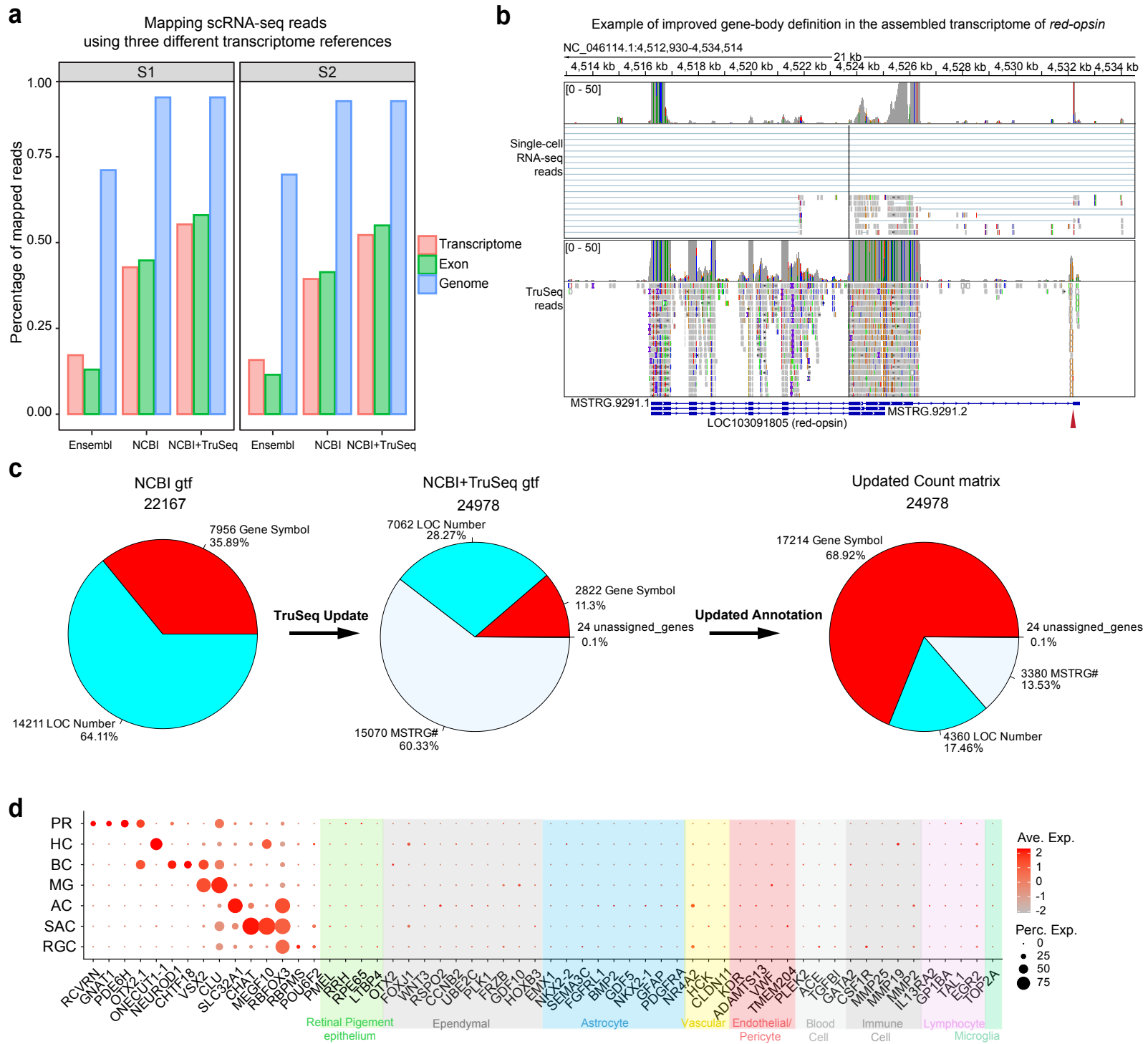


Supplementary Information

Supplementary Figure 1-11

Supplementary Table 1-5

Figure S1



Supplementary Figure 1. Assembly and annotation of the retina-specific transcriptome.

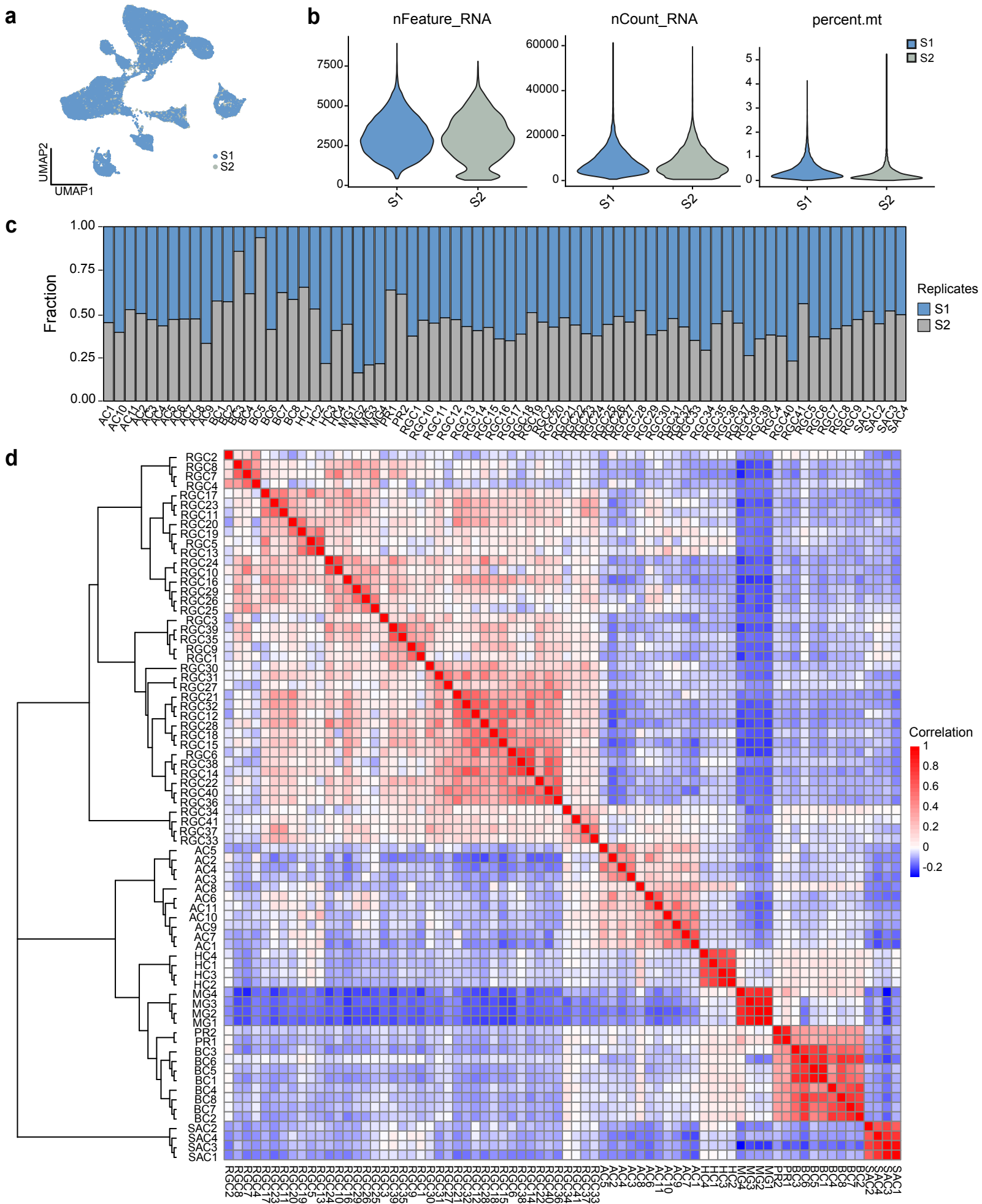
(a) Bar plots showing mapping percentages of scRNA-seq reads to “Transcriptome,” “Exon,” and “Genome” from two biological replicates (S1 and S2) with Ensembl (Pmarinus_7.0), NCBI (kPetMar1.pri), or the updated NCBI+TruSeq transcriptome reference. See Source Data.

(b) Improved gene-body definition for the *red-opsin* gene in the NCBI+TruSeq transcriptome. The alignment of scRNA-seq (top panel) and TruSeq (bottom panel) reads to the “*red-opsin*” locus was visualized with the Integrated Genomics Viewer (IGV). Red arrowhead indicates a newly identified exon region of the *red-opsin* gene.

(c) Pie charts showing the proportions of genes annotated with LOC numbers, MSTRG numbers, or gene symbols in the raw count matrices. Three different gtf files are compared: the NCBI gtf, the NCBI+TruSeq gtf, and the NCBI+TruSeq gtf with updated gene annotation. The NCBI+TruSeq gtf file contains more genes than the NCBI gtf file due to the inclusion of newly identified transcripts following the TruSeq update. Most unassigned genes are mitochondrial tRNAs. See Source Data.

(d) Dot plot showing the expression patterns of canonical markers for retinal cell classes and non-neuronal cell classes, such as retinal pigmented epithelium, ependymal cells, astrocyte, etc. See Source Data.

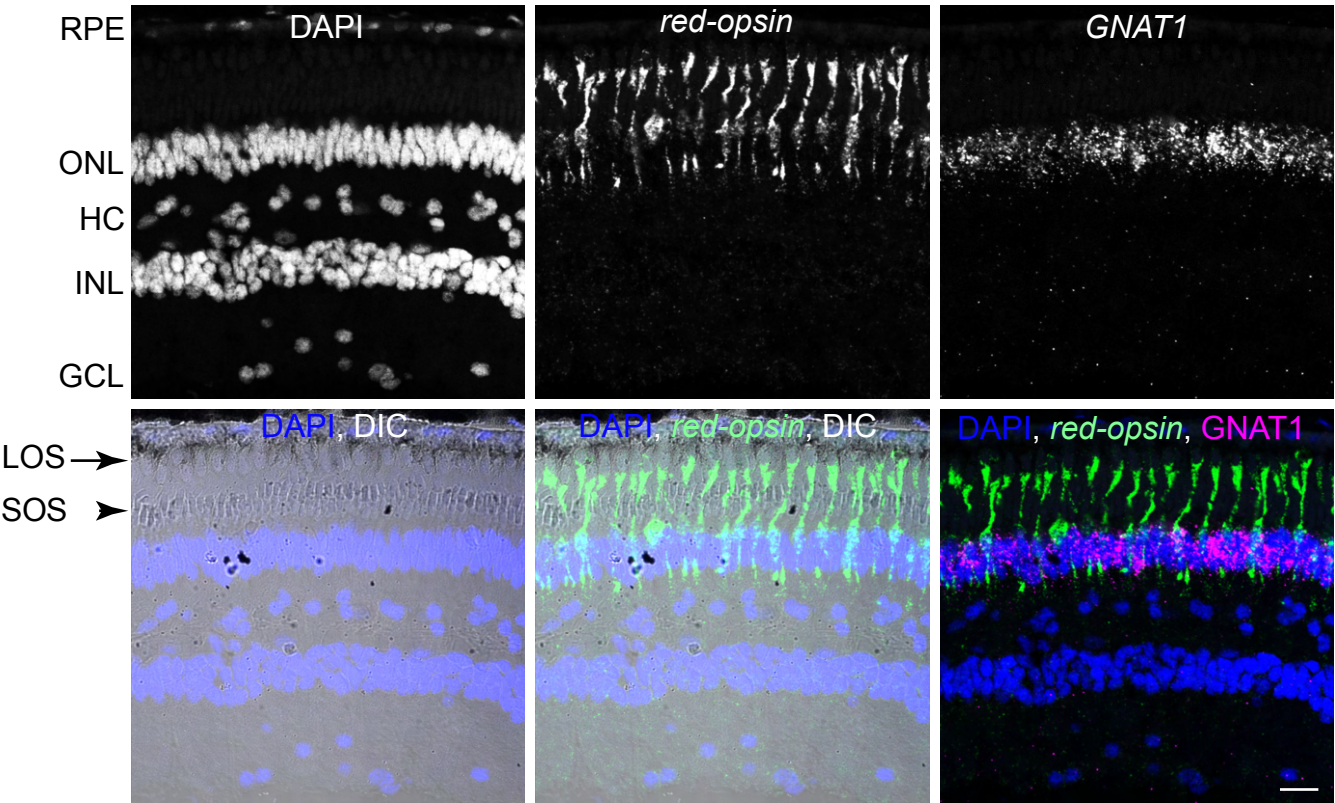
Figure S2



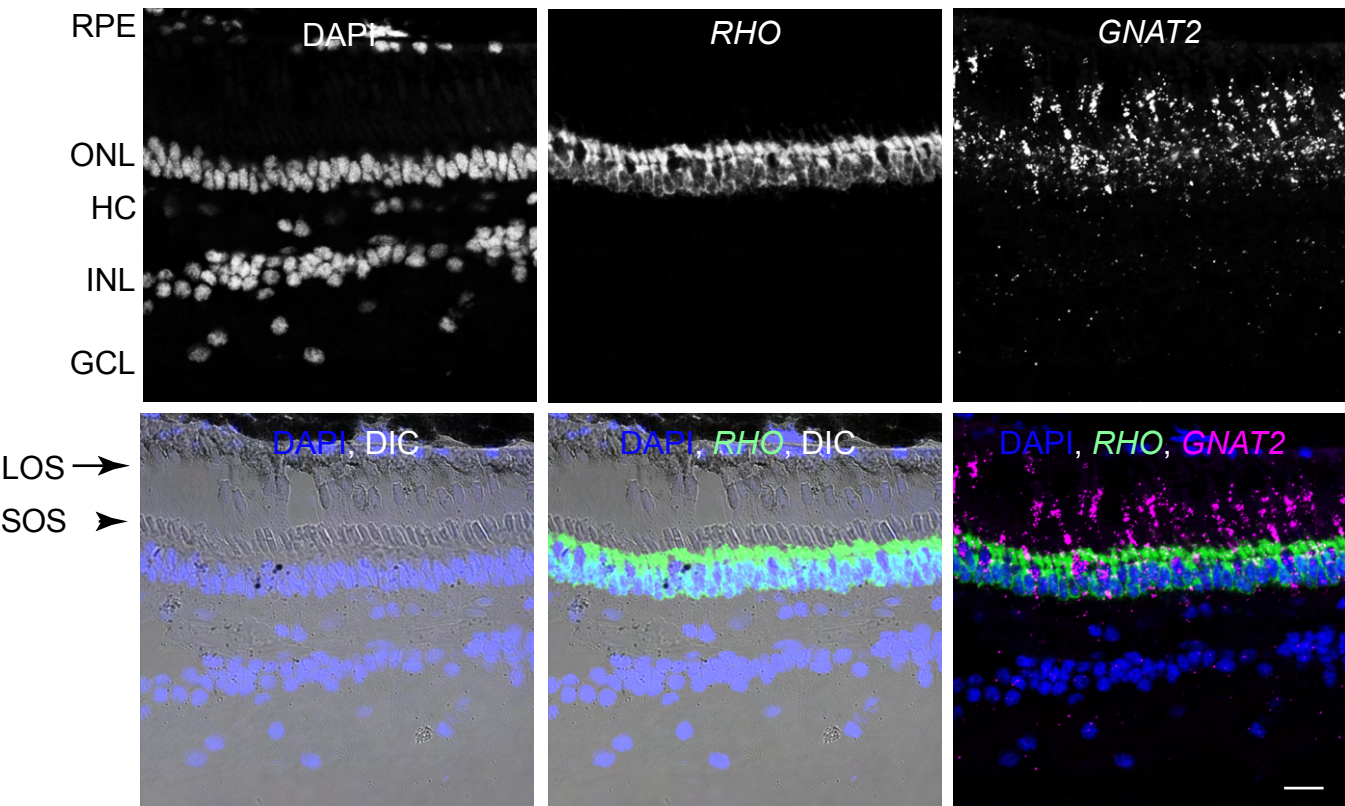
Supplementary Figure 2. Quality metrics of lamprey scRNA-seq data and cell type hierarchical relationships.

- (a) UMAP visualization of all lamprey cells, as depicted in Figure 1c, but here colored by distinct replicates to demonstrate lack of bias associated with different replicates.
- (b) Violin plots showing distributions of the number of expressed genes (nFeature_RNA), RNA counts, and percentages of mitochondrial genes (percent.mt) detected in each replicate.
- (c) Bar plot displaying fractions of cells from each replicate across individual cell types. See Source Data.
- (d) Hierarchical clustering and heatmap showing the Pearson correlation coefficients calculated for each pair of cell types, using the top 3,000 highly variable genes. See Source Data.

a



b

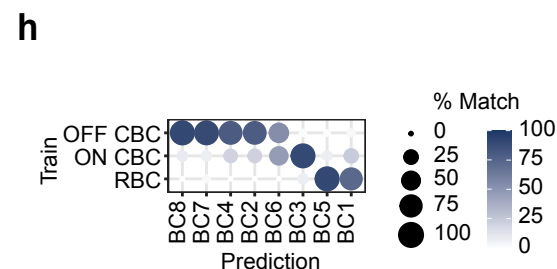
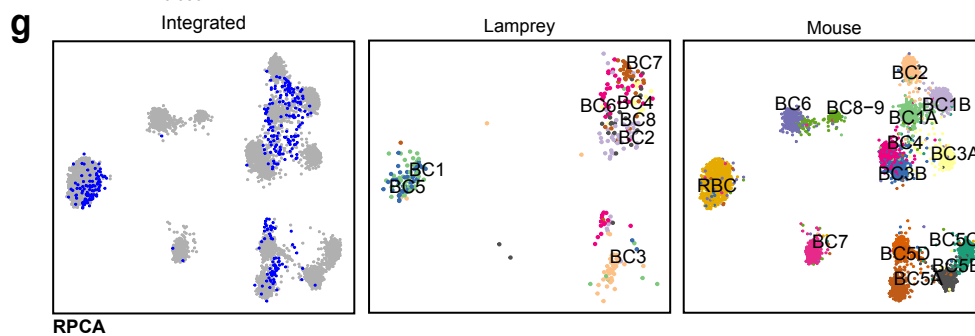
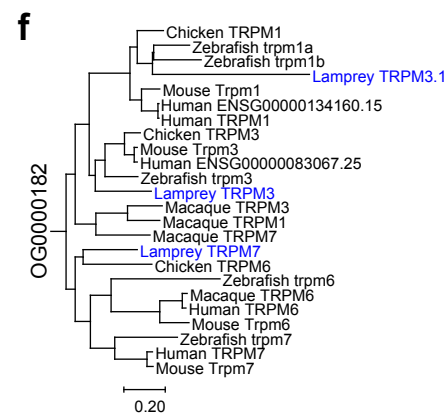
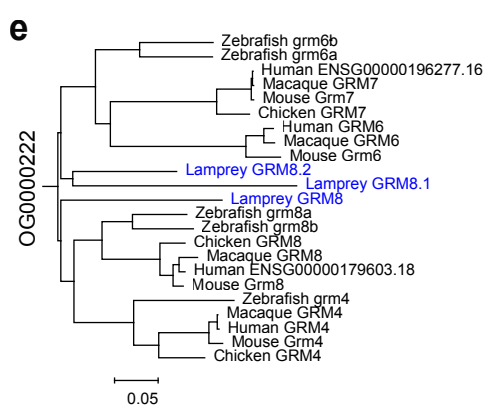
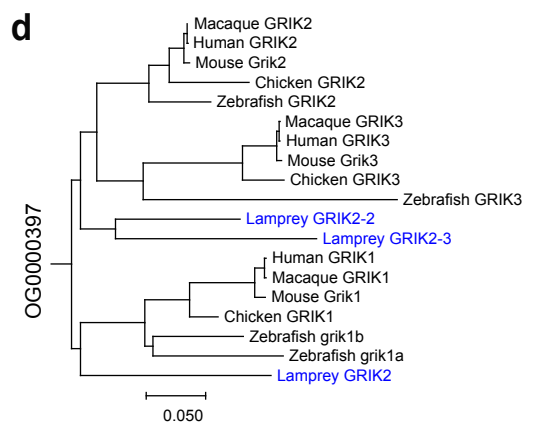
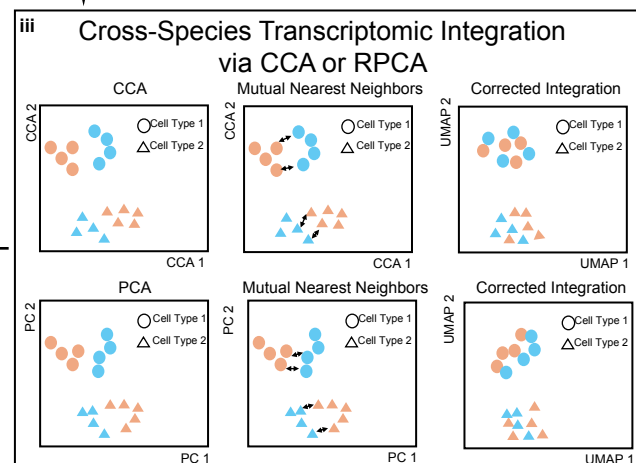
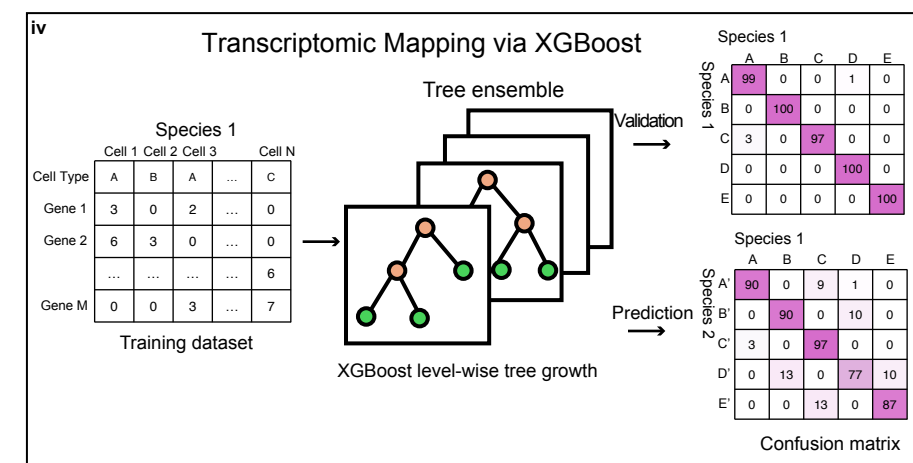
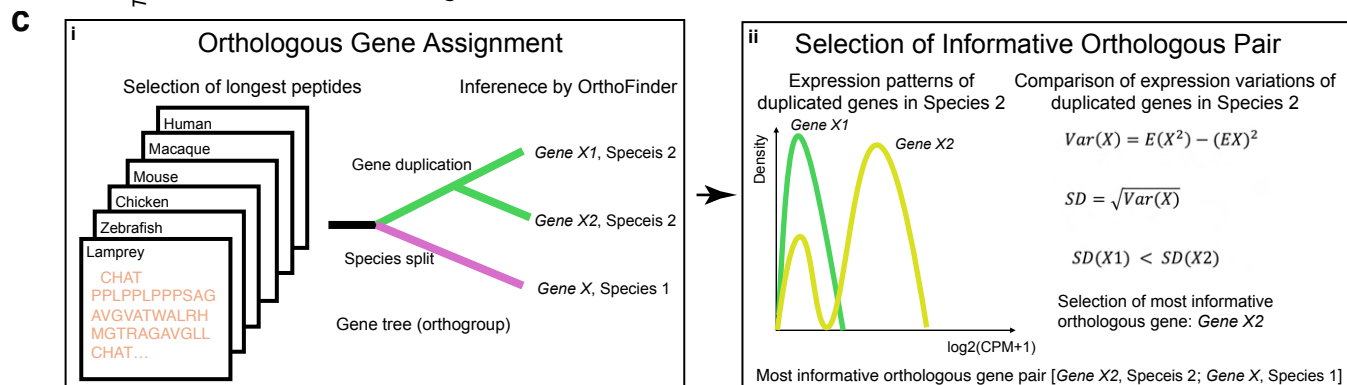
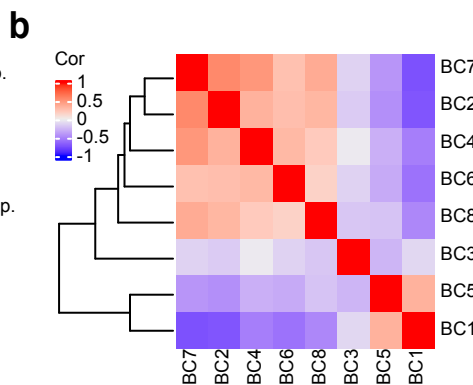
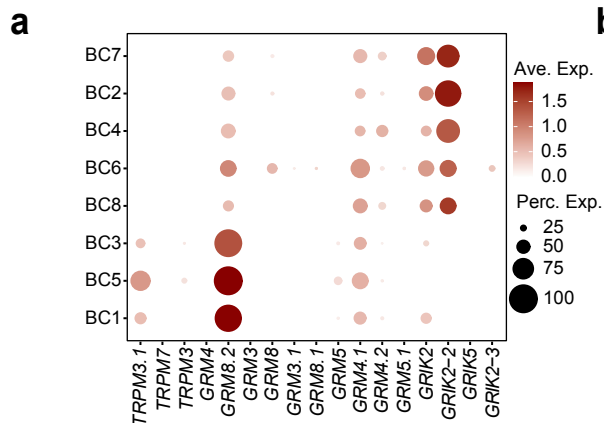


Supplementary Figure 3. Fluorescence in situ validation of marker genes for lamprey PR types

(a) Fluorescence in situ hybridization (FISH) validations confirming exclusive expression patterns of *red-opsin* and *GNAT1* in PR2 and PR1. *Red-opsin* (green in the merged image) is expressed by PRs with long outer segments (LOS, indicated by the arrow); while *GNAT1* (magenta in the merged image) is expressed by PRs with short outer segments (SOS, indicated by the arrowhead). The LOS and SOS structures are more clearly visualized in the differential interference contrast (DIC) image.

(b) FISH validation showing the exclusive expression of *rhodopsin* (*RHO*) and *GNAT2* in PR1 and PR2. *RHO* (green in the merged image) is expressed by PRs with short outer segments (SOS, arrowhead); while *GNAT2* (magenta in the merged image) is expressed by PRs with long outer segments (LOS, arrow).

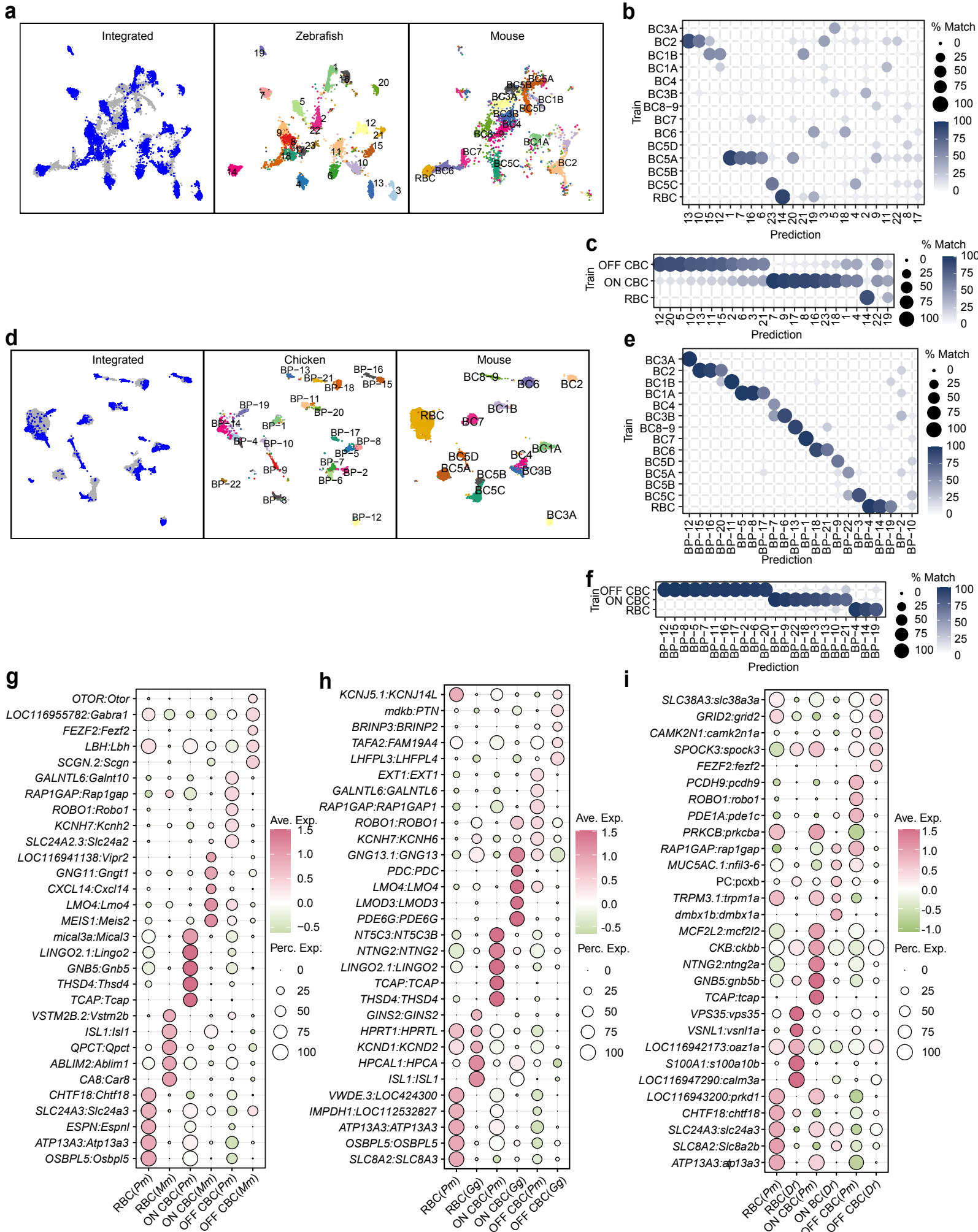
Nuclei stained with DAPI are in blue. Scale bar, 20 μm .



Supplementary Figure 4. Gene expression in lamprey BC types and cross-species integration of BC types.

- (a) Dot plot showing the expression patterns of genes from the *TRPM*, *GRM*, and *GRIK* families across lamprey BC types. See Source Data.
- (b) Hierarchical clustering and heatmap displaying Pearson correlation coefficients calculated between each pair of BC types using the top 3000 highly variable genes. The dendrogram on the left shows their hierarchical relationships, constructed from agglomerative hierarchical clustering based on correlation distance. See Source Data.
- (c) The analysis workflow for cross-species comparison of cell types. i) Identification of orthologous relationships of genes across lamprey, zebrafish, chicken, mouse, macaque, and human via OrthoFinder. ii) Identification of the most informative orthologous genes when multiple orthologous genes are present in the dataset. iii) Transcriptomic integration via CCA or RPCA. iv) Transcriptomic mapping via XGBoost.
- (d) Phylogenetic tree of ionotropic glutamate receptor genes among the selected five species, inferred from OrthoFinder. Scale bar refers to a phylogenetic distance of 0.05 substitutions per site. The orthogroup name is shown on the left.
- (e) Phylogenetic tree of metabolic glutamate receptor genes among the five species.
- (f) Phylogenetic tree of transient receptor potential TRPM genes among the five species.
- (g) Transcriptomic integration of lamprey and mouse BC types via RPCA.
- (h) Confusion matrix of the correspondence of lamprey BC types to mouse BC subclasses. Mouse BC subclasses were used as the training dataset. See Source Data.

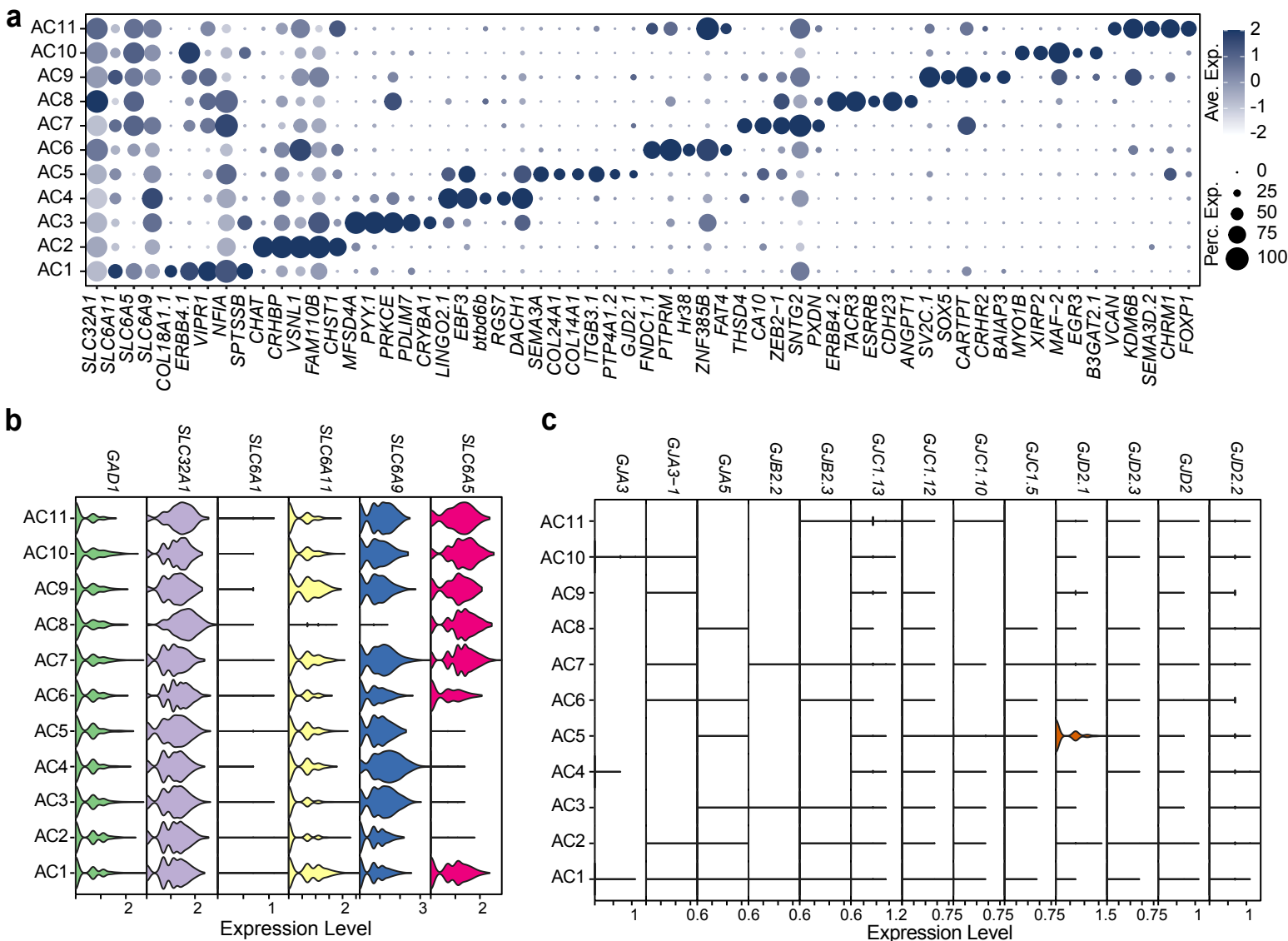
Figure S5



Supplementary Figure 5. Cross-species comparison of BC types.

- (a) Transcriptomic integration of zebrafish and mouse BC types via RPCA.
- (b) Confusion matrix of the correspondence of zebrafish BC types to mouse BC types. Mouse BC types were used as a training dataset. See Source Data.
- (c) Confusion matrix of the correspondence of zebrafish BC types to mouse BC subclasses. Mouse BC subclasses were used as a training dataset. See Source Data.
- (d) Transcriptomic integration of chicken and mouse BC types via RPCA.
- (e) Confusion matrix of the correspondence of chicken BC types to mouse BC types. Mouse BC types were used as a training dataset. See Source Data.
- (f) Confusion matrix of the correspondence of chicken BC types to mouse BC subclasses. Mouse BC subclasses were used as a training dataset. See Source Data.
- (g-i) Dot plots showing species-specific markers of BC subclasses between lamprey and mouse (g), lamprey and chicken (h), and lamprey and zebrafish (i). See Source Data.

Figure S6



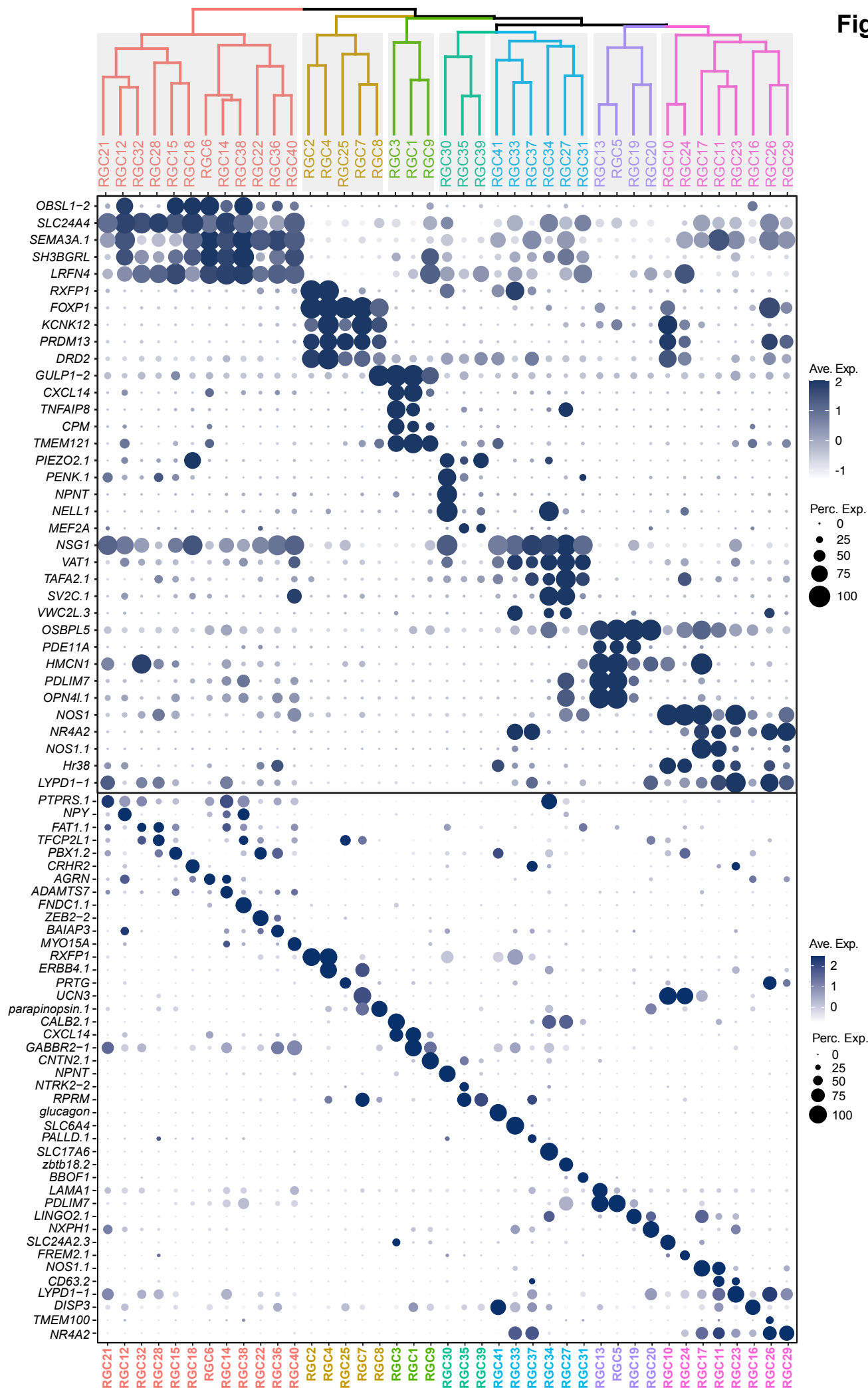
Supplementary Figure 6. Gene expression in lamprey AC types

(a) Dot plot showing the expression of genes for GABA transporters (*SLC32A1* and *SLC6A11*) and glycine transporters (*SLC6A5* and *SLC6A9*) together with marker genes for individual AC clusters. See Source Data.

(b) Stacked violin plot showing the expression patterns of all GABA and glycine transporters in lamprey AC types.

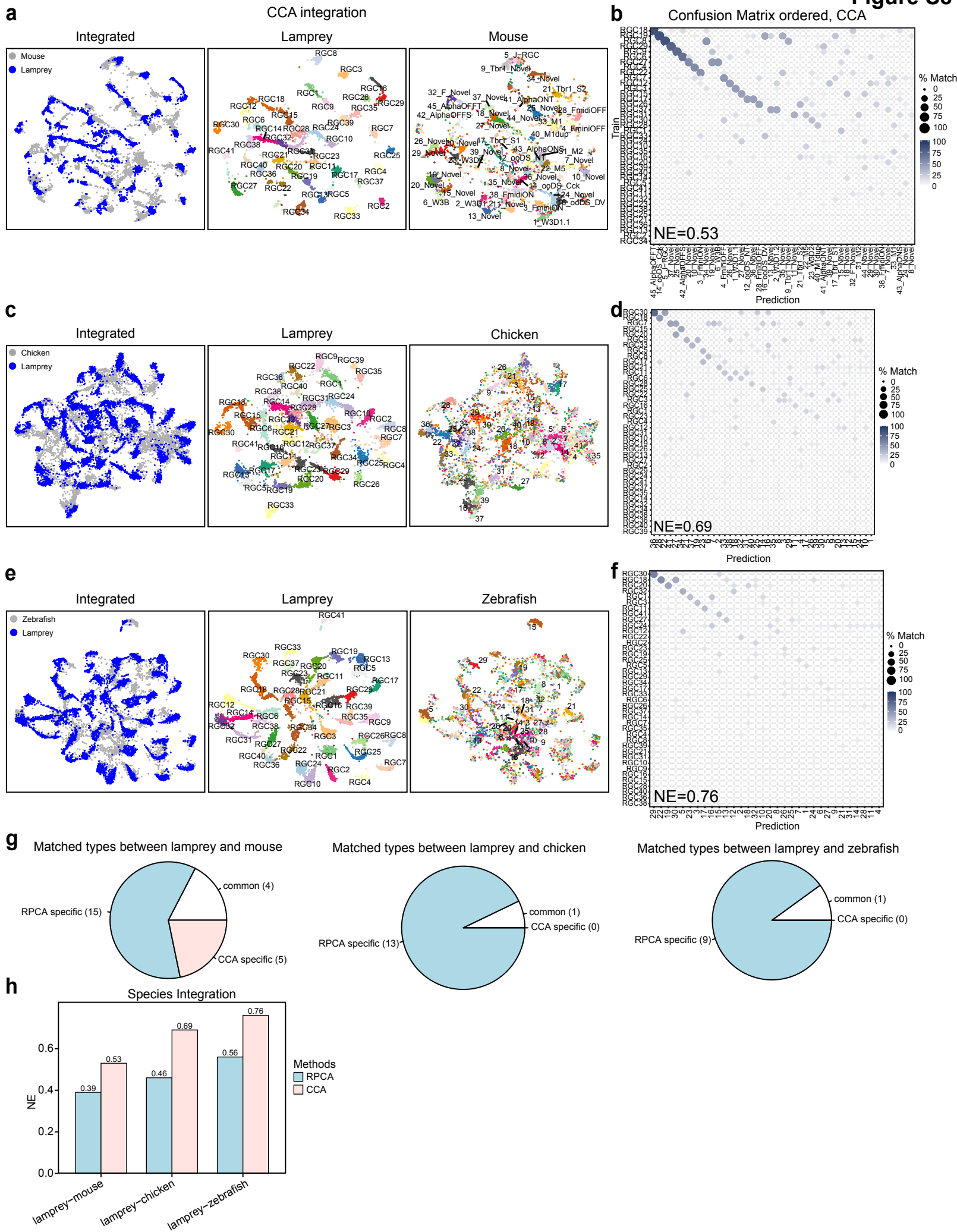
(c) Stacked violin plot showing the expression patterns of all gap junction genes in lamprey AC types.

Figure S7



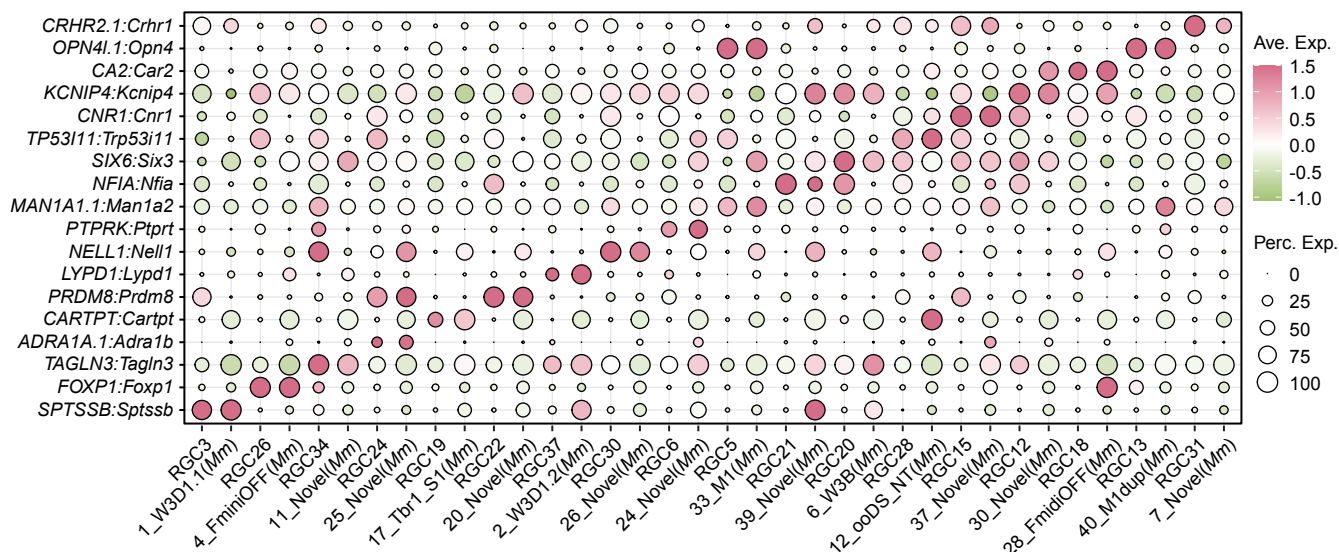
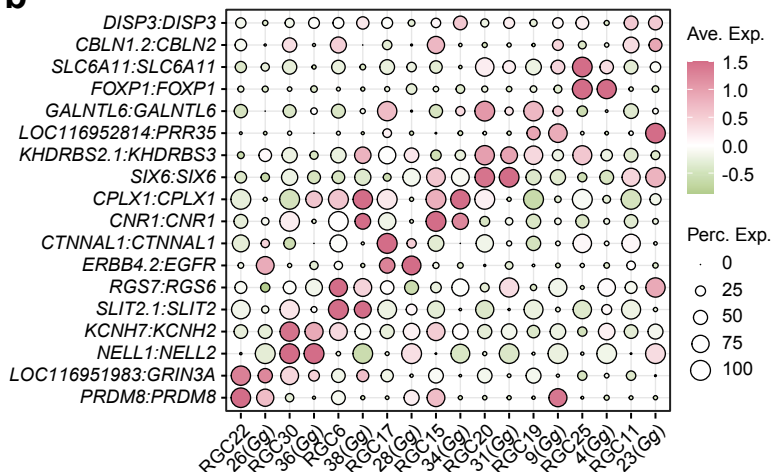
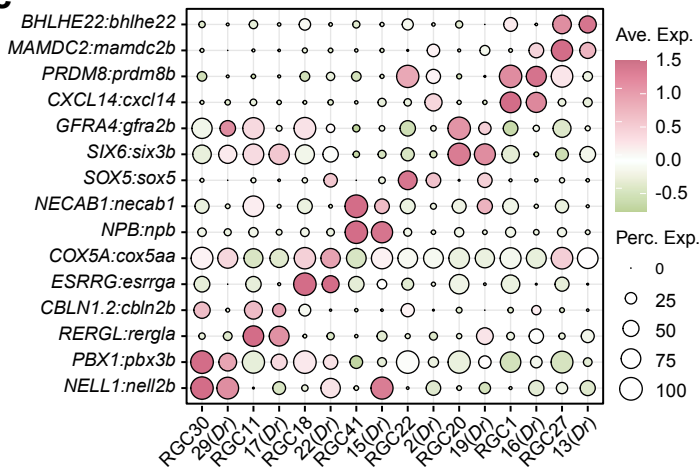
Supplementary Figure 7. Expression patterns of marker genes for RGC subgroups and types.

The dendrogram at the top shows the hierarchical relationships among RGC types, constructed from agglomerative hierarchical clustering based on correlation distance. RGC types are grouped into seven subgroups based on Pearson correlation coefficients calculated between each pair of RGC types from the top 3,000 highly variable genes. These seven RGC subgroups are visually distinguished by different colors and enclosed within shadow boxes. Subgroup-specific markers are shown in the top dot plot, while the expression patterns of markers for individual RGC types are presented in the bottom dot plot. See Source Data.



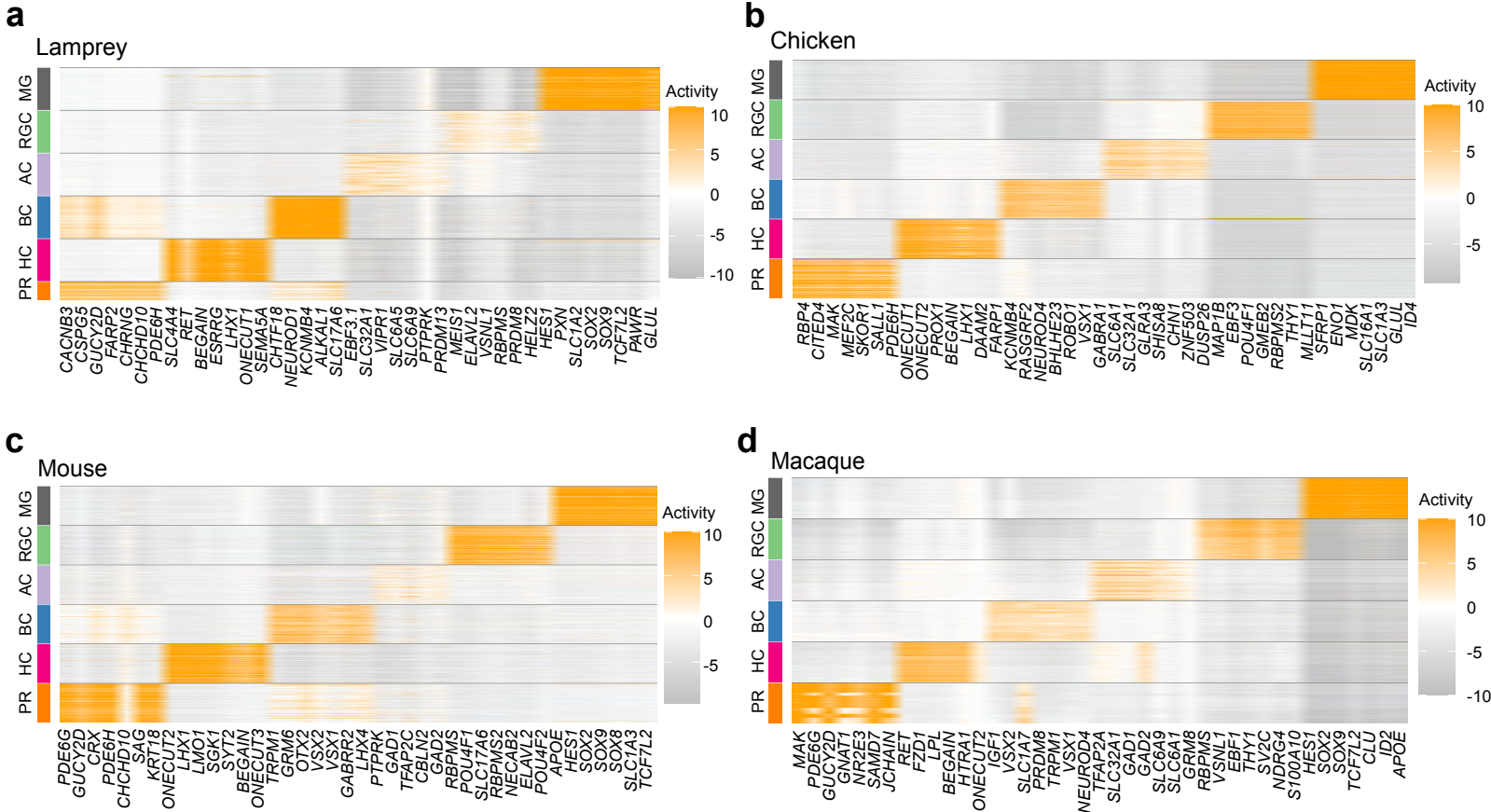
Supplementary Figure 8. Cross-species integration of RGC types.

- (a) Transcriptomic integration of lamprey and mouse RGC types via CCA, with both integrated and species-specific clusters presented in separated UMAP plots.
- (b) Confusion matrix of the correspondence of mouse RGC types to lamprey ones. Lamprey RGC types were used as a training dataset. NE, Normalized Entropy. See Source Data.
- (c) Transcriptomic integration of lamprey and chicken RGC types via CCA, with both integrated and species-specific clusters presented in separated UMAP plots.
- (d) Confusion matrix of the correspondence of chicken RGC types to lamprey ones. Lamprey RGC types were used as a training dataset. See Source Data.
- (e) Transcriptomic integration of lamprey and zebrafish RGC types via CCA, with both integrated and species-specific clusters presented in separated UMAP plots.
- (f) Confusion matrix of the correspondence of zebrafish RGC types to lamprey ones. Lamprey RGC types were used as a training dataset. See Source Data.
- (g) Pie charts showing the comparison of matched cell type numbers between CCA and RPCA methods. See Source Data.
- (h) Bar plot showing the comparison of NE values between CCA and RPCA. A lower NE value indicates higher prediction confidence. See Source Data.

a**b****c**

Supplementary Figure 9. Conserved markers showed conserved RGC types between species.

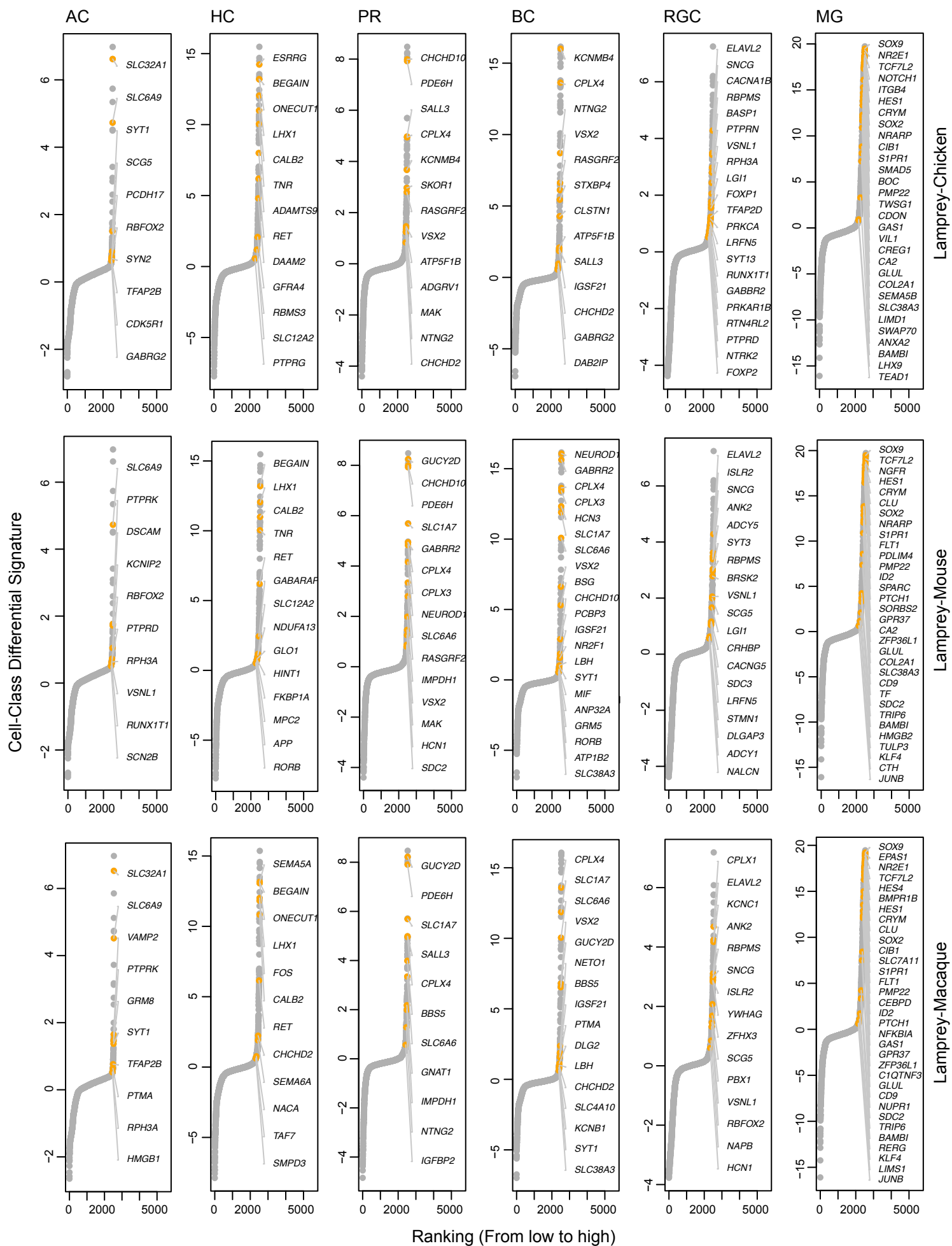
- (a) Dot plot showing conserved markers between matched lamprey and mouse RGC types.
 - (b) Dot plot showing conserved markers between matched lamprey and chicken RGC types.
 - (c) Dot plot showing conserved markers between matched lamprey and zebrafish RGC types.
- See Source Data.



Supplementary Figure 10. Identification of class-specific regulators in four vertebrate species.

- (a) Heatmap showing activities of class-specific regulators in lamprey.
- (b) Heatmap showing activities of class-specific regulators in chicken.
- (c) Heatmap showing activities of class-specific regulators in mouse.
- (d) Heatmap showing activities of class-specific regulators in macaque.

Figure S11



Supplementary Figure 11. Comparison of class-specific regulators between the lamprey and three jawed species.

Waterfall plots depicting all inferred proteins in the lamprey, ranked by their cell-class specificity. Top active regulators in the lamprey that are also activated in chicken (top), mouse (middle), and macaque (bottom) are highlighted in yellow and shown with the gene name.

Supplementary Table 1. Published Datasets Used in This Study

Dataset	GEO	Dataset URL	Usage	Reference
Mouse PR	GSE135406	https://github.com/jiewang/Single-cell-retinal-regeneration	Cross-Species Integration	Hoang et al. (<i>Ref 31</i>)
Mouse BC	GSE81905	https://singlecell.broadinstitute.org/single_cell/study/SCP3/retinal-bipolar-neuron-drop-seq	Cross-Species Integration	Shekhar et al. (<i>Ref 33</i>)
Chicken BC	GSE159107	https://singlecell.broadinstitute.org/single_cell/study/SCP1159/a-cell-atlas-of-the-chick-retina-based-on-single-cell-transcriptomics	Cross-Species Integration	Yamagata et al. (<i>Ref 19</i>)
Zebrafish BC	GSE237214	https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE237214	Cross-Species Integration	Hellevik et al. (<i>Ref 35</i>)
Chicken HC	GSE159107	https://singlecell.broadinstitute.org/single_cell/study/SCP1159/a-cell-atlas-of-the-chick-retina-based-on-single-cell-transcriptomics	Cross-Species Integration	Yamagata et al. (<i>Ref 19</i>)
Mouse SAC	GSE132555	https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSM3872341	Cross-Species Integration	Peng et al. (<i>Ref 30</i>)
Mouse AC	GSE149715	https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE149715	Cross-Species Integration	Yan et al. (<i>Ref 34</i>)
Chicken AC	GSE159107	https://singlecell.broadinstitute.org/single_cell/study/SCP1159/a-cell-atlas-of-the-chick-retina-based-on-single-cell-transcriptomics	Cross-Species Integration	Yamagata et al. (<i>Ref 19</i>)
Mouse RGC	GSE137400	https://singlecell.broadinstitute.org/single_cell/study/SCP509/mouse-retinal-ganglion-cell-adult-atlas-and-optic-nerve-crush-time-series	Cross-Species Integration	Tran et al. (<i>Ref 52</i>)

Chicken RGC	GSE159107	https://singlecell.broadinstitute.org/single_cell/study/SCP1159/a-cell-atlas-of-the-chick-retina-based-on-single-cell-transcriptomics	Cross-Species Integration	Yamagata et al. (Ref 19)
Zebrafish RGC	GSE152842	https://drive.google.com/drive/folders/1baRKtDkD4d8-6tG8P9v8VcjtUDpWeq5m	Cross-Species Integration	Kolsch et al. (Ref 74)
Mouse	GSE149715	https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE149715	Protein Activity Analysis	Yan et al. (Ref 34)
Chicken	GSE159107	https://singlecell.broadinstitute.org/single_cell/study/SCP1159/a-cell-atlas-of-the-chick-retina-based-on-single-cell-transcriptomics	Protein Activity Analysis	Yamagata et al. (Ref 19)
Macaque	GSE118480	https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE118480	Protein Activity Analysis	Peng et al. (Ref 30)

The Dataset URL links to the processed dataset if it is used.

For the raw dataset, we preprocessed the data following the Seurat guided clustering tutorial (https://satijalab.org/seurat/articles/pbm3k_tutorial).

Supplementary Table 2: The Information of RNA Probes

Target Genes	Primers (The T7 promoter sequences are placed in the reverse primer and indicated with lower cases)	Probe Length (bp)	Tag	Sequence Reference
<i>NEUROD1</i>	Forward: GGAAGTGCAGTCTCAACACG; reverse: gaaattaatacgaactcactatagggCCGCGTAGTGAAACCGATAG	1037	DIG	XM_032946356.1
<i>SLC32A1</i>	Forward: CAATTCTCTCGTCATCGGCTA; reverse: gaaattaatacgaactcactatagggACAGAGGCTGCATTGGAACAA	1074	DIG	XM_032952818
<i>GLUL</i>	Forward: GTTCGGGAAGCACTCTCTCC; reverse: gaaattaatacgaactcactatagggGCGGAGAACTCGTGTATGCT	1020	DIG	XM_032973232.1
<i>RHO</i>	Forward: TGAACGGCACAGAGGGAGAA; reverse: gaaattaatacgaactcactatagggGAGGCTCCCGAGTCTTCATC	1006	DIG Flu	XM_032971488.1
<i>Red-opsin</i>	Forward: GGCAGGGGGCGATGTTC; reverse: gaaattaatacgaactcactatagggCGTCGTCCACTTTCTTCCCA	1014	DIG Flu	XM_032971650.1
<i>GNAT1</i>	Forward: AACGTCAAGTTCGTGTTTCGAC; reverse: gaaattaatacgaactcactatagggCCCATCAGTTTCCCCAAACAC	1018	DIG Flu	XM_032954230
<i>GNAT2</i>	Forward: AGAAGAAGCTCGCCGAAGATG; reverse: gaaattaatacgaactcactatagggAGAGGCCGAGTCTTTGAGG	1002	DIG Flu	XM_032971636
<i>SLC17A6</i>	Forward: GCGAGCTGACTGCAATGTTT; reverse: gaaattaatacgaactcactatagggAGCACGTACCAACAGGACAG	1041	DIG	XM_032960051.1
<i>OPN4L1</i>	Forward: ATGGTCTCCCTCTCGCAGAT; reverse: gaaattaatacgaactcactatagggAGGATGGACTCCTCTCGTCC	1126	DIG	XM_032944209.1
<i>CHAT</i>	Forward: TCCCCCACTTCGAGTGTAGT; reverse: gaaattaatacgaactcactatagggTAAACCCCAAGCAATGCCCCA	1041	DIG	XM_032959092.1
<i>MEGF10</i>	Forward: TTCTGTGAGGAGGTTTGCCC; reverse: gaaattaatacgaactcactatagggTTCTCTGGCATGTGGTTCCC	1183	DIG	XM_032953473.1

DIG: Digoxigenin; Flu: Fluorescein

Supplementary Table 3. Parameters for Removing Low-quality Cells in Lamprey Cell Classes.

Class	nFeature_RNA	nCount_RNA
AC	>700 & <5000	>950 & <15000
BC	>300 & <7000	>500 & <32000
HC	>1500 & <7200	>2500 & <40000
MG	>330 & <9000	>500 & <61500
PR	>350 & <4350	>500 & <14800
RGC	>1000 & <6000	>1400 & <29500

Supplementary Table 4. Cross-species Integration Parameters and Hyperparameters

Integration	Integration method	Down-sample Parameter (# cells per type)	k. anchor
Lamprey-mouse PR (Fig. 2d)	CCA	NA	Default
Lamprey-mouse BC (Fig. 3f)	CCA	NA	Default
Lamprey-mouse BC (Supplementary Fig. 4g)	RPCA	Lamprey 50; Mouse nonRBC 300, RBC 800	10
Lamprey-mouse BC (Fig. 3h)	RPCA	Lamprey 50; Mouse nonRBC 300, RBC 1000	20
Lamprey-chicken BC (Fig. 3h)	RPCA	Lamprey 50; Chicken 200	20
Lamprey-zebrafish BC (Fig. 3h)	RPCA	Lamprey 300*; Zebrafish 200	20
Mouse-chicken BC (Supplementary Fig. 5d)	RPCA	Mouse NA; Chicken 200	20
Mouse-zebrafish BC (Supplementary Fig. 5a)	RPCA	Lamprey 300; Zebrafish 300	20
Lamprey-chicken HC (Fig. 4d)	CCA	NA	Default
Lamprey-mouse SAC (Fig. 4j)	CCA	NA	Default
Lamprey-mouse AC (Fig. 5a)	CCA	Lamprey NA; Mouse 200	Default
Lamprey-chicken AC (Fig. 5b)	CCA	Lamprey 100; Chicken 100	Default
Lamprey-mouse RGC (Fig. 6a)	RPCA	Lamprey 200; Mouse 200	20
Lamprey-chicken RGC (Fig. 6a)	RPCA	Lamprey 200; Chicken 200	20
Lamprey-zebrafish RGC (Fig. 6a)	RPCA	Lamprey 200; Zebrafish 200	20
Lamprey-mouse RGC (Supplementary Fig. 8a)	CCA	Lamprey 100; Mouse 100	Default
Lamprey-chicken RGC (Supplementary Fig. 8c)	CCA	Lamprey 200; Chicken 200	Default
Lamprey-zebrafish RGC (Supplementary Fig. 8e)	CCA	Lamprey 200; Zebrafish 200	Default

NA, Not Applicable. All the cells were used.

* The number of cells per subclass.

Supplementary Table 5. Downsampling Parameters for Generating the ARACNe-AP Network.

Species	Down-sampled Cell Number per Class per Sample	Number of Sample
Lamprey	NonRGC 200; RGC 400	2
Chicken	150	1
Mouse	100	1
Macaque	200	3