



# **Concept of an Active Amplification Mechanism in the Infrared Organ of Pyrophilous** *Melanophila* **Beetles**

Erik S. Schneider<sup>1†</sup>, Anke Schmitz<sup>2†</sup> and Helmut Schmitz<sup>2\*†</sup>

<sup>1</sup> Institute of Zoology, University of Graz, Graz, Austria, <sup>2</sup> Institute of Zoology, University of Bonn, Bonn, Germany

Jewel beetles of the genus Melanophila possess a pair of metathoracic infrared (IR) organs. These organs are used for forest fire detection because Melanophila larvae can only develop in fire killed trees. Several reports in the literature and a modeling of a historic oil tank fire suggest that beetles may be able to detect large fires by means of their IR organs from distances of more than 100 km. In contrast, the highest sensitivity of the IR organs, so far determined by behavioral and physiological experiments, allows a detection of large fires from distances up to 12 km only. Sensitivity thresholds, however, have always been determined in non-flying beetles. Therefore, the complete micromechanical environment of the IR organs in flying beetles has not been taken into consideration. Because the so-called photomechanic sensilla housed in the IR organs respond bimodally to mechanical as well as to IR stimuli, it is proposed that flying beetles make use of muscular energy coupled out of the flight motor to considerably increase the sensitivity of their IR sensilla during intermittent search flight sequences. In a search flight the beetle performs signal scanning with wing beat frequency while the inputs of the IR organs on both body sides are compared. By this procedure the detection of weak IR signals could be possible even if the signals are hidden in the thermal noise. If this proposed mechanism really exists in Melanophila beetles, their IR organs could even compete with cooled IR quantum detectors. The theoretical concept of an active amplification mechanism in a photon receptor innervated by highly sensitive mechanoreceptors is presented in this article.

Keywords: *Melanophila*, pyrophilous insect, infrared receptor, photomechanic receptor, fire detection, active amplification

# INTRODUCTION

With 13 recent species jewel beetles of the genus *Melanophila* mainly can be found in the boreal and temperate forests of the holarctic zone (Bellamy, 2008). According to the current state of knowledge, males and females approach forest fires because their larvae can only develop in wood of freshly burnt trees (Linsley, 1943; Apel, 1988, 1989, 1991). As an adaptation to the pyrophilous way of life the 1 cm long black beetles (**Figure 1A**) are equipped with special antennal smoke receptors (Schütz et al., 1999) and one pair of metathoracic IR organs (Evans, 1964; Vondran et al., 1995; Schmitz et al., 1997). An IR organ consists of a little array of dome-shaped sensilla which is situated at the bottom of a little pit (**Figures 1B, 2A,B**). The inner spherule of each sensillum is innervated by a ciliary mechanosensitive cell (**Figure 2C**; Vondran et al., 1995; Schmitz et al., 2007). Thus, the

#### **OPEN ACCESS**

#### Edited by:

Sylvia Anton, Institut National de la Recherche Agronomique, France

#### Reviewed by:

Maria Hellwig, University of Vienna, Austria Daniel Robert, University of Bristol, UK

> \*Correspondence: Helmut Schmitz

h.schmitz@uni-bonn.de <sup>†</sup>These authors have contributed

equally to this work.

#### Specialty section:

This article was submitted to Invertebrate Physiology, a section of the journal Frontiers in Physiology

Received: 09 October 2015 Accepted: 30 November 2015 Published: 21 December 2015

#### Citation:

Schneider ES, Schmitz A and Schmitz H (2015) Concept of an Active Amplification Mechanism in the Infrared Organ of Pyrophilous Melanophila Beetles. Front. Physiol. 6:391. doi: 10.3389/fphys.2015.00391

1

IR sensilla do not only respond to IR radiation but also to mechanical stimuli. A bimodality has already been demonstrated by electrophysiological experiments: single sensilla respond to weak vibratory stimuli with distinct action potentials (see Figure 3C in Schmitz and Bleckmann, 1998).

There are several hints in the literature that beetles are not only able to detect forest fires but also fires and heat sources of anthropogenic origin from distances of 30 km, 60 km, and even more than 100 km (Van Dyke, 1926; Linsley, 1943; Linsley and



FIGURE 1 | (A) *Melanophila cuspidata* dorsally glued to a needle flying in front of a wind tunnel. Red box encircles area shown in (B). (B) Boundary between meso- and metathorax. Cx, coxa; EPI, episternite; VE, ventrite with IR organ (IR) close to the boundary to the mesothorax.

Hurd, 1957). Currently it seems not very likely that beetles use the smell of smoke to detect fires from greater distances. It could not be demonstrated that Melanophila beetles could be lured by the smell of smoke (Evans, 1964) or that beetles resting at temperatures of 25°C could be aroused by smoke (unpublished data). A recent study, however, shows that beetles can be attracted by certain volatiles emitted by burning or smoldering wood (Paczkowski et al., 2013). In the study crawling beetles were tested in a two arm olfactometer at a temperature of 30°C. No information about the sex and mating state of the beetles is provided in this study. So these data are more likely suited to show that beetles (e.g., mated females?), once having landed on a burnt tree, can detect a suitable spot for oviposition by olfactory cues. Evaluations of satellite images very often yielded the result that the large smoke plume from a forest fire initially is driven away from the fire by the wind in a narrow angle over distances of many kilometers and finally gradually ascends to higher altitudes. So only beetles inside the smoke plume have a chance to become aware of the fire by olfactory cues. In contrast, beetles that are already close to the fire but outside the smoke plume most probably can see the plume but are not able to smell the smoke. Also the light of the flames-generally only visible at night-may not play an important role, because Melanophila beetles, as nearly all jewel beetles, are diurnal (Evans et al., 2007).

All threshold sensitivities published so far were measured in non-flying beetles (highest sensitivity 60  $\mu$ W/cm<sup>2</sup>, see Table 1).





TABLE 1	Sensitivity	thresholds	of biological	and	technical	IR sensors.
---------	-------------	------------	---------------	-----	-----------	-------------

	Threshold	Source/Explanation
<i>Melanophila</i> Behavioral experiments	$60\mu\text{W/cm}^2$	Evans, <i>Ecology</i> (1966); Determined by behavioral experiments with non-flying beetles
Pyroelectric sensor (room temperature) (DIAS 256 LTI)	$8.4\mu\text{W/cm}^2$	Sensorarray DIAS Infrared, Dresden
Rattlesnake Crotalus atrox	$3.3 \mu\text{W/cm}^2$	Ebert and Westhoff, JCP A (2006)
Microbolometer (room temperature)	2.3 µW/cm <sup>2</sup>	Sensorarray Information provided by Dr. N. Hess, DIAS Infrared GmbH, Dresden. Compiled after: annual SPIE-Conferences "Infrared Technology," Orlando, USA, 1999–2011
Cooled quantum detector (AIM HgCdTe)	79 nW/cm <sup>2</sup>	Sensorarray AIM Infrarotmodule GmbH, Heilbronn AIM Data Sheet 2nd-3rd-Generation
<i>Melanophila</i> Coalinga-Simulation	40 nW/cm <sup>2</sup>	Schmitz and Bousack, <i>PLoS ONE</i> (2012)
Cryogenic cooled quantum detector (Hamamatsu P5274 Serie)	250 pW/cm <sup>2</sup>	High sensitivity single element quantum detector Hamamatsu MCT photoconductive detectors Data sheet P2748/P5274 series

Theoretical calculations, however, show that these sensitivities only allow a detection of a large fire from a distance of about 12 km (Evans, 1966; Schmitz and Bleckmann, 1998).

The results of a simulation of a huge man-made fire provide further evidence that *Melanophila* beetles might be able to detect IR radiation emitted by remote fires from much larger distances (Schmitz and Bousack, 2012). In this study a big oil-tank fire was modeled that burnt in 1925 for 3 days in Coalinga (California) and attracted "untold numbers" of Melanophila consputa. This event has also been documented in the entomological literature (Van Dyke, 1926). The site of the fire in the woodless Central Valley of California suggests that most beetles detected the fire by IR radiation from distances of 130 km. This would imply a threshold sensitivity of only 40 nW/cm<sup>2</sup> (Table 1), corresponding to an energy at a single sensillum of  $1.3 \times 10^{-17}$  J. If the threshold of the Melanophila IR organ should really be in this range, the biological IR receptor would be two orders of magnitude more sensitive than all current uncooled technical IR sensors and would be able to compete with much more expensive cooled quantum detectors (cf. Table 1).

However, a sensitivity three orders of magnitude higher than the highest sensitivity ever published (**Table 1**) is only explainable by active amplification mechanisms. Until now, active amplification of very weak input signals has only be reported in the context of hearing: in the cochlear amplifier in the inner ear of vertebrates (Hudspeth, 1989, 1997; Gillespie and Müller, 2009), in antennal ears of mosquitoes and the fly *Drosophila* (Göpfert and Robert, 2001, 2003; Göpfert et al., 2005, 2006; Mhatre, 2015) and recently discovered also in the tympanal ears of a tree cricket (Mhatre and Robert, 2013). An amplification of 1000-fold can be achieved by electromotile outer hair cells in the mammalian cochlea (Robles and Ruggero, 2001; Ashmore et al., 2010). Like hair cells and scolopidia in



FIGURE 3 | (A) Morphology of the pleural region of the metathorax in beetles shown here for the rhinoceros beetle *Oryctes rhinoceros* (after Darwin and Pringle, 1959). The episternite with the fulcrum and the apodem of the basalar muscle (ApoB) are connected by a posterior hinge (red arrow) to the ventrite. In the region of the sternopleural suture (SPS, green) a hinge membrane allows movement of the episternite against the ventrite (Pringle, 1957). (B)
Dorsoventrally oriented section through episternite and ventrite in the anterior region (indicated by red line in A) in the buprestid beetle *Chrysobothris solieri*.
(C) Corresponding section about 150 µm in posterior direction.

vertebrate and insect ears, respectively, the mechanosensitive sensory cells that innervate the IR sensilla in *Melanophila* are ciliary mechanoreceptors. Therefore, the search for active amplification mechanisms appears highly challenging.

## MORPHOLOGICAL PREREQUISITES

Starting point for the development of the concept was the consideration that the IR organs are situated on the metathorax just below the wing hinges (fulcra) of the alae (Figures 1A, 4A,B). Thus, the IR organs are located at a site strongly subjected to vibrations during flight. Additionally, a detail so far not understood was considered: the sphere of a sensillum is attached by a little stalk to the outer cuticular dome (Figures 2C,D). These stalks are missing in the photomechanic IR sensilla of pyrophilous *Aradus* bugs which are quite similar to the *Melanophila* IR sensilla but are not enclosed in a pit organ (Schmitz et al., 2010). By this constellation, vibrations of the spheres in flying *Melanophila* beetles can be proposed to affect the receptor potentials of the mechanosensory cells.

To realize genuine amplification, however, a mechanism has to be implemented permitting a precise regulation of the depolarization amplitude. The morphological prerequisites for such a regulation mechanism were found in the two hitherto investigated species Melanophila acuminata and M. cuspidata. The structural preadaptation for the evolution of an adjustable beat mechanism most probably was a special feature of the metathoracic pleural region in beetles: the sternopleural suture (SPS, Figures 3A-C). Episternite and ventrite are tightly connected only by a posterior hinge and thus can be moved against each other-especially at their anterior edges (Pringle, 1957). Amongst other purposes this mechanism mainly serves for lifting the fulcrum during flight. When the basalar muscle contracts, the episternite is pulled against the ventrite and the leading edge of the wing is pronated during the downstroke (Darwin and Pringle, 1959). The opposing edges of the episternite and the ventrite are formed in a manner so that the episternite can glide over the ventrite (as in the non IR-sensitive jewel beetle Chrysobothris solieri, closely related to the Melanophila-species, Figures 3B,C).

It turned out that in IR-sensitive *Melanophila*-species especially the morphology of the opposing anterior edges of episternite and ventrite has been modified. The slim ventral edge of the episternite can be beaten in a trench on the dorsal edge of the ventrite, thus a kind of impact edge is realized (**Figures 4D**, **5A–C**). According to the current conception beating of the two sclerites against one another is accomplished by contractions of the basalar muscle, which extends from the dorsal apodeme (ApoB; shown in **Figure 4B**) posteriorly to the basal region of the ventrite. By regulating the power of the muscle contractions during a search flight sequence (see below), the vibrations of the spheres caused by the proposed mechanism can be controlled. Therefore, also the magnitude of receptor depolarizations could be regulated.

In this context it is of great importance that the IR sensilla are innervated by ciliary mechanoreceptors. Specialized arthropod mechanoreceptors innervated by ciliary mechanosensory cells are the most sensitive receptors known. This could be shown for trichobothria in spiders, where energies of  $1.5 \times 10^{-19}$  J to  $2.5 \times 10^{-20}$  J are still sufficient for a suprathreshold stimulation of the receptors (Humphrey et al., 2003; Barth, 2004) and also for filiform hairs in insects (Thurm, 1982). Filiform hairs in crickets serving for detection of faint airflows can already generate an action potential if energies are still in the range of k<sub>B</sub>T (k<sub>B</sub>: Boltzmann constant; T: temperature), i.e., about  $4 \times 10^{-21}$  J (Shimozawa et al., 2003).

At the threshold, these ultrasensitive mechanoreceptors already work within the range of thermal noise of Brownian





molecular motion and therewith close to the physical limit (Barth, 2004). It can be concluded that for a subthreshold depolarization of the mechanosensitive sensory cell innervating the IR sensilla most probably vibration amplitudes of the spheres of less than one nanometer are already sufficient.

In the two Melanophila species investigated so far, M. acuminata and M. cuspidata, morphological differences were found. It appears that the IR detection system (i.e., IR-organ plus the structures used for the proposed beat mechanism) in M. cuspidata is more ancient and relatively simple: IR organs contain less sensilla into which the wax glands are fully integrated (Figure 2A). So an unrestrained vibration of the sphere most probably may be somewhat hampered and less precise. An explanation could be that M. cuspidata is distributed in the Mediterranean region where fires are more frequent than in northern Europe (San-Miguel and Camia, 2009). Thus, the necessity to detect fires also from very large distances seems not to be predominant. In contrast, M. acuminata is distributed in the boreal forests of the northern hemisphere (in Europe northern distribution up to Fennoscandia; Horion, 1955), where forest fires are less frequent. Accordingly, a higher evolutionary pressure with respect to the sensitivity of the IR organs can be proposed. In M. acuminata, the IR organs contain significantly more IR sensilla from which wax glands are clearly separated (**Figures 2B–D**). It can be concluded that unobstructed vibrations of the spheres are possibles. Furthermore, an additional damping system obviously has developed. This system may allow a precise "fine tuning" of the depolarizations of the IR sensilla caused by the beat mechanism. By a system of at least two muscles of hitherto unknown origin a damping cushion of about 300  $\mu$ m lengths can be brought down into the inner trench of the ventrite (**Figures 5A–D**). Thus, at a given contraction power of the basalar muscle a very precise adjustment of the beat intensity and consequently of the evoked pre-depolarizations could be adjusted. Despite intense search in two further specimens such a damping cushion could not be found in the Mediterranean *M. cuspidata* (cf. **Figure 4**).

# HOW IT COULD WORK

According to the present idea how *Melanophila* beetles may be able to become aware of a fire from large distances, beetles use a combination of visual cues (view of a big cloud against the horizon) and IR radiation. To make sure that a smoke plume and not a cloud bank is approached over distances of many kilometers a zone of IR emission has to exist at the base of the cloud above tree top level. IR sensitive *Melanophila* beetles, therefore, will conduct search flights for fire





detection. While doing so, beetles especially examine potential smoke plumes in view of additional IR emission. By the beat mechanism a cyclic depolarization of the IR sensilla with wing beat frequency appears possible (**Figures 6A,B**). In beetles, the basalar muscle serves as a well-developed direct wing depressor, which, together with the indirect dorso-longitudinal muscles (main muscle for propulsion), is used for wing downstroke. In principle the basalar muscle can be classified as a steering muscle (Nachtigall, 2003). By adjusting the contraction power of the basalar muscle, the wing inclination angle during the downstroke of the wing (pronation) and in this way propulsion



FIGURE 6 | Schematic depiction of the proposed beating mechanism. (A) Coupling in muscular energy (blue double arrows) from the flight motor results in vibration of the cuticular spheres. This in turn causes the stimulation of the mechanoreceptive sensory cells by which the spheres are innervated. By sensory feedback the sensory cells can be systematically depolarized with wing beat frequency. In contrast, energy for active amplification in hearing organs is generated by molecular motors of the sensory cells itself (green double arrows at the sensory cells). (B) In non-flying beetles the receptor potential of the IR sensilla is set at about  $-70\,\mathrm{mV}.$  Slight fluctuations are caused by thermal and mechanical noise. When the beetles take off, IR sensilla become depolarized by the beating mechanism: (1) by increasing beat intensity, the beetle is able to release action potentials in the absence of IR radiation. (2) by tuning the beat intensity and - in Melanophila acuminate-by additional fine tuning of the damping system, depolarization can be adjusted in a way that the generator potentials of most sensilla remain just below the threshold. (3) in turn IR radiation superimposed to the thermal noise at the peaks of the receptor potentials causes the generation of (additional) action potentials in exposed sensilla.

and buoyancy is adjusted. During a supposed search flight sequence, which may last for a few seconds only, the beetles could tune the contractions of the basalar muscles exactly so that the peak amplitudes of the oscillatory receptor potential almost reach the spike-triggering threshold. At the same time a slight reduction in propulsion and buoyancy would not be disadvantageous. To tune the IR organs to maximal sensitivity in anticipation of arriving IR radiation, the beetles should be able to adjust the intensity of the beat mechanism and therewith the probability of impulse generation by sensory feedback. As a result only a certain, most probably very low, percentage of sensilla in both organs generate action potentials.

With respect to symmetry the inputs of both IR organs could be permanently compared by central comparator neurons. Such central units enable acoustically communicating insects to approach, e.g., a sound source by paired hearing organs (von Helversen and von Helversen, 1995; Stumpner and von Helversen, 2001). A mechanical prestimulation of only a few sensilla in the Melanophila IR organ could be explainable by the fact that the sensilla show minute differences in their dimensions (cf. Figures 2A,B). In case of an oscillatory mechanical stimulation with constant intensity some sensilla will already generate first action potentials whereas most others will remain just below the threshold. At the peak of a given subthreshold depolarization additional IR radiation will slightly increase the height of the amplitude (Figure 6B). This will result in a few more spiking sensilla in the organ exposed to IR radiation. The asymmetry in the inputs of both organs immediately could be detected by comparator neurons. Therewith the beetle gets information about the spatial direction from which IR radiation arrives. This information could be combined with visual cues (e.g., a smoke plume) and the beetle should be able to directly approach a fire.

Based on theoretical considerations it seems essential that the vibrations of the spheres caused by the proposed beat mechanism have to be strongly damped. By appropriate damping an uncontrolled soaring up of the system can be suppressed and it can be ensured that the spheres all are in a defined initial state before the next impact impulse arrives. Ideally a creeping case (i.e., damping so strong that no oscillation can arise) or at least an aperiodic limit case (i.e., strong damping ensures oscillation of the sphere with only one zero crossing) has to be proposed. In this way beat impulses of constant intensity will always cause monotonic depolarization amplitudes. Most probably damping is realized by a slender margin of fluid surrounding each sphere (Figure 2C). This margin with a thickness of about  $0.3 \,\mu m$ consists of the apical extensions of the two outer enveloping cells (Vondran et al., 1995). By this specific feature a fluidic damping system is build. Subthreshold depolarizations of the receptors most probably are already evoked by sub-nanometer vibrations of the spheres. The necessary small scale dislocation of water is allowed by compensatory air sacs below the IR organ (Figure 4D, AS).

Provided that the cuticular apparatus (i.e., mainly the spheres) is able to convert the energies of absorbed IR photons effectively into mechanical energy (so-called photomechanic mechanism of IR reception, Schmitz and Bleckmann, 1998) it could be possible that the sensitivity threshold of a *Melanophila* IR organ is about thousand fold lower (analogous to the mammalian cochlear amplifier) than the hitherto published lowest threshold of  $60 \,\mu$ W/cm<sup>2</sup>.

# **CLOSING REMARKS**

Among biological IR sensory systems it is a unique feature of Melanophila beetles that IR sensilla serve as photon receptors although they are innervated by mechanosensitive neurons. The cuticular apparatus absorbs incoming IR radiation and transforms photon energy into a micromechanical event measured by a dedicated mechanoreceptor. In principle this constellation provides the possibility of active amplification of faint mechanical input signals. As mentioned in the Introduction active amplification has been shown in the context of hearing: for the hair cells in the cochlear amplifier in vertebrates (Hudspeth, 1997; LeMasurier and Gillespie, 2005; Fettiplace and Hackney, 2006; Ashmore et al., 2010) but also for the chordotonal organs in the ears of certain flies (Göpfert and Robert, 2001; Göpfert et al., 2005; Nadrowski et al., 2008) and a tree cricket (Mhatre and Robert, 2013). In hearing, however, the energy required for amplification is expended by the sensory cells themselves whereas in the proposed active IR receptors in Melanophila beetles the energy originates from the flight motor. Thus, in principle, the proposed mechanism to achieve a high sensitivity in a receptor for electromagnetic radiation is new. We further suggest that three different mechanisms are involved: (i) as proposed active signal amplification, (ii) active sensing which means that activity

## REFERENCES

- Apel, K.-H. (1988). Befallsverteilung von Melanophila acuminata DEG., Phaenops cyanea F. und Ph. formaneki JACOB. (Col., Buprestidae) auf Waldbrandflächen. Beitr. Forstwirtschaft 22, 45–48.
- Apel, K.-H. (1989). Zur Verbreitung von Melanophila acuminata DEG. (Col., Buprestidae). Entomol. Nach. Ber. 33, 278–280.
- Apel, K.-H. (1991). Die Kiefernprachtkäfer. Eberswalde: Forschungsanstalt für Forst- und Holzwirtschaft Eberswalde, Merkblatt Nr. 50, 1–30.
- Ashmore, J., Avan, P., Brownell, W. E., Dallos, P., Dierkes, K., Fettiplace, R., et al. (2010). The remarkable cochlear amplifier. *Hear. Res.* 266, 1–17. doi: 10.1016/j.heares.2010.05.001
- Barth, F. G. (2004). Spider mechanoreceptors. Curr. Opin. Neurobiol. 14, 415–422. doi: 10.1016/j.conb.2004.07.005
- Bellamy, C. L. (2008). A World Catalogue and Bibliography of the Jewel Beetels (Coleoptera: Buprestidae). Pensoft Series Faunistica No. 78.
- Darwin, F. W., and Pringle, J. W. S. (1959). The physiology of insect fibrillar muscle. I. Anatomy and innervation of the basalar muscle of lamellicorn beetles. *Proc. R. Soc. London B* 151, 194–203. doi: 10.1098/rspb.1959.0059
- Evans, H. F., Moraal, L. G., and Pajares, J. A. (2007). "Biology, ecology and economic importance of Buprestidae and Cerambycidae," in *Bark and Wood Boring Insects in Living Trees in Europe, A Synthesis*, eds K. R. D. F. Lieutier, A. Battisti, J.-C. Grégoire, and H. F. Evans (Dordrecht: Springer), 447–474.
- Evans, W. G. (1964). Infrared receptors in *Melanophila acuminata* De Geer. *Nature* 202, 211. doi: 10.1038/202211a0
- Evans, W. G. (1966). Perception of infrared radiation from forest fires by Melanophila acuminata De Geer (Buprestidae, Coleoptera). Ecology 47, 1061–1065. doi: 10.2307/1935658
- Fettiplace, R., and Hackney, C. M. (2006). The sensory and motor roles of auditory hair cells. *Nat. Rev. Neurosci.* 7, 19–29. doi: 10.1038/nrn1828

of the sensor system already starts in anticipation of a stimulus (Nelson and MacIver, 2006), and (iii) stochastic resonance: noise—in this case self-generated—is used for better signal detection (Harmer et al., 2002; Moss et al., 2004; McDonnell and Abbott, 2009).

The proposed beat mechanism, however, shows some marked differences compared to the mechanisms mentioned. No energy used for target analysis is emitted into the surrounding (difference to conventional *active sensing*), muscular energy is used for signal amplification (fundamental difference to the cellular molecular motors of the sensory cells in ears) and the crucial part of the "noise" is produced by self-generated oscillations. For the purpose of ultrasensitive stimulus detection the probability of action potential generation can be adjusted by altering the overall noise amplitude (difference to stochastic resonance that only works at an optimal noise intensity, which can hardly be influenced by the sensor system).

If the proposed high sensitivity of the IR organ could be demonstrated, the biological IR sensor would advance into the sensitivity gap currently existing between relatively cheap uncooled thermal IR sensors and expensive cooled quantum detectors requiring much more effort during operation and also more costly service (see **Table 1**). Thus, the demonstration of the postulated amplification mechanism would also be of technical interest for the development of new active IR sensors.

# ACKNOWLEDGMENTS

We are indebted to Horst Bleckmann for useful discussion and improvements of the manuscript.

- Gillespie, P. G., and Müller, U. (2009). Mechanotransduction by hair cells: models, molecules, and mechanisms. *Cell* 139, 33-44. doi: 10.1016/j.cell.2009. 09.010
- Göpfert, M. C., Albert, J. T., Nadrowski, B., and Kamikouchi, A. (2006). Specification of auditory sensitivity by *Drosophila* TRP channels. *Nat. Neurosci.* 9, 999–1000. doi: 10.1038/nn1735
- Göpfert, M. C., Humphris, A. D. L., Alber, J. T., Robert, D., and Hendrich, O. (2005). Power gain exhibited by motile mechanosensory neurons in *Drosophila* ears. *Proc. Natl. Acad. Sci. U.S.A.* 102, 325–330. doi: 10.1073/pnas.04057 41102
- Göpfert, M. C., and Robert, D. (2001). Active auditory mechanics in mosquitoes. *Proc. R. Soc. Lond. B* 268, 333–339. doi: 10.1098/rspb.2000.1376
- Göpfert, M. C., and Robert, D. (2003). Motion generation by Drosophila mechanosensory neurons. Proc. Natl. Acad. Sci. U.S.A. 100, 5514–5519. doi: 10.1073/pnas.0737564100
- Harmer, G. P., Davis, B. R., and Abbott, D. (2002). A review of stochastic resonance: circuits and measurement. *IEEE Trans. Instrum. Meas.* 51, 299–309. doi: 10.1109/19.997828
- Horion, A. (1955). Faunistik der mitteleuropäischen Käfer, Bd. 4: Sternoxia (Buprestidae), Fossipedes, Macrodactylia, Brachymera. Entomol. Arb. Mus. Frey, Tutzing, Sonderband.
- Hudspeth, A. J. (1989). How the ear's works work. *Nature* 341, 397-404. doi: 10.1038/341397a0
- Hudspeth, A. J. (1997). Mechanical amplification of stimuli by hair cells. Curr. Opin. Neurobiol. 7, 480–486. doi: 10.1016/S0959-4388(97)80026-8
- Humphrey, J. A. C., Barth, F. G., Reed, M., and Spak, A. (2003). "The physics of arthropod medium-flow sensitive hairs: biological models for artificial sensors," in *Sensors and Sensing in Biology and Engineering*, eds J. H. FG Barth, and T. W. Secomb (Wien; New York, NY: Springer-Verlag), 129-144.

- LeMasurier, M., and Gillespie, P. G. (2005). Hair-cell mechanotransduction and cochlear amplification. *Neuron* 48, 403–415. doi: 10.1016/j.neuron.2005.10.017
- Linsley, E. G. (1943). Attraction of *Melanophila* beetles by fire and smoke. J. Econ. Entomol. 36, 341–342. doi: 10.1093/jee/36.2.341
- Linsley, E. G., and Hurd, P. D. (1957). *Melanophila* beetles at cement plants in southern California (*Coleoptera, Buprestidae*). *Coleopt. Bull.* 11, 9–11.
- McDonnell, M. D., and Abbott, D. (2009). What is stochastic resonance? Definitions, misconceptions, debates, and its relevance to biology. *PLoS Computat. Biol.* 5:e1000348. doi: 10.1371/journal.pcbi.1000348
- Mhatre, N. (2015). Active amplification in insect ears: mechanics, models and molecules. J. Comparat. Physiol. A 201, 19–37. doi: 10.1007/s00359-014-0969-0
- Mhatre, N., and Robert, D. (2013). A tympanal insect ear exploits a critical oscillator for active amplification and tuning. *Curr. Biol.* 23, 1–6. doi: 10.1016/j.cub.2013.08.028
- Moss, F., Ward, L. M., and Sannita, W. G. (2004). Stochastic resonance and sensory information processing: a tutorial and review of application. *Clin. Neurophysiol.* 115, 267–281. doi: 10.1016/j.clinph.2003.09.014
- Nachtigall, W. (2003). Insektenflug. Berlin; Heidelberg: Springer-Verlag.
- Nadrowski, B., Albert, J. T., and Göpfert, M. C. (2008). Transducer-based force generation explains active process in *Drosophila* hearing. *Curr. Biol.* 18, 1365–1372. doi: 10.1016/j.cub.2008.07.095
- Nelson, M. E., and MacIver, M. A. (2006). Sensory acquisition in active sensing systems. J. Comp. Physiol. A 192, 573–586. doi: 10.1007/s00359-006-0099-4
- Paczkowski, S., Paczkowska, M., Dippel, S., Schulze, N., Schutz, S., Sauerwald, T., et al. (2013). The olfaction of a fire beetle leads to new concepts for early fire warning systems. *Sens. Actuators. B. Chem.* 183, 273–282. doi: 10.1016/j.snb. 2013.03.123
- Pringle, J. W. S. (1957). Inset Flight. London: Cambridge University Press.
- Robles, L., and Ruggero, M. A. (2001). Mechanics of the mammalian cochlea. *Physiol. Rev.* 81, 1305–1352. Available online at: http://physrev.physiology.org/ content/81/3/1305.short
- San-Miguel, J., and Camia, A. (2009). "Forest fires at a glance: facts, figures and trends in the EU," in *Living with Wildfires: What Science Can Tell Us. EFI Discussion Paper 15*, ed Y. Birot (European Forest Institute), 11–18.
- Schmitz, A., Schätzel, H., and Schmitz, H. (2010). Distribution and functional morphology of photomechanic infrared sensilla in flat bugs of the genus Aradus (Heteroptera, Aradidae). Arthropod Struct. Dev. 39, 17–25. doi: 10.1016/j.asd.2009.10.007
- Schmitz, A., Sehrbrock, A., and Schmitz, H. (2007). The analysis of the mechanosensory origin of the infrared sensilla in *Melanophila acuminata* (*Coleoptera; Buprestidae*) adduces new insight into the transduction mechanism. *Arthropod Struct. Dev.* 36, 291–303. doi: 10.1016/j.asd.2007.02.002

- Schmitz, H., and Bleckmann, H. (1998). The photomechanic infrared receptor for the detection of forest fires in the buprestid beetle *Melanophila acuminata. J. Comp. Physiol. A* 182, 647–657. doi: 10.1007/s0035900 50210
- Schmitz, H., and Bousack, H. (2012). Modelling a historic oil-tank fire allows an estimation of the sensitivity of the infrared receptors in pyrophilous *Melanophila* beetles. *PLoS ONE* 7:e37627. doi: 10.1371/journal.pone.0037627
- Schmitz, H., Mürtz, M., and Bleckmann, H. (1997). Infrared detection in a beetle. *Nature* 386, 773–774. doi: 10.1038/386773a0
- Schütz, S., Weissbecker, B., Hummel, H. E., Apel, K.-H., Schmitz, H., and Bleckmann, H. (1999). Insect antennae as a smoke detector. *Nature* 398, 298–299. doi: 10.1038/18585
- Shimozawa, T., Murakami, J., and Kumagai, T. (2003). "Cricket wind receptors: Thermal noise for the highest sensitivity known," in *Sensors and Sensing in Biology and Engineering*, eds F. G. Barth, J. A. C. Humphrey, and T. W. Secomb (New York, NY: Springer), 145–157.
- Stumpner, A., and von Helversen, D. (2001). Evolution and function of auditory systems in insects. *Naturwissenschaften* 88, 159–170. doi: 10.1007/s001140100223
- Thurm, U. (1982). "Grundzüge der Transduktionsmechanismen," in Sinneszellen. Mechano-elektrische Transduktion, eds W. Hoppe, W. Lohmann, H. Markl, and H. Ziegler (Berlin: Springer), 681–696.
- Van Dyke, E. C. (1926). Buprestid swarming. Pan-Pac. Ent. 3, 41.
- Vondran, T., Apel, K. H., and Schmitz, H. (1995). The infrared receptor of *Melanophila acuminata* De Geer (*Coleoptera: Buprestidae*): ultrastructural study of a unique insect thermoreceptor and its possible descent from a hair mechanoreceptor. *Tissue Cell* 27, 645–658. doi: 10.1016/S0040-8166(05) 80020-5
- von Helversen, D., and von Helversen, O. (1995). Acoustic pattern recognition and orientation in orthopteran insects: parallel or serial processing? *J. Comp. Physiol. A* 177, 767–774. doi: 10.1007/BF00187635

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Schneider, Schmitz and Schmitz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.