

AEG-1–AKT2: A novel complex controlling the aggressiveness of glioblastoma

Luni Emdad^{1,2,3,*}, Bin Hu¹, Swadesh K Das^{1,2,3}, Devanand Sarkar^{1,2,3}, and Paul B Fisher^{1,2,3,*}

¹Department of Human and Molecular Genetics; ²VCU Institute of Molecular Medicine; ³VCU Massey Cancer Center; Virginia Commonwealth University; School of Medicine; Richmond, VA USA

Keywords: AEG-1, AKT2, glioblastoma, MTDH

Abbreviations: AEG-1, astrocyte elevated gene 1; AKT, protein kinase B; GBM, glioblastoma multiforme; RTK, receptor tyrosine kinase; PI3K, phosphatidylinositol 3-kinase; NF- κ B, nuclear factor kappa-light-chain-enhancer of activated B cells.

Expression of AEG-1 (also known as MTDH or LYRIC) is elevated in many cancers including glioblastoma multiforme (GBM), in which it functions as an oncogene. AEG-1 activates AKT signaling and physically interacts with AKT2 in GBM. Disruption of this interaction reduces glioma cell survival and invasion, uncovering a novel potential target for development of an effective therapy against GBM.

Glioblastoma multiforme (GBM), graded by the World Health Organization (WHO) as a grade IV malignant glioma, is the most lethal form of glioma. Most patients who are diagnosed with GBM survive only 12 to 15 months.¹ Treatment options remain multimodal, including surgery, radiation, and chemotherapy, and impart only incremental changes in cognitive functions and survival.² Perhaps the most challenging complications in GBM therapy are a consequence of the highly invasive nature and complex heterogeneous molecular makeup of this cancer, which contribute to the aggressiveness of this disease. Over the past decade, many multitargeted therapies have been evaluated with only limited success. As a result, defining novel targets and developing targeted therapies remains a priority for this invariably fatal neoplasm.

Astrocyte-elevated gene-1 (*AEG-1*) was identified using subtraction hybridization from primary human fetal astrocytes as a human immunodeficiency virus (HIV)-1- and tumor necrosis factor (TNF)- α -inducible gene that is upregulated in the majority of cancers analyzed to date.³ Subsequent studies identified *AEG-1* as

metadherin (*MTDH*) in the mouse and lysine-rich CEACAM1 coisolated (*LYRIC*) in the rat.³ *AEG-1* promotes tumor growth via multiple oncogenic signaling pathways and regulates a wide array of key cellular processes involved in cancer progression including migration, invasion, angiogenesis, and metastasis. As a multifunctional protein, *AEG-1* interacts with a plethora of proteins in different cancer types and these defined interactions facilitate tumor progression.⁵

AKT, also known as protein kinase B, acts as a nexus-signaling molecule in the receptor tyrosine kinase/phosphatidylinositol 3-kinase (RTK/PI3K) pathway and functions as an essential regulator of cell proliferation, survival, migration, and invasion.⁶ The *AKT* gene family consists of 3 homologous isoforms, *AKT1*, *AKT2*, and *AKT3*. A recent study comprehensively analyzed the expression levels of all 3 AKT isoforms in malignant glioma.⁷ In high-grade gliomas both *AKT2* mRNA and protein levels were increased, whereas *AKT3* mRNA and protein levels were reduced. *AKT2* knockdown in glioma cell lines reduced colony formation and induced apoptosis; however, knockdown

of *AKT1* did not alter cell growth or promote apoptosis, suggesting that *AKT2* and *AKT3* may be the key regulators of GBM cell growth.⁷

Our previous studies demonstrated that *AEG-1* functions as a downstream target of *Ha-Ras*, and that induction of *AEG-1* was attenuated by treatment with the PI3K inhibitor LY294002 or overexpression of phosphatase and tensin homolog (PTEN).⁸ Promoter mapping suggested that *Ha-Ras* enhanced the binding of *c-Myc* to the E-box elements in the promoter region of *AEG-1* via the PI3K/AKT/*c-Myc* pathway, thereby regulating *AEG-1* transcription.⁸ In another study, *AEG-1* overexpression inhibited serum starvation-induced apoptosis by activating the *Ras* and PI3K-AKT signaling pathways.⁹ Subsequently, *AEG-1* regulation of AKT signaling pathways was shown to be critical in cell proliferation, invasion, apoptosis resistance, chemoresistance, and angiogenesis in multiple cancer contexts by several groups.⁵ However, the mechanism by which *AEG-1* regulates AKT remained unclear.

In our recent study,¹⁰ we demonstrated a novel specific interaction between *AEG-*

© Luni Emdad, Bin Hu, Swadesh K Das, Devanand Sarkar, and Paul B Fisher

*Correspondence to: Luni Emdad; Email: lemdad@vcu.edu; Paul B. Fisher; Email: pbfisher@vcu.edu

Submitted: 11/25/2014; Revised: 11/26/2014; Accepted: 11/27/2014

<http://dx.doi.org/10.4161/23723556.2014.995008>

This is an Open Access article distributed under the terms of the Creative Commons Attribution-Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The moral rights of the named author(s) have been asserted.

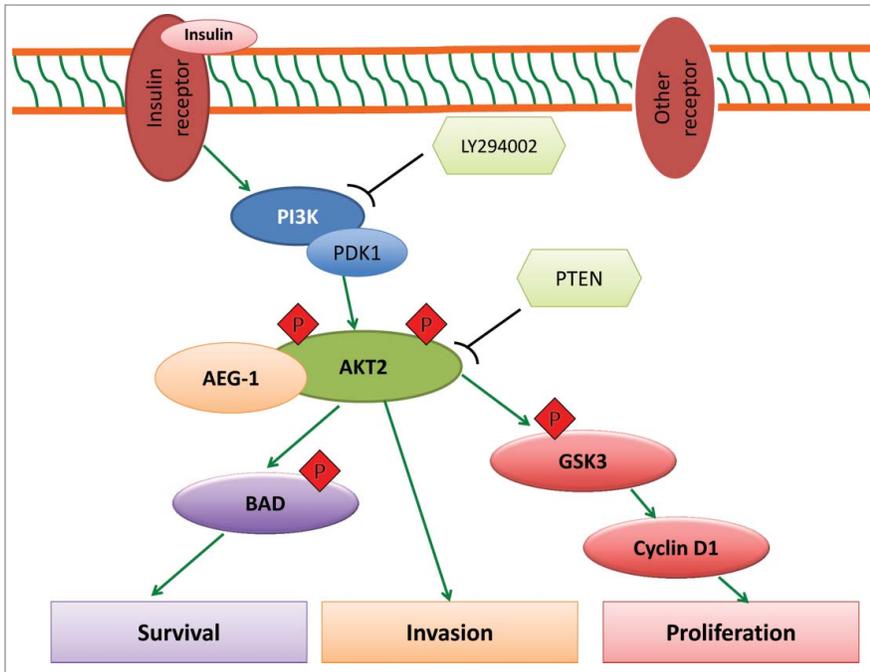


Figure 1. AEG-1–AKT2 signaling in glioma. Upon binding of insulin or other ligands to their respective receptors, PI3K, PDK1, or other kinases become activated and then phosphorylate AKT at T308 or S473/474. By forming a complex with AKT2, AEG-1 stabilizes the phosphorylation of AKT at S473/474. Phospho-AKT then activates downstream target genes and regulates a cascade of events implicated in tumor progression including unlimited cell proliferation, survival, and invasion. AEG-1, astrocyte elevated gene 1; BAD, Bcl-2-associated death promoter; GSK3, glycogen synthase kinase 3; PDK1, phosphoinositide-dependent kinase-1; PI3K, phosphatidylinositol 3-kinase; PTEN, phosphatase and tensin homolog; RTK, receptor tyrosine kinase.

1 and AKT2 in glioma. We showed that AEG-1 specifically interacts with AKT2, but not with AKT1 or AKT3 isoforms, in GBM. Immunohistochemical analysis revealed a high correlation between the expression of AEG-1 and AKT2 in clinical glioma samples. Using TCGA data analysis we also confirmed the clinical significance of these 2 key molecules for survival of glioma patients—higher expression was associated with poorer survival. Results from our functional assays indicated that disruption of the AEG-1–AKT2 interaction reduced AEG-1–mediated invasion. This observation was consistent with a previous report that AKT2 promotes breast cancer cell migration and invasion. Interestingly, we found that AEG-1–AKT2 interaction prolonged stabilization of AKT2 phosphorylation at S474. To explore the upstream regulators of AEG-1–AKT signaling, we examined AKT signaling upon insulin stimulation when

AEG-1 expression was knocked down with siRNA and found that AEG-1 knockdown effectively blocked insulin-induced activation of AKT2-S474. In contrast, AEG-1 overexpression maintained S474 phosphorylation when activation of PI3K–AKT was inhibited by PTEN and LY294002, and this was associated with parallel changes in the AKT downstream molecules p-glycogen synthase kinase 3 (p-GSK3 β) and cyclin D1. We also showed that phosphorylation of the proapoptotic protein Bcl-2-associated death promoter (BAD) was regulated by the AEG-1–AKT2 interaction, suggesting that the AEG-1–AKT2 complex protects glioma cells from the classic intrinsic mitochondrial apoptosis pathway. Together, these data highlight the importance of AEG-1–AKT2 interaction in glioma proliferation, survival, and invasion (Fig. 1).

AEG-1 was previously shown to crosstalk with several signaling pathways to regulate cancer cell proliferation and promote glioma survival.⁵ For that reason, we investigated the potential crosstalk between AEG-1–AKT2 and nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signaling using an I kappa B kinase (IKK) inhibitor, caffeic acid phenyl ester (CAPE). CAPE modestly affected AEG-1–AKT2 signaling, suggesting that NF- κ B activation by AEG-1 is independent of AKT activation. Finally, we showed that conditional expression of the AKT2-PH domain combined with AEG-1 silencing significantly enhanced survival in an orthotopic mouse model of human GBM. These findings further confirm that AEG-1–AKT2 is a critical protein–protein signaling complex in glioma. We are now exploring this interaction in other cancer contexts in which both of these molecules are highly upregulated.

We confirmed that the AKT2-PH domain is responsible for interacting with AEG-1 in glioma and mapped the interacting regions in AEG-1. This allowed us to define a target site for the development of candidate small-molecule drugs capable of disrupting this interaction. We predict that targeting AEG-1 or AKT2 alone will not be sufficient to engender a cure for this aggressive cancer. Rather, small-molecule inhibitors designed to disrupt AEG-1–AKT2 interactions in combination with conventional treatment modalities might represent a viable strategy to develop novel therapeutics for GBM.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed

Funding

This research was supported in part by the NIH/NCI grant R01 CA134721 and the National Foundation for Cancer Research. PBF holds the Thelma Newmeyer Corman Chair in Cancer Research in the MCC.

References

1. Ostrom QT, Gittleman H, Farah P, Ondracek A, Chen Y, Wolinsky Y, Stroup NE, Kruchko C, Barnholtz-Sloan JS. CBTRUS statistical report: Primary brain and central nervous system tumors diagnosed in the United States in 2006-2010. *Neuro Oncol* 2013; 15 Suppl 2:ii1-56; PMID:24137015; <http://dx.doi.org/10.1093/neuonc/not151>
2. Wen PY, Kesari S. Malignant gliomas in adults. *N Engl J Med* 2008; 359:492-507; PMID:18669428; <http://dx.doi.org/10.1056/NEJMra0708126>
3. Lee SG, Kang DC, DeSalle R, Sarkar D, Fisher PB. AEG-1/MTDH/LYRIC, the beginning: initial cloning, structure, expression profile, and regulation of expression. *Adv Cancer Res* 2013; 120:1-38; PMID:23889986; <http://dx.doi.org/10.1016/B978-0-12-401676-7.00001-2>
4. Sarkar D, Fisher PB. AEG-1/MTDH/LYRIC: clinical significance. *Adv Cancer Res* 2013; 120:39-74; PMID:23889987; <http://dx.doi.org/10.1016/B978-0-12-401676-7.00002-4>
5. Emdad L, Das SK, Dasgupta S, Hu B, Sarkar D, Fisher PB. AEG-1/MTDH/LYRIC: signaling pathways, downstream genes, interacting proteins, and regulation of tumor angiogenesis. *Adv Cancer Res* 2013; 120:75-111; PMID:23889988; <http://dx.doi.org/10.1016/B978-0-12-401676-7.00003-6>
6. Chautard E, Ouedraogo ZG, Biau J, Verrelle P. Role of Akt in human malignant glioma: from oncogenesis to tumor aggressiveness. *J Neuro Oncol* 2014; 117:205-15; PMID:24477623; <http://dx.doi.org/10.1007/s11060-014-1382-9>
7. Mure H, Matsuzaki K, Kitazato KT, Mizobuchi Y, Kuwayama K, Kageji T, Nagahiro S. Akt2 and Akt3 play a pivotal role in malignant gliomas. *Neuro Oncol* 2010; 12:221-32; PMID:20167810; <http://dx.doi.org/10.1093/neuonc/nop026>
8. Lee SG, Su ZZ, Emdad L, Sarkar D, Fisher PB. Astrocyte elevated gene-1 (AEG-1) is a target gene of oncogenic Ha-ras requiring phosphatidylinositol 3-kinase and c-Myc. *Proc Natl Acad Sci U S A* 2006; 103:17390-5; PMID:17088530
9. Lee SG, Su ZZ, Emdad L, Sarkar D, Franke TF, Fisher PB. Astrocyte elevated gene-1 activates cell survival pathways through PI3K-Akt signaling. *Oncogene* 2008; 27:1114-21; PMID:17704808; <http://dx.doi.org/10.1038/sj.onc.1210713>
10. Hu B, Emdad L, Bacolod MD, Kegelman TP, Shen XN, Alzubi MA, Das SK, Sarkar D, Fisher PB. Astrocyte elevated gene-1 (AEG-1) interacts with Akt isoform 2 to control glioma growth, survival and pathogenesis. *Cancer Res* 2014; 74:7321-32; PMID:25304263; <http://dx.doi.org/10.1158/0008-5472.CAN-13-2978>