrimidinyl- and biphenyl methyl-β-D-C-glucopyranosides. *Mol. Div.* 2011, 15, 759–768. [CrossRef]
- 42. Johnson, C.R.; Johns, B.A. Suzuki cross-coupling of carbohydrates: Synthesis of β-arylmethyl-C-glycosides and aryl-scaffolded trisaccharide mimics. *Synlett* **1997**, 1406–1408. [CrossRef]
- 43. Pearce, A.J.; Ramaya, S.; Thorn, S.N.; Bloomberg, G.B.; Walter, D.S.; Gallagher, T. C-glycosyl tyrosines. Synthesis and incorporation into C-glycopeptides. J. Org. Chem. 1999, 64, 5453–5462. [CrossRef] [PubMed]
- 44. Tóth, M.; Somsák, L. *Exo*-glycals from glycosyl cyanides. First generation of C-glycosylmethylene carbenes from 2,5- and 2,6-anhydroaldose tosylhydrazones. *J. Chem. Soc. Perkin. Trans.* 1 2001, 942–943. [CrossRef]
- 45. Tóth, M.; Kun, S.; Somsák, L.; Goyard, D. Preparation of *exo*-glycals from 2,6-anhydro-aldose-tosylhydrazones. In *Carbohydrate Chemistry: Proven Synthetic Methods*; Kováč, P., Ed.; CRC Press: Boca Raton, FL, USA, 2012; Volume 1, pp. 367–375.
- Kim, I.S.; Kim, S.J.; Lee, J.K.; Li, Q.R.; Jung, Y.H. Synthesis of (2R,5S)-dihydroxymethyl-(3R,4R)-dihydroxypyrrolidine (DGDP) via stereoselective amination using chlorosulfonyl isocyanate. *Carbohydr. Res.* 2007, 342, 1502–1509. [CrossRef] [PubMed]
- 47. Schlubach, H.H.; Huntenburg, W. Zwei neue pentabenzoyl-glucosen. Ber. Dtsch. Chem. Ges. 1927, 60, 1487–1488. [CrossRef]
- Pourceau, G.; Valle-Carrandi, L.d.; Di Gianvincenzo, P.; Michelena, O.; Penadés, S. On the chiroptical properties of Au(i)-thiolate glycoconjugate precursors and their influence on sugar-protected gold nanoparticles (glyconanoparticles). *RSC Adv.* 2014, 4, 59284–59288. [CrossRef]
- 49. Tamura, Y.; Kanomata, K.; Kitaoka, T. Interfacial hydrolysis of acetals on protonated TEMPO-oxidized cellulose nanofibers. *Sci. Rep.* **2018**, *8*, 5021. [CrossRef]
- Jensen, H.H.; Nordstrøm, L.U.; Bols, M. The disarming effect of the 4,6-acetal group on glycoside reactivity: Torsional or electronic? J. Am. Chem. Soc. 2004, 126, 9205–9213. [CrossRef]
- Sipos, S.; Jablonkai, I. Preparation of 1-C-glycosyl aldehydes by reductive hydrolysis. *Carbohydr. Res.* 2011, 346, 1503–1510. [CrossRef]
- 52. Ali, M.H.; Collins, P.M.; Overend, G. Titanium-mediated methylene transfer reactions on sugar esters, lactones, and uloses. *Carbohydr. Res.* **1990**, 205, 428–434. [CrossRef]
- 53. Walker, J.R.; Alshafie, G.; Nieves, N.; Ahrens, J.; Clagett-Dame, M.; Abou-Issa, H.; Curley, R.W. Synthesis and preliminary chemotherapeutic evaluation of the fully *C*-linked glucuronide of *N*-(4-hydroxyphenyl)retinamide. *Bioorg. Med. Chem.* **2006**, *14*, 3038–3048. [CrossRef]