JACC: BASIC TO TRANSLATIONAL SCIENCE © 2020 THE AUTHOR. PUBLISHED BY ELSEVIER ON BEHALF OF THE AMERICAN COLLEGE OF CARDIOLOGY FOUNDATION. THIS IS AN OPEN ACCESS ARTICLE UNDER THE CC BY-NC-ND LICENSE (http://creativecommons.org/licenses/by-nc-nd/4.0/).

STATE-OF-THE-ART REVIEW

Mutual Antagonism of Hypoxia-Inducible Factor Isoforms in Cardiac, Vascular, and Renal Disorders

Milton Packer, MD^{a,b}

HIGHLIGHTS

- HIF-1a and HIF-2a promote cellular adaptation to acute hypoxia, but during prolonged activation, these isoforms exert mutually antagonistic effects on the redox state and on proinflammatory pathways.
- Imbalances in HIF-1α and HIF-2α may contribute to the evolution and progression of chronic cardiac, vascular, and renal disorders.
- Selective activation of HIF-2α can be achieved with drugs that inhibit isoform-selective PHDs or that promote the redox sensor, SIRT-1 (e.g., SGLT2 inhibitors).

SUMMARY

Hypoxia-inducible factor (HIF)-1 α and HIF-2 α promote cellular adaptation to acute hypoxia, but during prolonged activation, these isoforms exert mutually antagonistic effects on the redox state and on proinflammatory pathways. Sustained HIF-1 α signaling can increase oxidative stress, inflammation, and fibrosis, actions that are opposed by HIF-2 α . Imbalances in the interplay between HIF-1 α and HIF-2 α may contribute to the progression of chronic heart failure, atherosclerotic and hypertensive vascular disorders, and chronic kidney disease. These disorders are characterized by activation of HIF-1 α and suppression of HIF-2 α , which are potentially related to mitochondrial and peroxisomal dysfunction and suppression of the redox sensor, sirtuin-1. Hypoxia mimetics can potentiate HIF-1 α and/or HIF-2 α ; ideally, such agents should act preferentially to promote HIF-2 α while exerting little effect on or acting to suppress HIF-1 α . Selective activation of HIF-2 α signaling through sirtuin-1 activation may explain the effect of sodium-glucose cotransporter 2 inhibitors). Selective HIF-2 α signaling through sirtuin-1 activation may explain the effect of sodium-glucose cotransporter 2 inhibitors to simultaneously promote erythrocytosis and ameliorate the development of cardiomyopathy and nephropathy. (J Am Coll Cardiol Basic Trans Science 2020;5:961-8) © 2020 The Author. Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.O/).

Manuscript received March 30, 2020; revised manuscript received May 7, 2020, accepted May 7, 2020.

From the ^aBaylor Heart and Vascular Institute, Baylor University Medical Center, Dallas, Texas; and the ^bImperial College, London, United Kingdom. Dr. Packer has consulted for Abbvie, Actavis, Akcea, Amarin, Amgen, AstraZeneca, Boehringer Ingelheim, Cardiorentis, Daiichi Sankyo, Johnson & Johnson, Novartis, NovoNordisk, Pfizer, Relypsa, Sanofi, Synthetic Biologics, and Theravance.

The author attests they are in compliance with human studies committees and animal welfare regulations of the authors' institutions and Food and Drug Administration guidelines, including patient consent where appropriate. For more information, visit the *JACC: Basic to Translational Science* author instructions page.

ABBREVIATIONS AND ACRONYMS

HIF = hypoxia inducible factor PHD = prolyl hydroxylase SGLT2 = sodium-glucose cotransporter 2

SIRT1 = sirtuin-1

tor there a substitution the set of the se

nisms of regulation and spatial expression patterns and differ meaningfully in their cellular actions (1,2).

Despite an identical DNA consensus recognition sequence, DNA binding does not correspond to increased transcriptional activity, and each isoform loads at a distinct repertoire of cell type-specific sites across the genome (3). HIF-1 α is a universal master regulator for hypoxia-inducible gene expression and is expressed in a wide range of cell types (2,4). In contrast, HIF-2a is expressed selectively, that is, primarily in alveolar epithelial cells in the lung, specialized peritubular interstitial cells in the kidney, in hepatic parenchymal cells, and in endothelial cells in the heart (4-7). Furthermore, the expression of HIF-1 α is highly sensitive to environmental oxygen, and inactivation of HIF-1a completely abolishes induction of HIF target genes (2). In contrast, the expression of HIF-2a is less influenced by acute changes in oxygen tension (4-6,8) but its activity is upregulated by cellular stress and hypoxia mimetics. Although both HIF-1a and HIF-2a can modulate the transcription of the same genes, the 2 isoforms often do so in an opposing manner (9,10). Thus, the ability of HIF-1 α and HIF-2 α to activate specific target genes is highly context-dependent, particularly with respect to the inciting stimulus and cell type (2).

The activities of HIF-1 α and HIF-2 α are upregulated both by hypoxia and by drugs that mimic hypoxia under normoxic conditions (e.g., cobalt chloride) (4,6,11). Oxygen influences the activity of these isoforms by directly modulating the activation of a family of 3 prolyl hydroxylases (PHD1, PHD2, and PHD3) that function to degrade the 2 transcription factors; the PHDs are sensitive to environmental oxygen, and thus, act as oxygen sensors (12-14). Inhibition of these PDHs by hypoxia leads to stabilization of one or both HIFs, which leads to enhanced HIF signaling. However, PHD1, PHD2, and PHD3 differ meaningfully with respect to their expression profiles, subcellular localization, and their effects on HIF-1 α or HIF-2 α (15,16).

Therefore, despite the structural homology of HIF-1 α and HIF-2 α , differences in cellular and

subcellular expression patterns, upstream regulators, and sensitivity to degradative enzymes explain why HIF-1a and HIF-2a exert highly distinctive effects under a range of specific conditions. In general, HIF-1 α promotes the transcription of proteins that decrease oxygen use and increase angiogenesis, whereas HIF-2a is the primary stimulus for erythropoietin synthesis (17,18). In addition, under conditions of prolonged cellular stress, HIF-1a and HIF-2a exert diametrically opposing actions on critical pathways that determine the balance between health and disease, especially in the development of chronic cardiac, vascular, and renal disorders (9,10). As a result, switching from the expression of one isoform to another appears to play a role in the pathogenesis of many diseases and may mediate the effects of treatment (19).

OXYGEN-RELATED ACTIVATION OF HIF-1α AND HIF-2α AND THE INTERPLAY OF THESE ISOFORMS TO MODULATING THE CELLULAR REDOX STATE AND THE INFLAMMATORY SET POINT

During commonly encountered physiological states, a reduction in environmental oxygen enhances signaling through both HIF-1a and HIF-2a. However, under the pathological conditions that prevail in many chronic cardiac, renal, and vascular disorders, HIF signaling is also stimulated by abnormalities in mitochondria and peroxisomes, the most important oxygen-consuming organelles in cells. Derangements in these cellular constituents redirects the use of oxygen toward the generation of reactive oxygen species, which (like hypoxia) has a direct effect to inhibit PDHs (20). The activation of HIFs by oxidative stress is dependent on the presence of mitochondria that are capable of consuming oxygen and generating reactive oxygen species (20,21). Therefore, the organellar dysfunction that characterizes many chronic diseases appears to play a critical role in enhancing HIF signaling.

Once activated by oxygen-related organellar stresses, HIF-1 α and HIF-2 α act to mute these stresses by reducing the amount of oxygen consumed by mitochondria and peroxisomes, both directly and indirectly. HIF-1 α directly inhibits both the biogenesis and oxidative functions of mitochondria (22). In addition, both HIF-1 α and HIF-2 α promote autophagy, a lysosome-dependent degradative pathway that mediates the clearance of dysfunctional organelles. HIF-1 α enhances the autophagic clearance of damaged mitochondria (mitophagy) (23), whereas HIF-2 α stimulates the autophagic disposal of injured peroxisomes (pexography) (24). Therefore, once

activated by oxygen-related stress, HIF-1 α and HIF-2 α signaling can act to reduce this stress, particularly that caused by the organellar dysfunction characteristic of chronic heart and renal disease.

MUTUAL ANTAGONISM BETWEEN HIF-1α AND HIF-2α IN THE REGULATION OF REDOX STATE AND IN THE MODULATION OF PROINFLAMMATORY **PROFIBROTIC PATHWAYS.** Although the actions of HIF-1 α and HIF-2 α can be concordant, these isoforms exert mutually antagonistic effects on many aspects of cellular homeostasis (Figure 1). In particular, the interplay of HIF-1 α and HIF-2 α helps to determine the set point for the redox state of cells. HIF-2a hypomorphic mice showed increased HIF-1 α and an oxidized intracellular redox state, which was accompanied by exaggerated hypoxic sensitivity and was reversed by a HIF-1a inhibitor or a superoxide scavenger (25). Conversely, HIF-1a hypomorphic mice demonstrated increased levels of HIF-2a and a reduced intracellular redox state, which was accompanied by blunted oxygen sensing and was corrected by a HIF-2 α inhibitor (25). HIF-2 α may reduce oxidative stress not only by promoting pexophagy but also by transactivating genes that encode for antioxidant enzymes (26). Disorders characterized regional and transient derangements in oxygen delivery can cause striking imbalances between HIF-1a and HIF-2a signaling; for example, chronic intermittent hypoxia activates HIF-1 α (27) but downregulates HIF-2 α (28).

HIF-1 α and HIF-2 α also have mutual antagonistic effects on inflammation and fibrosis in diverse organs, including the heart, kidney, vasculature, and adipose tissue (29-31). HIF-1 α promotes the activation of inducible nitric oxide synthase (a proinflammatory mediator) and enhances M1 macrophage polarization (a proinflammatory phenotype); both effects are opposed by the actions of HIF-2 α (31). HIF-1 α promotes (whereas HIF-2 α inhibits) the actions of proinflammatory cytokines (32,33). The genes that encode for profibrotic chemokines and collagen deposition are activated by HIF-1 α (34), whereas HIF-2 α promotes collagen matrix degradation (35). Therefore, the balance between HIF-1 α and HIF-2 α may determine the set point for inflammation and fibrosis.

What influences the point of equilibrium between HIF-1 α and HIF-2 α ? An important modulator of both isoforms is sirtuin-1 (SIRT1) (19), a nicotinamide adenine dinucleotide responsive deacetylase that serves as both as a redox rheostat and a nutrient and/or oxygen sensor. SIRT1 activation leads to downregulation of HIF-1 α (36) but upregulation of HIF-2 α (37); the latter effect is likely mediated by SIRT1-mediated deacetylation of HIF-2 α . In many



chronic disorders (e.g., obesity, diabetes, chronic heart failure, and chronic kidney disease), the activity of SIRT1 is suppressed (38,39), thus shifting the balance toward upregulation of HIF-1 α and downregulation of HIF-2 α . Conversely, activation of SIRT1 activation favors HIF-2 α over HIF-1 α , and SIRT1 has direct effects to preserve the integrity of mitochondria and peroxisomes similar to the actions of HIF-2 α (40). For these reasons, SIRT1 is poised to act as a HIF-1 α to HIF-2 α switch (19).

MODULATION OF HIF-1 α /HIF-2 α SIGNALING IN THE ISCHEMIC AND FAILING HEART

Following an abrupt decline in oxygen tension, activation of HIF-1 α and HIF-2 α promotes the adaptation of the myocardium to hypoxia; HIF upregulation ameliorates ischemia–reperfusion injury, whereas HIF downregulation exacerbates hypoxic dysfunction (41). Suppression of HIF-1 α causes inadequate vascularization, and thus, decompensation of acute pressure overload or ischemic states (42).

However, enhanced HIF-1 α signaling may not be beneficial in chronic heart failure, which is characterized by oxidative stress and SIRT1 suppression (Figure 2) (38). The action of HIF-1 α to shift glucose metabolism from oxidative to glycolytic pathways limits cardiac performance because glycolysis is an inefficient mechanism of generating adenosine triphosphate; this shift also promotes lipid



accumulation in cardiomyocytes, which leads to contractile dysfunction (43). Moreover, HIF-1a impairs mitochondrial biogenesis, further limiting the generation of adenosine triphosphate (22). HIF-1 α can also promote hypertrophy and inflammation in cardiomyocytes (44-46) and may activate the sympathetic nervous system in chronic heart failure (47). For all of these reasons, marked, sustained, and isolated activation of HIF-1a acts to impair cardiac performance. Stabilization of HIF-1α at high levels leads to worsening myocardial function (48), and genetic overexpression of HIF-1a results in cardiomyopathy (49). Conversely, inhibition of HIF-1a slows the transition from cardiac hypertrophy to dilated cardiomyopathy (50). Therefore, it is interesting that expression of HIF-1 α is increased in the failing heart, both experimentally and clinically (48,51). High circulating levels of HIF-1 α are associated with a poor prognosis in patients with chronic heart failure (52), and drugs that are effective in treating heart failure can diminish the activation of HIF-1 α (53).

In contradistinction to HIF-1 α , activation of HIF-2 α exerts protective effects in cardiomyocytes (54), and decreased HIF-2 α expression is accompanied by inflammasome activation in the heart (55). Upregulation of HIF-2 α occurs in myocardial regions most susceptible to ventricular remodeling, perhaps as an adaptive response (56). Despite these observations, little is known about the role of HIF-2 α in chronic heart failure.

MODULATION OF HIF-1α/HIF-2α SIGNALING IN VASCULAR DISORDERS

Hypertension and atherosclerosis involve increased sheer stress as well as hypoxia and inflammation in the arterial wall. Increased intravascular pressures and the medial infiltration by inflammatory cells promotes the activation of HIF-1a (Figure 2) (57). Upregulation of HIF-1 α in macrophages, vascular smooth muscle, and endothelial cells has been implicated in the pathogenesis of neointimal proliferation, medial hypertrophy, and activation of proinflammatory pathways, and thus, the progression of atherosclerosis (58-60). Genetic silencing of HIF-1a in macrophages and vascular smooth muscle cells attenuates vascular inflammation, mitigating the deleterious actions of oxidized lipoproteins and the formation of foam cells, as well as slowing development of atherosclerotic plaque (58,61). HIF-1α hypomorphic mice showed decreased production of proinflammatory cytokines and a shift in the polarization of macrophages from the M1 to the M2 phenotype, leading to amelioration of vascular remodeling (62). The proinflammatory and mitogenic effects of inducible nitric oxide synthase may be mediated by HIF-1a (63). Vascular wall hypoxia can promote arterial thrombus formation through activation of HIF-1 α (64).

In contrast, upregulation of HIF-2 α suppresses the development of neointimal proliferation and atherosclerosis (65); however, oxidized lipoproteins abolish the hypoxic induction of HIF-2 α (66), raising the possibility that atherosclerosis represents a HIF-2 α -deficient state. Activation of HIF-2 α in the endothelium and media of coronary and renal arteries reduces the magnitude of tissue injury following experimental ischemia (67,68).

MODULATION OF HIF-1 α **/HIF-2** α **SIGNALING IN THE DIABETIC AND NONDIABETIC KIDNEY**. During acute renal ischemia or hypoxia, the activation of HIF-1 α mitigates renal injury, potentially by promoting angiogenesis and tissue repair (69,70). Silencing of HIF-1 α during low-oxygen tension states exacerbates renal tubular epithelial cell necrosis, and potentiation of HIF-1 α prevents this injurious effect (71). Upregulation of endothelial HIF-2 α can exert a similar renoprotective action during acute ischemia (68).

However, prolonged HIF-1 α activation exerts deleterious effects on the kidney, even if the causal

mechanism is hypoxia or ischemia (**Figure 2**) (72). Sustained upregulation of HIF-1 α promotes epithelial to mesenchymal transition in renal tubular epithelial cells (73). In chronic kidney disease, sustained signaling through HIF-1 α promotes proinflammatory and profibrotic pathways in the glomerulus and renal tubules (72,74,75), which is characterized by activation of inflammation-related cytokines, profibrotic gene transcription, macrophage infiltration and collagen deposition, mesangial cell proliferation, and tubulo-interstitial inflammation (74–77). Experimental suppression of HIF-1 α attenuates mesangial matrix expansion and glomerulosclerosis and alleviates tubulointerstitial fibrosis (75,76,78).

Although HIF-1a promotes inflammation, activation of HIF-2a mutes inflammation and reduces injury in renal tissues (68,79). Chronic kidney disease is characterized by upregulation of HIF-1a but downregulation of HIF-2 α (30,79,80), and the deficiency in HIF-2a likely explains why chronic kidney disease is accompanied by blunted production of erythropoietin and anemia (81), and why impaired erythropoietin synthesis is accompanied by increased inflammatory and angiogenic markers (82). The interplay between HIF-1α and HIF-2α may be particularly important in diabetes (78), because hyperglycemia and advanced glycation end products directly promote the transcription of HIF-1a in glomerular and renal tubular cells (83,84). In addition, SIRT1 signaling (along with its ability to inhibit HIF-1 α and activate HIF-2 α) is impaired in the diabetic kidney (38,39).

EFFECT OF HYPOXIA MIMETICS (PHD INHIBITORS AND SIRT1 ACTIVATORS) ON CHRONIC CARDIAC, RENAL, AND VASCULAR INJURY

Hypoxia mimetics enhance the activity of HIF-1a and/or HIF-2 α , most commonly by inhibiting 1 or some of the 3 PHDs that are responsible for degradation of the isoforms. PHDs differ with respect to their expression profiles, subcellular localization, and effects on HIF-1a or HIF-2a. Inhibition of PHD2 acts primarily to boost the activity of HIF-1 α (85), whereas PHD3 acts preferentially on HIF-2 α (16,86,87). Hypoxia mimetics that increase the production of erythropoietin and that can treat anemia (e.g., cobalt chloride and roxadustat) inhibit PHD3 (3,85,88), although they may also suppress other PHDs (depending on dose). Drugs that selectively inhibit PHD1, PHD2, or PHD3 have been developed (Central Illustration), but studies of PHD inhibition have not often assessed the degree of selectivity. For example, most reports of the effects of



CENTRAL ILLUSTRATION Mechanisms of Selective HIF-2α Activation by Drugs That Enhance Sirtuin-1 Signaling or Exert Preferential Inhibitory Effects on Certain Prolyl Hydroxlyases

cobalt chloride in experimental organ injury have been difficult to interpret, because they often failed to measure the effect of the drug on both HIF isoforms.

Selective changes in the relative activities of HIF-1 α and HIF-2 α can also be achieved by modulating the activity of SIRT(1,19) because SIRT1 upregulation enhances signaling through HIF-2a, but suppresses the activity of HIF-1a (Central Illustration) (36,37). Resveratrol, a SIRT1 activator, inhibits HIF-1α in diverse tissues (89,90). Suppression of HIF-1 α in the kidney by resveratrol diminishes inflammation and fibrosis in glomerular mesangial cells and ameliorates tubulointerstitial injury (90,91). Furthermore, the action of resveratrol to downregulate HIF-1α in cardiomyocytes reduces hypoxic injury (92). Resveratrol also increases erythropoietin (93), which is presumably related to an action on HIF-2a, although the effect of resveratrol on HIF-2a signaling has been directly studied.

Similarly, sodium-glucose cotransporter 2 (SGLT2) inhibitors that are used to treat type 2 diabetes have important cardioprotective and reno-protective effects, both experimentally and clinically (38,39). These drugs appear to activate SIRT1 and its downstream effectors by virtue of their ability to induce a fasting-like transcriptional paradigm (94-97); the

effects of SGLT2 inhibitors on SIRT1 signaling have been proposed to mediate their favorable effects on the heart and kidneys (38,39). SIRT1 upregulation may explain the effect of SGLT2 inhibitors to suppress HIF-1 α in the kidney (98,99), as well as their effect to enhance the production of erythropoietin and promote erythrocytosis (100) (which is potentially related to SIRT1-mediated activation of HIF-2 α 37). The importance of this latter effect is reinforced by the results of statistical mediation analyses, which showed that changes in hemoglobin produced by SGLT2 inhibitors are a powerful predictor of their ability to reduce the risk of cardiovascular death and hospitalizations for heart failure (101,102).

CONCLUSIONS

HIF-1 α and HIF-2 α exert mutually antagonistic effects on the redox state and on proinflammatory pathways. Although the short-term actions of HIF-1 α can reduce hypoxia-related injury, prolonged signaling through HIF-1 α leads to increases in oxidative stress, inflammation, and fibrosis, but these deleterious effects are opposed by the actions of HIF-2 α . The interplay between HIF-1a and HIF-2a may contribute to the evolution and progression of chronic heart failure, atherosclerotic and hypertensive vascular disorders, and chronic kidney disease. These disorders are characterized by activation of HIF-1a and suppression of HIF-2a; the latter effect explains why many chronic inflammatory diseases decrease the production of erythropoietin and cause anemia. Hypoxia mimetics are capable of potentiating both HIF-1 α and HIF-2 α ; ideally, such agents should act preferentially to promote HIF-2a while exerting little effect on or acting to suppress HIF-1a. Selective upregulation of HIF-2a can be achieved with drugs that inhibit isoform-selective PHDs or that promote the actions of SIRT1 (i.e., SGLT2 inhibitors). Augmentation of SIRT1 and HIF-2a signaling may explain the link between the proerythrocytic action of SGLT2 inhibitors and their cardioprotective effects.

ADDRESS FOR CORRESPONDENCE: Dr. Milton Packer, Baylor Heart and Vascular Institute, 621 North Hall Street, Dallas, Texas 75226. E-mail: milton. packer@baylorhealth.edu.

REFERENCES

1. Semenza GL. Hypoxia-inducible factors in physiology and medicine. Cell 2012;148:399–408.

2. Loboda A, Jozkowicz A, Dulak J. HIF-1 and HIF-2 transcription factors-similar but not identical. Mol Cells 2010;29:435-42.

3. Smythies JA, Sun M, Masson N, et al. Inherent DNA-binding specificities of the HIF-1 α and HIF-2 α transcription factors in chromatin. EMBO Rep 2019;20:e46401.

4. Park SK, Dadak AM, Haase VH, Fontana L, Giaccia AJ, Johnson RS. Hypoxia-induced gene expression occurs solely through the action of hypoxia-inducible factor 1alpha (HIF-1alpha): role of cytoplasmic trapping of HIF-2alpha. Mol Cell Biol 2003;23:4959-71.

5. Mole DR, Blancher C, Copley RR, et al. Genomewide association of hypoxia-inducible factor (HIF)-1alpha and HIF-2alpha DNA binding with expression profiling of hypoxia-inducible transcripts. J Biol Chem 2009;284:16767-75.

6. Wiesener MS, Jürgensen JS, Rosenberger C, et al. Widespread hypoxia-inducible expression of HIF-2alpha in distinct cell populations of different organs. FASEB J 2003;17:271-3.

7. Bernhardt WM, Schmitt R, Rosenberger C, et al. Expression of hypoxia-inducible transcription factors in developing human and rat kidneys. Kidney Int 2006;69:114–22.

8. Uchida T, Rossignol F, Matthay MA, et al. Prolonged hypoxia differentially regulates hypoxiainducible factor (HIF)-1alpha and HIF-2alpha expression in lung epithelial cells: implication of natural antisense HIF-1alpha. J Biol Chem 2004; 279:14871-8.

9. Chae KS, Kang MJ, Lee JH, et al. Opposite functions of HIF- α isoforms in VEGF induction by TGF- β 1 under non-hypoxic conditions. Oncogene 2011;30:1213-28.

10. Ahmad A, Ahmad S, Malcolm KC, et al. Differential regulation of pulmonary vascular cell growth by hypoxia-inducible transcription factor- 1α and hypoxia-inducible transcription factor- 2α . Am J Respir Cell Mol Biol 2013;49:78-85.

11. Yuan Y, Hilliard G, Ferguson T, Millhorn DE. Cobalt inhibits the interaction between hypoxiainducible factor-alpha and von Hippel-Lindau protein by direct binding to hypoxia-inducible factor-alpha. J Biol Chem 2003;278:15911-6.

12. Sridharan V, Guichard J, Bailey RM, Kasiganesan H, Beeson C, Wright GL. The prolyl hydroxylase oxygen-sensing pathway is cytoprotective and allows maintenance of mitochondrial membrane potential during metabolic inhibition. Am J Physiol Cell Physiol 2007;292:C719-28.

13. Tarhonskaya H, Hardy AP, Howe EA, et al. Kinetic investigations of the role of factor inhibiting hypoxia-inducible factor (FIH) as an oxygen sensor. J Biol Chem 2015;290:19726-42.

14. Berchner-Pfannschmidt U, Tug S, Trinidad B, et al. Nuclear oxygen sensing: induction of endogenous prolyl-hydroxylase 2 activity by hypoxia and nitric oxide. J Biol Chem 2008;283:31745-53.

15. Vengellur A, Woods BG, Ryan HE, Johnson RS, LaPres JJ. Gene expression profiling of the hypoxia

signaling pathway in hypoxia-inducible factor 1alpha null mouse embryonic fibroblasts. Gene Expr 2003;11:181–97.

16. Appelhoff RJ, Tian YM, Raval RR, et al. Differential function of the prolyl hydroxylases PHD1, PHD2, and PHD3 in the regulation of hypoxia-inducible factor. J Biol Chem 2004;279:38458-65.

17. Kapitsinou PP, Liu Q, Unger TL, et al. Hepatic HIF-2 regulates erythropoietic responses to hypoxia in renal anemia. Blood 2010;116:3039-48.

18. Warnecke C, Zaborowska Z, Kurreck J, et al. Differentiating the functional role of hypoxiainducible factor (HIF)-1alpha and HIF-2alpha (EPAS-1) by the use of RNA interference: erythropoietin is a HIF-2alpha target gene in Hep3B and Kelly cells. FASEB J 2004;18:1462-4.

19. Koh MY, Powis G. Passing the baton: the HIF switch. Trends Biochem Sci 2012;37:364-72.

20. Chandel NS, McClintock DS, Feliciano CE, et al. Reactive oxygen species generated at mitochondrial complex III stabilize hypoxia-inducible factorlalpha during hypoxia: a mechanism of O₂ sensing. J Biol Chem 2000;275:25130–8.

21. Pan Y, Mansfield KD, Bertozzi CC, et al. Multiple factors affecting cellular redox status and energy metabolism modulate hypoxia-inducible factor prolyl hydroxylase activity in vivo and in vitro. Mol Cell Biol 2007;27:912-25.

22. Zhang H, Gao P, Fukuda R, et al. HIF-1 inhibits mitochondrial biogenesis and cellular respiration in VHL-deficient renal cell carcinoma by repression of C-MYC activity. Cancer Cell 2007;11:407-20.

23. Schönenberger MJ, Kovacs WJ. Hypoxia signaling pathways: modulators of oxygen-related organelles. Front Cell Dev Biol 2015;3:42.

24. Walter KM, Schönenberger MJ, Trötzmüller M, et al. Hif- 2α promotes degradation of mammalian peroxisomes by selective autophagy. Cell Metab 2014;20:882–97.

25. Yuan G, Peng YJ, Reddy VD, et al. Mutual antagonism between hypoxia-inducible factors 1α and 2α regulates oxygen sensing and cardio-respiratory homeostasis. Proc Natl Acad Sci U S A 2013;110:E1788–96.

26. Scortegagna M, Ding K, Oktay Y, et al. Multiple organ pathology, metabolic abnormalities and impaired homeostasis of reactive oxygen species in Epas1-/- mice. Nat Genet 2003;35:331-40.

27. Yuan G, Nanduri J, Khan S, Semenza GL, Prabhakar NR. Induction of HIF-1alpha expression by intermittent hypoxia: involvement of NADPH oxidase, Ca2+ signaling, prolyl hydroxylases, and mTOR. J Cell Physiol 2008;217:67-85.

28. Nanduri J, Wang N, Yuan G, et al. Intermittent hypoxia degrades HIF-2alpha via calpains resulting in oxidative stress: implications for recurrent apnea-induced morbidities. Proc Natl Acad Sci U S A 2009;106:1199-204.

29. Ban JJ, Ruthenborg RJ, Cho KW, Kim JW. Regulation of obesity and insulin resistance by hypoxia-inducible factors. Hypoxia (Auckl) 2014;2: 171-83.

30. Yu X, Fang Y, Liu H, et al. The balance of beneficial and deleterious effects of hypoxiainducible factor activation by prolyl hydroxylase inhibitor in rat remnant kidney depends on the timing of administration. Nephrol Dial Transplant 2012;27:3110–9.

31. Takeda N, O'Dea EL, Doedens A, et al. Differential activation and antagonistic function of HIF-{alpha} isoforms in macrophages are essential for NO homeostasis. Genes Dev 2010;24:491-501.

32. Tashiro N, Segawa R, Tobita R, et al. Hypoxia inhibits TNF-α-induced TSLP expression in keratinocytes. PLoS One 2019;14:e0224705.

33. Charron CE, Chou PC, Coutts DJ, et al. Hypoxia-inducible factor 1alpha induces corticosteroid-insensitive inflammation via reduction of histone deacetylase-2 transcription. J Biol Chem 2009;284:36047-54.

34. Wang Z, Tang L, Zhu Q, et al. Hypoxia-inducible factor-1α contributes to the profibrotic action of angiotensin II in renal medullary interstitial cells. Kidney Int 2011;79:300-10.

35. Ishizuka S, Sakai T, Hiraiwa H, et al. Hypoxiainducible factor- 2α induces expression of type X collagen and matrix metalloproteinases 13 in osteoarthritic meniscal cells. Inflamm Res 2016; 65:439-48.

36. Yoon H, Shin SH, Shin DH, Chun YS, Park JW. Differential roles of Sirt1 in HIF-1 α and HIF-2 α mediated hypoxic responses. Biochem Biophys Res Commun 2014;444:36-43.

37. Chen R, Xu M, Hogg RT, et al. The acetylase/ deacetylase couple CREB-binding protein/sirtuin 1 controls hypoxia-inducible factor 2 signaling. J Biol Chem 2012;287:30800-11. **38.** Packer M. Role of deranged energy deprivation signaling in the pathogenesis of cardiac and renal disease in states of perceived nutrient overabundance. Circulation 2020;141: 2095-105.

39. Packer M. Role of impaired nutrient and oxygen deprivation signaling and deficient autophagic flux in diabetic CKD development: implications for understanding the effects of sodium-glucose cotransporter 2-inhibitors. J Am Soc Nephrol 2020;31:907-19.

40. Tang BL. Sirt1 and the mitochondria. Mol Cells 2016;39:87-95.

41. Hyvärinen J, Hassinen IE, Sormunen R, et al. Hearts of hypoxia-inducible factor prolyl 4hydroxylase-2 hypomorphic mice show protection against acute ischemia-reperfusion injury. J Biol Chem 2010;285:13646-57.

42. Wei H, Bedja D, Koitabashi N, et al. Endothelial expression of hypoxia-inducible factor 1 protects the murine heart and aorta from pressure overload by suppression of TGF- β signaling. Proc Natl Acad Sci U S A 2012;109. E841–50.

43. Krishnan J, Suter M, Windak R, et al. Activation of a HIF1alpha-PPARgamma axis underlies the integration of glycolytic and lipid anabolic pathways in pathologic cardiac hypertrophy. Cell Metab 2009;9:512-24.

44. Wei Q, Bian Y, Yu F, et al. Chronic intermittent hypoxia induces cardiac inflammation and dysfunction in a rat obstructive sleep apnea model. J Biomed Res 2016;30:490-5.

45. Pu J, Zhu S, Zhou D, et al. Propofol alleviates apoptosis induced by chronic high glucose exposure via regulation of HIF-1 α in H9c2 cells. Oxid Med Cell Longev 2019;2019:4824035.

46. Kumar S, Wang G, Liu W, et al. Hypoxiainduced mitogenic factor promotes cardiac hypertrophy via calcium-dependent and hypoxiainducible factor- 1α mechanisms. Hypertension 2018;72:331-42.

47. Sharma NM, Cunningham CJ, Zheng H, Liu X, Patel KP. Hypoxia-inducible factor-1a mediates increased sympathoexcitation via glutamatergic N-Methyl-d-aspartate receptors in the paraventricular nucleus of rats with chronic heart failure. Circ Heart Fail 2016;9:e003423.

48. Hölscher M, Schäfer K, Krull S, et al. Unfavourable consequences of chronic cardiac HIF-1 α stabilization. Cardiovasc Res 2012;94:77-86.

49. Bekeredjian R, Walton CB, MacCannell KA, et al. Conditional HIF-1alpha expression produces a reversible cardiomyopathy. PLoS One 2010;5: e11693.

50. Guo J, Mihic A, Wu J, et al. Canopy 2 attenuates the transition from compensatory hypertrophy to dilated heart failure in hypertrophic cardiomyopathy. Eur Heart J 2015;36:2530-40.

51. Pohl J, Hendgen-Cotta UB, Stock P, et al. Myocardial expression of macrophage migration inhibitory factor in patients with heart failure. J Clin Med 2017;6:95.

52. Li G, Lu WH, Wu XW, et al. Admission hypoxiainducible factor 1α levels and in-hospital mortality in patients with acute decompensated heart failure. BMC Cardiovasc Disord 2015;15:79. **53.** Shyu KG, Lu MJ, Chang H, Sun HY, Wang BW, Kuan P. Carvedilol modulates the expression of hypoxia-inducible factor-1alpha and vascular endothelial growth factor in a rat model of volume-overload heart failure. J Card Fail 2005;11: 152-9.

54. Martin CM, Ferdous A, Gallardo T, et al. Hypoxia-inducible factor-2alpha transactivates Abcg2 and promotes cytoprotection in cardiac side population cells. Circ Res 2008;102:1075-81.

55. Mastrocola R, Collino M, Penna C, et al. Maladaptive modulations of NLRP3 inflammasome and cardioprotective pathways are involved in diet-induced exacerbation of myocardial ischemia/ reperfusion injury in mice. Oxid Med Cell Longev 2016;2016:3480637.

56. Jürgensen JS, Rosenberger C, Wiesener MS, et al. Persistent induction of HIF-1alpha and -2alpha in cardiomyocytes and stromal cells of ischemic myocardium. FASEB J 2004;18:1415-7.

57. Wu D, Huang RT, Hamanaka RB, et al. HIF-1 α is required for disturbed flow-induced metabolic reprogramming in human and porcine vascular endothelium. Elife 2017;6:e25217.

58. Liu D, Lei L, Desir M, et al. Smooth muscle hypoxia-inducible factor 1α links intravascular pressure and atherosclerosis-brief report. Arterioscler Thromb Vasc Biol 2016;36:442–5.

59. Aarup A, Pedersen TX, Junker N, et al. Hypoxia-inducible factor- 1α expression in macrophages promotes development of atherosclerosis. Arterioscler Thromb Vasc Biol 2016;36:1782–90.

60. Schultz K, Fanburg BL, Beasley D. Hypoxia and hypoxia-inducible factor-1alpha promote growth factor-induced proliferation of human vascular smooth muscle cells. Am J Physiol Heart Circ Physiol 2006;290:H2528-34.

61. Na TY, Lee HJ, Oh HJ, Huh S, Lee IK, Lee MO. Positive cross-talk between hypoxia inducible factor-1 α and liver X receptor α induces formation of triglyceride-loaded foam cells. Arterioscler Thromb Vasc Biol 2011;31:2949-56.

62. Nakayama T, Kurobe H, Sugasawa N, et al. Role of macrophage-derived hypoxia-inducible factor (HIF)-1 α as a mediator of vascular remodelling. Cardiovasc Res 2013;99:705-15.

63. Natarajan R, Jones DG, Fisher BJ, Wallace TJ, Ghosh S, Fowler AA 3rd. Hypoxia inducible factor-1: regulation by nitric oxide in posthypoxic microvascular endothelium. Biochem Cell Biol 2005;83:597-607.

64. Matsuura Y, Yamashita A, Iwakiri T, et al. Vascular wall hypoxia promotes arterial thrombus formation via augmentation of vascular thrombogenicity. Thromb Haemost 2015;114:158-72.

65. Zhang X, Zhang Y, Wang P, et al. Adipocyte hypoxia-inducible factor 2α suppresses atherosclerosis by promoting adipose ceramide catabolism. Cell Metab 2019;30:937-51.

66. Poitz DM, Augstein A, Weinert S, Braun-Dullaeus RC, Strasser RH, Schmeisser A. OxLDL and macrophage survival: essential and oxygenindependent involvement of the Hif-pathway. Basic Res Cardiol 2011;106:761-72.

67. Zhang S, Zhao L, Wang J, Chen N, Yan J, Pan X. HIF-2 α and Oct4 have synergistic effects on

survival and myocardial repair of very small embryonic-like mesenchymal stem cells in infarcted hearts. Cell Death Dis 2017;8:e2548.

68. Kapitsinou PP, Sano H, Michael M, et al. Endothelial HIF-2 mediates protection and recovery from ischemic kidney injury. J Clin Invest 2014; 124:2396-409.

69. Xu X, Song N, Zhang X, et al. Renal protection mediated by hypoxia inducible factor-1α depends on proangiogenesis function of miR-21 by targeting thrombospondin 1. Transplantation 2017;101: 1811-9.

70. Conde E, Giménez-Moyano S, Martín-Gómez L, et al. HIF-1α induction during reperfusion avoids maladaptive repair after renal ischemia/reperfusion involving miR127-3p. Sci Rep 2017;7:41099.

71. Fang Y, Zhang H, Zhong Y, Ding X. Prolyl hydroxylase 2 (PHD2) inhibition protects human renal epithelial cells and mice kidney from hypoxia injury. Oncotarget 2016;7:54317-28.

72. Lourenço BN, Coleman AE, Schmiedt CW, et al. Profibrotic gene transcription in renal tissues from cats with ischemia-induced chronic kidney disease. Am J Vet Res 2020;81:180-9.

73. Luo L, Luo G, Fang Q, Sun Z. Stable expression of hypoxia-inducible factor-1α in human renal proximal tubular epithelial cells promotes epithelial to mesenchymal transition. Transplant Proc 2014;46:130–4.

74. Li ZL, Lv LL, Tang TT, et al. HIF-1 α inducing exosomal microRNA-23a expression mediates the cross-talk between tubular epithelial cells and macrophages in tubulointerstitial inflammation. Kidney Int 2019;95:388-404.

75. Deng W, Ren Y, Feng X, et al. Hypoxia inducible factor-1 alpha promotes mesangial cell proliferation in lupus nephritis. Am J Nephrol 2014; 40:507-15.

76. Baumann B, Hayashida T, Liang X, Schnaper HW. Hypoxia-inducible factor-1α promotes glomerulosclerosis and regulates COL1A2 expression through interactions with Smad3. Kidney Int 2016;90:797-808.

77. Kimura K, Iwano M, Higgins DF, et al. Stable expression of HIF-1alpha in tubular epithelial cells promotes interstitial fibrosis. Am J Physiol Renal Physiol 2008;295:F1023–9.

78. Nayak BK, Shanmugasundaram K, Friedrichs WE, et al. HIF-1 mediates renal fibrosis in OVE26 type 1 diabetic mice. Diabetes 2016;65: 1387-97.

79. Kong KH, Oh HJ, Lim BJ, et al. Selective tubular activation of hypoxia-inducible factor- 2α has dual effects on renal fibrosis. Sci Rep 2017;7: 11351.

80. Kojima I, Tanaka T, Inagi R, et al. Protective role of hypoxia-inducible factor-2alpha against

ischemic damage and oxidative stress in the kidney. J Am Soc Nephrol 2007;18:1218-26.

81. Landau D, London L, Bandach I, Segev Y. The hypoxia inducible factor/erythropoietin (EPO)/EPO receptor pathway is disturbed in a rat model of chronic kidney disease related anemia. PLoS One 2018;13:e0196684.

82. Olmos G, Muñoz-Félix JM, Mora I, et al. Impaired erythropoietin synthesis in chronic kidney disease is caused by alterations in extracellular matrix composition. J Cell Mol Med 2018;22: 302-14.

83. Bondeva T, Heinzig J, Ruhe C, Wolf G. Advanced glycated end-products affect HIF-transcriptional activity in renal cells. Mol Endocrinol 2013;27:1918-33.

84. Isoe T, Makino Y, Mizumoto K, et al. High glucose activates HIF-1-mediated signal transduction in glomerular mesangial cells through a carbohydrate response element binding protein. Kidney Int 2010;78:48–59.

85. Cioffi CL, Liu XQ, Kosinski PA, Garay M, Bowen BR. Differential regulation of HIF-1 alpha prolyl-4-hydroxylase genes by hypoxia in human cardiovascular cells. Biochem Biophys Res Commun 2003;303:947-53.

86. Aprelikova O, Chandramouli GV, Wood M, et al. Regulation of HIF prolyl hydroxylases by hypoxia-inducible factors. J Cell Biochem 2004; 92:491-501.

87. Miikkulainen P, Högel H, Seyednasrollah F, Rantanen K, Elo LL, Jaakkola PM, et al. Hypoxiainducible factor (HIF)-prolyl hydroxylase 3 (PHD3) maintains high HIF2A mRNA levels in clear cell renal cell carcinoma. J Biol Chem 2019;294: 3760-71.

88. Fukui K, Shinozaki Y, Kobayashi H, et al. JTZ-951 (enarodustat), a hypoxia-inducible factor prolyl hydroxylase inhibitor, stabilizes HIF- α protein and induces erythropoiesis without effects on the function of vascular endothelial growth factor. Eur J Pharmacol 2019;859: 172532.

89. Li X, Li J, Wang L, et al. The role of metformin and resveratrol in the prevention of hypoxiainducible factor 1α accumulation and fibrosis in hypoxic adipose tissue. Br J Pharmacol 2016;173: 2001-15.

90. Ryu DR, Yu MR, Kong KH, et al. Sirt1-hypoxiainducible factor-1 α interaction is a key mediator of tubulointerstitial damage in the aged kidney. Aging Cell 2019;18:e12904.

91. Shao Y, Lv C, Wu C, Zhou Y, Wang Q. Mir-217 promotes inflammation and fibrosis in high glucose cultured rat glomerular mesangial cells via Sirt1/HIF-1 α signaling pathway. Diabetes Metab Res Rev 2016;32:534–43.

92. Wang S, Qian Y, Gong D, Zhang Y, Fan Y. Resveratrol attenuates acute hypoxic injury in

cardiomyocytes: correlation with inhibition of iNOS-NO signaling pathway. Eur J Pharm Sci 2011; 44:416-21.

93. Pektaş MB, Sadi G, Koca HB, et al. Resveratrol ameliorates the components of hepatic inflammation and apoptosis in a rat model of streptozotocin-induced diabetes. Drug Dev Res 2016;77:12–9.

94. Osataphan S, Macchi C, Singhal G, et al. SGLT2 inhibition reprograms systemic metabolism via FGF21-dependent and -independent mechanisms. JCI Insight 2019;4:e123130.

95. Swe MT, Thongnak L, Jaikumkao K, Pongchaidecha A, Chatsudthipong V, Lungkaphin A. Dapagliflozin not only improves hepatic injury and pancreatic endoplasmic reticulum stress, but also induces hepatic gluconeogenic enzymes expression in obese rats. Clin Sci (Lond) 2019:133:2415–30.

96. Packer M. SGLT2 inhibitors produce cardiorenal benefits by promoting adaptive cellular reprogramming to induce a state of fasting mimicry: a paradigm shift in understanding their mechanism of action. Diabetes Care 2020;43: 508–11.

97. Mohamed HE, Asker ME, Keshawy MM, Hasan RA, Mahmoud YK. Inhibition of tumor necrosis factor-α enhanced the antifibrotic effect of empagliflozin in an animal model with renal insulin resistance. Mol Cell Biochem 2020;466: 45-54.

98. Li J, Liu H, Takagi S, Nitta K, et al. Renal protective effects of empagliflozin via inhibition of EMT and aberrant glycolysis in proximal tubules. JCI Insight 2020;5:e129034.

99. Bessho R, Takiyama Y, Takiyama T, et al. Hypoxia-inducible factor- 1α is the therapeutic target of the SGLT2 inhibitor for diabetic nephropathy. Sci Rep 2019;9:14754.

100. Mazer CD, Hare GMT, Connelly PW, et al. Effect of empagliflozin on erythropoietin levels, iron stores and red blood cell morphology in patients with type 2 diabetes and coronary artery disease. Circulation 2020; 141:704-7.

101. Inzucchi SE, Zinman B, Fitchett D, et al. How does empagliflozin reduce cardiovascular mortality? Insights from a mediation analysis of the EMPA-REG OUTCOME trial. Diabetes Care 2018; 41:356-63.

102. Li JW, Woodward M, Perkovic V, et al. Mediators of the effects of canagliflozin on heart failure in patients with type 2 diabetes. J Am Coll Cardiol HF 2020;8:57-66.

KEY WORDS cobalt chloride, hypoxiainducible factor-1α, hypoxia-inducible factor-2α, roxadustat, sodium-glucose cotransporter 2 inhibitors