

RESEARCH ARTICLE

# Automatic microarray image segmentation with clustering-based algorithms

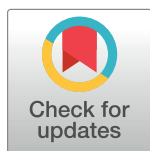
Guifang Shao<sup>\*</sup>, Dongyao Li, Junfa Zhang<sup>†</sup>, Jianbo Yang, Yali Shangguan

Department of Automation, Xiamen University, Xiamen, China

\* [gfshao@xmu.edu.cn](mailto:gfshao@xmu.edu.cn)

## Abstract

Image segmentation, as a key step of microarray image processing, is crucial for obtaining the spot expressions simultaneously. However, state-of-art clustering-based segmentation algorithms are sensitive to noises. To solve this problem and improve the segmentation accuracy, in this article, several improvements are introduced into the fast and simple clustering methods (K-means and Fuzzy C means). Firstly, a contrast enhancement algorithm is implemented in image preprocessing to improve the gridding precision. Secondly, the data-driven means are proposed for cluster center initialization, instead of usual random setting. The third improvement is that the multi features, including intensity features, spatial features, and shape features, are implemented in feature selection to replace the sole pixel intensity feature used in the traditional clustering-based methods to avoid taking noises as spot pixels. Moreover, the principal component analysis is adopted for various feature extraction. Finally, an adaptive adjustment algorithm is proposed based on data mining and learning for further dealing with the missing spots or low contrast spots. Experiments on real and simulation data sets indicate that the proposed improvements made our proposed method obtains higher segmented precision than the traditional K-means and Fuzzy C means clustering methods.



## OPEN ACCESS

**Citation:** Shao G, Li D, Zhang J, Yang J, Shangguan Y (2019) Automatic microarray image segmentation with clustering-based algorithms. PLoS ONE 14(1): e0210075. <https://doi.org/10.1371/journal.pone.0210075>

**Editor:** Zhaoqing Pan, Nanjing University of Information Science and Technology, CHINA

**Received:** June 21, 2018

**Accepted:** November 20, 2018

**Published:** January 22, 2019

**Copyright:** © 2019 Shao et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Funding:** This work was supported by the National Natural Science Foundation of China with grant no.61403318 to G.S. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

## Introduction

Microarray is a kind of useful biotechnological tool and has been widely applied in the field of life sciences, such as cancer research, pharmacology, disease diagnosis, and environmental engineering [1]. Microarray technology has the advantage of allowing scientists to measure the expression levels of thousands of genes simultaneously. It involves sample preparing, microarray designing, scanning, image processing, and data analyzing [2, 3]. Among them, image processing plays an important role in extracting the gene expressions. Microarray image processing is mainly included by four parts: 1) pre-processing, 2) gridding, 3) segmentation, and 4) intensity extraction [3]. Pre-processing is mainly aimed at reducing the noises and enhancing the image quality. Gridding is implemented to find out a series of horizontal and vertical lines so as to separate one slide image into sub-grids and individual spots areas. The procedure of segmentation divides the spot pixels into foreground, background, or noise. Finally, the intensity extraction is aimed at obtaining the gene expression levels according to

the previous operational results. During the four procedures, the segmentation is quite crucial for accurately extracting gene expressions. However, it is a challenging task because the real microarray image usually contains noises, poor contrast and various spot shapes.

(In the past decades, lots of state-of-art tools, such as GenePix [4], Imagen [5], QuantArray [6], ScanAlyze [7], Magic Tool [8], Spot [9], Dapple [10], Spotfinder [11], P-Scan [12], UCSF Spot [13], have been proposed for microarray image processing. They are originally manual and now semi-automatic. In addition, since 1997, microarray image segmentation has attracted increasing attention due to the fact that it is an essential step for extracting gene expression values, and many segmentation algorithms have been proposed for microarray image segmentation. These segmentation methods can be classified into the following seven categories:

1. Shape-based segmentation, including fixed circle or adaptive circle, adaptive shape [14], active contours [15–17] and Snake Fisher model [18]. It can obtain the boundary and region information of spots based on the relativity of target shape;
2. Model-based segmentation, involving Markov Random Filed [19], 3D spot modeling [20] and total variation (TV)-based regularization method [21]. It segments spots by building spot model or modeling the image as a function according to a series of parameter estimation;
3. Region-based segmentation, containing seeded region growing (SRG) algorithm [22] and watershed algorithm [23]. The spots is segmented by splitting image into areas depend on the image topology;
4. Threshold-based segmentation, including global and local threshold [24], soft-thresholding [25]. This method separates foreground and background based on fluorescence intensities.
5. Morphology-based segmentation, involving mathematical morphological method [26–27]. It segments spots using morphological operations with various structuring element.
6. Supervised learning-based segmentation, containing neural networks [28] and support vector machines [29]. These methods segment the spots depended on prepared training dataset.
7. Unsupervised learning-based segmentation, including K-means clustering [30–31], improved K-means clustering [32–34], Fuzzy C means clustering [35–39], and other clustering methods, such as expectation-maximization [40], model-base clustering [41], combined clustering [3,42], genetic algorithm combined clustering[43]. This kind of methods segment spots based on the natural relationship between intensities.

However, among these methods, the clustering-based methods are prone to be affected by noises. Meanwhile, real microarray images usually contain poor quality problem such as low contrast, noises, artifacts, shape-varied spots, and so on. All these deficiencies make the clustering-based segmentation become a challenging task. Therefore, in this article, an improved clustering-based scheme is proposed to improve the segmentation efficiency. Firstly, an improvement of automatic contrast enhancement and noise reduction is introduced into the image preprocessing. Subsequently, an initial clustering center design scheme is proposed for improving the clustering performance instead of random points selecting. Because the single feature of intensity cannot reflect all characters of spots, the factors of spatial, intensity and shape features are considered. In addition, the principal component analysis (PCA) is implemented for feature selection. Finally, an adaptive adjustment strategy is adopted for improving the segmentation accuracy. This article is organized as follows. Section 2 introduces the

improved clustering-based method. Section 3 presents the comparison experiments on different data sets. The main conclusions are described in Section 4.

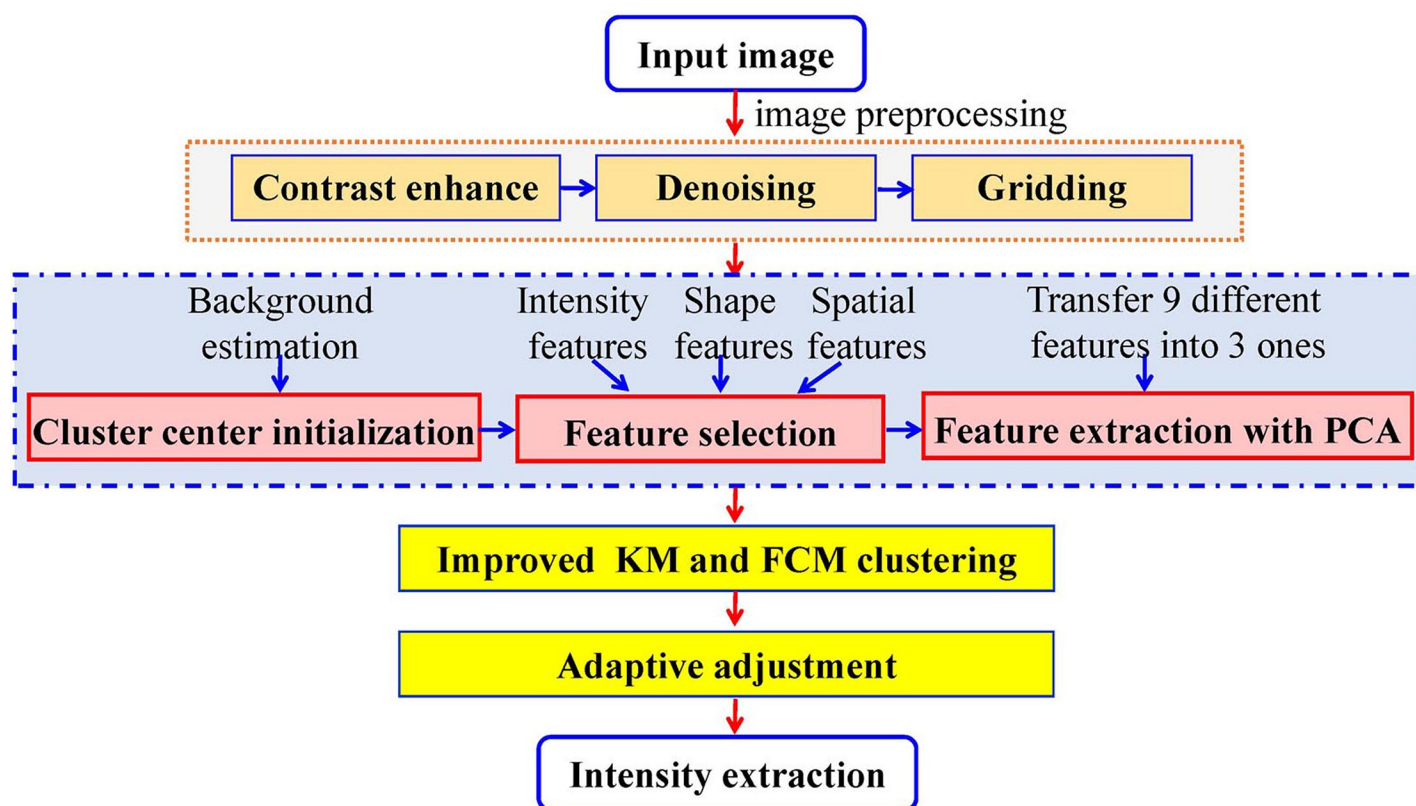
## The improved clustering-based method

As aforementioned, a great number of clustering methods have been proposed for cDNA microarray image segmentation. However, only a portion of them perform well on simulation images. In addition, single intensity feature is adopted for clustering in almost all the algorithms. To improve the segmentation accuracy and take full advantage of traditional clustering method, we improved the KM and FCM clustering algorithms. The proposed method (online code is in [S1 Algorithm](#)) includes six parts: 1) image preprocessing, 2) cluster center initialization, 3) feature selection, 4) feature extraction with PCA, 5) improved KM and FCM clustering, 6) adaptive adjustment, as shown in [Fig 1](#).

### Image preprocessing

To enhance the image quality, the contrast enhancement and gridding are conducted during the image preprocessing step. Our previous method has been adopted for contrast enhancement [3], and it can be executed according to the following steps:

1. Computing image contrast by using the four-order moment  $C = Sd/[Fom/Mse^2]^{1/4}$ , in which  $Sd = \sqrt{\frac{1}{N}(\sum (p - \bar{p})^2)}$  denotes the standard deviation,  $Mse = \frac{1}{N}\sum (p - \bar{p})^2$  represents the mean square error,  $Fom = \frac{1}{N}\sum (p - \bar{p})^4$  is the four-order moment and  $\bar{p} = \frac{1}{N}\sum p$



**Fig 1. Flowchart of improved clustering-based image segmentation method.**

<https://doi.org/10.1371/journal.pone.0210075.g001>

is the mean value of the image,  $N$  is the number of image pixels. For example, the cDNA microarray images drawn from GEO data set are generally low contrast, and their  $C$  values are all under 1000.

2. Estimating background gray value with the proposed method, that is  $k = \frac{1}{m} \sum_{i=0}^m \min \{ \max_j A_j \} (j \in [1, 12])$ , in which  $m$  is the repeated experiments times,  $A_j$  is a  $10^*10$  window randomly selected from four borders of the image.

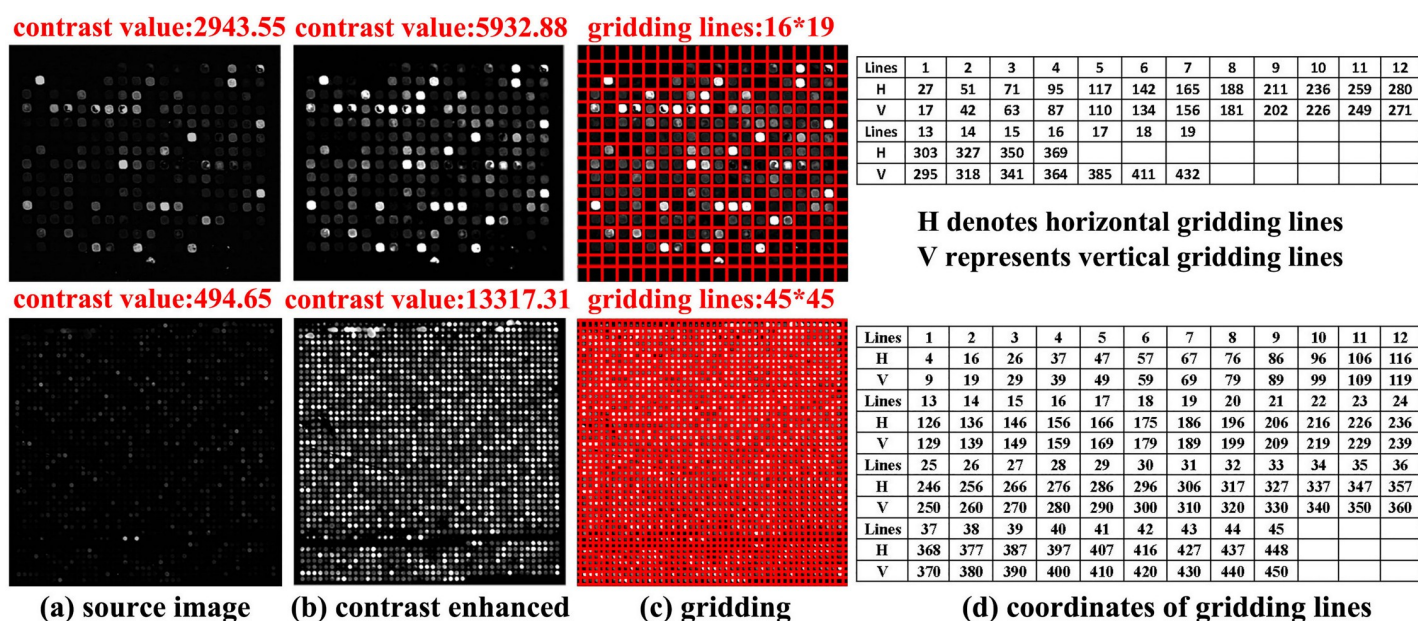
3. Enhancing the image contrast by using the following operation

$$g(x, y) = \begin{cases} f(x, y) \times (10000/C) & f(x, y) \leq k \\ f(x, y) & otherwise \end{cases}$$

After contrasting the enhancement operation, a median filter is applied for noise reduction. In addition, the maximum between-class variance based gridding is conducted according to the following steps [3]:

1. Computing the horizontal or vertical projection signal;
2. Filtering the projection signal by morphological reconstruction;
3. Finding the optimal threshold by maximum between-class variation operation;
4. Obtaining the horizontal or vertical grid lines according to the thresholding;
5. Revising the gridding result based on the heuristic rules;
6. Acquiring the final coordinate vector  $H$  and  $V$  for all grid lines, meanwhile obtaining the horizontal grid line number  $h$  and the vertical grid line number  $v$ .

Examples of image preprocessing are shown in Fig 2. It can be seen from Fig 2 that the image contrast is greatly enhanced and the correct gridding results are obtained.



**Fig 2.** Examples of image preprocessing with (a) source image, (b) contrast enhanced image, (c) gridding image, and (d) coordinates of gridding lines.

<https://doi.org/10.1371/journal.pone.0210075.g002>



## Cluster center initialization

The class number and the initial cluster center are two vital factors for obtaining good clustering performance. Here we set the number of clustering classes to 2. For the initial cluster center of one group, we put forward to adopt the background gray value  $k$  estimated in the image pre-processing step instead of random initialization. As for another group, the gray mean of a  $3 \times 3$  window in each spot center is selected as the initial cluster center.

## Feature selection

Generally, the pixel intensity features are adopted by various clustering algorithms and the classification is realized according to the Euclidian distance between the pixel and cluster center. If the intensity of a pixel is high, it will have a great probability to belong to the spot and vice versa. However, there exists the missing classification situation since the noises may contain higher intensities. Therefore, here we implemented other features, including intensity features, spatial features and shape feature, as shown in Table 1. To be specific, the intensity features are composed of the pixel intensity, the mean intensity, the intensity standard deviation and the entropy [29]. The spatial features include the coordinates of each pixel, the Euclidean distance and the city block distance between the pixel and the clustering center. In addition, to describe the similarity between the pixels centered area and the theoretical spot, the shape feature which computes the correlation coefficient is adopted. In the shape feature, the neighborhood size of

$$\text{pixel and the size of Gaussian template is } d = \frac{1}{2} \left( \frac{1}{v} \sum_{i=1}^v (V_{i+1} - V_i) + \frac{1}{h} \sum_{j=1}^h (H_{j+1} - H_j) \right),$$

obtained from gridding step,  $H$  and  $V$  denotes the coordinates of horizontal and vertical gridding lines, respectively.

According to the definition above, a feature vector  $F = [r, i, j, D_M, D_{Eud}, I(i, j), \bar{I}(i, j), \delta, E]^T$  of each pixel can be obtained. However, how to use these different features is quite important and no literatures discussed the multi-features fusion scheme for microarray image segmentation to date. Therefore, we deal with these features by using PCA which transfers a set of correlation variables into another irrelevance one by linear transformation. According to the

Table 1. Features used in our approach.

Type	Form	Description
Shape feature	$r$	$r = \frac{\sum_i \sum_j (I(i, j) - \bar{I})(T(i, j) - \bar{T})}{\sqrt{(\sum_i \sum_j (I(i, j) - \bar{I})^2)(\sum_i \sum_j (T(i, j) - \bar{T})^2)}}, T(i, j)$ is a Gaussian template with variable size of $d \times d$ , $d$ represents the estimated spot diameter
Spatial features	$i$	Row of the pixel
	$j$	Column of the pixel
	$D_M =  i - i_c  +  j - j_c $	City block distance of each pixel to clustering center, $i_c, j_c$ are row and column of clustering center
	$D_{Eud} = \sqrt{(i - i_c)^2 + (j - j_c)^2}$	Euclidean distance of each pixel to clustering center
Intensity features	$I(i, j)$	Intensity of pixel $(i, j)$
	$\bar{I}(i, j) = (1/9) \sum_{i=1}^3 \sum_{j=1}^3 I(i, j)$	Mean intensity value of the $3 \times 3$ pixel centered window
	$\delta = \sqrt{\frac{1}{9} \sum_{i=1}^3 \sum_{j=1}^3 (I(i, j) - \bar{I})^2}$	Intensity standard deviation of the $3 \times 3$ pixel centered window
	$E = \sum_{m=0}^{I_{\max}} p_m \log_2(1/p_m)$	Entropy of a $3 \times 3$ pixel centered window, $p_m$ is histogram counts, $I_{\max}$ is maximum gray value of the image

<https://doi.org/10.1371/journal.pone.0210075.t001>

covariance eigenvalue output from PCA operation, the top three transformed principal components are selected. Finally, the Euclidean distance is adopted for evaluating the relationship between the clustering center and each pixel.

### Improved KM and FCM clustering

As KM clustering is a kind of general algorithm, therefore, we mainly introduce the operations of FCM clustering here. There are four main steps for conducting FCM clustering.

Step1: Initializing the membership grade function according to  $\sum_{i=1}^c u_{ij} = 1, \forall j = 1, \dots, N$ .

$u_{ij}$  represents the membership degree of pixel  $j$  to cluster  $i$ .

Step 2: Updating the membership values for each pixel

$$u_{ij} = 1 / \left( \sum_{k=1}^c (d_{ij} / d_{kj})^{2/(m-1)} \right). \quad (1)$$

Where  $d_{ij} = \|x_j - c_i\|$  and  $d_{kj} = \|x_j - c_k\|$  denote the Euclidean distance between the feature vectors of pixel  $j$  to cluster  $i$  and  $k$ .

Step 3: Updating the cluster centroid by

$$c_i = \left( \sum_{j=1}^N u_{ij}^m x_j \right) / \left( \sum_{j=1}^N u_{ij}^m \right), \quad (2)$$

$m$  is the fuzziness parameter.

Step 4: Computing the cost function by the following equation

$$J(U, c_1, \dots, c_c) = \sum_{i=1}^c J_i = \sum_{i=1}^c \sum_j^N u_{ij}^m d_{ij}^2. \quad (3)$$

Step 5: Repeating step 2–4 until the cost function is minimized.

The detail Pseudo of the proposed FCM algorithm can be seen as following.

#### Algorithm 1

The pseudo code of improved FCM algorithm

**Input:**  $c$ : number of clusters. Max: maximum number of iterations.

$X$ : data matrix with selected features. tol: termination threshold.

$U$ : membership grade function.  $m$ : the fuzzy coefficient.

**Output:** Predicted labels of data and cluster centers.

Initializing the cluster centers;

**for**  $j \leftarrow 1$  **to**  $N$  **do**

    Normalizing  $U$ ;

**end**

$C_{old} \leftarrow C$ ;

**for**  $k \leftarrow 1$  **to** Max **do**

**for**  $i \leftarrow 1$  **to**  $c$  **do**

**for**  $j \leftarrow 1$  **to**  $N$  **do**

            update membership grade function  $U$  by Eq (1);

**end**

**end**

**for**  $i \leftarrow 1$  **to**  $c$  **do**

        update the cluster centers  $c_{new}$  based on Eq (2);

**end**

```

compute  $\delta \leftarrow \max(|C_{new} - C_{old}|)$ ;
 $C_{old} \leftarrow C_{new}$ ;
if  $\delta < tol$ 
    quit;
end
end
predicted labels  $\leftarrow U$ ;
return;

```

## Adaptive adjustment

After clustering by using the method of IKM and IFCM, there may exist some situations as shown in Fig 3. On one hand, the segmented spots may be surrounded by noises, for example, the top line shown in Fig 3. On the other hand, the missing spots appeared with different situations, as the bottom line depicted in Fig 3.

To avoid the deficiencies described above, an adaptive adjustment is a crucial step. Therefore, we put forward a method of noise removal and missing spot estimation based on the spot size. Firstly, an approximate spot size is estimated according to the microarray image data. To realize this, we utilize the boundary lines information computed in the gridding step [3]. Let  $BH^0 = [H_1^0, \dots, H_{bh}^0]$  and  $BV^0 = [V_1^0, \dots, V_{bv}^0]$  represent the boundary grid lines coordinates on horizontal and vertical, respectively. Then we can estimate one spot size by  $s_i = \frac{1}{2} [(H_{2i}^0 - H_{2i-1}^0) + (V_{2i}^0 - V_{2i-1}^0)]$ . Therefore, the rough spot size for one microarray sub-grid image can be obtained by  $s = \frac{2}{bh} \sum_{i=1}^{bh/2} (H_{2i}^0 - H_{2i-1}^0) + \frac{2}{bv} \sum_{j=1}^{bv/2} (V_{2j}^0 - V_{2j-1}^0)$ .

The adaptive adjustment algorithm can be executed according to the following steps:

*Step 1.* Estimate the rough spot diameter  $s$  by using the results drawn from the gridding step.

*Step 2.* Draw a circle in diameter  $s$  within each spot area.

*Step 3.* Consider the recognized foreground pixels (the white pixel) outside the circle as noises and remove them.

*Step 4.* Count the total pixel number  $n_s$  and the recognized foreground pixel number  $n_f$  during the rectangular spot area.

*Step 5.* Count the number of recognized foreground pixels  $n_c$  inside the circle area.

*Step 6.* A spot is considered as a missing spot when  $n_c < 0.5 \times (3.14 \times s)$  or  $(n_c \geq 3.14 \times s) \& (n_f > 0.9 \times n_s)$ . The former indicates that the recognized spot pixels are less than half of the circle area pixel number. The latter one describes the situation that the recognized spot pixels are more than the pixel number in a circle area and the recognized foreground pixel



Fig 3. Deficiency examples of different segmentation results.

<https://doi.org/10.1371/journal.pone.0210075.g003>

number is more than 90% of pixel number in a rectangular area (see Fig 3). All these illustrate that the segmented results are false.

## Intensity extraction

When the spots are segmented into foreground and background parts by our proposed clustering method, the spot expression value can be obtained according to  $E = \log_2(I_{cy3}/I_{cy5})$ . In which  $I_{cy3} = R_{fore} - R_{back}$ ,  $I_{cy5} = G_{fore} - G_{back}$  indicates the background corrected spot intensities of Red and Green channels, and  $R_{fore}$ ,  $G_{fore}$  represent the mean intensities of the foreground pixels, yet  $R_{back}$ ,  $G_{back}$  denote the mean intensities of the background pixels.

## Image quality assessment

To estimate the performance of our proposed methods, two means are adopted. One is conducting experiments on both the simulation and real images. The other is introducing various quantitative measurements.

## Testing data sets

On the simulation and real images, six real data sets and one simulation data set are adopted, as shown in Table 2. In addition, we used a microarray simulation model (see <http://www.gnu.org/copyleft/gpl.html> [44]) to generate various quality images.

First, the ScanAlyze toolbox is used to generate real microarray image, and some parameters (gray mean of spot and background intensity) are obtained. Then the gray mean of each spot is taken as the input of simulation model. Meanwhile, the cDNA microarray data can be generated through simulating the hybridization behavior of probe on slide surface and modifying the general options. To control the quality of the simulated image, some model parameters such as different noise model selecting, slide parameter setting, hybridization parameter controlling, and virtual scanner parameter setting, are tuned. Based on the steps above, three types (good, normal and poor) of microarray images are generated.

## Quantitative measurement

To comparative analyze the segmentation results, some quantitative measurements are conducted on the expression level of spots. Firstly, the log differential expression ratio  $M = \log_2(I_{cy3}/I_{cy5})$  and the log intensity  $A = \frac{1}{2} \log_2(I_{cy3} \times I_{cy5})$  are introduced. In addition, a quality index [49]  $q_{index} = (q_{sig-noise} \cdot q_{bkg1} \cdot q_{bkg2})^{1/3}$  is used and  $q_{sig-noise} = F_{mean}/(F_{mean} + B_{mean})$  represents the signal to noise ratio, the level of the local background is expressed as  $q_{bkg2} = \frac{1}{\max \left[ \frac{b_{bkg1}}{b_{bkg1} + B_{mean}} \right]} \cdot \frac{1}{b_{bkg1} + B_{mean}}$ , the local background variability is defined as  $q_{bkg1} = \frac{1}{\max \left[ \frac{BSD}{B_{mean}} \right]} \cdot \frac{1}{B_{mean}}$ . In which  $F_{mean}$  denotes the

Table 2. Description of six real microarray data sets.

Data set	Full name	No. of blocks	Spot layout in each block	Spot resolution	Image resolution
SMD	Stanford Microarray Database	48	14*18 or 44*44	18*18 or 8*8	460*451
GEO	Gene Expression Omnibus	48	13*14	12*12	451*391
BCM	Bachelor College of Medicine	48	22*22	25*25	943*949
DeRisi	Joe DeRisi Individual	4	40*40	8*8	463*455
SIB	Institute for Experiment Cancer Research	4	5*7 or 7*7	18*18	366*350
UCSF	University of California, San Francisco	36	14*15	8*8	218*209

<https://doi.org/10.1371/journal.pone.0210075.t002>



mean value of the spot,  $B_{mean}$  is the mean value of the local background,  $BSD$  indicates the standard deviation of the local background, and  $bgk_0$  represents the global average of the background. In addition, the number of pixels clustered as foreground and background for each spot is represented by  $N_{fore}, N_{back}$ , respectively. Furthermore, mean of the expression value  $MI_{cy3}, MI_{cy5}$  for each sub-grid are also introduced.

For the simulation data, owing to its corresponding annotate image, hence we can compute the pixel to pixel accuracy  $acc = (TP+TN)/(TP+TN+FP+FN)$ , the sensitivity  $se = TP/(TP+FN)$  and the specificity  $sp = TP/(TP+FP)$ . In which,  $TP$  denotes the correct number of pixels segmented as spot,  $TN$  represents the correct number of pixels segmented as background, yet  $FP$  and  $FN$  indicate the false number that spot pixels are recognized as background and background pixels are considered as spot, respectively.

## Results and discussion

The proposed clustering based algorithms (IKM and IFCM) are compared with traditional KM, FCM methods and moving K-means (MKM) [3] on six different data sets and one simulation data set.

### Image segmentation

Performance of four methods is compared on 188 real sub-grids drawn from 6 data sets, and the randomly selected original images and their corresponding segmented results are shown in Fig 4. It can be seen that a majority of spots are separated from background by these four methods. Segmentation results on GEO and SMD data sets are poor due to the low contrast of original images. In contrast, segmentation results on SIB data set are better owing to their high contrast of original images and FCM performs worst due to its sensitiveness to noises. It can be concluded that the performance of these methods is dependent on image quality. Moreover, IKM and IFCM algorithms can extract more low contrast spots than KM and FCM methods. Especially, IKM and IFCM methods can recognize missing spots by our proposed adaptive adjustment method.

To further compare the performance of four methods, the corresponding gene expression values for images in Fig 4 are presented in Fig 5. Generally, gene expression values between -2 to +2 are considered as normal data, yet those less than -2 or higher than +2 may represent noises or special genes. Therefore, we use a bigger size symbol to describe those special gene expressions as shown in Fig 5.

From Fig 5A–5C, it reveals that gene expression values varies greatly on GEO, SMD and BCM data sets owing to the low contrast of original images. At the same time, there is more gene expressions exceeding +2 or lowering -2 and most of them can be correctly obtained by IKM and IFCM methods, indicating that the improved methods perform better on poor quality image than KM and FCM algorithms. For Fig 5D–5F, a majority of gene expressions are between +2 to -2 due to the high quality of original images. In other words, almost all the spots are correctly segmented by these four methods.

In addition, the number of gene expression beyond +2 and -2 are counted for different methods on 6 data sets, as shown in Table 3. It can be seen that IKM and IFCM algorithms obtain more special gene expressions than KM and FCM ones. Especially for SMD and GEO data sets, partial spots cannot be extracted by KM and FCM methods due to their poor quality. Therefore, there is a lot of special gene expressions obtained by IKM and IFCM.

### Quantitative analysis

Scatter plot method [45] is usually used for describing the correlation between two objects. In general, a straight line in scatter plot indicates that the data points are too closer, i.e., it is a

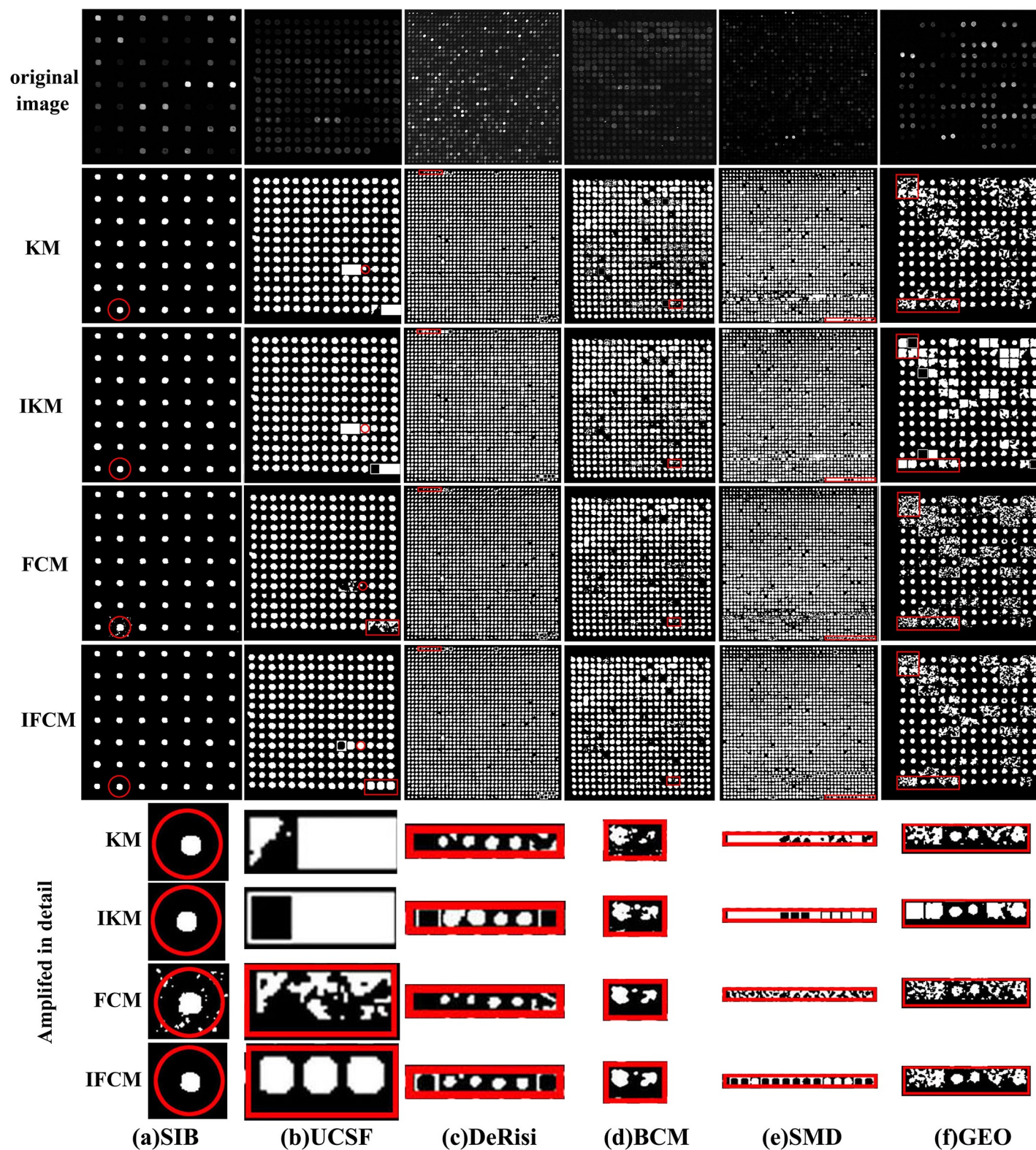


Fig 4. Segmentation results of four methods on microarray images drawn from six data sets.

<https://doi.org/10.1371/journal.pone.0210075.g004>

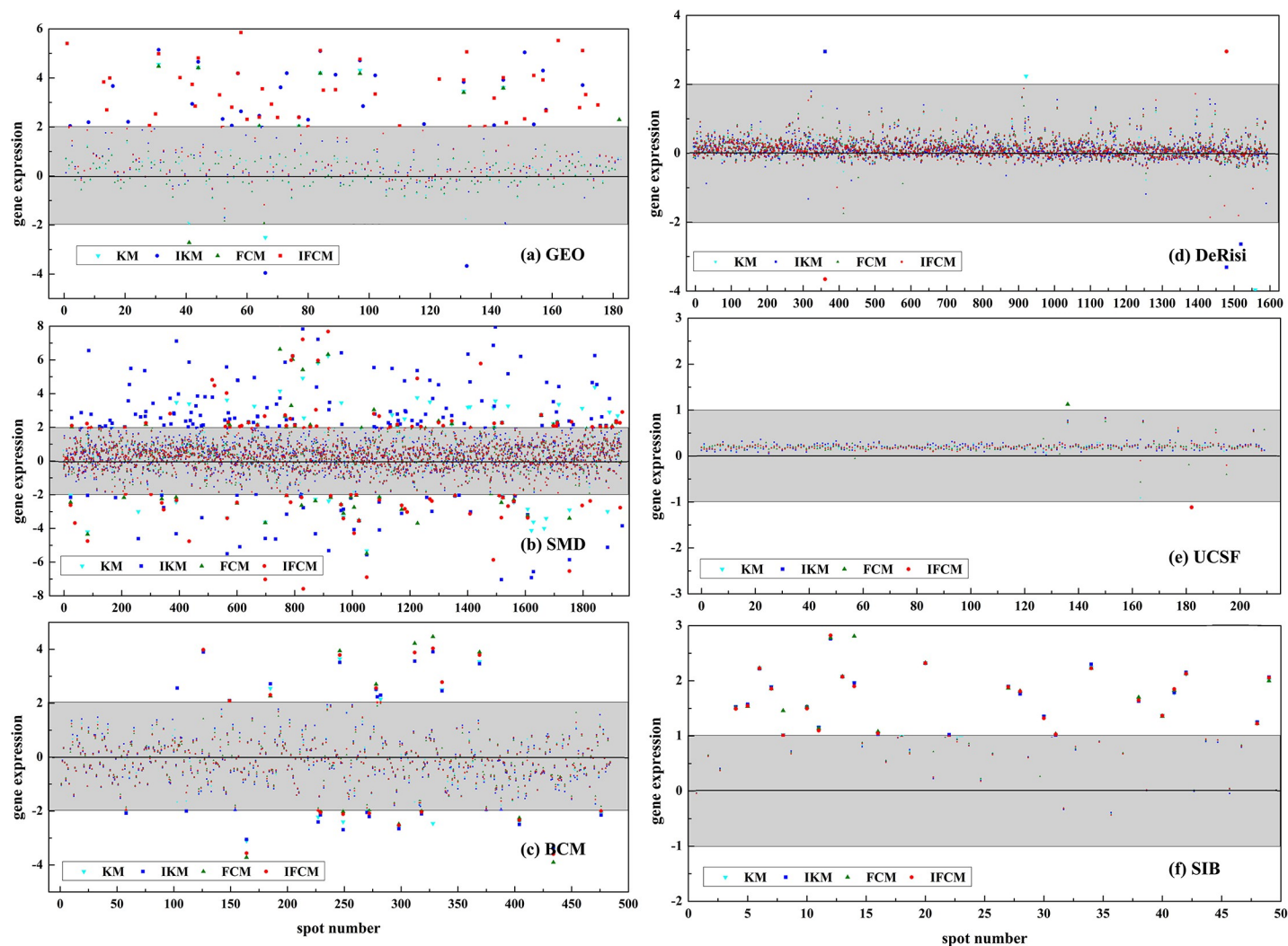


Fig 5. Gene expression for images in Fig 4 with four different methods.

<https://doi.org/10.1371/journal.pone.0210075.g005>

perfect correlation with their ratio equal to 1. Here, we adopt the scatter plot to compare the background-corrected spot intensities for red channel  $I_{cy3}$  and green channel  $I_{cy5}$ . The scatter plot of four methods on DeRisi and SMD datasets results are shown in Figs 6 and 7, respectively. Meanwhile, M-A plot is utilized to diagram the log differential expression ratio M and

Table 3. Special gene expression comparison of four methods on 6 data sets.

Data set	No. of gene expression > 2				No. of gene expression < -2			
	KM	IKM	FCM	IFCM	KM	IKM	FCM	IFCM
GEO	8	31	9	50	1	3	1	0
SMD	51	129	30	55	29	45	28	42
BCM	8	12	7	11	12	12	5	8
DeRisi	1	1	0	1	1	4	0	1
UCSF	0	0	0	1	1	1	0	0
SIB	7	7	8	7	0	0	0	0

<https://doi.org/10.1371/journal.pone.0210075.t003>



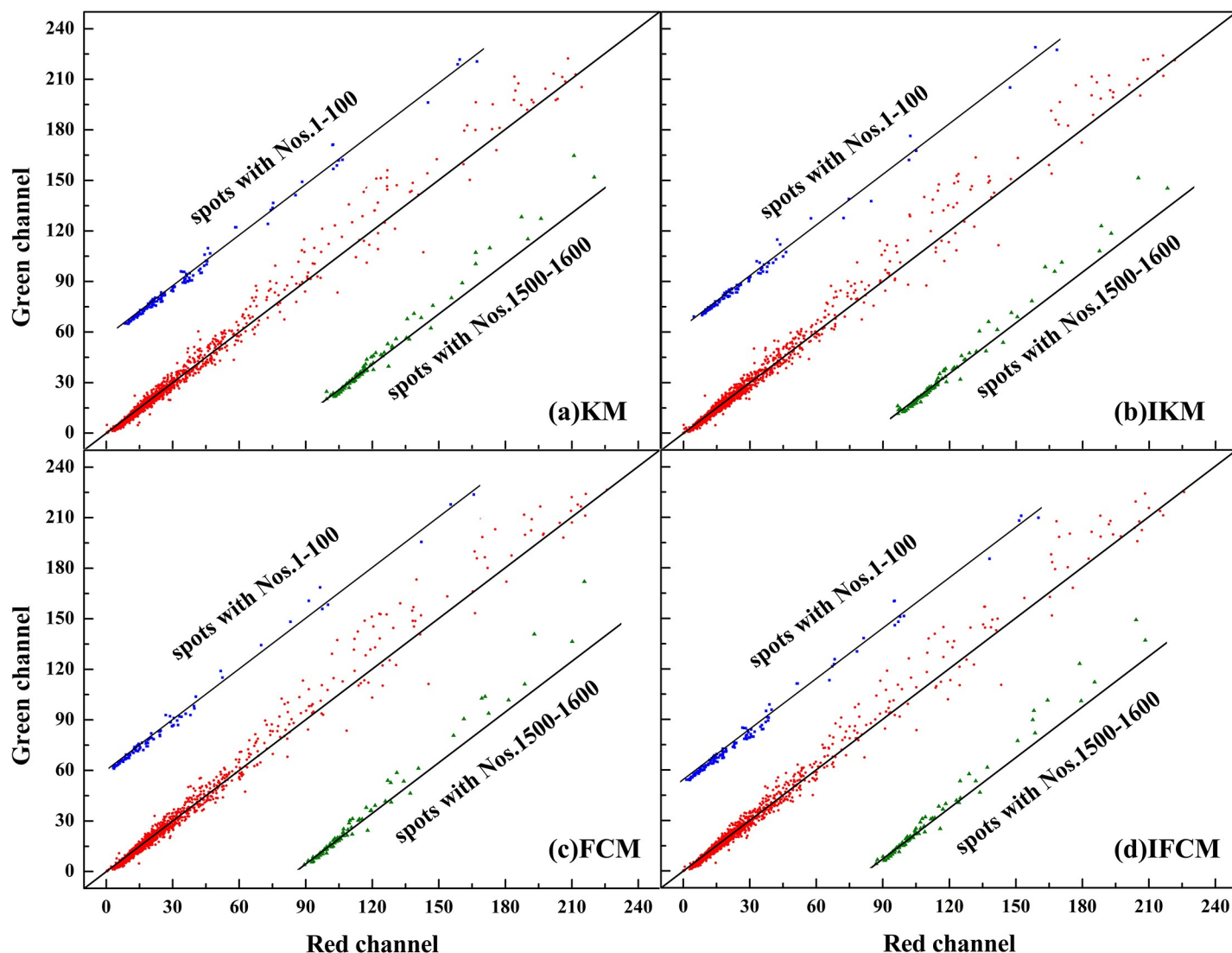


Fig 6. Scatter plot of two channel intensities for four methods on image drawn from DeRisi dataset.

<https://doi.org/10.1371/journal.pone.0210075.g006>

the log intensity  $A$  of each spot, results of four methods on DeRisi and SMD datasets are shown in Figs 8 and 9. Here, to display the results clearly, we convert the original 16-bit gray values into 8-bits on dividing the original data by 256.

In Figs 6 and 7, all data points are exhibited in Red, partial selected data points (to clearly display the correlation between  $I_{cy3}$  and  $I_{cy5}$ ) are in blue and green, respectively. From Fig 6, it can be seen that the results on the four methods are quite similar and a majority of data points are close to the diagonal line. This phenomenon is in agreement with the analysis on gene expression presented in Fig 5. However, Fig 7 reveals quite different results. First of all, the background corrected intensities of two channels on SMD dataset are much lower than that of DeRisi dataset (Fig 6), that is, most spot intensities are confined around 15 for SMD, yet 60 for DeRisi. In addition, IKM and IFMC algorithms obtain more spots with lower intensities compared to KM and FCM methods (see the circle in Fig 7). In other words, these four methods possess similar performance on spots with higher intensities, while on low contrast spots extracting, only IKM and IFMC methods perform well.

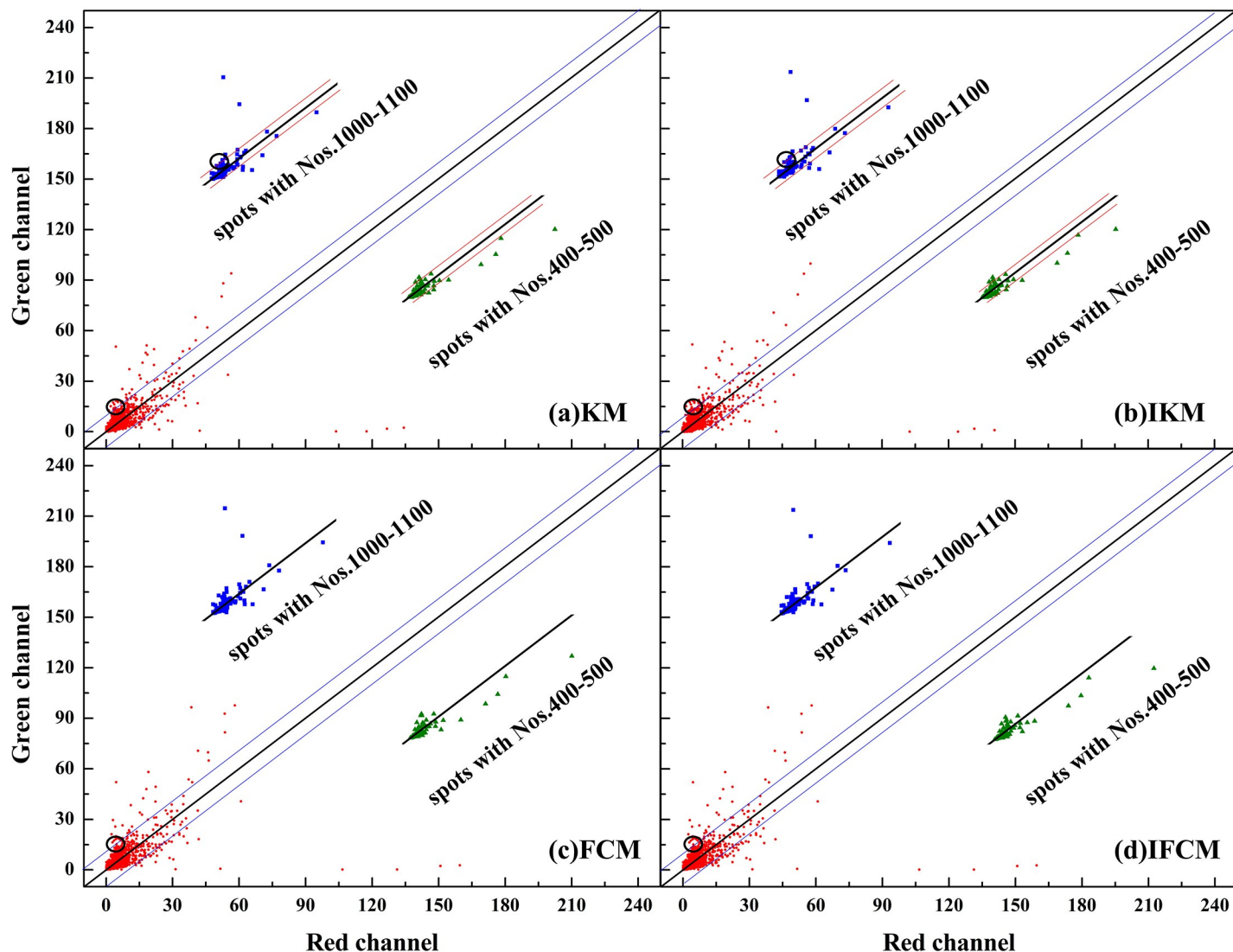


Fig 7. Scatter plot of two channel intensities for four methods on image drawn from SMD dataset.

<https://doi.org/10.1371/journal.pone.0210075.g007>

To further prove the effectiveness of our proposed methods, the mean ( $MI_{cy3}, MI_{cy5}$ ), minimum and maximum background corrected intensities for two channels are calculated, as shown in Table 4. To display the data clearly, we also transferred the data from 16-bit to 8-bit by dividing 256.

One can see from Table 4 that almost all mean background corrected intensities (see  $MI_{cy3}$  and  $MI_{cy5}$ ) obtained by IKM and IFCM are smaller than those by KM and FCM on six data-sets. The reason is that the proposed IKM and IFCM methods can extract more spots with lower contrast than KM and FCM ones. Meanwhile, the minimum and maximum intensities obtained by IKM and IFCM also exhibit a wider range, implies that our proposed methods perform better on different quality images.

In addition, the M-A plot of the four methods on DeRisi and SMD data set are shown in Figs 8 and 9, respectively. From these two figures, it can be seen that the performance of the four methods are similar for log intensity A, all higher than their mean values, but there is remarkably different for the log intensity A, smaller than its mean. In other words, for the low



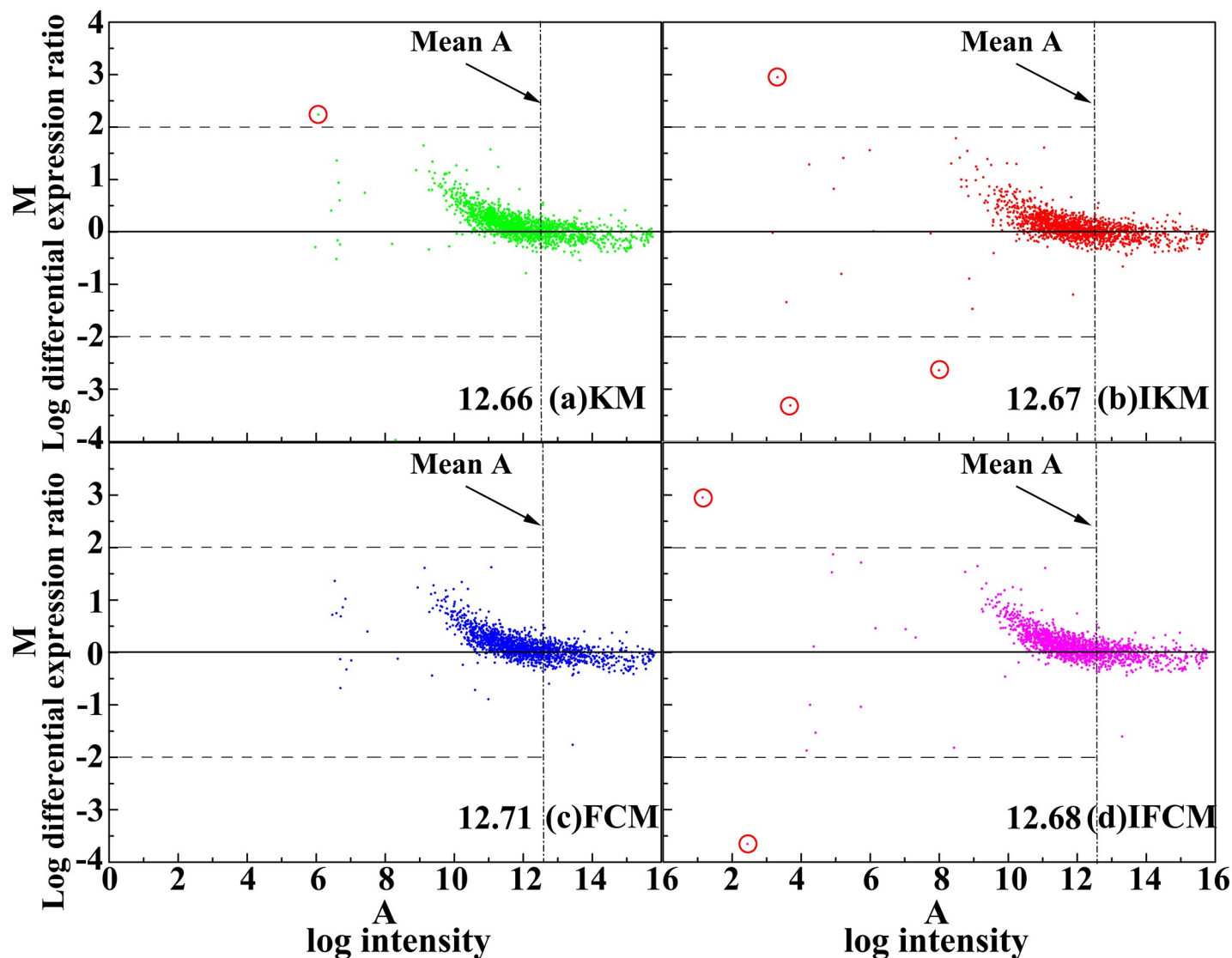


Fig 8. M-A plot of four methods on image drawn from DeRisi dataset.

<https://doi.org/10.1371/journal.pone.0210075.g008>

log intensity part, IKM and IFCM can obtain more spots with their  $M$  beyond +2 and -2 than KM and FCM (see circled spots in Fig 8 and spots beyond the dotted line in Fig 9). Actually, spots with higher intensities are easy to extract, yet those spots with lower intensities are difficult to be found. Therefore, IKM and IFCM algorithms may obtain more spots with lower intensities, revealing that these two methods own superior performance. In summary, all these results indicate that the methods of IKM and IFCM perform better than KM and FCM, which proves the effectiveness of our improved strategies.

### Spot segmentation

As we know, real spots in a microarray image usually contain various shapes instead of circles. Therefore, we choose some spots randomly according to their quality and shape. Examples of these spots and their corresponding segmented binary images are shown in Fig 10. For those good quality spots, no matter what their expression level is, IKM and IFCM algorithms can

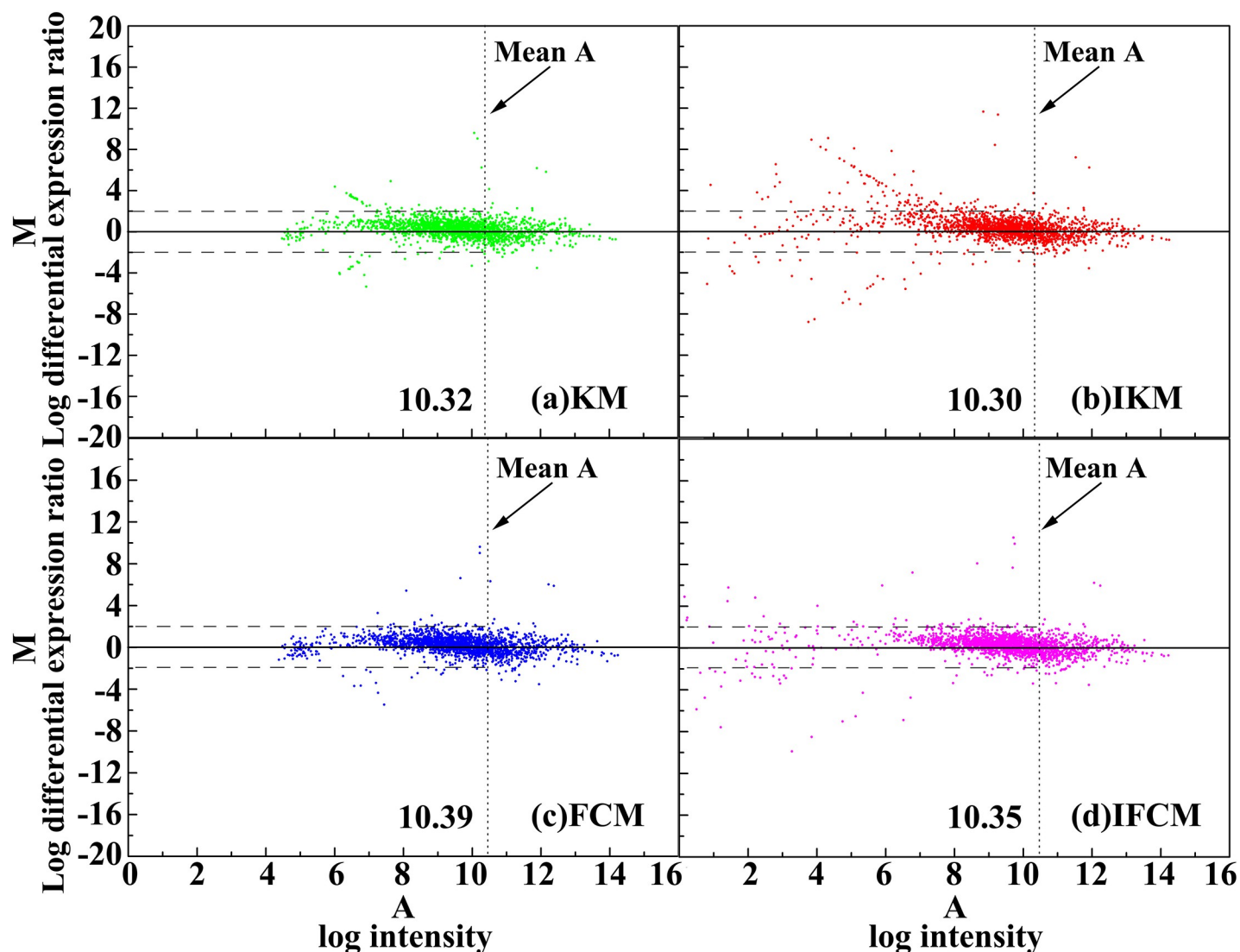


Fig 9. M-A plot of four methods on image drawn from SMD dataset.

<https://doi.org/10.1371/journal.pone.0210075.g009>

extract the spots perfectly, yet KM and FCM methods cannot segment the spots completely under the low gene expression level. Obviously, the proposed methods perform better than KM and FCM for both normal and poor quality spots. For MKM, it performs better than KM, but worse than IKM. Similarly, our proposed methods also possess optimal segmented results than KM and FCM on various shape spots.

To further evaluate the effectiveness of our proposed methods, quantitative analysis (pixels segmented as foreground or background, expression level and signal to noise ratio) on above different spots are displayed in Table 5. From Table 5, one can observe that our improved methods (IKM, IFCM) classify more required pixels into foreground ( $N_{fore}$ ) than KM and FCM algorithms, so as to be closer to the real spot shape. Furthermore, spot intensities for two channels ( $I_{cy3}, I_{cy5}$ ) obtained by IFCM and IKM are generally lower than those gained from KM and FCM, indicating that IKM and IFCM find more pixels with lower gray values around the spot edge. In other words, KM and FCM algorithms preferentially extract pixels with higher gray values. What's more, the number of foreground pixels obtained by MKM is similar

**Table 4.** The mean, minimum and maximum background-corrected intensities of four methods for two channels on six datasets.

Method	MI <sub>cy3</sub>	MI <sub>cy5</sub>	Min I <sub>cy3</sub>	Max I <sub>cy3</sub>	Min I <sub>cy5</sub>	Max I <sub>cy5</sub>
<b>Data set</b>		<b>block name</b>		<b>Spots</b>		
<b>GEO</b>		<b>GSM15898_CH1-1</b>		<b>182</b>		
KM	23.78	24.01	0.54	170.06	0.38	189.80
IKM	23.41	17.74	-0.13	176.32	-0.07	199.57
FCM	25.32	25.55	0.69	178.40	0.50	144.39
IFCM	24.53	18.86	0.71	176.32	-0.02	152.61
<b>SMD</b>		<b>49_ch1-4</b>		<b>1936</b>		
KM	5.20	4.81	0.06	134.34	0.06	94.00
IKM	5.14	4.74	-0.07	140.71	-0.13	99.82
FCM	5.43	5.07	0.05	149.79	0.08	95.62
IFCM	5.29	4.90	-0.01	159.58	-0.02	97.65
<b>BCM</b>		<b>129cy3-1</b>		<b>484</b>		
KM	12.22	13.70	-1.55	132.86	2.75	82.94
IKM	12.32	13.61	2.09	139.62	1.73	81.17
FCM	13.26	14.58	3.88	147.94	3.27	106.93
IFCM	13.03	14.12	3.82	154.32	3.40	130.09
<b>DeRisi</b>		<b>1303_ch1-1</b>		<b>1600</b>		
KM	25.29	25.22	0.22	214.43	0.12	222.41
IKM	25.28	25.53	-0.11	221.59	0.14	224.15
FCM	26.17	26.20	0.32	225.85	0.23	226.55
IFCM	25.72	25.72	-0.006	225.85	0.00	225.05
<b>UCSF</b>		<b>Cy3-4</b>		<b>210</b>		
KM	11.66	10.09	0.03	52.86	0.07	43.95
IKM	11.68	10.10	-0.00	54.01	0.02	44.61
FCM	12.20	10.62	0.03	53.63	0.02	44.54
IFCM	11.91	10.34	0.00	52.26	0.00	45.44
<b>SIB</b>		<b>Def661cy5-1</b>		<b>49</b>		
KM	51.48	89.55	3.06	226.41	9.18	227.90
IKM	51.45	89.23	3.04	226.90	9.16	221.26
FCM	53.21	93.90	3.20	231.62	9.50	236.74
IFCM	53.34	92.46	1.76	233.72	8.61	240.74



































































<https://doi.org/10.1371/journal.pone.0210075.t004>

to IKM and more than KM. Finally, the higher value of signal to noise ratio  $q_{sig-noise}$  also indicate the better performance of our proposed methods.

## Segmentation results on simulation images

Because the real images lack of annotation image, we choose simulation images for further comparison. Fig 11 displays the segmented results of four methods. It can be seen that all the spots are extracted from the background for all four methods owing to the fact that simulation image is simple and high quality compared to real image. Meanwhile, as the local magnified parts shown, it is obvious to see that segmentation results of KM and FCM contain lots of noises.

Furthermore, the pixel to pixel accuracy, specificity and sensitivity are computed for each spot and their corresponding means on images in Fig 11, as shown in Table 6. In general, IKM and IFCM own higher values of acc, sp and se than the original KM and FCM, indicating that these two improved algorithms perform better. In addition, the ratio that they cluster the

Quality	Original	KM	IKM	FCM	IFCM	MKM	Data set
High level of gene expression, Low level of noises							
Good							SIB
Normal							DeRisi
Poor							
Normal level of gene expression							
Good							SIB
Normal							GEO
Low level of gene expression, High level of noises							
Good							GEO
Normal							SMD
Poor							DeRisi
Various shape							
Square							SMD
Split							
Half-moon							

**Fig 10. Segmentation results of various shape spots.**

<https://doi.org/10.1371/journal.pone.0210075.g010>

correct pixels into foreground (see *se* in Table 6) are both more than 99%. All these results prove that our proposed IKM and IFCM algorithms outperform than KM, FCM and MKM methods.

### Computational complexity analysis

As we know, the computational complexity of KM is  $O(n)$ , here  $n$  represents the number of spot pixels. Compared to KM, FCM involves computing the Centroid values of each cluster according to its fuzzy membership relation and updating the membership values, so its computational complexity is about several times of KM, As for MKM, there is only one more pixel reassignment operation compared to KM, so their computational complexity is similar. For our improved algorithms, the image contrast enhancement, the multi-features computing, feature selection by PCA and refinement are extra operations compared to KM and FCM, so our methods may be about two or three times over than its original method. Table 7 illustrates

Table 5. Quantum analysis of the segmentation results showed in Fig 10.

method	$N_{fore}$	$N_{back}$	$I_{cy3}$	$I_{cy5}$	$I_{cy3}/I_{cy}$	$q_{sig-noise}$
KM	149	1853	58342	57961	1.01	0.9887
IKM	157	1845	56093	58086	0.96	0.9891
FCM	140	1862	61630	59832	1.03	0.9888
IFCM	144	1858	60606	59294	1.02	0.9893
MKM	156	1846	56087	58082	0.96	0.9890
KM	151	2404	16483	10325	1.59	0.9710
IKM	154	2401	16295	10210	1.59	0.9715
FCM	135	2417	17831	11089	1.60	0.9728
IFCM	142	2410	17173	11089	1.54	0.9724
MKM	152	2403	16338	10289	1.59	0.9713
KM	88	294	2680.5	2370.4	1.13	0.8196
IKM	135	247	1852.5	2266.8	0.81	0.8533
FCM	71	311	3451.7	3020.6	1.14	0.8259
IFCM	142	240	1843.7	2569.4	0.71	0.8823
MKM	128	254	2178.4	2298.7	0.95	0.8368
KM	50	84	46692	40819	1.16	0.9152
IKM	54	80	46694	36234	1.28	0.9388
FCM	51	83	47387	41684	1.13	0.9252
IFCM	66	68	40021	34795	1.15	0.9440
MKM	52	82	46693	38174	1.22	0.9217
KM	102	280	3036	2338	1.29	0.8517
IKM	139	243	2244.4	2161.5	1.03	0.8804
FCM	73	309	4117.5	3285.3	1.25	0.8502
IFCM	141	241	2212.5	2782.9	0.79	0.9025
MKM	128	254	2794.7	2297.6	1.21	0.8638
KM	55	47	1788.6	1807.1	0.98	0.8189
IKM	56	46	1791.8	1828.4	0.98	0.8234
FCM	52	50	1724.8	1828.5	0.94	0.7965
IFCM	53	49	1680	1797.6	0.93	0.7919
MKM	55	47	1788.6	1807.1	0.98	0.8189
KM	60	74	3033.1	3672.3	0.82	0.6920
IKM	60	74	2754.4	3198.1	0.86	0.6751
FCM	51	83	3117.4	3890.9	0.80	0.6882
IFCM	69	65	2753.2	3231.7	0.85	0.6827
MKM	60	74	2936.8	3469.5	0.85	0.6931
KM	85	49	976.8	1233	0.79	0.5885
IKM	87	47	609	1136.3	0.53	0.5532
FCM	74	60	943.4	1477.5	0.63	0.5842
IFCM	79	55	877.5	1233	0.71	0.5785
MKM	86	48	863.4	1194.6	0.72	0.5893
KM	19	83	507.3	607.9	0.83	0.7322
IKM	23	79	422	92.2	4.5	0.7083
FCM	13	89	733.4	367.7	1.99	0.7814
IFCM	16	86	605	284.6	2.13	0.7558
MKM	21	81	478.2	524.8	0.91	0.7568
KM	28	74	13343	14022	0.95	0.9296
IKM	39	63	10757	12273	0.87	0.9451

(Continued)



Table 5. (Continued)

method	$N_{fore}$	$N_{back}$	$I_{cy3}$	$I_{cy5}$	$I_{cy3}/I_{cy}$	$q_{sig-noise}$
FCM	29	73	14178	12935	1.09	0.9529
IFCM	32	70	13121	12184	1.07	0.9546
MKM	32	70	12866	13242	0.97	0.9362
KM	49	83	2781.2	1958.7	1.41	0.8879
IKM	49	83	2781.2	1911.6	1.45	0.8879
FCM	47	85	2864.5	2871.4	0.99	0.8880
IFCM	47	85	2864.5	1800.1	1.59	0.8880
MKM	49	83	2781.2	1958.7	1.41	0.8879

<https://doi.org/10.1371/journal.pone.0210075.t005>

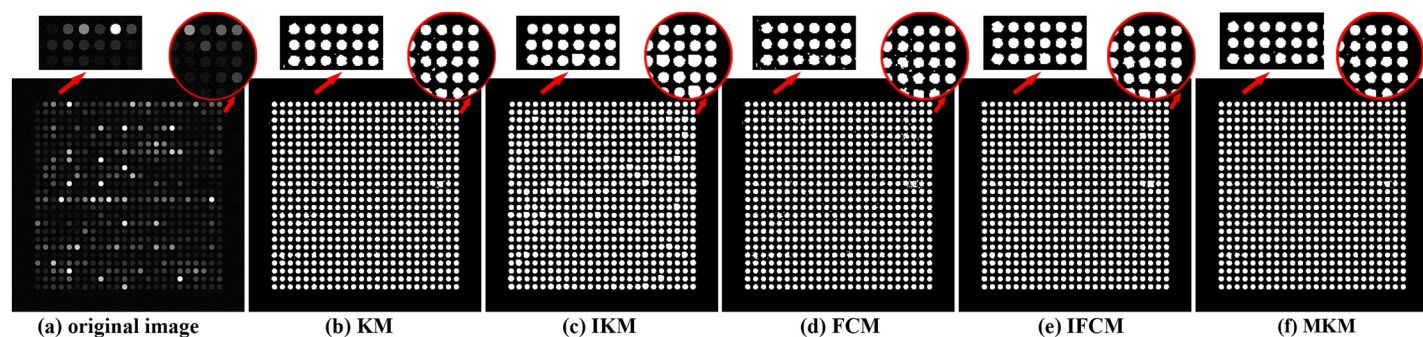


Fig 11. Examples of segmentation results on simulation images.

<https://doi.org/10.1371/journal.pone.0210075.g011>

Table 6. Quantitative evaluation for four methods on simulation image.

method	KM	IKM	FCM	IFCM	MKM
sp	0.7839	0.8343	0.7819	0.8388	0.8226
se	0.9825	0.9917	0.9775	0.9949	0.9903
acc	0.8565	0.8854	0.8540	0.8898	0.8785

<https://doi.org/10.1371/journal.pone.0210075.t006>

Table 7. Computational time comparison of five methods on blocks drawn from six data sets (seconds).

method	GEO	SMD	BCM	DeRisi	UCSF	SIB
KM	1.829	7.298	16.476	7.111	1.107	2.853
IKM	4.004	15.121	36.107	16.258	2.336	5.896
FCM	10.159	29.655	99.188	29.064	4.913	15.907
IFCM	27.585	79.515	288.847	82.411	13.327	36.804
MKM	1.882	7.684	17.099	7.506	1.147	16.472

<https://doi.org/10.1371/journal.pone.0210075.t007>

the average computational time of above five methods on six data sets. From Table 7 we can see that the same algorithm required various computational times for different data sets due to the distinguished image layout and resolution as displayed in Table 2, in which blocks in UCSF data set take minimum time, yet blocks among BCM data set require maximum time.

According to all above analysis, we can draw the following conclusions: for good quality image with high contrast, less noises and sparse spot distribution, the IKM can be used for its

segmentation. For poor quality image with low contrast and more noises, IFCM can be utilized to segment the spots. For image contains lots of special shape spots, IKM performs better than IFCM. Finally, IKM and IFCM are both suitable for normal quality image segmentation.

## Conclusions

In conclusion, although current state-of-art clustering-based segmentation methods (KM, FCM) are easy, fast and effective, they are prone to be affected by noises and mostly be conducted based on one feature. Therefore, we proposed the IKM and IFCM algorithms by introducing multi-features such as intensity, spatial, and shape features. Meanwhile, an adaptive adjustment method for segmentation results is introduced and a cluster center initialization strategy is considered. Experiments on six real data sets and one simulation data set verify that our proposed IKM and IFCM methods perform better than the original KM and FCM. In addition, the quantitative analysis on gene expression, background corrected intensities and log-differential expression ratio versus log intensity, further proves the effectiveness of our proposed methods.

## Supporting information

**S1 Algorithm.**  
(ZIP)

## Acknowledgments

We would like to thank Professor Yuhua Wen for his help in revising the manuscript.

## Author Contributions

**Conceptualization:** Guifang Shao.

**Formal analysis:** Junfa Zhang, Yali Shangguan.

**Methodology:** Jianbo Yang.

**Resources:** Yali Shangguan.

**Validation:** Guifang Shao.

**Writing – original draft:** Guifang Shao, Dongyao Li.

**Writing – review & editing:** Guifang Shao.

## References

1. Rajashekhar C.M., Varun S.S. (2015) DNA microarray spot detection using statistical image segmentation. *International Journal of Innovative Research in Computer and Communication Engineering* 3 (5):140–147.
2. Katsigiannis S., Zacharia E., Maroulis D. (2017) MIGS-GPU: microarray image gridding and segmentation on the GPU. *IEEE Journal of Biomedical and Health Informatics* 21(3):867–874. <https://doi.org/10.1109/JBHI.2016.2537922> PMID: 26960232
3. Shao G.F., Li T.J., Zuo W.D., Wu S.X., Liu T.D. (2015) A combinational clustering based method for cDNA microarray image segmentation. *PLOS ONE* 10(8):1–23.
4. Axon Instruments Inc: GenePix 4000A User's Guide. 1997.
5. ImaGene, Biodiscovery Inc. [<http://www.biodiscovery.com/imagene/>]. 1997.
6. GSI Lumonics: QuantArray Analysis Software, Operator's Manual. 1999. [<http://www.bioprocessonline.com/doc/quantarray-analysis-software-0001>]
7. Eisen M.B. (1997) ScanAlyze. [<http://rana.Stanford.EDU/software/>]

8. Heyer L.J., Moskowitz D.Z., Abele J.A., Karnik P., Choi D., Campbell A.M., Oldham E.E., Akin B.K. (2005) Magic tool: Integrated microarray data analysis. *Bioinformatics* 21(9):2114–2115. [<http://www.bio.davidson.edu/MAGIC/>] <https://doi.org/10.1093/bioinformatics/bti247> PMID: 15647303
9. Buckley MJ, The Spot user's guide. CSIRO Mathematical and Information Sciences, 2000. [<http://www.cmis.csiro.au/IAP/Spot/spotmanual.htm>].
10. Buhler J., Ideker T., Haynor D. (2000) Dapple: improved techniques for finding spots on DNA microarrays. UWCSE Tech Report, UWTR Department of Computer Science and Engineering, University of Washington. [<http://www.cs.wustl.edu/~jbuhler/research/dapple/>]
11. Spotfinder Online Manual, The Insitute for Genomics Research (TIGR), 2004. [<http://www.tm4.org/spotfinder.html>]
12. Carlisle A. J., Prabhu V. V., Elkahouloun A., Hudson J., Trent J. M., Linehan W. M., et al.(2000) Development of a prostate cDNA microarray and statistical gene expression analysis package. *Molecular Carcinogenesis* 28: 12–22. [<https://abs.cit.nih.gov/pscan/>] PMID: 10820484
13. Jain A.N., Tokuyasu T.A., Snijders A.M., Segreaves R., Albertson D.G., Pinkel D. (2002) Fully automatic quantification of microarray image data. *Genome Res.* 12: 325–332. [<http://cc.ucsf.edu/jain/public>] <https://doi.org/10.1101/gr.210902> PMID: 11827952
14. Kashyap Ramgopal and Gautam Pratima (2013) Microarray image segmentation using improved GOGAC method. *International Journal of Computer Science and Engineering* 2(4):67–74.
15. Ni S.H., Wang P., Paun M., Dai W.Z., Paun A. (2009) Spotted cDNA microarray image segmentation using ACWE. *Romanian Journal of Information Science and Technology* 12(2):249–263.
16. Li Y., P'aun A., P'aun M.(2017) Improvements on contours based segmentation for DNA microarray image processing. *Theoretical Computer Science* 701:174–189.
17. Nguyen H.N., Paveau V., Cauchois C., Kervrann C. (2018) ATMAD: robust image analysis for Automatic Tissue MicroArray De-arraying. *BMC Bioinformatics* 19:148. <https://doi.org/10.1186/s12859-018-2111-8> PMID: 29673310
18. Ho J.,Hwang W.L. (2008) Automatic microarray spot segmentation using a snake-fisher model. *IEEE Transactions on Medical Imaging* 27(6): 847–57. <https://doi.org/10.1109/TMI.2008.915697> PMID: 18541491
19. Athanasiadis E., Cavouras D., Kostopoulos S., Glotsos D., Kalatzis I., Nikiforidis G. (2011) A wavelet-based markov random field segmentation model in segmenting microarray experiments. *Computer methods and programs in biomedicine* 104(3):307–315. <https://doi.org/10.1016/j.cmpb.2011.03.007> PMID: 21531035
20. Zacharia Eleni and Maroulis Dimitris (2011) A spot modeling evolutionary algorithm for segmenting microarray images. in *Evolutionary Algorithms*, InTech, 459–479.
21. Khalilabad N.D., Hassanpour H. (2017) Employing image processing techniques for cancer detection using micro array images. *Computers in Biology and Medicine* 81:139–147. <https://doi.org/10.1016/j.combiomed.2016.12.012> PMID: 28061369
22. Deepa J., Thomas T. (2009) Automatic segmentation of DNA microarray images using an improved seeded region growing method. *World Congress on Nature & Biologically Inspired Computing, Coimbatore, INDIA*, 1468–1473.
23. Saadalgawady A., Eltoukhy M.M., Eltawel G., Wahed M.E. (2015) Segmentation of complementary DNA microarray images using marker-controlled watershed technique. *International Journal of Computer Applications* 110(12):30–34.
24. Thamaraيمانalan P., Kumar D.D., Nirmalakumari K. (2014) Efficient gridding and segmentation for microarray images. *International Journal of Computer Science and Mobile Computing* 3(2):353–360.
25. Rajkumar P., Vennila I., Nirmalakumari K. (2013) An intelligent segmentation algorithm for microarray image processing. *International Journal on Computer Science and Engineering* 5(6):528–537.
26. Angulo J. (2008) Polar modeling and segmentation of genomic microarray spots using mathematical morphology. *Image Analysis & Stereology* 27(2):107–124.
27. Manjunath S.S., Shreenidhi B.S., Nagaraja J., Pradeep B.S. (2013) Morphological spot detection and analysis for microarray images. *International Journal of Innovative Technology and Exploring Engineering* 2(5):189–93.
28. Farouk R.M., Badr E.M., SayedElahl M.A. (2014) Recognition of cDNA microarray image using feed forward artificial Neural Network. *International Journal of Artificial Intelligence & Applications* 5(5):21–31.
29. Giannakeas N., Karvelis P.S., Exarchos T.P., Kalatzi F.G., Fotiadis D.I. (2013) Segmentation of microarray images using pixel classification -Comparison with clustering-based methods. *Computers in Biology and Medicine* 43(6):705–716. <https://doi.org/10.1016/j.combiomed.2013.03.003> PMID: 23668346

30. Bozinov D., Rahnenfuhrer J. (2002) Unsupervised technique for robust target separation and analysis of DNA microarray spots through adaptive pixel clustering. *Bioinformatics* 18(5):747–756. PMID: [12050071](https://pubmed.ncbi.nlm.nih.gov/12050071/)
31. Belean B., Borda, Ackermann M., Koch J., Balacescu O.I. (2015) Unsupervised image segmentation for microarray spots with irregular contours and inner holes. *BMC Bioinformatics* 16:1–12. <https://doi.org/10.1186/s12859-014-0430-y>
32. Nagarajan R. (2003) Intensity-based segmentation of microarray images. *IEEE Trans. On Medical Imaging* 22(7): 882–889. <https://doi.org/10.1109/TMI.2003.815063> PMID: [12906242](https://pubmed.ncbi.nlm.nih.gov/12906242/)
33. Rahnenfuhrer J., Bozinov D. (2004) Hybrid clustering for microarray image analysis combining intensity and shape features. *BMC Bioinformatics* 5: 1–11. <https://doi.org/10.1186/1471-2105-5-1>
34. Maguluri L.P., Rajapanthula K., Srinivasu P.N. (2013) A comparative analysis of clustering based segmentation algorithms in microarray images. *International Journal of Emerging Science and Engineering* 1(5):27–32.
35. Giannakeas N., Fotiadis D.I. (2009) An automated method for gridding and clustering-based segmentation of cDNA microarray images. *Computerized Medical Imaging and Graphics* 33(1):40–49. <https://doi.org/10.1016/j.compmedimag.2008.10.003> PMID: [19046850](https://pubmed.ncbi.nlm.nih.gov/19046850/)
36. Uslan V., Bucak I.Ö. (2010) Microarray image segmentation using clustering methods. *Mathematical & Computational Applications* 15(2): 240–247.
37. Harikiran J., RamaKrishna D., Phanendra M.L., Lakshmi P.V., Kiran Kumar R. (2012) Fuzzy c-means with bi-dimensional empirical mode decomposition for segmentation of microarray image. *International Journal of Computer Science Issues* 9(3):316–321.
38. Biju V.G., Mythili P. (2015) Fuzzy clustering algorithms for cDNA microarray image spots segmentation. *International Conference on Information and Communication Technologies* 46:417–424.
39. Biju V. G. Myth P. (2017) Possibilistic reformed fuzzy local information clustering technique for noisy microarray image spots segmentation. *Current Science* 113:1072–1080.
40. Blekas K., Galatsanos N., Likas A., Lagaris I.E. (2005) Mixture model analysis of DNA microarray images. *IEEE Transactions on Medical Imaging* 24(7):901–909. PMID: [16011320](https://pubmed.ncbi.nlm.nih.gov/16011320/)
41. Li Q., Fraley C., Bumgarner R.E., Yeung K.Y., Raftery A.E. (2005) Donuts, scratches and blanks: robust model-based segmentation of microarray images. *Bioinformatics* 21: 2875–2882. <https://doi.org/10.1093/bioinformatics/bti447> PMID: [15845656](https://pubmed.ncbi.nlm.nih.gov/15845656/)
42. Raghavarao S., Madhanmohan M.S., Prasad G.M.V. (2011) Segmentation of microarray image using information bottleneck. *Global Journal of Computer Science and Technology* 11(19):30–33.
43. Biju V.G. and Mythili P. (2012) A genetic algorithm based fuzzy c mean clustering model for segmenting microarray images. *International Journal of Computer Applications* 52(11):42–48.
44. National Human Genome Research Institute, <http://www.genome.gov>, 2009.
45. Battiato S., Blasi G. D., Farinella G., Gallo G., and Guarnera G. (2007) Adaptive techniques for microarray image analysis with related quality assessment. *Journal of Electronic Imaging* 16(4):043013.