Article

Water Dynamics at the Solid–Liquid Interface to Unveil the Textural Features of Synthetic Nanosponges

Paolo Lo Meo,* Federico Mundo, Samuele Terranova, Pellegrino Conte,* and Delia Chillura Martino



ABSTRACT: A fast-field-cycling NMR investigation was carried out on a set of polyurethane cyclodextrin nanosponges, in order to gain information on their textural properties, which have been proven to be quite difficult to assess by means of ordinary porosimetric techniques. Experiments were performed on both dry and wet samples, in order to evaluate the behavior of the "nonexchangeable" C-bound ¹H nuclei, as well as the one of the mobile protons belonging to the skeletal hydroxyl groups and the water molecules. The results acquired for the wet samples accounted for the molecular mobility of water molecules within the channels of the nanosponge network, leading back to the possible pore size distribution. Owing to the intrinsic difficulties involved in a quantitative assessment of the textural properties, in the present study we alternatively propose an extension to nanosponges of



the concept of "connectivity", which has been already employed to discuss the properties of soils.

INTRODUCTION

Nanosponges (NSs)¹⁻³ are an emerging class of smart materials. Their tunable absorption and release abilities toward both organic⁴ and inorganic⁵ species make them ideal candidates as platforms for drug carrier/delivery systems,^{6,7} environmental remediation devices,^{8,9} and supports for metal nanoparticle catalysts.^{10–13} These materials are obtained by reacting supramolecular host units (e.g., cyclodextrins,¹⁴ calixarenes,¹⁵ pillararenes¹⁶) with suitable reticulating agents that afford the linker units. The properties of NSs can be widely tuned by either premodification of the host monomers or postsynthesis chemical modification of the obtained product.¹⁷ Moreover, reticulating agents bearing ionizable (polyamines,¹⁸ triazoles,¹⁹ pyromellytic anhydride^{20,21}) or other stimuli-sensitive²² groups can be used; hence, functional tailored materials can be obtained.

A reliable evaluation of the textural features, such as average pore size (*D*), specific surface area (*S*), and specific pore volume (*V*), constitutes one of the main issues in the characterization of NSs, in order to rationalize their properties. Because of their peculiar hyper-reticulated highly disordered structure, NSs are supposed to present a thick network of nanosized channels between the host monomers. However, it has been reported on several occasions that the ordinary methodologies based on the N₂ gas adsorption isotherms, analyzed by the well-known BET²⁻³ and BJH²⁻⁴ approaches, result in abnormally low values (sometimes even below the instrumental sensitivity limits), unless particularly long and rigid structural units are used as the linkers. For instance, *S* values ranging up to 263 m² g⁻¹ were claimed for a series of

materials obtained from β -cyclodextrin (β CD) and tetrafluoroterephtalonitrile.²⁵ Comparable results were found by using decafluorobiphenyl,²⁶ whereas an outstanding area of 1225 m² g^{-1} and a pore volume of 1.71 cm³ g^{-1} were found for a material obtained by reacting a *per*-benzyloxy-cyclodextrin with formaldehyde dimethylacetal.²⁷ Nevertheless, beyond these few exceptions, S values by far below 10 m² g⁻¹ are usually found for materials prepared from cyclodextrins and typical reticulating agents such as diisocyanates,^{28,29} epichlorohy-drin,^{30,31} or poly(carboxylic acid)s.^{32,33} Similarly, *S* values not larger than 8 m^2 g⁻¹ were found for a series of cyclodextrincalixarene copolymers, together with V values smaller than 0.03 $cm^3 g^{-1}$ and \tilde{S} values smaller than 4 nm.³⁴ Wilson et al. recently examined a set of polyurethane β CD polymers obtained with diverse reticulating diisocyanates.³⁵ They studied the relevant textural properties by both the ordinary BET method and a different approach based on the absorption of a suitable probe dye, namely, p-nitrophenol, having a known molecular area (0.525 nm²) and volume (0.0908 nm³). The latter methodology relies on the evaluation of the maximum absorption capacity of the material, obtained by analyzing the absorption isotherms by either the Langmuir or the Sips model. It led to the estimation of S values on the order of several hundreds of m² g⁻¹, which undoubtedly appears as a much more sensible

Received:December 26, 2019Revised:February 3, 2020Published:February 18, 2020



result, in comparison to N_2 adsorption. However, even this method is not devoid of criticism (see later), because the results obtained critically depend on the choice of the probe molecule and can be unpredictably affected by rearrangements of the microscopic structure due to swelling.³⁵ Therefore, it seems that the concept itself of "surface" has conceivably a somehow elusive or deceptive meaning in the case of NS materials, so that new paradigms should be possibly explored.

Since the seminal work by Brownstein and Tarr,³⁶⁻³⁸ fastfield-cycling (FFC) NMR relaxometry has been used as a valuable tool to assess water mobility at the liquid-solid interface (and also to study the microscopic dynamics of polymers $^{39-44}$ and proteins $^{45-48}$). Data interpretation can be related to the textural features of solid systems such as clays and microporous materials in general.⁴⁹⁻⁵⁹ Very briefly (the bases of FFC-NMR relaxometry are summarized in the Supporting Information), this technique relies on the simple though counterintuitive idea that the tighter a water molecule is bound to the surface of a porous wet system (that is the more restricted its motion is), the faster the longitudinal relaxation rate (R_1) is that the water ¹H nuclei will experience (the same is true, indeed, also for transverse relaxation, which we do not consider here). Thus, as long as water is trapped into micropores, mesopores, or macropores, the observed R_1 values decrease in the same order.⁶⁰ Therefore, FFC-NMR relaxometry can also be used as a valid alternative to the traditional porosimetry investigations to obtain pore size distributions.⁶⁰ Very recently, FFC-NMR relaxometry has been also applied in soil science⁶¹ to quantitatively measure the hydrological connectivity inside the soil (HCS).^{62,63} In general, connectivity refers to the processes involving a transfer of matter, energy, and/or organisms within or between elements of an ecological system. This implies the presence of a transport vector, such as water, and accounts for how mobility within the spatial patterns inside the soil (referred to as the "structural" connectivity) affects the occurrence of physicochemical processes (subsurface flow, sediment transport, etc., which is referred to as the "functional" connectivity).^{62,6}

Due to the presence of channels and sinks in the structure of NSs, we reasoned that the concept of connectivity developed in ecology and soil science might also be suitable for the description of these materials. In order to develop a different methodology for assessing their texture features, in the present work we performed an FFC-NMR study of three NS materials (referred to as NS1, NS2, and NS3) obtained by reacting β -cyclodextrin with hexamethylene-diisocyanate (HMDI, Figure 1).

Different mole-to-mole ratios (i.e., 1:4, 1:2 and 1:1, respectively) were applied, in order to achieve a different extent of reticulation. The reaction between the -OH groups of β CD and the -N=C=O groups of HMDI results in the



Figure 1. Structures of β CD and HMDI.

formation of urethane units. The relaxometric behavior of the nanosponges obtained was studied for both dry and wet samples, in order to consider the dynamics of water molecules into the nanochannels subjected to possible swelling.

EXPERIMENTAL SECTION

Materials and Instrumentation. All the reagents and solvents needed were used as purchased (Sigma-Aldrich, Cyclolab), without further purification. Anhydrous β -cyclodextrin was obtained by drying the commercial hydrated product in vacuo over phosphorus pentoxide at 90 °C overnight.

Spectrophotometric determinations of *p*-nitrophenol concentration (for obtaining absorption isotherms) were carried out with a Beckmann Coulter DU 800 apparatus. ATR-FTIR spectra were recorded on a PerkinElmer SPECTRUM TWO instrument. The N_2 absorption–desorption isotherms were registered at 77 K using a Quantachrome Nova 2200 multistation high speed gas sorption analyzer. Thermogravimetric analyses were performed on a Q5000 IR apparatus (TA Instruments) under nitrogen flow (25 cm³ min⁻¹).

Synthesis and FTIR Characterization. Preparation of materials NS1, NS2, and NS3 was accomplished according to literature reports.³⁵

NS1. Anhydrous β CD (568 mg, 5 × 10⁻⁴ mol) was mixed in a glass vial with a solution prepared by dissolving 320 μL of HMDI (336 mg, 2×10^{-3} mol) in 400 μ L of dry DMSO. The system was mechanically stirred with a tiny steel rod to ensure effective mixing and then was kept still at 60 °C for 18 h. The hard reaction crude was coarsely crunched and suspended into 40 mL of distilled water; the suspension was sonicated for ca. 10 min, and the solid residue was finally recovered by centrifugation (5500 rpm for 10 min). The residue was then suspended in 40 mL of methanol, sonicated, and centrifuged as described above. The same washing procedure was iterated with another portion of methanol (40 mL) and then with diethyl ether (40 mL). The solid was finally recovered by filtration, finely crunched, passed through a 150 μ m sieve, and dried overnight, by being kept in vacuo at 60 °C over phosphorus pentoxide. Yield 885 mg.

NS2. The same procedure as for NS1 was followed, starting from 568 mg (5×10^{-4} mol) of β CD and 160 μ L (168 mg, 1 × 10^{-3} mol) of HMDI. Yield 721 mg.

NS3. The same procedure as for NS1 was followed, starting from 568 mg (5 × 10⁻⁴ mol) of β CD and 80 μ L (84 mg 5 × 10⁻⁴ mol) of HMDI. Yield 455 mg.

In general, the accomplishment of the reticulation process was immediately verified by the mechanical hardness and lack of solubility of the reaction products, which were recovered almost quantitatively in the cases of NS1 and NS2. Conversely, NS3 was partly soluble in water, thereby providing a lesser reaction yield (ca. 70%). The latter observation can be justified by the insufficient amount of HMDI, which prevented a full reticulation.

The formation of the polymeric network was confirmed by ATR-FTIR analysis (spectra are synoptically shown in Figure 2). The main features in the spectrum of the starting β CD are the large O–H str (stretching) band centered at 3304 cm⁻¹, followed by a tiny C–H str signal at 2918 cm⁻¹ and by the typical β CD fingerprints system (various C–O str) in the range ~1200–900 cm⁻¹. On the other hand, the spectrum of HMDI shows a system of two signals at 2940 and 2863 cm⁻¹ (asymmetric and symmetric str of the methylene units) and an



Figure 2. ATR-FTIR spectra of β CD and HMDI and materials NS1, NS2, NS3.

intense band at 2276 cm⁻¹ (-N=C=O str). On passing to nanosponge materials, NS1 and NS2 spectra show the following main features: (i) two C-H str signals at 2928 and 2857 cm⁻¹, accounting for the presence of methylene groups of the linker bridges; (ii) a system of three signals at 1694, 1630 (amide-I-like carbonyl str), and 1548 cm^{-1} (amide-II-like N-H bend), accounting for the presence of the urethane groups formed during the reticulation; and (iii) a strong signal at 1251 cm⁻¹, conceivably due to the O-carbonyl str of the urethane groups. The same signals, though much weaker, can also be identified in the spectrum of NS3. Finally, all the NSs show the β CD fingerprints at ~1200-900 cm⁻¹ Thus, the actual accomplishment of the reticulation reaction (and the fact that the materials are not mere physical mixtures of the reactants) is positively assessed by the simultaneous presence of signals that can be traced back to the β CD and the hexamethylene chain backbones, and to the newly formed urethane functional groups as well, together with the absence of any signal relevant to unreacted isocyanate groups.

Relaxometry. Relaxometric experiments were performed on a Stelar Spinmaster FFC 2000 relaxometer (Stelar s.r.l., Mede, PV-Italy) at the constant temperature of 25 °C. The proton spins were polarized at a polarization field (B_{POL}) corresponding to a proton Larmor frequency ($\omega_{\rm L}$) of 10 MHz for a period of polarization corresponding to about 5 times the T_1 estimated at this frequency. After each B_{POL} application, the magnetic field intensity (indicated as B_{RLX}) was systematically changed through the proton Larmor frequency $\omega_{\rm L}$ range 0.015–35 MHz. The period τ_i , during which B_{RLX} was applied, was varied on 32 logarithmic spaced time sets, each of them adjusted at every relaxation field in order to optimize the sampling of the decay/recovery curves. FIDs were recorded following a single ¹H 90° pulse of 5.5 μ s applied at an acquisition field corresponding to the proton Larmor frequency of 7.20 MHz. A time domain of 100 μ s sampled with 1000 points was applied. The field-switching time was 3 ms, while the spectrometer dead time was 15 μ s. For all experiments, a recycle delay of 1 s was used. The nonpolarized FFC sequence was applied when the relaxation magnetic fields were in the range of the proton Larmor frequencies between 35 and 9 MHz. A polarized FFC sequence was applied in the proton Larmor frequencies B_{RLX} range 9-0.015 MHz.

RESULTS AND DISCUSSION

Porosimetry. Porosimetric determinations (i.e., N₂ absorption isotherms) performed on the fully insoluble materials NS1 and NS2 afforded, as expected, very poor results, partly beyond

the lower sensitivity limit of the applied instrumental apparatus. In particular, by means of the BET analysis, an S value of $0.134 \text{ m}^2 \text{ g}^{-1}$ for NS2 only was found. The use of the BJH analysis resulted in S values of 0.115 and 0.280 m² g⁻¹ for NS1 and NS2, respectively. This trend follows the reticulation degree expected on the grounds of the amount of reticulating agent used in the syntheses. A V value of 0.001 cm³ g⁻¹ was found for both NS1 and NS2, with an average pore diameter of 7.1 and 3.8 nm for the two systems, respectively. The latter finding does not appear, consistent with the expected reticulation degree. On the whole, these results agree with those reported elsewhere,³⁵ thereby confirming that N₂ absorption is not a suitable method for the measurement of nanosponge textures. The reasons of such a failure are unclear, and currently an object of debate.³⁵ Probably, the assumptions on which the BET/BJH analyses rely cannot be properly applied in the case of our materials. In particular, the equivalence of the binding sites onto the surface of the nanopores cannot be taken for granted on a microscopic scale; moreover, the system can hardly be modeled as a network of cylindrical pores, as provided by theory.

We also evaluated the specific surface areas and volumes of NS1 and NS2 by means of the p-nitrophenol absorption method. Absorption isotherms were studied at 25 °C in aqueous acetate buffer at pH 4.4. Data were subjected to regression analysis by means of the Sips equation in the form $q_e = q_{max}(Kc_{eq})^n / [1 + (Kc_{eq})^n]$, where q_e and q_{max} are the amount of guest absorbed at equilibrium and the maximum capacity of the material (mol g^{-1}), respectively; c_{eq} is the concentration of the guest at equilibrium; K is the apparent equilibrium constant; and n is an empirical coefficient. We found the following for NS1: $q_{\text{max}} = (7.8 \pm 0.6) \times 10^{-4}$ mol g^{-1} , $K = (2.1 \pm 0.2) \times 10^3$ M⁻¹, $n = 1.36 \pm 0.08$. For NS2, $q_{\text{max}} = (7.9 \pm 0.9) \times 10^{-4}$ mol g^{-1} , $K = (1.74 \pm 0.06) \times 10^3$ M⁻¹, and $n = 1.21 \pm 0.09$. As a consequence, very similar apparent S $(250 \pm 30 \text{ m}^2 \text{ g}^{-1})$ and $V (0.043 \pm 0.004 \text{ cm}^3 \text{ g}^{-1})$ values can be estimated for both materials. These results are comparable with those reported by Wilson et al.³⁵ for a material with the same composition as NS2 (i.e., $q_{\text{max}} = 11.5 \times 10^{-4} \text{ mol g}^{-1}$, S =364 m² g⁻¹). The discrepancy between the q_{max} values is likely due to the presence of the buffer, according to the well-known effect of electrolytes on the binding equilibria of free cyclodextrins in solution.^{64–66}

The previous results can be interestingly compared with the composition of NS1 and NS2, which contain 5.5×10^{-4} and 6.8×10^{-4} mol g⁻¹ of β CD, respectively. In general, psubstituted nitrobenzene derivatives form in solution only 1:1 complexes both with native and chemically modified β CDs;⁶⁷⁻⁷⁰ however, due to the smaller volume of these guests as compared to the host cavity, a simultaneous dynamic coinclusion of solvent molecules occurs.⁷⁰ Moreover, evidence has been reported (2D-LFSE solid-state NMR) that, in the case of the inclusion of a *p*-nitroaniline derivative in a polyamino-cyclodextrin NS, the guest specifically occupies the cyclodextrin cavities and does not reside in the nanochannels.¹⁸ Now, the q_{max} values found experimentally suggest that most of the *p*-nitrophenol guest should be included into the host cavities, whereas only a minor amount (i.e., ca. 30% for NS1, ca. 14% for NS2) may reside into the nanochannels. This implies that the channel surface, which represents the interspace between the cyclodextrin units, is not adequately sampled. Furthermore, taking into account the intrinsic volume of the β CD cavity (i.e., 0.262 nm³, which corresponds to 15.77

cm³ mol⁻¹ or 0.14 cm³ g⁻¹), from the composition of NS1 and NS2, one can calculate that, even neglecting the contribution from nanochannels, minimum *V* values as large as 0.087 and 0.107 cm³ g⁻¹, respectively, should be expected. The latter values are much larger than the experimental results. Thus, we can conclude that the use of the *p*-nitrophenol as a probe actually leads to underestimating both *S* and *V* values. Therefore, even the reliability of this approach appears seriously questionable.

FFC-NMR Relaxometry. Relaxometry experiments performed here aimed at studying the variations of the longitudinal relaxation rates (R_1) in the magnetic field range 0.1-35 MHz. We first investigated the dry NS1-NS3 samples; anhydrous β CD was also analyzed for useful comparison. Then, we considered the two fully insoluble materials, NS1 and NS2, after equilibration with water, added in a 1:2 w/w amount (the relevant samples are indicated hereinafter as NS1 + H_2O and NS2 + H_2O). Hence, information on the dynamics at the solid-liquid interface was achieved. Finally, the dynamics of the skeletal ¹H nuclei was also studied by equilibrating NS1 with D_2O (1:2.5 w/w, sample NS1 + D_2O). As a general remark, for the dry samples (β CD, NS1, NS2, NS3)¹H relaxation followed a simple first-order exponential trend. Conversely, relaxation of the wet samples (i.e., NS1 + H_2O_1 NS1 + D_2O_1 NS2 + H_2O_2 showed a more complex kinetic profile, which was suitably modeled as the sum of two distinct first-order processes: a "fast" and a "slow" one. This peculiar behavior is mirrored by the inverse-Laplace transform analysis of the relaxation kinetic curves (see later). For the sake of completeness, we must mention here that for the NS1 + D₂O sample the slow component was affected by very large errors, thereby showing a very scattered trend with respect to the proton Larmor frequency $(\omega_{\rm I})$; therefore, it was not further analyzed. NMR dispersion curves $(R_1 \text{ vs } \omega_1)$ for all the samples are synoptically shown in Figure 3 (the complete data set is reported in the Supporting Information, Table S1).

Interestingly, most of the dispersion curves show the presence of the typical dips located at $\omega_{\rm L}$ ca. 2.7 and 3.1 MHz, due to the quadrupolar coupling effect with the urethane N atoms (inset in Figure 3). Dips are very detectable not only in the curves relevant to dry materials NS1, NS2, and NS3, but also for the fast components of the NS1 + H₂O and NS2 +



Figure 3. NMR dispersion curves $(R_1 \text{ vs } \omega_L)$.

H₂O samples. The latter finding suggests the occurrence of strong long-range N–H interactions even under conditions that are able to favor proton exchange. Conversely, dips are absent in the dispersion curves relevant to the slow component of the wet samples, to the fast component of NS1 + D₂O and, obviously, to dry β CD.

NMR dispersion curves were subjected to regression analysis according to Halle's approach,^{71–73} as the sum of two or three "stretched" components, to which two additional Lorentzian-like contributions were added in order to account for the simultaneous occurrence of the quadrupolar dips (when needed):

$$R_{1} = \sum_{i} \frac{c_{i}\tau_{i}}{1 + (\omega_{\rm L}\tau_{i})^{2}} + \sum_{j_{\rm (dips)}} \left[\frac{2}{\pi} \times \frac{c_{j}s_{j}}{4(\omega_{\rm L} - \omega_{j})^{2} + s_{j}^{2}}\right]$$
(1)

From the fitting parameters (reported in the Supporting Information, Table S2), the relevant correlation times (τ_c 's) for each curve were calculated (Table 1), according to the relationship

$$\tau_{\rm c} = \frac{\sum_i c_i \tau_i}{\sum_i c_i} \tag{2}$$

Table 1. Correlation Times $(\tau_c s)$

sample	$\tau_{\rm c}~({\rm ns})$	sample	$\tau_{\rm c}~({\rm ns})$
β CD dry	600 ± 180		
NS1 dry	430 ± 40	NS2 dry	450 ± 50
NS1 + H ₂ O fast	68 ± 5	NS2 + H ₂ O fast	58 ± 4
NS1 + H_2O slow	37 ± 4	NS2 + H_2O slow	20 ± 4
$NS1 + D_2O$ fast	500 ± 90	NS3 dry	580 ± 240

For molecular compounds, τ_c is interpreted as the average time needed for a molecule to rotate one radian or to move within a distance equal to its length. Therefore, it can be considered as a suitable measure for structural mobility at a microscopic scale. Dry samples showed similar correlation times, within the limits of experimental uncertainties. By contrast, wet samples NS1 + H₂O and NS2 + H₂O showed much smaller τ_c values for the fast component, and even smaller values for the slow one. Both correlation times for NS1 + H_2O are significantly larger than those for NS2 + H_2O . Noticeably, sample NS1 + D_2O shows a comparable τ_c value with respect to the relevant dry material. These observations suggest that correlation times for samples with H₂O mainly keep into account the relaxometric behavior of water molecules. However, we can distinguish at least two different populations having a significantly different mobility. Trends for $\tau_{\rm c}$ values are consistent with the different reticulation degree expected for the materials. In fact, NS1 is supposed to present significantly narrower channels than NS2, having been prepared with a larger amount of reticulating agent. Therefore, we may reasonably relate the fast component (longer τ_c) of the relaxation kinetics to H₂O molecules which are tightly bound to the nanosponge network, thereby forming its immediate hydration shell/layer. This idea is supported by the residual presence of the quadrupolar dips observed in the relevant dispersion curve. Conversely, the slow component (shorter τ_c) may be associated with loosely bound molecules flowing within the lumen of the channels. For this purpose, for the sake of

information completeness, it is worth mentioning here that dry materials retain a small percentage of water, ca. $\sim 5-7\%$ (by TGA analysis). On the grounds of trivial stoichiometric calculations, this on average corresponds to 7 ± 2 water molecules per β CD unit. It is quite reasonable to assume that these water molecules are very tightly bound to the structure either via hydrogen bonding, or as small clusters placed inside the β CD cavities. Thus, keeping into account the theoretical molecular formulas (namely, $[(C_{42}H_{70}O_{35})\cdot(C_8H_{12}N_2O_2)_4]_n$ for NS1, $[(C_{42}H_{70}O_{35}) \cdot (C_8H_{12}N_2O_2)_2]_n$ for NS2), it can be estimated that nearly 75% of the H atoms present in the samples are stably bound to C atoms, whereas the remaining 25% are "exchangeable protons" bound to O or N atoms. Differently, for both the wet samples $NS1 + H_2O$ and NS2 +H₂O, it can be calculated that the exchangeable protons correspond to ca. 79% of the total H atoms population.

Close inspection of the dispersion curves tells a more articulated story. Seen on a logarithmic scale, curves for dry NS1 and NS2 appear to be almost superimposable at large $\omega_{\rm L}$ values and only slightly diverging at low $\omega_{\rm L}$. This suggests that relaxation mechanisms are very similar in the two cases. However, relaxation for anhydrous β CD is much slower in the whole $\omega_{\rm L}$ range, clearly indicating that lack of reticulation favors molecular motions on a microscopic scale. The curves for dry NS3 and NS1 + D₂O are in an intermediate position between β CD and dry NS1 at large ω_{L} , while they approach the latter ones at low $\omega_{\rm L}$ values. This suggests an intermediate structural mobility. In the former case, this can be justified with the incomplete reticulation due to the low combination ratio between β CD and HMDI (i.e., 1:1 mol/mol), whereas, in the latter case, it positively indicates a swelling occurrence. On passing to wet samples, curves for the fast component of NS1 + H₂O and NS2 + H₂O approach those for the corresponding dry materials only at the largest $\omega_{\rm L}$ values, but neatly diverge in the region at low $\omega_{L'}$ where the relaxation rates for both samples are very similar and almost independent of the Larmor frequency. This peculiar behavior suggests that different sections of the dispersion curves account for different aspects of the microscopic dynamism of the samples. In particular, curve trends suggest that the relaxometric response appears more sensitive to not-exchangeable C-bound protons and to tightly bound water molecules of the hydration layer at the larger $\omega_{\rm L}$ values. Conversely, larger sensitivity to less tight, easily exchangeable water molecules seems to exist at the lower $\omega_{\rm L}$ values. Finally, the quadrupolar dips should specifically account for the behavior of protons bound to the urethane N atoms

In order to clarify and support these hypotheses, we considered the inverse-Laplace transform curves of the relaxation kinetics data at five different $\omega_{\rm L}$ values (namely, 35, 10, 3, 1, and 0.3 MHz), obtained by means of the UPEN algorithm. Normalized transform curves (a representative example is shown in Figure 4) can be subjected to regression analysis as skewed log-normal distributions, the maximum and the full width at half-height (fwhh) values of which were considered (Table 2, the complete data set is reported in the Supporting Information, Table S3 and Figures S2–S6).

As long as dry samples are concerned, all inverse-Laplace transforms afford a unimodal distribution at any Larmor frequency, according to the observed first-order relaxation kinetics. Consistently with the discussion of dispersion curves reported hereinabove, the distribution maxima increase along the series NS2 \approx NS1 < NS3 < β CD. This trend fairly agrees





Figure 4. Normalized inverse-Laplace transforms (UPEN) at 35 MHz.

with the reticulation extent of the materials, but for the fact that values for NS1 are slightly larger than for NS2. The latter apparent anomaly might be justified with the much larger presence in NS1 of the hexamethylene linker chains, which benefit from a moderate conformational flexibility. Conversely, transforms for wet samples show bimodal distributions. Their major component is centered at larger T_1 values as compared to those found for dry materials, whereas the minor component is centered at comparable T_1 values. This finding, in turn, is consistent with the occurrence of the complex relaxation kinetics observed in these cases. More in detail, the two components correspond to the slow and fast counterparts of the relaxation kinetics, respectively. As long as the samples with H_2O are concerned, the two distribution maxima for NS1 occur at much lower T_1 values in comparison to NS2, consistent with its more extensive reticulation. On the other hand, maxima for the fast component occur at significantly larger values with respect to dry materials, accounting for larger molecular motions, which can be attributed to both the skeletal ¹H atoms and the hydration shell water molecules. The distribution maximum relevant to the fast component for the $NS1 + D_2O$ sample is in an intermediate position between the ones for dry NS1 and NS1 + H₂O. The latter detail confirms the effect of swelling in increasing molecular motions for the skeletal H atoms in the wet sample. Finally, the relative populations of the two components significantly change on varying the Larmor frequency. In particular, for NS1 + H₂O the fast component passes from a 20% population at 35 MHz down to 9% at 0.3 MHz, whereas for NS2 + H_2O , the fast component passes from 19% down to 14% population in the same $\omega_{\rm L}$ range. The latter observations strongly support the idea that the sensitivity of the relaxometric response to water molecules tightly interacting with the nanosponge framework decreases on decreasing the Larmor frequency.

Texture Features and Connectivity Indexes. At this point the question arises as to how the relaxometric behavior observed may enable researchers to gain information on the texture properties. According to theory, $^{54-56,74}$ for wet microor nanoporous materials the relaxation rates R_1 can be related to the fraction of water molecules interacting with the pore surface (f_s) by the equation

$$R_{1} = R_{w} + f_{S}(R_{S} - R_{w}) \tag{3}$$

sample	$\omega_{\rm L}$ (MHz)	max (ns)	fwhm (ns)	pop. (%)	sample	$\omega_{\rm L}~({\rm MHz})$	max (ns)	fwhm (ns)	pop. (%)
βCD	35	303	141	100	NS1 dry	35	142	47	100
	10	122	49	100		10	46	26	100
	3	36.2	12.1	100		3	10.7	2.8	100
	1	13.3	8.3	100		1	6.6	2.1	100
	0.3	5.1	1.8	100		0.3	2.8	0.7	100
NS2 dry	35	145	67	100	NS3 dry	35	234	62	100
	10	43	24	100		10	76	33	100
	3	9.5	3.8	100		3	16.3	7.3	100
	1	5.9	2.3	100		1	4.6	2.9	100
	0.3	2.5	1.2	100					
$NS1 + H_2O$	35	770	319	80	$NS2 + H_2O$	35	1442	1023	81
		231	205	20			129	117	19
	10	526	352	82		10	906	1164	83
		66	106	18			44	40	17
	3	394	461	84		3	860	1175	82
		19.0	22.6	16			11.2	16.0	18
	1	392	382	85		1	793	888	83
		13.2	12.9	15			9.9	11.7	17
	0.3	314	328	91		0.3	715	1003	86
		9.8	11.9	9			5.8	13.5	14
$NS1 + D_2O$	35	160	124	100					
	10	734	714	37					
		52	52	63					
	3	14.3	32	100					
	1	210	226	9					
		10.6	14.6	91					
	0.3	179	274	21					
		3.0	71	79					

pubs.acs.org/JPCB

Table 2. Parameters for the	Inverse-Laplace	Transforms	(UPEN))
-----------------------------	-----------------	------------	--------	---

where R_w is the relaxation rate of bulk water (ca. 0.33 s^{-1,55} almost independent of ω_L in the frequency range analyzed in this study) and R_s is the intrinsic relaxation rate for "surface" water. Hence, R_1 can lead back to the porosimetric parameters of the material according to the relationships

$$R_1 = R_w + \rho(S/V) \tag{4}$$

$$R_1 = R_w + \rho(m/D) \tag{5}$$

$$R_1 = R_w + \lambda (m/D)(R_s - R_w) \tag{6}$$

where ρ is a parameter defined as the "surface relaxivity" (in the case of clay materials or natural soils, for instance, ρ values have been reported^{55,56,74} in the range ca. ~5–60 μ m s⁻¹), λ represents the thickness of the hydration shell of pore walls (usually set as large as 0.3 nm), and *m* is a geometry parameter, the value of which is a function of pore shape (*m* = 4 for cylindrical pores, 6 for spherical ones). By combining eqs 3–6, the two relationships $D = \lambda(m/f_S)$ and $f_S = \lambda(S/V)$ can be deduced with few trivial passages.

According to the previous discussion, in our case we can assume that the fast and slow components of the relaxation kinetics for wet samples adequately describe the behavior of surface and pore water molecules, respectively. Consequently, we deduce

$$R_{\rm slow} = R_{\rm w} + f_{\rm S} \cdot (R_{\rm fast} - R_{\rm w}) \tag{7}$$

and then:

$$f_{\rm S} = (R_{\rm slow} - R_{\rm w})/(R_{\rm fast} - R_{\rm w})$$
(8)

Interestingly, the apparent values of $f_{\rm S}$ calculated from our data vary as a function of $\omega_{\rm L}$ (Figure 5, see data in the

Article



Figure 5. Apparent dependence of $f_{\rm S}$ on $\omega_{\rm L}$ for NS1 and NS2.

Supporting Information, Table S4), passing through a minimum located between 1 and 2 MHz. This implies that even the apparent thickness λ of the hydration shell of the pore surface depends on $\omega_{\rm L}$ as well, according to the previous relationship $f_{\rm S} = \lambda(S/V)$. In fact, the ratio S/V must be a constant as an intrinsic feature of the material.

We may reason that the value of 0.3 nm assumed for λ in the case of clays and soils is conceivably unsuitable for nanosponges. In fact, water molecules can form clusters into the

 β CD cavities (0.65 nm). It is reasonable to assume that a similar behavior may also be shown by water molecules constrained into channels of similar width. Moreover, the secondary rim of cyclodextrins is able to exert a highly ordering effect on the water molecules located in its immediate surroundings, in such a way as to form a so-called "expanded hydrophobic sphere". $^{75-77}$ The presence of this peculiar arrangement of water molecules is able to significantly affect the thermodynamics of binding of long-chain guests in solution. In particular, there is positive evidence that the size of the "expanded hydrophobic sphere" is roughly as large as an extended propylene diamine chain (i.e., ca. 0.5 nm).⁷⁸ Finally, the hydrophobic hexamethylene chains may induce an extensive clusterization of water molecules, due to the hydrophobic effect. This, in turn, affects the long-range solvent organization in the nearby. On the whole, consistent with the discussion of the R_1 vs ω_L curves reported hereinabove, our results can be rationalized assuming that the relaxometric response is able to sample at the largest $\omega_{\rm L}$ values a long-range structuration of the pore solvent shell, that is somehow less perceived at the lowest $\omega_{\rm L}$ values.

From eq 8, the average pore diameters of the materials may be related to relaxation rates according to the following expressions:

$$D = \lambda m / f_{\rm S}$$

= $\lambda m (R_{\rm fast} - R_{\rm w}) / (R_{\rm slow} - R_{\rm w})$
= $\lambda m (T_{\rm fast}^{-1} - R_{\rm w}) / (T_{\rm slow}^{-1} - R_{\rm w})$ (9)

Unfortunately, on the basis of the previous discussion, eq 9 cannot be applied in our case, because neither λ nor *m* values can be satisfactorily set. In fact, λ is $\omega_{\rm L}$ -dependent; moreover, it is hardly possible to provide a reasonable value for the pore-shape-dependent *m* parameter, because of both the disordered NS microscopic structure and the unpredictable swelling effects. Nevertheless, it is interesting to notice that (as can be easily verified from the data in Table S4) the ratio between the $f_{\rm S}$ values for NS1 and NS2 is independent of $\omega_{\rm L}$. More in detail, we found $f_{\rm S,NS1}/f_{\rm S,NS2} = 2.2 \pm 0.3$, which hence provides a reasonable estimation of the ratio between the average pore diameters for the two materials.

Furthermore, the existence of a distribution of relaxation times, mirroring pore size distribution, must be considered. Indeed, the normalized inverse-Laplace transforms provide a frequency distribution function $(P(T_1))$ for the T_1 values, so that we can define an average relaxation time $\langle T_1 \rangle$ as

$$\left\langle T_{1}\right\rangle = \int_{0}^{\infty} P(T_{1})T_{1} dT_{1}$$
(10)

In order to apply eq 9, average values for T_{fast} and T_{slow} should be first calculated according to eq 10, by considering independently the normalized fitting expressions for the fast and the slow components in place of $P(T_1)$. Noticeably, because *D* can be expressed as a function of T_{fast} and T_{slow} by eq 9, the normalized frequency distribution $P(T_1)$ for the isolated slow component may be transformed in a frequency distribution function for pore diameters (P'(D)) by a trivial variable exchange, i.e.

$$P'(D) = P([R_{\rm w} + \lambda m(\langle T_{\rm fast} \rangle^{-1} - R_{\rm w})/D])$$
(11)

Then, the average pore size could be calculated as

$$D = \int_0^\infty P'(D)D \, \mathrm{d}D \tag{12}$$

Once again, lack of reliable values for λ and *m* forbids defining the function P'(D); so, even calculation of *D* according to eq 12 is not possible.

The intrinsic difficulties in establishing a satisfactory method to evaluate the proper textural parameters, which are apparent from the whole of the previous discussions, moved us to explore some possible alternative approaches. In particular, we reasoned that the features of soils are somehow comparable to those of nanosponges and may constitute a suitable study model. As we mentioned in the Introduction, soil properties have been adequately described, on the basis of the relaxometric responses, by introducing the concept of "connectivity", intended both in a "structural" (i.e., the through-space interconnection of the pore network) and in a "functional" (i.e., the ability to mediate physical, chemical, and biological phenomena) sense. It is worth stressing that the key point here is the fact that "connectivity" provides an elegant and viable way to describe the functional role of water in mediating the transport phenomena through the channel network, and how this affects the overall interaction/exchange abilities of the porous system. It is immediately apparent that such a role is perfectly mirrored in NS materials. Accordingly, two connectivity indexes have been defined,^{62,63} namely, a "functional connectivity index" (FCI) and a "structural connectivity index" (SCI), able to provide a suitable quantitative assessment of these properties. More in detail, starting from the probability function $P(T_1)$ defined in eq 10, two arbitrary reference T_1 values (T_A and T_B) are first set according to the conditions

$$\int_{0}^{T_{\rm A}} P(T_{\rm l}) \, \mathrm{d}T_{\rm l} = 0.01 \text{ and } \int_{0}^{T_{\rm B}} P(T_{\rm l}) \, \mathrm{d}T_{\rm l} = 0.99 \tag{13}$$

in such a way that the interval $T_{\rm A}-T_{\rm B}$ comprises the most significant part (98%) of the T_1 values distribution, accounting for water molecules' mobility. Then, the aforementioned connectivity indexes are simply defined as FCI = $T_{\rm B}/T_{\rm A}$ and SCI = $T_{\rm B} - T_{\rm A}$.

In order to apply these ideas in our case, we preliminarily chose to adopt a slightly more restrictive significance criterion, keeping into account only 95% of the overall ¹H population. Therefore, we defined T_A and T_B by fixing the upper integration limits in eqs 13 at 0.025 and 0.975, respectively. Then, considering that f_S linearly depends on the average pore diameter *D* by a λm factor (and to the *S/V* ratio as well, eqs 3–6), in analogy with eq 9, we decided to define a further "pore connectivity index" (PCI) according to the relationship

$$PCI = \frac{\left(\int_{T_{A,fast}}^{T_{B,fast}} P(T_{fast}) T_{fast} \, \mathrm{d}T_{fast}\right)^{-1} - R_{w}}{\left(\int_{T_{A,slow}}^{T_{B,slow}} P(T_{slow}) T_{slow} \, \mathrm{d}T_{slow}\right)^{-1} - R_{w}}$$
(14)

Thus, PCI is in fact directly related to *D*, through the parameters λ and *m*. The values of FCI, SCI, and PCI calculated from the relaxometric data relevant to the wet samples are collected in Table 3.

At any Larmor frequency, the values of the three indexes are larger for NS2 than for NS1. This is perfectly consistent with the different reticulation extent. In agreement with the findings of previous works on soils,⁶² neither FCI nor SCI values show

		NS1			NS2	
$\omega_{\rm L}~({\rm MHz})$	FCI	SCI	PCI	FCI	SCI	PCI
35	1.98	547	4.3	3.30	1713	13.1
10	2.98	652	6.2	7.05	2750	28.0
3	6.36	953	18.2	7.15	2132	69.3
1	5.25	595	27.7	5.98	1708	87.5
0.3	5.58	565	26.0	8.74	2088	46.4

regular variations as a function of $\omega_{\rm L}$. However, we can notice that FCI tends to decrease on increasing $\omega_{\rm L}$, whereas SCI might be considered roughly constant. Differently, PCI values for both materials pass through a maximum, centered at 1 MHz. Again, the ratio between the PCI values for the two materials does not depend on $\omega_{\rm L}$, within the reasonable statistical variations (PCI_{NS2}/PCI_{NS1} = 3.2 ± 0.6). Therefore, PCI can be considered as a measure of the different mobilities of water molecules within the nanochannels of the two different materials. On the whole, these findings suggest that PCI can be considered an excellent alternative parameter (with respect to the textural D, S, and V values), providing a satisfactory assessment of the textural features of nanosponges from a functional standpoint. In particular, PCI has the advantage to bypass the intrinsic difficulties deriving from the poor definition of the λ and m, the values of which critically depend on the highly disordered and swellable microscopic NS structure. Moreover, in comparison with the previous FCI and SCI indexes, PCI benefits from the fact that its trends depend on $\omega_{\rm L}$ in a way that appears easier to be rationalized.

As a final remark, it is worth stressing that, from a general reconsideration of the entire topic based on the analysis of the whole of the experimental results, the definition of the ordinary textural properties (D, S, V) appears unsatisfactory and elusive in the case of nanosponges. Indeed, the very fact that the structure of these materials may undergo fair swelling in the presence of an aqueous medium, with the consequent effect on the overall structure, causes unpredictable variations of pore sizes; for the same reason, even the concept itself of surface seems somehow to fade away at such a microscopic scale. Therefore, it makes much more sense, in our opinion, to define a porosity parameter in functional terms. This requirement seems adequately fulfilled by the connectivity indexes such as PCI.

CONCLUSIONS

This study reports for the first time the application of fast-fieldcycling NMR relaxometry in order to gain information on the textural features of nanosponges. In particular, the study of wet samples revealed a complex behavior, which was attributed to the existence of two different water molecule populations, having significantly different relaxation rates and molecular mobility. This, in turn, led back to the pore size distribution in the materials. Moreover, inspired by the results from soil science, we adapted to nanosponges the concepts of connectivity indexes. From the inverse-Laplace T_1 distributions functions, a new "pore connectivity index" (PCI) was suitably defined (eq 14), the values of which are positively correlated to the reticulation degree of the synthesized NSs. Although the PCI index is positively related to the pore size distribution, it provides more than a mere assessment of structural features but rather accounts for the functional properties of the nanosponge materials related to the mobility of water

throughout the pore network. In this sense, it is noteworthy that the PCI definition trespasses the intrinsic conceptual difficulties in assessing the texture features deriving from the unpredictable structural effects of swelling.

Finally, even though it cannot directly provide a quantitative measure of textural parameters, FFC NMR relaxometry proved to offer an easy and viable methodology to assess the functional features of nanosponges. It is undoubtedly more reliable and less time- and material-consuming than ordinary porosimetric methods based on the analysis of absorption isotherms. This, in turn, makes it a valuable tool for rationalizing the supramolecular behavior and abilities of nanosponges. Therefore, it may be helpful for clarifying the molecular mechanisms involved in their controlled absorption and release properties, and, in perspective, for a rational design of tailored systems.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.jpcb.9b11935.

Short outline of FFC-NMR theory, R_1 vs ω_L data, Halle's regression parameters R_1 vs ω_L curves, normalized inverse-Laplace (UPEN) T_1 distribution curves, log-normal regression parameters for inverse-Laplace (UPEN) T_1 distribution curves, and f_S vs ω_L data for NS1 and NS2 (PDF)

AUTHOR INFORMATION

Corresponding Authors

- Paolo Lo Meo Department of Biological, Chemical and Pharmaceutical Sciences and Technologies (STEBICEF), University of Palermo, 90128 Palermo, Italy; orcid.org/ 0000-0001-5089-0556; Email: paolo.lomeo@unipa.it
- Pellegrino Conte Department of Agricultural, Food and Forest Sciences (SAAF), University of Palermo, 90128 Palermo, Italy; orcid.org/0000-0002-2211-1225; Email: pellegrino.conte@unipa.it

Authors

- **Federico Mundo** Department of Biological, Chemical and Pharmaceutical Sciences and Technologies (STEBICEF), University of Palermo, 90128 Palermo, Italy
- Samuele Terranova Department of Biological, Chemical and Pharmaceutical Sciences and Technologies (STEBICEF), University of Palermo, 90128 Palermo, Italy
- **Delia Chillura Martino** Department of Biological, Chemical and Pharmaceutical Sciences and Technologies (STEBICEF), University of Palermo, 90128 Palermo, Italy; orcid.org/ 0000-0001-5141-7285

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jpcb.9b11935

Author Contributions

All authors have given approval to the final version of the manuscript.

Funding

University of Palermo (FFR funding) is gratefully acknowledged for financial support.

Notes

The authors declare no competing financial interest.

pubs.acs.org/JPCB

ACKNOWLEDGMENTS

Prof. G. Lazzara and Dr. S. Cataldo (Department of Physics and Chemistry, University of Palermo) are gratefully acknowledged for TGA determinations. Prof. M. L. Saladino and Dr. G. Polito (Dept. STEBICEF, University of Palermo) are gratefully acknowledged for BET/BJH measurements.

ABBREVIATIONS

ATR, attenuated total reflectance; BET, Braunauer–Emmett– Teller theory; BJH, Barrett–Joyner–Halenda theory; FFC, fast-field-cycling; fwhm, full width at half-maximum

REFERENCES

(1) Caldera, F.; Tannous, M.; Cavalli, R.; Zanetti, M.; Trotta, F. Evolution of Cyclodextrin Nanosponges. *Int. J. Pharm.* **2017**, *531*, 470–479.

(2) Nanosponges:Synthesis and Applications; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2019.

(3) Pawar, S.; Shende, P.; Trotta, F. Diversity of β -Cyclodextrin-Based Nanosponges for Transformation of Actives. *Int. J. Pharm.* **2019**, 565, 333–350.

(4) Sherje, A. P.; Dravyakar, B. R.; Kadam, D.; Jadhav, M. Cyclodextrin-based Nanosponges: A Critical Review. *Carbohydr. Polym.* **201**7, 173, 37–49.

(5) Taka, A. L.; Fosso-Kankeu, E.; Pillay, K.; Mbianda, X. Y. Removal of Cobalt and Lead Ions from Wastewater Samples Using an Insoluble Nanosponge Biopolymer Composite: Adsorption Isotherm, Kinetic, Thermodynamic, and Regeneration Studies. *Environ. Sci. Pollut. Res.* **2018**, *25*, 21752–21767.

(6) Fontana, R. M.; Milano, N.; Barbara, L.; Di Vincenzo, A.; Gallo, G.; Lo Meo, P. Cyclodextrin-Calixarene Nanosponges as Potential Platforms for pH-Dependent Delivery of Tetracycline. *ChemistrySelect* **2019**, *4*, 9743–9747.

(7) Allahyari, S.; Trotta, F.; Valizadeh, H.; Jelvehgari, M.; Zakeri-Milani, P. Cyclodextrin-Based Nanosponges as Promising Carriers for Active Agents. *Expert Opin. Drug Delivery* **2019**, *16*, 467–479.

(8) Sikder, M. T.; Rahman, M. M.; Jakariya, M.; Hosokawa, T.; Kurasaki, M.; Saito, T. Remediation of water pollution with native cyclodextrins and modified cyclodextrins: A comparative overview and perspectives. *Chem. Eng. J.* **2019**, 355, 920–941.

(9) Morin-Crini, N.; Winterton, P.; Fourmentin, S.; Wilson, L. D.; Fenyvesi, É.; Crini, G. Water-Insoluble β -Cyclodextrin–Epichlorohydrin Polymers for Removal of Pollutants from Aqueous Solutions by Sorption Processes Using Batch Studies: A Review of Inclusion Mechanisms. *Prog. Polym. Sci.* **2018**, 78, 1–23.

(10) Russo, M.; Spinella, A.; Di Vincenzo, A.; Lazzara, G.; Correro, M. R.; Shahgaldian, P.; Lo Meo, P.; Caponetti, E. Synergistic Activity of Silver Nanoparticles and Polyaminocyclodextrins in Nanosponge Architectures. *ChemistrySelect* **2019**, *4*, 873–879.

(11) Sadjadi, S.; Heravi, M. M.; Malmir, M. Pd(0) Nanoparticle Immobilized on Cyclodextrin-Nanosponge-Decorated Fe_2O_3 @SiO₂ Core-Shell Hollow Sphere: An Efficient Catalyst for C–C Coupling Reactions. J. Taiwan Inst. Chem. Eng. **2018**, 86, 240–251.

(12) Martin-Trasanco, R.; Cao, R.; Esparza-Ponce, H. E.; Montero-Cabrera, M. E.; Arratia-Pérez, R. Reduction of Au(III) by a β -Cyclodextrin Polymer in Acid Medium. A Stated Unattainable Reaction. *Carbohydr. Polym.* **201**7, 175, 530–537.

(13) Vasconcelos, D. A.; Kubota, T.; Santos, D. C.; Araujo, M. V. G.; Teixeira, Z.; Gimenez, I. F. Preparation of Au_n Quantum Clusters with Catalytic Activity in β -Cyclodextrin Polyurethane Nanosponges. *Carbohydr. Polym.* **2016**, 136, 54–62.

(14) Cyclodextrin Nanosponges and Their Applications. In *Cyclodextrins in Pharmaceutics, Cosmetics, and Biomedicine*; Bilensoy, E., Ed.; John Wiley & Sons, Inc.: Hokboken, NJ, 2011; pp 323–342.

(15) Spinella, A.; Russo, M.; Di Vincenzo, A.; Chillura Martino, D.; Lo Meo, P. Hyper-Reticulated Calixarene Polymers: a New Example of Entirely Synthetic Nanosponge Materials. Beilstein J. Org. Chem. 2018, 14, 1498-1507.

(16) Lu, P.; Cheng, J.; Li, Y.; Li, L.; Wang, Q.; He, C. Novel Porous β -Cyclodextrin/Pillar[5]arene Copolymer for Rapid Removal of Organic Pollutants from Water. *Carbohydr. Polym.* **2019**, 216, 149–156.

(17) Cinà, V.; Russo, M.; Lazzara, G.; Chillura Martino, D.; Lo Meo, P. Pre- and Post-Modification of Mixed Cyclodextrin-Calixarene Co-Polymers: A Route towards Tunability. *Carbohydr. Polym.* **2017**, *157*, 1393–1403.

(18) Russo, M.; Saladino, M. L.; Chillura Martino, D.; Lo Meo, P.; Noto, R. Polyaminocyclodextrin Nanosponges: Synthesis, Characterization and pH-Responsive Sequestration Abilities. *RSC Adv.* **2016**, *6*, 49941–49953.

(19) Di Vincenzo, A.; Russo, M.; Cataldo, S.; Milea, D.; Pettignano, A.; Lo Meo, P. Effect of pH Variations on the Properties of Cyclodextrin-Calixarene Nanosponges. *ChemistrySelect* **2019**, *4*, 6155–6161.

(20) Castiglione, F.; Crupi, V.; Majolino, D.; Mele, A.; Rossi, B.; Trotta, F.; Venuti, V. Effect of Cross-Linking Properties on the Vibrational Dynamics of Cyclodextrins-Based Polymers: An Experimental-Numerical Study. *J. Phys. Chem. B* **2012**, *116*, 7952–7958.

(21) Crupi, V.; Fontana, A.; Giarola, M.; Longeville, S.; Majolino, D.; Mariotto, G.; Mele, A.; Paciaroni, A.; Rossi, B.; Trotta, F.; Venuti, V. Vibrational Density of States and Elastic Properties of Cross-Linked Polymers: Combining Inelastic Light and Neutron Scattering. *J. Phys. Chem. B* **2014**, *118*, 624–633.

(22) Trotta, F.; Caldera, F.; Dianzani, C.; Argenziano, M.; Barrera, G.; Cavalli, R. Glutathione Bioresponsive Cyclodextrin Nanosponges. *ChemPlusChem* **2016**, *81*, 439–443.

(23) Brunauer, S.; Emmett, P. H.; Teller, E. Adsorption of Gases in Multimolecular Layers. J. Am. Chem. Soc. **1938**, 60, 309–319.

(24) Kruk, M.; Antochshuk, V.; Jaroniec, M.; Sayari, A. New Approach to Evaluate Pore Size Distributions and Surface Areas for Hydrophobic Mesoporous Solids. *J. Phys. Chem. B* **1999**, *103*, 10670–10678.

(25) Alsbaiee, A.; Smith, B. J.; Xiao, L. L.; Ling, Y. H.; Helbling, D. E.; Dichtel, W. R. Rapid Removal of Organic Micropollutants from Water by a Porous β -Cyclodextrin Polymer. *Nature* **2016**, *529*, 190–194.

(26) Li, Y.; Lu, P.; Cheng, J.; Zhu, X.; Guo, W.; Liu, L.; Wang, Q.; He, C.; Liu, S. Novel Microporous β -Cyclodextrin Polymer as Sorbent for Solid-Phase Extraction of Bisphenols in Water Samples and Orange Juice. *Talanta* **2018**, *187*, 207–215.

(27) Li, H.; Meng, B.; Chai, S.-H.; Liu, H.; Dai, S. Hyper-Crosslinked β -Cyclodextrin Porous Polymer: an Adsorption-Facilitated Molecular Catalyst Support for Transformation of Water-Soluble Aromatic Molecules. *Chemical Science* **2016**, *7*, 905–909.

(28) Xiao, P.; Dudal, Y.; Corvini, P. F. X.; Shahgaldian, P. Polymeric Cyclodextrin-Based Nanoparticles: Synthesis, Characterization and Sorption Properties of Three Selected Pharmaceutically Active Ingredients. *Polym. Chem.* **2011**, *2*, 120–125.

(29) Wilson, L. D.; Mohamed, M. H.; Berhaut, C. L. Sorption of Aromatic Compounds with Copolymer Sorbent Materials Containing β -Cyclodextrin. *Materials* **2011**, *4*, 1528–1542.

(30) Pratt, D. Y.; Wilson, L. D.; Kozinski, J. A.; Mohart, A. M. Preparation and Sorption Studies of β -Cyclodextrin/Epichlorohydrin Copolymers. *J. Appl. Polym. Sci.* **2010**, *116*, 2982–2989.

(31) Gholibegloo, E.; Mortezazadeh, T.; Salehian, F.; Ramazani, A.; Amanlou, M.; Khoobi, M. Improved Curcumin Loading, Release, Solubility and Toxicity by Tuning the Molar Ratio of Cross-Linker to β -Cyclodextrin. *Carbohydr. Polym.* **2019**, *213*, 70–78.

(32) Junthip, J.; Promma, W.; Sonsupap, S.; Boonyanusith, C. Adsorption of Paraquat from Water by Insoluble Cyclodextrin Polymer Crosslinked with 1,2,3,4-Butanetetracarboxylic Acid. *Iran. Polym. J.* **2019**, *28*, 213–223.

(33) Junthip, J. Water-Insoluble Cyclodextrin Polymer Crosslinked with Citric Acid for Paraquat Removal from Water. J. Macromol. Sci., Part A: Pure Appl.Chem. 2019, 56, 555–563.

(34) Lo Meo, P.; Lazzara, G.; Liotta, L.; Riela, S.; Noto, R. Cyclodextrin-Calixarene Co-Polymers as a New Class of Nanosponges. *Polym. Chem.* **2014**, *5*, 4499–4510.

(35) Wilson, L. D.; Mohamed, M. H.; Headley, J. V. Surface Area and Pore Structure Properties of Urethane-Based Copolymers Containing β -Cyclodextrin. J. Colloid Interface Sci. **2011**, 357, 215– 222.

(36) Brownstein, K. R.; Tarr, C. E. Relaxation Time versus Water Content: Linear or Nonlinear? *Science* **1976**, *194*, 213–214.

(37) Brownstein, K. R.; Tarr, C. E. Spin-Lattice Relaxation in a System Governed by Diffusion. J. Magn. Reson. (1969-1992) 1977, 26, 17–24.

(38) Brownstein, K. R.; Tarr, C. E. Importance of Classical Diffusion in NMR Studies of Water in Biological Cells. *Phys. Rev. A: At., Mol., Opt. Phys.* **1979**, *19*, 2446–2453.

(39) Mattea, C.; Fatkullin, N.; Fischer, E.; Beginn, U.; Anoardo, E.; Kroutieva, M.; Kimmich, R. The "Corset Effect" of Spin-Lattice Relaxation in Polymer Melts Confined in Nanoporous Media. *Appl. Magn. Reson.* **2004**, *27*, 371–381.

(40) Phani Kumar, B. V. N.; Stapf, S.; Mattea, C. Molecular Dynamics in the Lyophases of Copolymer P123 Investigated with FFC NMR Relaxometry. *Langmuir* **2019**, *35*, 435–445.

(41) Ferrante, G.; Sykora, S. Technical Aspects of Fast Field Cycling. *Adv. Inorg. Chem.* **2005**, *57*, 405–470.

(42) Kruk, D.; Herrmann, A.; Rössler, E. A. Field-Cycling NMR Relaxometry of Viscous Liquids and Polymers. *Prog. Nucl. Magn. Reson. Spectrosc.* **2012**, *63*, 33–64.

(43) Rössler, E. A.; Stapf, S.; Fatkullin, N. Recent NMR Investigations on Molecular Dynamics of Polymer Melts in Bulk and in Confinement. *Curr. Opin. Colloid Interface Sci.* **2013**, *18*, 173– 182.

(44) Flämig, M.; Hofmann, M.; Lichtinger, A.; Rössler, E. A. Application of Proton Field-Cycling NMR Relaxometry for Studying Translational Diffusion in Simple Liquids and Polymer Melts. *Magn. Reson. Chem.* **2019**, *57*, 805–817.

(45) Parigi, G.; Rezaei-Ghaleh, N.; Giachetti, A.; Becker, S.; Fernandez, C.; Blackledge, M.; Griesinger, C.; Zweckstetter, M.; Luchinat, C. Long-Range Correlated Dynamics in Intrinsically Disordered Proteins. J. Am. Chem. Soc. **2014**, *136*, 16201–16209.

(46) Di Gregorio, E.; Ferrauto, G.; Lanzardo, S.; Gianolio, E.; Aime, S. Use of FCC-NMRD Relaxometry for Early Detection and Characterization of Ex-Vivo Murine Breast Cancer. *Sci. Rep.* **2019**, *9*, 4624.

(47) Steele, R. M.; Korb, J. P.; Ferrante, G.; Bubici, S. New Applications and Perspectives of Fast Field Cycling NMR Relaxometry. *Magn. Reson. Chem.* **2016**, *54*, 502–509.

(48) Araya, Y. T.; Martínez-Santiesteban, F.; Handler, W. B.; Harris, C. T.; Chronik, B. A.; Scholl, T. J. Nuclear Magnetic Relaxation Dispersion of Murine Tissue for Development of T_1 (R_1) Dispersion Contrast Imaging. *NMR Biomed.* **2017**, *30*, No. e3789.

(49) Davies, S.; Packer, K. J.; Roberts, D. R.; Zelaya, F. O. Pore-Size Distributions from NMR Spin-Lattice Relaxation Data. *Magn. Reson. Imaging* **1991**, *9*, 681–685.

(50) Korb, J. P.; Whaley Hodges, M.; Bryant, R. Translational diffusion of liquids at surface of microporous materials: New theoretical analysis of field cycling magnetic relaxation measurements. *Magn. Reson. Imaging* **1998**, *16*, 575–578.

(51) Howard, J. J. Quantitative Estimates of Porous Media Wettability from Proton NMR Measurements. *Magn. Reson. Imaging* **1998**, *16*, 529–533.

(52) Todoruk, T. R.; Langford, C. H.; Kantzas, A. Pore-Scale Redistribution of Water during Wetting of Air-Dried Soils As Studied by Low-Field NMR Relaxometry. *Environ. Sci. Technol.* **2003**, *37*, 2707–2713.

(53) Mikutta, C.; Lang, F.; Kaupenjohann, M. Soil Organic Matter Clogs Mineral Pores. *Soil Sci. Soc. Am. J.* **2004**, *68*, 1853–1862.

(54) Bird, N. R. A.; Preston, A. R.; Randall, E. W.; Whalley, W. R.; Whitmore, A. P. Measurement of the Size Distribution of Water-Filled Pores at Different Matric Potentials by Stray Field Nuclear Magnetic Resonance. *Eur. J. Soil Sci.* 2005, *56*, 135–143.

(55) Hinedi, Z. R.; Chang, A. C.; Anderson, M. A.; Borchardt, D. B. Quantification of Microporosity by Nuclear Magnetic Resonance Relaxation of Water Imbibed in Porous Media. *Water Resour. Res.* **1997**, 33, 2697–2704.

(56) Pohlmeier, A.; Haber-Pohlmeier, S.; Stapf, S. A Fast Field Cycling Nuclear Magnetic Resonance Relaxometry Study of Natural Soils. *Vadose Zone J.* **2009**, *8*, 735–742.

(57) Belorizky, E.; Fries, P. H.; Guillermo, A.; Poncelet, O. Almost Ideal 1D Water Diffusion in Imogolite Nanotubes Evidenced by NMR Relaxometry. *ChemPhysChem* **2010**, *11*, 2021–2026.

(58) Fleury, M.; Kohler, E.; Norrant, F.; Gautier, S.; M'Hamdi, J.; Barré, L. Characterization and Quantification of Water in Smectites with Low-Field NMR. *J. Phys. Chem. C* **2013**, *117*, 4551–4560.

(59) Buchmann, C.; Bentz, J.; Schaumann, G. E. Intrinsic and Model Polymer Hydrogel-Induced Soil Structural Stability of a Silty Sand Soil as Affected by Soil Moisture Dynamics. *Soil Tillage Res.* **2015**, *154*, 22–33.

(60) Conte, P. In Field-Cycling NMR Relaxometry: Instrumentation, Model Theories and Applications; The Royal Society of Chemistry, 2019; Chapter 10, pp 229–254.

(61) Conte, P.; Alonzo, G. Environmental NMR: Fast-Field-Cycling Relaxometry. *eMagRes.* **1996**, *2*, 389–398.

(62) Conte, P.; Ferro, V. Standardizing the Use of Fast-Field Cycling NMR Relaxometry for Measuring Hydrological Connectivity Inside the Soil. *Magn. Reson. Chem.* **2020**, *58*, 41–50.

(63) Conte, P.; Ferro, V. Measuring Hydrological Connectivity Inside a Soil by Low Field Nuclear Magnetic Resonance Relaxometry. *Hydrol. Processes* **2018**, *32*, 93–101.

(64) Mochida, K.; Kagita, A.; Matsui, Y.; Date, Y. Effects of Inorganic Salts on the Dissociation of a Complex of β -Cyclodextrin with an Azo Dye in an Aqueous Solution. *Bull. Chem. Soc. Jpn.* **1973**, 46, 3703–3707.

(65) Al Omari, M. M.; El-Barghouthi, M. I.; Zughul, M. B.; Davies, J. E. D.; Badwan, A. A. Dipyridamol/ β -Cyclodextrin Complexation: Effect of Buffer Species, Thermodynamics, and Guest–Host Interactions Probed by ¹H-NMR and Molecular Modeling Studies. *J. Inclusion Phenom. Mol. Recognit. Chem.* **2009**, *64*, 305–315.

(66) Ghosh, M.; Zhang, R.; Lawler, R. G.; Seto, C. T. The Effects of Buffers on the Thermodynamics and Kinetics of Binding between Positively-Charged Cyclodextrins and Phosphate Ester Guests. *J. Org. Chem.* **2000**, *65*, 735–741.

(67) Lo Meo, P.; D'Anna, F.; Gruttadauria, M.; Riela, S.; Noto, R. Binding Properties of Mono-(6-Deoxy-6-Amino)-β-Cyclodextrin towards p-Nitroaniline Derivatives: a Polarimetric Study. *Tetrahedron* **2009**, 65, 10413–10417.

(68) Lo Meo, P.; D'Anna, F.; Riela, S.; Gruttadauria, M.; Noto, R. Spectrophotometric Determination of Binding Constants Between Some Aminocyclodextrins and Nitrobenzene Derivatives at Various pH Values. *Tetrahedron* **2002**, *58*, 6039–6045.

(69) Russo, M.; La Corte, D.; Pisciotta, A.; Riela, S.; Alduina, R.; Lo Meo, P. Binding Abilities of Polyaminocyclodextrins: Polarimetric Investigations and Biological Assays. *Beilstein J. Org. Chem.* **2017**, *13*, 2751–2763.

(70) Lo Meo, P.; D'Anna, F.; Riela, S.; Gruttadauria, M.; Noto, R. Binding Equilibria Between β -Cyclodextrin and *p*-Nitro-Nniline Derivatives: the First Systematic Study in Mixed Water-Methanol Solvent Systems. *Tetrahedron* **2009**, *65*, 2037–2042.

(71) Halle, B.; Jóhannesson, H.; Venu, K. Model-Free Analysis of Stretched Relaxation Dispersions. *J. Magn. Reson.* **1998**, *135*, 1–13.

(72) Halle, B. The Physical Basis of Model-Free Analysis of NMR Relaxation Data from Proteins and Complex Fluids. *J. Chem. Phys.* **2009**, *131*, 224507.

(73) Bertini, I.; Fragai, M.; Luchinat, C.; Parigi, G. H NMRD Profiles of Diamagnetic Proteins: A Model-Free Analysis. *Magn. Reson. Chem.* **2000**, *38*, 543–550.

(74) Stingaciu, L. R.; Weihermüller, L.; Haber-Pohlmeier, S.; Stapf, S.; Vereecken, H.; Pohlmeier, A. Determination of Pore Size

Distribution and Hydraulic Properties Using Nuclear Magnetic Resonance Relaxometry: A Comparative Study of Laboratory Methods. *Water Resour. Res.* **2010**, *46*, W11510.

(75) Rekharsky, M. V.; Inoue, Y. Complexation Thermodynamics of Cyclodextrins. *Chem. Rev.* **1998**, *98*, 1875–1918.

(76) Matsui, Y.; Mochida, K. Binding Forces Contributing to the Association of Cyclodextrin with Alcohol in an Aqueous Solution. *Bull. Chem. Soc. Jpn.* **1979**, *52*, 2808–2814.

(77) Saenger, W. Cyclodextrin Inclusion Compounds in Research and Industry. Angew. Chem., Int. Ed. Engl. 1980, 19, 344-362.

(78) Lo Meo, P.; D'Anna, F.; Gruttadauria, M.; Riela, S.; Noto, R. Thermodynamics of Binding Between α - and β -Cyclodextrins and Some *p*-Nitro-Aniline Derivatives: Reconsidering the Enthalpy–Entropy Compensation Effect. *Tetrahedron* **2004**, *60*, 9099–9111.