









Melatonin and the cardiovascular system in animals: systematic review and meta-analysis

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Melatonin, a hormone released by the pineal gland, demonstrates several effects on the cardiovascular system. Herein, we performed a systematic review and meta-analysis to verify the effects of melatonin in an experimental model of myocardial infarction. We performed a systematic review according to PRISMA recommendations and reviewed MEDLINE, Embase, and Cochrane databases. Only articles in English were considered. A systematic review of the literature published between November 2008 and June 2019 was performed. The meta-analysis was conducted using the RevMan 5.3 program provided by the Cochrane Collaboration. In total, 858 articles were identified, of which 13 were included in this review. The main results of this study revealed that melatonin benefits the cardiovascular system by reducing infarct size, improving cardiac function according to echocardiographic and hemodynamic analyses, affords antioxidant effects, improves the rate of apoptosis, decreases lactate dehydrogenase activity, enhances biometric analyses, and improves protein levels, as analyzed by western blotting and quantitative PCR. In the meta-analysis, we observed a statistically significant decrease in infarct size (mean difference [MD], -20.37 [-23.56, -17.18]), no statistical difference in systolic pressure (MD, -1.75 [-5.47, 1.97]), a statistically significant decrease in lactate dehydrogenase in animals in the melatonin group (MD, -4.61 [-6.83, -2.40]), and a statistically significant improvement in the cardiac ejection fraction (MD, -8.12 [-9.56, -6.69]). On analyzing potential bias, we observed that most studies presented a low risk of bias; two parameters were not included in the analysis, and one parameter had a high risk of bias. Melatonin exerts several effects on the cardiovascular system and could be a useful therapeutic target to combat various cardiovascular diseases.

KEYWORDS: Cardiology; Melatonin; Meta Analysis; Review; Systematic Review.

■ BACKGROUND

Melatonin (N-acetyl-5-methoxytryptamine) is a hormone produced by the pineal gland exclusively at night and is released into the bloodstream and cerebrospinal fluid in a circadian manner to regulate several physiological and neuroendocrine functions (1-3). The effects of melatonin are dependent on non-receptor- and receptor-mediated mechanisms of action. Membrane melatonin receptors (MT1, MTNR1A, MT1, and MTRN1B) are G-protein-coupled receptors, signaling through G_i-G₀ or G_q-G₁₁ transduction

pathways, depending on the target organ. Melatonin secreted at night might interact with its effector and produce immediate effects when melatonin is present in the circulation (e.g., nighttime blood pressure dipping). Moreover, during the night and through several mechanisms of action, melatonin primes prospective effects (such as controlling autonomic nervous system activity) that can be observed only during the day when no pineal melatonin production occurs (3-6).

Over the last 20 years, several studies have suggested that melatonin influences the cardiovascular system (7,8). Melatonin may have significant anti-inflammatory and cardio-protective properties by directly eliminating free radicals, as well as indirectly via antioxidant activity. In addition, melatonin may be involved in blood pressure regulation and have significant anti-atherogenic effects (8-13).

In this systematic review, cardiovascular diseases such as hypertension, myocardial infarction, ischemia, and reperfusion were selected to verify the action of melatonin, as we believe that these cardiopathies currently represent a large number of cardiovascular diseases (13,14). Our study aimed

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to verify the effects of melatonin in an experimental model of myocardial infarction.

SEARCH STRATEGIES

In the present study, the search strategy was performed as described by Tawfik et al. (15). We used MEDLINE, Google Scholar, and Cochrane databases and reviewed literature published from November 2008 to June 2019; we restricted this systematic review to the last ten years, covering the latest and most relevant articles worldwide. First, we selected keywords from related articles, using Medical Subject Headings (MeSH) to identify more related keywords with similar meanings as follows: (“melatonin”) [MeSH Terms] AND (“cardiovascular system”) [MeSH Terms] [All Fields]. We then searched the three databases. Accordingly, we identified 2096 articles in PubMed using the “other animals” filter, 602 articles using Google Scholar

filtering for keywords only in the title, and three articles using a Cochrane Library advanced search; the terms used were “melatonin and cardiovascular system” In addition, we reviewed retrieved articles to identify additional studies (Figure 1). This review was conducted according to the recommendations of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) (16,17).

We excluded studies with cell culture experiments, as well as pre- and post-conditioning studies. The inclusion criteria were animal studies, cell culture studies, and *in vivo* experiments. The control group was the melatonin group in this study. The melatonin group varied in each article, as studies persistently experimented with a melatonin group related to a drug or an event.

The process of paper retrieval and titles and abstract evaluation was conducted by two independent blinded researchers capable of compiling systematic reviews (ECV and RS), following the inclusion and exclusion criteria

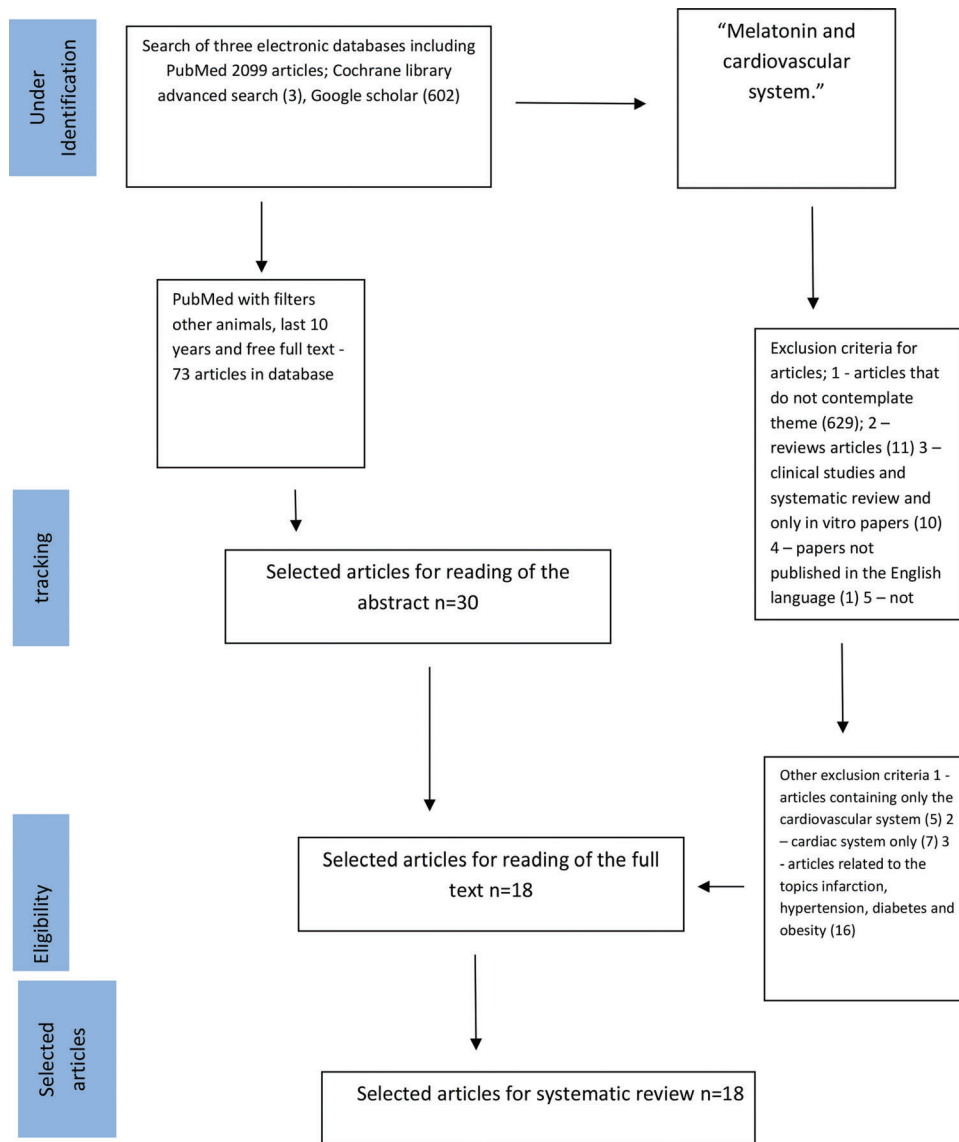


Figure 1 - Flow chart of experimental design.



according to the tenets of PICO (16-19). The PICO was defined as patients in case the systematic review was performed in animals, interventions considering the administration of melatonin in animals using an experimental model of myocardial infarction, comparison, to compare the melatonin group with the control group receiving no melatonin, and outcome, which were results of administering melatonin. The selected articles were critically evaluated to determine their potential inclusion in the review. In the event of a disagreement between investigators regarding studies selected, a third reviewer was consulted (LCA).

In the present systematic review, data obtained from selected studies were tabulated, and the following characteristics were listed when present in the articles: authors' names, year of publication, animal type, sex (M/F), animal species, age (months), weight, induction model, and site injury (Table 1). Table 2 presents the following information: authors, sample size, number of groups, number of animals per group, melatonin administration, melatonin doses, and dependent variables. Table 3 lists the most frequent recommendations in preclinical research guidelines for *in vivo* animal experiments (18). Table 4 evaluates the study characteristics of selected controlled animal studies, with prior exercise and myocardial infarction as variables that showed a significant difference between the melatonin control group and the study group. These were classified as S for "significant difference," and variables that did not present a significant difference were classified as NS (not significant).

RevMan (version 5.3; Cochrane Collaboration, Oxford, UK) was used to perform the meta-analysis. The random-effects model was used to account for the heterogeneity.

Statistical analysis

Mean values and standard deviation between studies, presented as the mean difference (MD) of post-intervention values after calculating the inverse variance, were employed to verify the magnitude of the protection afforded by melatonin (19). In addition, heterogeneity was assessed using Cochran's Q and I² tests, followed by visual inspection of the graph. The analyses were performed using the RevMan software (version 3.3.1) (20).

RESULTS

Figure 1 presents the search process, identification, and selection of articles. Based on our search strategies, 73 articles were retrieved from 2099 identified articles in PubMed using the "other animals" filter; among these, 18 were selected after reading the title and abstract. In addition, we selected three articles from BIREME and 0 articles from the Cochrane database. The inclusion and exclusion criteria are described in Figure 1 (21-38). Articles were primarily excluded when assessments were performed in human subjects, apart from being unrelated to components of PICO; we mainly focused on experimental animal studies.

In Table 4, we employed the criteria of Henderson et al. (18). We found that 61.11% (21,22,25-27,30,31,33,35,36,38) of selected studies had an appropriate sample size and 61.11% (21,23,25,26,27,30,31,33-36) had randomized animals, according to their materials and methods. All articles were blinded to the outcome assessment (21-38). We could not determine the criterion underlying the flow of animals through experiments, as no explicit statement regarding the same was available in the materials and methods. We observed that

Table 1 - Study characteristics of selected control experimental studies assessing melatonin and the cardiovascular system.

Authors	Animal type	Animal race	Age (months)	Weight	Induction model	Site injury
Zhang et al. (21)	Mice	C57/B6	-	-	Sepsis-induced cardiac dysfunction	Cardiovascular system
Benova et al. (22)	Rat	Wistar	9 months	-	Obesity	Cardiovascular system
Chen et al. (23)	Rat	Sprague-Dawley	-	200-250 g	Myocardial ischemia reperfusion	Myocardial tissue
Liu et al. (24)	Mice	C57/B6	6 months	-	Myocardial infarction	Heart
Simko et al. (25)	Rat	Wistar	3 months	-	hypertension	Cardiovascular system
Simko et al. (26)	Rat	Wistar	3 months	-	hypertension	Cardiovascular system
Stacchiotti et al. (27)	Mice	B6.V ^{LEAN} /OlaHsd and B6.V ^{Lep^{ob}} /OlaHsd	4 weeks	-	Obesity	Mitochondria of cardiomyocyte
Chaudagar et al. (28)	Rat	Wistar	8 months	-	hypertension	Cardiovascular system
Salmanoglu et al. (29)	Rat	Wistar	-	250-350 g	Diabetic	Liver tissue
Cheng et al. (30)	Rabbits	New Zealand	4 weeks	2.0-2.5 Kg	Atherosclerosis	Aorta
Liu et al. (31)	Rat	Sprague-Dawley	3 months	280-360 g	Myocardial ischemia reperfusion	Myocardial tissue
Zhu et al. (32)	Rat	Sprague-Dawley	10 weeks	250 g	Myocardial infarction	Heart
Liu et al. (33)	Rat	-	-	350-400 g	Myocardial ischemia reperfusion	Heart
Drobnik et al. (34)	Rat	Wistar	4 weeks	290-320 g	Myocardial infarction	Heart
Repova et al. (35)	Rat	Wistar	3 months	300-330 g	hypertension	Cardiovascular system
Drobnik et al. (36)	Rat	Wistar	-	-	Myocardial infarction	Heart
Chen et al. (37)	Mice	Mice Gpx ^{-/-} C57BL/6	-	-	Myocardial ischemia reperfusion <i>in vitro</i>	Heart
Petrosillo et al. (38)	Rat	Wistar	-	250-330 g	Myocardial ischemia reperfusion	Heart



Table 2 - Characteristics (samples size, number of groups, number of animals/groups, dependent variables) of selected experimental studies assessing the effects of melatonin and the cardiovascular system.

Authors	Sample size	Number of groups	Number of animals/groups	Melatonin administration	Melatonin doses	Dependent variables
Zhang et al. (21)	24	4	6	Intraperitoneal injection	30 mg/kg	Echo, histological analysis, creatinine kinase measurement, TUNEL analysis, western blotting.
Benova et al. (22)	48	4	12	Drinking water	10 mg of melatonin was dissolved in 100 mL of water for 8 weeks	Heart function in Langendorff perfusion, western blot, real-time PCR,
Chen et al. (23)	30	5		Intraperitoneal at the reperfusion	20 mg/kg	Echo, IS2, lactate dehydrogenase release, CMEC measurement <i>in vitro</i> IRI assay, western blotting, qRT-PCR, and detection of autophagosomes.
Liu et al. (24)	18	3	6	Gavage	50 mg/kg	Echo, histological analysis, PCR, western blot, CTRP3 detection.
Simko et al. (25)	40	4	10	Water consumption was 12-13 mL/100 g of body weight	10 mg of melatonin was dissolved in 100 mL of water for 4 weeks	Hemodynamics measures, biometric analysis, determination of hydroxyproline, angiotensin, and aldosterone analysis.
Simko et al. (26)	66	6	11	Drinking water adjustment to daily water consumption to ensure the correct dosage	10 mg/kg/day for 6 weeks	Hemodynamics measures, determination of hydroxyproline, NO synthase activity, oxidative load measurement, and western blotting of NF- κ B.
Stacchiotti et al. (27)	40	4	10	5 th to 13 th weeks of life/drinking water	100 mg/kg/day for 8 weeks	Histomorphometric evaluations, nuclear cardiomyocyte morphometry, mitochondrial and immunohistochemical analysis.
Chaudagar et al. (28)	24	4	6	Drinking water	10 mg/kg/day for 67 days	Hemodynamics measures, biometric analysis, and NO assays.
Saimanoglu et al. (29)	35	6	-	Oral gavage	10 mg/kg/day for 2 weeks	Vasocontractile response, measurement of total cholesterol, LDL, HDL, glucose, NO, and insulin, MDA assay, and tissue antioxidant levels.
Cheng et al. (30)	60	3	20	-	20 mg/kg for 4 weeks	Immunohistochemical analysis, HE staining, western blot analysis, and qRT-PCR.
Liu et al. (31)	60	5	12	Intravenous injection immediately after reperfusion	10 mg/kg	IF2, myocardial ultrastructure, western blotting and determination of the opening degree of MPTPs.
Zhu et al. (32)	-	-	-	Melatonin stem cells were treated for 24 hours	5 μ M	Measurements of cell culture antioxidant properties, apoptosis, analysis of paracrine factors, LV functions, histology.
Liu et al. (33)	60	6	12	Intraperitoneal injection	Group I: 2.5 mg/kg, Group II: 5 mg/kg, Group III: 10 mg/kg	Hemodynamics measures, apoptosis, electron microscope examination, analysis on mitochondria.
Drobnik et al. (34)	21	3	7	Drinking water for 6 weeks	10 mg/kg	Collagen determination, estimation of glycosaminoglycans, electron microscope examination.
Repova et al. (35)	40	4	10	Drinking water for 6 weeks	10 mg/kg	Collagen determination, hemodynamics measures.
Drobnik et al. (36)	60	5	12	Intraperitoneal injection for 4 weeks	Group 1: 300 μ g/100 g b.w. Group 4: 3 mg/100 g b.w. Group 5: 1.5 mg/100 g b.w.	Estimation of lipid peroxidation, collagen determination, estimation of glycosaminoglycans.
Chen et al. (37)	-	-	-	Intraperitoneal injection 30 min before harvesting the hear for <i>in vitro</i> preparation	150 μ g/kg	Cardiac function, hemodynamics measures, lactate dehydrogenase released, apoptosis, immunohistochemistry.
Petrosillo et al. (38)	42	6	7	Krebs-Henseleit solution for isolated heart	50 μ M	Infarct size, lactate dehydrogenase released, hemodynamics measures, analysis on mitochondria.

IS1, measurement of infarct size by echocardiography; IS2, measurement of infarct size by Evans Blue or tetrazolium; echo, echocardiography measurements; CMEC, cardiac microvascular endothelial cells; IRI, ice recrystallization inhibition; CTRP3, C1q TNF Related Protein 3; NO, nitric oxide; LDL, low-density lipoprotein; HDL, high-density lipoprotein; MDA, malondialdehyde; qRT-PCR, quantitative reverse transcription-polymerase chain reaction; LV, left ventricular; MPTP, mitochondrial permeability transition pore; NF- κ B, Nuclear factor-kappa B; g.b.w., gross body weight; HE, hematoxylin-eosin.



Table 3 - Most frequent recommendations appearing in preclinical research guidelines for *in vivo* animal experiments [Hendersen et al. (18)].

Validity type	Recommendation Category	Examples
Internal	Choice of sample size	Power calculation, larger samples sizes
	Randomized allocation of animals to treatment	Various methods of randomization
	Blinding of outcome assessment	Blinded measurement or analysis
	Flow of animals through an experiment	Recording animals excluded from treatment through to analysis
Construct	Selection of appropriate control groups	Using negative, positive, concurrent, or vehicle control groups
	Study of dose-response relationships	Testing above and below optimal therapeutic dose
	Characterization of animal properties at baseline	Characterizing inclusion/exclusion criteria, disease severity, age or sex
	Matching model to the human manifestation of the disease	Matching mechanism, chronicity or symptoms
External	Treatment response along a mechanistic pathway	Characterizing pathway in terms of molecular biology, histology, physiology or behavior
	Matching outcome measures to clinical settings	Using functional or non-surrogate outcome measures
	Matching model to the age of patients in clinical settings	Using aged or juvenile animals
	Replication in different models of the same disease	Different transgenic strains or lesion techniques
Research program	Independent replication	Different investigators or research groups
	Replication in different species	Rodents and nonhuman primates
	Inter-study standardization of experimental design	Coordination between independent research groups

77.77% of articles selected appropriate control groups (21,23,25-31,34,36) and 61.11% (23,25,26,28,30-32,34,36,38) had well-defined dose-response relationships. Moreover, all studies analyzed (21-38) had standard characterizations of animal properties at baseline. Overall, 89% of studies had employed an appropriate animal model that simulated the human manifestation of the disease (21-23,25-27,29-38). All studies that examined treatment responses according to a known mechanism (21-38) had characteristics within the requested standards. Only 67% (23-25,31,33,35,37,38) of selected manuscripts were within the range of the standard model of patient age in clinical settings. All studies did not follow other standard application models. In replicating different models of the same disease, 78% of studies (21-23,25-30,32,34-38) were independently replicated, whereas 88.88% (21-23,25-29,31-38) were replicated in different species. For studies where the objective was inter-study standardization of an experimental design, 39% (25-28,32,35,37) reached this standard (Table 4).

In Table 5, we analyzed the study characteristics of selected controlled animal studies. Accordingly, we obtained the following results according to each experiment performed in articles examined in this systematic review. Table 5 presents experiments in which melatonin significantly improved the investigated variable (marked as S), as well as those where melatonin showed no significant improvements (NS). Chen et al. (23), Liu et al. (31), and Petrosillo et al. (38) reported that melatonin significantly decreased infarct size. Zhang et al. (21) and Liu et al. (24) reported that melatonin improved echocardiographic measurements. Furthermore, studies by Benova et al. (22), Liu et al. (24), Simko et al. (25), Simko et al. (26), Liu et al. (33), Repova et al. (35), and Chen et al. (37) showed that melatonin had a positive effect on hemodynamic variables. In addition, we observed that the effects of melatonin were not significantly different from those reported in the study by Chaudagar et al. (28). These findings indicated the substantial benefit of using melatonin to stabilize hemodynamic parameters. Moreover, Zhu et al. (32) and Chen et al. (37) revealed that melatonin improved left ventricular cardiac function (Figure 2, Table 5).

Zhang et al. (21), Liu et al. (24), Simko et al. (26), Salmanoğlu et al. (29), Zhu et al. (32), and Chen et al. (37) revealed that melatonin had positive effects on the rate of apoptosis (Table 5). Melatonin showed positive effects in studies examining western blotting of various proteins and quantitative reverse transcription-polymerase chain reaction (qRT-PCR), including those by Zhang et al. (21), Benova et al. (22), Chen et al. (37), Liu et al. (31), and Liu et al. (24) (Table 5). Drobnik et al. (34) and Repova et al. reported that melatonin reportedly reduced collagen deposition (35) (Table 5). Immunohistochemical analyses were performed by Zhang et al. (21), Stacchiotti et al. (27), Cheng et al. (30), Liu et al. (33), Drobnik et al. (34), and Cheng et al. (37), revealing that melatonin consistently yielded a positive result (Table 5). Melatonin showed benefits in biometric analyses, as determined by Benova et al. (22), Simko et al. (25), and Chaudagar et al. (28) (Table 5).

Furthermore, melatonin showed beneficial effects on autophagosome evaluation, lactose dehydrogenase measurements, angiotensin, and aldosterone, nitric oxide levels, and mitochondrial analysis, as determined by Zhang et al. (21), Chen et al. (23), Simko et al. (25), Liu et al. (31), Liu et al. (33), Chen et al. (37), and Petrosillo et al. (38) (Table 5).



Table 4 - Most frequent recommendations in preclinical research guidelines for *in vivo* animal experiments [Henderson et al. (18)].

Validity type	Recommendation Category	Studies	n (Percent of guidelines Citing)
Internal	Choice of sample size	Zhang et al. (21); Benova et al. (22); Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Cheng et al. (30); Liu et al. (31); Liu et al. (33); Repova et al. (35); Drobnik et al. (36); Petrosillo et al. (38).	61.11%
	Randomized allocation of animals to treatment	Zhang et al. (21); Chen et al. (23); Simko et al. (25); Simko et al. (26); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36).	61.11%
	Blinding of outcome assessment	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Liu et al. (24); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Zhu et al. (32); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	100%
	Flow of animals through an experiment	-	-
	Selection of appropriate control groups	Zhang et al. (21); Chen et al. (23); Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Petrosillo et al. (38).	77.77%
	Study of dose-response relationships	Chen et al. (23); Simko et al. (25); Simko et al. (26); Chaudagar et al. (28); Cheng et al. (30); Liu et al. (31); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Petrosillo et al. (38).	61.11%
	Characterization of animal properties at baseline	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Liu et al. (24); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Zhu et al. (32); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	100%
	Matching model to the human manifestation of the disease	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Simko et al. (26); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Zhu et al. (32); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Petrosillo et al. (38).	88.88%
	Treatment response along a mechanistic pathway	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Liu et al. (24); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Zhu et al. (32); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	100%
	Matching outcome measures to clinical settings	Chen et al. (23); Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Liu et al. (33); Repova et al. (36); Chen et al. (37); Petrosillo et al. (38).	66.66%
External	Matching model to the age of patients in clinical settings	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Liu et al. (24); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Liu et al. (31); Zhu et al. (32); Liu et al. (33); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	100%
	Replication in different models of the same disease	-	-
Research program	Independent replication	Zhang et al. (21); Chen et al. (23); Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Cheng et al. (30); Zhu et al. (32); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	77.77%
	Replication in different species	Zhang et al. (21); Benova et al. (22); Chen et al. (23); Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Salimanoglu et al. (29); Liu et al. (31); Zhu et al. (32); Drobnik et al. (34); Repova et al. (35); Drobnik et al. (36); Chen et al. (37); Petrosillo et al. (38).	88.88%
	Inter-study standardization of experimental design	Simko et al. (25); Simko et al. (26); Stacchiotti et al. (27); Chaudagar et al. (28); Zhu et al. (32); Repova et al. (35); Chen et al. (37).	38.88%



Type of bias	Domain	Description of domain	Review authors judgment
Selection bias	Sequence generation	Describe the methods used, if any, to generate the allocation sequence in sufficient detail to allow an assessment of whether it should produce comparable groups.	Was the allocation sequence adequately generated and applied?
Selection bias	Baseline characteristics	Describe all the possible prognostic factors or animal characteristics, if any, that are compared to judge whether intervention and control groups were similar at the start of the experiment.	Were the groups similar at baseline, or were they adjusted for confounders in the analysis?
Selection bias	Allocation concealment	Describe the method used to conceal the allocation sequence in sufficient detail to determine whether intervention allocations could have been foreseen before or during enrolment.	Was the allocation adequately concealed?
Performance bias	Random housing	Describe all measures used, if any, to house animals randomly within the animal room.	Were the animals randomly housed during the experiment?
Performance bias	Blinding	Describe all measurements, if any, to blind trial caregivers and researchers from knowing which intervention each animal received. Provide any information relating to whether the intended blinding was effective.	Were the caregivers and/or investigators blinded from knowledge regarding the intervention each animal received during the experiment?
Detection bias	Random outcome assessment	Describe whether animals were selected at random for outcome assessment and which methods to select the animals, if any, were used.	Were animals selected at random for outcome assessment?
Detection bias	Blinding	Describe all measures used, if any, to blind outcome assessors from knowing which intervention each animal received. Provide any information relating to whether the intended blinding was effective.	Was the outcome assessor blinded?
Attrition bias	Incomplete outcome data	Describe the completeness of outcome data for each main outcome, including attrition and exclusions from the analysis. State whether attrition and exclusions were reported, the numbers in each intervention group (compared with total randomized animals), reasons for attrition or exclusions, and any re-inclusions in analysis for the review.	Were incomplete outcome data adequately addressed?
Reporting bias	Selective outcome reporting	State how selective outcome reporting was examined and what was determined.	Are reports of the study free of selective outcome reporting?
Other	Other sources of bias	State any important concerns regarding bias not covered by other domains in the tool.	Was the study free of other problems that could result in a high risk of bias?

Figure 2 - Representation of the SYRCLÉ's risk of bias tool for animal studies. Hooijmans et al. (43).



Table 5 - Study characteristics of selected controlled animal studies assessing melatonin and the cardiovascular system.

Authors	Assessments					
Zhang et al. (21)	S Echocardiography measurements	S Apoptosis analysis	S Western blotting	S Creatinine kinase measurement	S Immunohistochemical analysis	S Detection of autophagosomes
Benova et al. (22)	S Biometric analysis	S Western blotting	S qRT-PCR	S Hemodynamics measures	NS	
Chen et al. (23)	S Measurements of the infarct size	S Measurement of lactate dehydrogenase	S Measures of CMEC <i>in vitro</i> IRI assay	S Western blotting	S qRT-PCR	S Detection of autophagosomes
Liu et al. (24)	S Echocardiography measurements	S Hemodynamics measurements	S Apoptosis analysis	S Western blotting	S qRT-PCR	
Simko et al. (25)	S Biometric analysis	S Hemodynamics measures	NS Determination of hydroxyproline	S Angiotensin analysis	NS Aldosterone analysis	
Simko et al. (26)	NS Hemodynamics measures	S Determination of hydroxyproline	S NO synthase activity	S Oxidative load	S Measurement and western blotting of NF- κ B	
Stacchiotti et al. (27)	S Histomorphometrically evaluations	S Nuclear cardiomyocyte morphometric	S Mitochondrial analysis	S Immunohistochemical analysis		
Chaudagar et al. (28)	NS Hemodynamics measures	S Biometric analysis	S NO assays			
Salmanoglu et al. (29)	NS Vasocontractile response	NS Measures of total cholesterol, LDL, HDL	S NO assays	S MDA assay	NS Measurements of tissue antioxidant levels	
Cheng et al. (30)	S Immunohistochemical analysis	S Western blotting	S qRT-PCR			
Liu et al. (31)	S Measurements of the infarcted size	S Western blotting	S Determination of the opening degree of MPTPs			
Zhu et al. (32)	S Measurements of cell cultures antioxidant properties	S Apoptosis analysis	S LV functions			
Liu et al. (33)	S Hemodynamics measures	S Apoptosis analysis	S Electron microscope examination	S Analysis on mitochondria		
Drobnik et al. (34)	S Determination of collagens	S Determination of glycosaminoglycans	S Electron microscope examination			
Repova et al. (35)	S Hemodynamics measures	S Determination of collagen				
Drobnik et al. (36)	S Estimation of lipid peroxidation	NS Determination of collagen	S Determination of glycosaminoglycans			

(Continued)



Table 5 - Continued.

Authors	Assessments			
Chen et al. (37)	S	S	S	S
Petrosillo et al. (38)	S	S	S	S
	Cardiac function	Hemodynamics measures	Lactate dehydrogenase	Apoptosis analysis
	Measurements of the infarct size	Lactate dehydrogenase	Hemodynamics measures	Analysis on mitochondria
				Immunohistochemistry

S, statistically significant; NS, not significant; CMEC, cardiac microvascular endothelial cells; IRI, ice recrystallization inhibition; qRT-PCR, quantitative reverse transcription-polymerase chain reaction; NO, nitric oxide; NF-κB, Nuclear factor-kappa B; LDL, low-density lipoprotein; HDL, high-density lipoprotein; MDA, malondialdehyde; LV, left ventricular.

The meta-analysis revealed a statistically significant decrease in infarct size (MD -20.37 [-23.56, -17.18]). However, there was no statistical difference in systolic pressure between articles analyzed (MD -1.75 [-5.47, 1.97]). In articles analyzing lactate dehydrogenase, a statistically significant decrease in the levels of this enzyme was noted in animals in melatonin groups (MD -4.61 [-6.83, -2.40]). With regard to the ejection fraction, two articles showed improvement in melatonin-treated groups. Another study analyzed the influence of melatonin in infarcted animals with the same ejection fraction; however, this parameter was not statistically significant in the meta-analysis (MD -8.12 [-9.56, -6.69]) (Figure 3).

In terms of selection bias, the results were well-balanced between low risk, no clear risk, and high risk of bias. All studies presented a low risk of bias in the baseline variable characteristics. On analyzing allocation concealment, most selected articles had a high risk, and a little less than half presented a low risk of bias. The randomization parameter was also fairly balanced between low risk, no clear risk, and high risk of bias. On analyzing random outcome assessment, most studies (more than 50%) had a low risk of bias, and some presented an unclear risk of bias. On analyzing blinding bias, most articles were unclear as to whether investigators were blinded. The articles presented a low risk of bias in the results of incomplete outcome data (Figure 4 and 5).

DISCUSSION

This systematic review revealed that melatonin has various beneficial effects on the cardiovascular system; these effects include decreased infarct size, improved cardiac function and cellular oxidation functions, reduced apoptosis, and healthier cellular histomorphology.

In the present review, studies that analyzed echocardiographic measures exhibited melatonin benefits such as decreased infarct size, improved ejection fractions, improved systolic and diastolic diameters, and ameliorated recovery rates of cardiac function (24), while also favoring the treatment of cardiac hypertrophy and hearts that experienced myocardial infarction, ischemia, or reperfusion (21-38). On analyzing hemodynamic and biometric variables, melatonin appeared to confer significant benefits, such as improvements in systolic pressure, positive pressure derivative, lower left ventricular end-diastolic pressure, reduced left ventricular weight in relation to the total heart weight, and improved lung water content (22-25,28,35,38). Experimental models of obesity, hypertension, and other cardiovascular diseases reinforce the scientific practice of adopting animal models and assessing results prior to human application. These pre-clinical results indicate the effect of melatonin on the examined cardiovascular diseases.

Reportedly, melatonin is an important anti-apoptotic agent in various tissues, reducing calcium uptake, mitigating reactive oxygen species generation, and decreasing the levels of pro-apoptotic proteins, such as Bax (39). In addition, melatonin destabilizes hypoxia-induced hypoxia-inducible factor (HIF)-1α protein expression. Moreover, melatonin suppresses HIF-1α transcriptional activity under hypoxic conditions, resulting in vascular endothelial growth factor expression (40). Melatonin also confers anti-inflammatory effects on the cardiovascular system (41). Furthermore, a systematic review and recent meta-analysis have identified

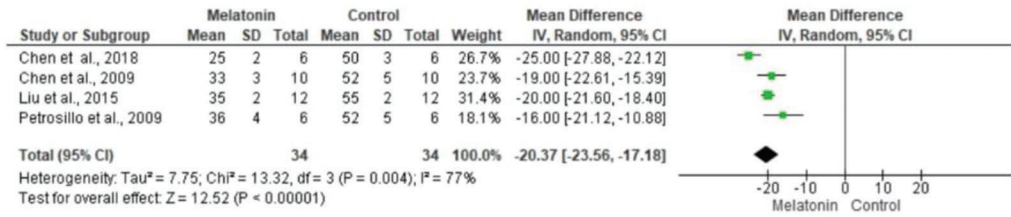


Figure 3a - Metanalysis of infarct size measurement by echocardiography (% left ventricular).

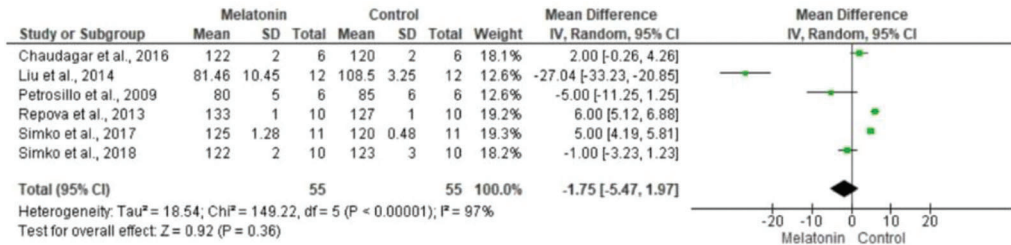


Figure 3b - Metanalysis of systolic blood pressure (mmHg).

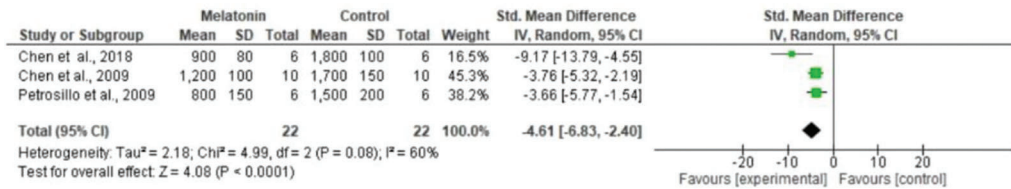


Figure 3c - Metanalysis of lactate dehydrogenase (U/L).

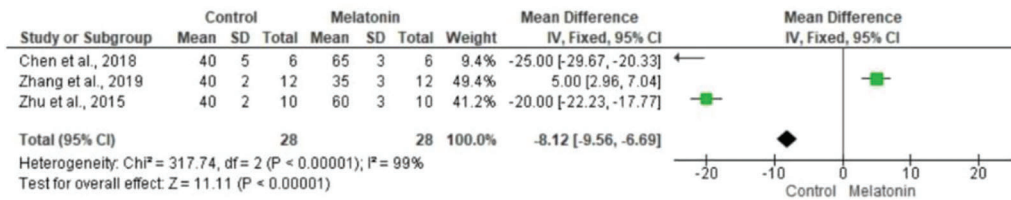


Figure 3d - Metanalysis of ejection fraction measured by echocardiography (% left ventricular).

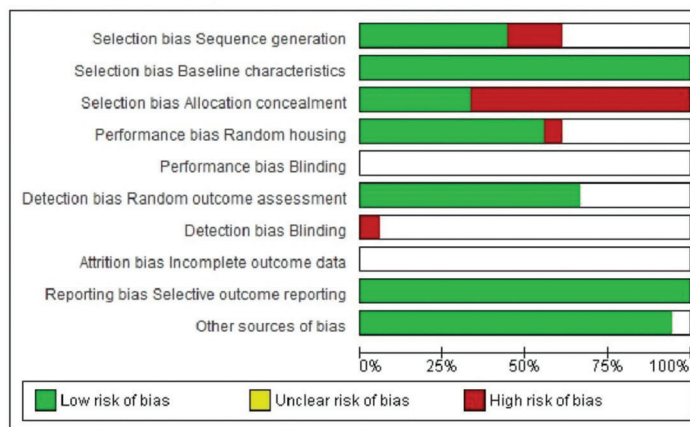


Figure 4 - Risk of bias graph: review of authors' judgment regarding each risk of bias item presented as percentages across all included studies.



	Selection bias Sequence generation	Selection bias Baseline characteristics	Selection bias Allocation concealment	Performance bias Random housing	Performance bias Blinding	Detection bias Random outcome assessment	Detection bias Blinding	Attrition bias Incomplete outcome data	Reporting bias Selective outcome reporting	Other sources of bias
Benova et al., 2019	+	+	-	+			-		+	
Chaudagar et al., 2016		+	-						+	+
Chen et al., 2009		+	+	-					+	+
Chen et al., 2018	+	+	-			+			+	+
Cheng et al., 2015		+	-			+			+	+
Drobnik et al., 2011	+	+	+	+		+			+	+
Drobnik et al., 2013	+	+	+			+			+	+
Liu et al., 2014	+	+	-	+					+	+
Liu et al., 2015	+	+	-	+		+			+	+
Liu et al., 2018	-	+	-	+					+	+
Petrosillo et al., 2009	+	+	+			+			+	+
Repova et al., 2013	-	+	-	+					+	+
Salmanoglu et al., 2016	+	+	-	+		+			+	+
Simko et al., 2017		+	-	+		+			+	+
Simko et al., 2018		+	-			+			+	+
Stacchiotti et al., 2017	-	+	+			+			+	+
Zhang et al., 2019	+	+	-	+		+			+	+
Zhu et al., 2015		+	+	+		+			+	+

Figure 5 - Risk of bias summary: review of authors' judgment regarding each risk of bias item for each included study.

that melatonin supplementation facilitates blood pressure regulation (42).

Melatonin has substantial benefits in the heart, involving various proteins (including superoxide dismutase [SOD], catalase [CAT], and glutathione peroxidase [Gpx]), while also improving the apoptosis rate. These findings were determined using several techniques, including western blotting analysis of BCL and Bx expression and the TUNEL assay, which measured the decrease in the level of

apoptosis in myocardial cells when melatonin was added (23-25,29,32). Other important variables analyzed following melatonin administration in the cardiovascular system were lactate dehydrogenase levels, mitochondrial analysis, lipid peroxidation, glycosaminoglycan, collagen level reduction, culture measurements, the antioxidant action of cells, opening gradient of mitochondrial channels, improvement in vasoconstriction, low-density lipoprotein (LDL), high-density lipoprotein (HDL), and cholesterol level measurements, nitric oxide



synthase level measurements, histomorphometric evaluations, determination of hydroxyproline levels, and assessment of autophagosomes (21-38).

The main novelty of this study is that it highlights the benefits assimilated by melatonin in experimental models of myocardial infarction, such as improved ejection fraction. Apart from limitations such as differences between animal organisms and humans, experimental research in Brazil is often restricted due to limited funding for animal studies when compared with human trials. In addition, results from animal studies fail to precisely correlate with the experience of testing melatonin or other substances in an environment that differs from the human body. Another limitation that must be considered is the nature of systematic reviews, which examine non-published data and data previously published by other authors, thus hindering novel scientific findings.

CONCLUSION

Notably, this systematic review is based on animal experiments. Melatonin may impact the cardiovascular system, including experimental myocardial infarction, and further studies are necessary to determine its use in clinical settings for treating cardiovascular diseases.

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AUTHOR CONTRIBUTIONS

Veiga ECA contributed substantially to the study conception and design, definition of intellectual content, was involved in literature search, data analysis, statistical analysis, and manuscript preparation, drafting and critical review for important intellectual content, and approved the final manuscript version to be published. Simões RS, Caviola LL, Abreu LC and Cavalli RC were involved in data analysis and statistical analysis, manuscript drafting and critical review for important intellectual content, and approved the final manuscript version to be published. Cipolla-Neto J, Baracat EC and Soares Junior JM substantially contributed to the study conception and design, definition of intellectual content, were involved in manuscript preparation, drafting and critical review for important intellectual content, and approved the final manuscript version to be published.

REFERENCES

1. Tordjman S, Chokron S, Delorme R, Charrier A, Bellissant E, Jaafari N, et al. Melatonin: Pharmacology, Functions and Therapeutic Benefits. *Curr Neuropharmacol*. 2017;15(3):434-43. <https://doi.org/10.2174/1570159X14666161228122115>
2. Amaral FGD, Cipolla-Neto J. A brief review about melatonin, a pineal hormone. *Arch Endocrinol Metab*. 2018;62(4):472-9. <https://doi.org/10.20945/2359-3997000000066>
3. Cipolla-Neto J, Amaral FGD. Melatonin as a Hormone: New Physiological and Clinical Insights. *Endocr Rev*. 2018;39(6):990-1028. <https://doi.org/10.1210/er.2018-00084>
4. Sharma S, Singh H, Ahmad N, Mishra P, Tiwari A. The role of melatonin in diabetes: therapeutic implications. *Arch Endocrinol Metab*. 2015;59(5):391-9. <https://doi.org/10.1590/2359-3997000000098>
5. Favero G, Franceschetti L, Buffoli B, Moghadasian MH, Reiter RJ, Rodella LF, et al. Melatonin: Protection against age-related cardiac pathology. *Ageing Res Rev*. 2017;35:336-49. <https://doi.org/10.1016/j.arr.2016.11.007>
6. Baltatu OC, Amaral FG, Campos LA, Cipolla-Neto J. Melatonin, mitochondria and hypertension. *Cell Mol Life Sci*. 2017;74(21):3955-64. <https://doi.org/10.1007/s00018-017-2613-y>

7. Rodella LF, Favero G, Rossini C, Foglio E, Bonomini F, Reiter RJ, et al. Aging and vascular dysfunction: beneficial melatonin effects. *Age (Dordr)*. 2013;35(1):103-15. <https://doi.org/10.1007/s11357-011-9336-z>
8. Dominguez-Rodriguez A, Abreu-Gonzalez P. Pharmacological cardioprotection in the acute myocardial infarction: potential of β -blockers and melatonin as forgotten cardioprotective agents. *Int J Cardiol*. 2014;172(2):e354-5. <https://doi.org/10.1016/j.ijcard.2013.12.303>
9. Dziegial P, Murawska-Ciałowicz E, Jethon Z, Januszewska L, Podhorska-Okołów M, Surowiak P, et al. Melatonin stimulates the activity of protective antioxidant enzymes in myocardial cells of rats in the course of doxorubicin intoxication. *J Pineal Res*. 2003;35(3):183-7. <https://doi.org/10.1034/j.1600-079X.2003.00079.x>
10. Dominguez-Rodriguez A, Abreu-Gonzalez P, Sanchez-Sanchez JJ, Kaski JC, Reiter RJ. Melatonin and circadian biology in human cardiovascular disease. *J Pineal Res*. 2010;49(1):14-22. <https://doi.org/10.1111/j.1600-079X.2010.00773.x>
11. Dominguez-Rodriguez A, Abreu-Gonzalez P, Reiter RJ. Melatonin and cardiovascular disease: myth or reality? *Rev Esp Cardiol (Engl Ed)*. 2012;65(3):215-8. <https://doi.org/10.1016/j.recsep.2011.10.009>
12. Paredes SD, Forman KA, García C, Vara E, Escames G, Tresguerres JA. Protective actions of melatonin and growth hormone on the aged cardiovascular system. *Horm Mol Biol Clin Investig*. 2014;18(2):79-88. <https://doi.org/10.1515/hmbci-2014-0016>
13. Sun H, Gusdon AM, Qu S. Effects of melatonin on cardiovascular diseases: progress in the past year. *Curr Opin Lipidol*. 2016;27(4):408-13. <https://doi.org/10.1097/MOL.0000000000000314>
14. McSweeney JC, Rosenfeld AG, Abel WM, Braun LT, Burke LE, Daugherty SL, et al. Preventing and Experiencing Ischemic Heart Disease as a Woman: State of the Science: A Scientific Statement From the American Heart Association. *Circulation*. 2016;133(13):1302-31. <https://doi.org/10.1161/CIR.0000000000000381>
15. Tawfik GM, Dila KAS, Mohamed MYF, Tam DNH, Kien ND, Ahmed AM, et al. A step by step guide for conducting a systematic review and meta-analysis with simulation data. *Trop Med Health*. 2019;47:46. <https://doi.org/10.1186/s41182-019-0165-6>
16. Shamseer L, Moher D, Clarke M, Ghersi D, Liberati A, Petticrew M, et al. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. *BMJ*. 2015;350:g7647. <https://doi.org/10.1136/bmj.g7647>
17. PRISMA harms checklist: improving harms reporting in systematic reviews. *BMJ*. 2016;353:i2229. <https://doi.org/10.1136/bmj.i157>
18. Henderson VC, Kimmelman J, Fergusson D, Grimshaw JM, Hackam DG. Threats to validity in the design and conduct of preclinical efficacy studies: a systematic review of guidelines for in vivo animal experiments. *PLoS Med*. 2013;10(7):e1001489. <https://doi.org/10.1371/journal.pmed.1001489>
19. Adriaenssens P, Eggermont E, Smeyers L. Een regionaal vangnet voor kindermishandeling: Het netwerk kind en gezin in nood. *Tijdschrift voor Kindergeneeskunde*. 1996.
20. Rhodes KM, Turner RM, Savovi J, Jones HE, Mawdsley D, Higgins JPT. Between-trial heterogeneity in meta-analyses may be partially explained by reported design characteristics. *J Clin Epidemiol*. 2018;95:45-54. <https://doi.org/10.1016/j.jclinepi.2017.11.025>
21. Zhang WX, He BM, Wu Y, Qiao JF, Peng ZY. Melatonin protects against sepsis-induced cardiac dysfunction by regulating apoptosis and autophagy via activation of SIRT1 in mice. *Life Sci*. 2019;217:8-15. <https://doi.org/10.1016/j.lfs.2018.11.055>
22. Egan Benova T, Viczenczova C, Szeiffova Bacova B, Knezl V, Dosenko V, Rauchova H, et al. Obesity-associated alterations in cardiac connexin-43 and PKC signaling are attenuated by melatonin and omega-3 fatty acids in female rats. *Mol Cell Biochem*. 2019;454(1-2):191-202. <https://doi.org/10.1007/s11010-018-3463-0>
23. Chen WR, Liu HB, Chen YD, Sha Y, Ma Q, Zhu PJ, et al. Melatonin Attenuates Myocardial Ischemia/Reperfusion Injury by Inhibiting Autophagy Via an AMPK/mTOR Signaling Pathway. *Cell Physiol Biochem*. 2018;47(5):2067-76. <https://doi.org/10.1159/000491474>
24. Liu Y, Li LN, Guo S, Zhao XY, Liu YZ, Liang C, et al. Melatonin improves cardiac function in a mouse model of heart failure with preserved ejection fraction. *Redox Biol*. 2018;18:211-21. <https://doi.org/10.1016/j.redox.2018.07.007>
25. Simko F, Baka T, Krajcirovicova K, Repova K, Aziriova S, Zorad S, et al. Effect of Melatonin on the Renin-Angiotensin-Aldosterone System in L-NAME-Induced Hypertension. *Molecules*. 2018;23(2):265. <https://doi.org/10.3390/molecules23020265>
26. Simko F, Pechanova O, Repova K, Aziriova S, Krajcirovicova K, Celec P, et al. Lactacystin-Induced Model of Hypertension in Rats: Effects of Melatonin and Captopril. *Int J Mol Sci*. 2017;18(8):1612. <https://doi.org/10.3390/ijms18081612>
27. Stacchiotti A, Favero G, Giugno L, Golic I, Korac A, Rezzani R. Melatonin Efficacy in Obese Leptin-Deficient Mice Heart. *Nutrients*. 2017;9(12):1323. <https://doi.org/10.3390/nu9121323>
28. Chaudagar KK, Viczenczova C, Szeiffova Bacova B, Egan Benova T, Barancik M, Tribulova N. Modulation of systemic and aortic nitric oxide



- by melatonin and n-3 polyunsaturated fatty acids in isoproterenol affected spontaneously hypertensive and normotensive Wistar rats. *Physiol Res.* 2016;65 Suppl 1:S109-18
29. Salmanoğlu DS, Gurpinar T, Vural K, Ekerbicer N, Dariverenli E, Var A. Melatonin and L-carnitin improves endothelial dysfunction and oxidative stress in Type 2 diabetic rats. *Redox Biol.* 2016;8:199-204. <https://doi.org/10.1016/j.redox.2015.11.007>
 30. Cheng X, Wan Y, Xu Y, Zhou Q, Wang Y, Zhu H. Melatonin alleviates myosin light chain kinase expression and activity via the mitogen-activated protein kinase pathway during atherosclerosis in rabbits. *Mol Med Rep.* 2015;11(1):99-104. <https://doi.org/10.3892/mmr.2014.2753>
 31. Liu LF, Qian ZH, Qin Q, Shi M, Zhang H, Tao XM, et al. Effect of melatonin on oncosis of myocardial cells in the myocardial ischemia/reperfusion injury rat and the role of the mitochondrial permeability transition pore. *Genet Mol Res.* 2015;14(3):7481-9. <https://doi.org/10.4238/2015.July.3.24>
 32. Zhu P, Liu J, Shi J, Zhou Q, Liu J, Zhang X, et al. Melatonin protects ADSCs from ROS and enhances their therapeutic potency in a rat model of myocardial infarction. *J Cell Mol Med.* 2015;19(9):2232-43. <https://doi.org/10.1111/jcmm.12610>
 33. Liu LF, Qin Q, Qian ZH, Shi M, Deng QC, Zhu WP, et al. Protective effects of melatonin on ischemia-reperfusion induced myocardial damage and hemodynamic recovery in rats. *Eur Rev Med Pharmacol Sci.* 2014;18(23):3681-6.
 34. Drobnik J, Tosik D, Piera L, Szczepanowska A, Olczak S, Zielinska A, et al. Melatonin-induced glycosaminoglycans augmentation in myocardium remote to infarction. *J Physiol Pharmacol.* 2013;64(6):737-44.
 35. Repová-Bednářová K, Aziriová S, Hrenák J, Krajcirovicova K, Adamcová M, Paulis L, et al. Effect of captopril and melatonin on fibrotic rebuilding of the aorta in 24 hour light-induced hypertension. *Physiol Res.* 2013;62(Suppl 1):S135-41
 36. Drobnik J, Slotwinska D, Olczak S, Tosik D, Pieniazek A, Matczak K, et al. Pharmacological doses of melatonin reduce the glycosaminoglycan level within the infarcted heart scar. *J Physiol Pharmacol.* 2011;62(1):29-35.
 37. Chen Z, Chua CC, Gao, J, Chua KW, Ho YS, Hamdy RC, et al. Prevention of ischemia/reperfusion-induced cardiac apoptosis and injury by melatonin is independent of glutathione peroxidase 1. *J Pineal Res.* 2009;46(2):235-41. <https://doi.org/10.1111/j.1600-079X.2008.00654.x>
 38. Petrosillo G, Colantuono G, Moro N, Ruggiero FM, Tiravanti E, Di Venosa N, et al. Melatonin protects against heart ischemia-reperfusion injury by inhibiting mitochondrial permeability transition pore opening. *Am J Physiol Heart Circ Physiol.* 2009;297(4):H1487-93. <https://doi.org/10.1152/ajpheart.00163.2009>
 39. Ferreira Cda S, Maganhin CC, Simões Rdos S, Girão MJ, Baracat EC, Soares JM Jr. [Melatonin: cell death modulator]. *Rev Assoc Med Bras (1992).* 2010;56(6):715-8. <https://doi.org/10.1590/S0104-42302010000600024>
 40. Park SY, Jang WJ, Yi EY, Jang JY, Jung Y, Jeong JW, et al. Melatonin suppresses tumor angiogenesis by inhibiting HIF-1alpha stabilization under hypoxia. *J Pineal Res.* 2010;48(2):178-84. <https://doi.org/10.1111/j.1600-079X.2009.00742.x>
 41. Paredes SD, Forman KA, Garcia C, Vara E, Escames G, Tresguerres JA. Protective actions of melatonin and growth hormone on the aged cardiovascular system. *Horm Mol Biol Clin Investig.* 2014;18(2):79-88. <https://doi.org/10.1515/hmbci-2014-0016>
 42. Hadi A, Ghaedi E, Moradi S, Pourmasoumi M, Ghavami A, Kafeshani M. Effects of Melatonin Supplementation On Blood Pressure: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Horm Metab Res.* 2019;51(3):157-64. <https://doi.org/10.1055/a-0841-6638>
 43. Hooijmans CR, Rovers MM, de Vries RB, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCLE's risk of bias tool for animal studies. *BMC Med Res Methodol.* 2014;14:43. <https://doi.org/10.1186/1471-2288-14-43>