

Nitrogen Deposition Reduces Plant Diversity and Alters Ecosystem Functioning: Field-Scale Evidence from a Nationwide Survey of UK Heathlands

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Abstract

Findings from nitrogen (N) manipulation studies have provided strong evidence of the detrimental impacts of elevated N deposition on the structure and functioning of heathland ecosystems. Few studies, however, have sought to establish whether experimentally observed responses are also apparent under natural, field conditions. This paper presents the findings of a nationwide field-scale evaluation of British heathlands, across broad geographical, climatic and pollution gradients. Fifty two heathlands were selected across an N deposition gradient of 5.9 to 32.4 kg ha⁻¹ yr⁻¹. The diversity and abundance of higher and lower plants and a suite of biogeochemical measures were evaluated in relation to climate and N deposition indices. Plant species richness declined with increasing temperature and N deposition, and the abundance of nitrophilous species increased with increasing N. Relationships were broadly similar between upland and lowland sites, with the biggest reductions in species number associated with increasing N inputs at the low end of the deposition range. Both oxidised and reduced forms of N were associated with species declines, although reduced N appears to be a stronger driver of species loss at the functional group level. Plant and soil biochemical indices were related to temperature, rainfall and N deposition. Litter C:N ratios and enzyme (phenol-oxidase and phosphomonoesterase) activities had the strongest relationships with site N inputs and appear to represent reliable field indicators of N deposition. This study provides strong. field-scale evidence of links between N deposition - in both oxidised and reduced forms - and widespread changes in the composition, diversity and functioning of British heathlands. The similarity of relationships between upland and lowland environments, across broad spatial and climatic gradients, highlights the ubiquity of relationships with N, and suggests that N deposition is contributing to biodiversity loss and changes in ecosystem functioning across European heathlands.

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Introduction

Human activities associated with the production of energy, fertilisers and leguminous crops have had a substantial effect on the global nitrogen (N) cycle [1]. Recent estimates suggest that anthropogenic contributions to the global reactive N pool have increased 10-fold since pre-industrial times and are predicted to double, relative to current day levels, by 2050 [2]. Deposition of N is spatially highly variable; in Europe, for example, rates currently range from $\sim 1~{\rm kg~ha^{-1}~yr^{-1}}$ in the relatively pristine areas of northern Scandinavia to $> 50~{\rm kg~ha^{-1}~yr^{-1}}$ in areas dominated by industry (e.g. Northern Italy) or intensive agriculture (e.g. Netherlands) [3].

The link between N deposition and changes in the structure and functioning of terrestrial ecosystems is now widely acknowledged, with N cited as one of the leading drivers of biodiversity loss at a global scale [4,5]. Manipulation experiments have demonstrated a wide range of plant responses to N, including changes in plant phenology, physiology, biochemistry, productivity and assimilate allocation [6–8]. Resulting changes in competition between

neighbouring species, as well as N-driven changes in soil pH and NH $_4$ ⁺ concentrations and/or increased sensitivity to environmental stresses, are believed to be responsible for changes in plant community composition observed in many ecosystems [9]. Similarly, N-driven changes in the activity and composition of the soil microbial community [10,11] have been linked with changes in key soil processes, such as rates of decomposition and nitrous oxide production [12–13], with implications for the functioning of affected ecosystems.

Although many of the responses reported in N-manipulation experiments are not specific to N and will be affected by factors such as climate and habitat management, several have been proposed as potential robust bioindicators of the effects and/or amounts of N deposition. These include foliar N accumulation, activities of enzymes involved in nutrient metabolism, soil C:N ratios and concentrations of N in soil solution and leachate [14,15]. Relatively few studies have explicitly tested whether the potential bioindicators of (effects of) N deposition observed in manipulation experiments are apparent and/or widespread under natural, field conditions. Those studies which have done so have

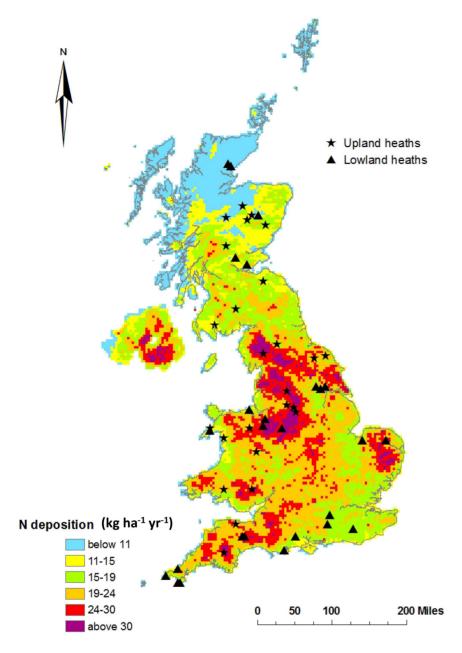


Figure 1. Site survey locations in relation to total N deposition (2006). (http://pollutantdeposition.defra.gov.uk/pollutant-maps). doi:10.1371/journal.pone.0059031.g001

tended to focus on relationships between N deposition and either species richness or biochemical indicators, but rarely both. For example, Remke et al. [16] reported a decline in species richness of coastal dune systems with increasing N deposition, while in a recent analysis of UK Countryside Survey data, Maskell et al. [17] suggest that acidification and eutrophication associated with N deposition are responsible for biodiversity loss in UK grasslands and heathlands. Surveys focusing on biochemical indicators of N deposition identified in manipulation experiments (e.g. tissue N concentrations, soil nutrient availability, litter chemistry and/or enzyme activities) have provided evidence of field-scale changes in ecosystem functioning, particularly in relation to nutrient cycling, along gradients of ambient N deposition [18–20]. A small number of studies have demonstrated links between diversity loss, changes in ecosystem function and N deposition, including a regional

survey of moorlands [18], and both a national [21] and pan-European [22] survey of acidic grasslands. To date, however, there has been no wide-scale evaluation of the relationships between community-level (compositional) bioindicators, biogeochemical (functional) indicators and N deposition in heathland ecosystems, despite the fact these have been shown to respond strongly to experimental N manipulation [23–26].

Atmospheric N occurs in both oxidised (NO_y) and reduced forms (NH_x) ; anthropogenic emissions are associated principally with the combustion of fossil fuels (oxidised N) and intensive agricultural practices (reduced N) processes [27]. Whilst ecosystem responses to increasing total N deposition have been extensively researched, far less is known about the sensitivity of soils and vegetation to the different forms of N [27]. Findings from the relatively few studies that have investigated the role of different

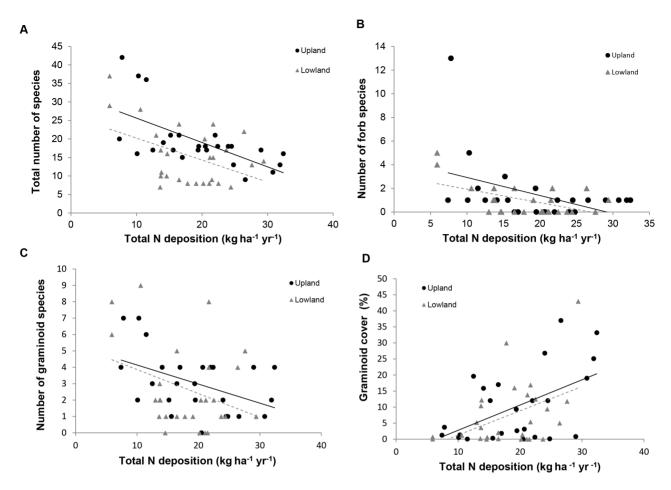


Figure 2. Relationships between N deposition and plant species richness and abundance. A decline of species richness across all plant groups correlates with increasing N deposition (A) and where N deposition and temperatures are higher (B). Whilst graminoid species richness declines in relation to increased N deposition inputs (C), graminoid abundance increases (D), indicating the promotion of fewer species at the higher end of the N gradient. doi:10.1371/journal.pone.0059031.q002

forms of N have reported differences in the strengths of relationships between oxidised and reduced forms in terms of plant tissue chemistry [28] species richness and community composition [29,30].

Upland and lowland heathlands are typically of low nutrient status and dominated by dwarf shrubs. However, their contrasting altitudes (lowlands typically <300m, uplands >300m [31]) result in characteristically different levels of soil organic matter, rates of nutrient cycling, plant species assemblages and management regimes [32] which might be expected to result in differing responses to elevated N inputs. Upland and lowland habitats also experience different climates (typically wetter and cooler in the former compared to the latter), which have the potential to modify their sensitivity to atmospheric N inputs [33]. Indeed, although N manipulation experiments have demonstrated similar qualitative responses to N in upland and lowland heathlands (e.g. increased Calluna productivity and reduced abundance of lower plants [24,34,35]), differences in terms of soil biogeochemical responses have been found [23,26]. The ubiquity of heathland responses to N across a range of altitudinal and climatic gradients has not yet been tested. This study, therefore, presents the first comprehensive, nationwide survey of biological and biogeochemical indicators of N deposition across lowland and upland heathlands. It explicitly aims to evaluate relationships between N deposition and 1) plant community composition and species richness and 2) ecosystem functioning, as measured by soil nutrient stocks, availabilities and enzyme activities. Based on results from earlier manipulation experiments and surveys, we hypothesise that similar relationships with N deposition will be evident in upland and lowland heaths, and that reduced forms of N are having a greater impact than oxidised N on the diversity and functioning of UK heathlands.

Methods

Site selection and field surveying

Fifty two *Calluna vulgaris* dominated heathland sites (representing building to mature phase stands, *sensu* Gimingham, [32]) were selected across a gradient of modelled N deposition (data provided by Ron Smith and Jane Hall, Centre for Ecology & Hydrology, Edinburgh, UK) spanning 5.9 kg N ha⁻¹ yr⁻¹ to 32.4 kg N ha⁻¹ yr⁻¹ (Figure 1). In addition, sites were chosen to be representative of the geographical, climatic and altitudinal (25 upland and 27

Table 1. Summary of optimal models for plant species richness in relation to N deposition and climate variables.

Response variable Total species	Model 1	(To	tal N	deposition	+ en	vironi	mental vari	ables)	Model 2 (oxidized/reduced N + environmental variables)											
	All sites	d.f.	Р	Uplands	d.f.	P	Lowlands	d.f.	P **	All sites	d.f.	P	Uplands	d.f.	Р	Lowlands	d.f.	P			
	(-) N	51	***	(-) N	23	***	(-) N	26		(-) NO _y	51	**	(-) NO _y	24	***	(-) NO _y	26	**			
	(-) gdd		***				(-) gdd		**	(-) gdd		***	(-) NH _x		*	(-) gdd		*			
	N x gdd		*							(-) NH _x		***									
Total lichen species	(-) N	51	**	(-) N	24	**	-	-	-	(-) NH _x	51	***	(–) NO _y	24	**	(-) NH _x	24	*			
	N x gdd		*	N x gdd		**				(-) NO _y		**	(-) NH _x		*						
										(-) gdd		***	NH _x x gdd		**						
													NH _x x rain		*						
Total bryophyte species	(-) gdd	51	***	-	-	-	(-) gdd	26	**	(-) gdd	51	***	-	-	-	(-) NOy	26	*			
																(-) gdd		,			
																(-) rain		×			
Total graminoid species	(-) N	51	**	(-) N	24	*	(-) N	26	*	(-) NO _y	51	*	(-) NH _x	24	*	(-) NO _y	26	*			
	(-) gdd		**				(-) gdd		**	(-) NH _x		*				(-) gdd		,			
										(-) gdd		**				NH _x x gdd		×			
Total forb specie	s (–) N	51	***	(-) N	24	**	(-) N	26	**	(-) NO _y	51	***	(–) NO _y	24	***	(-) NO _y	26	*			
				N x rain		*				(-) NH _x		*	(-) NH _x		*	NO _y x rain		*			
										(-) gdd		*				NH _x x rain		*			

(+/-) = direction of response; x = interaction; P values = * <0.05, **<0.001, ***<0.001; gdd = growing degree days. doi:10.1371/journal.pone.0059031.t001

lowland heaths) variation encountered in heathland ecosystems across the United Kingdom.

Sites were visited during the summer of 2009 and grid reference and altitudinal details were recorded at each site. Vascular and non-vascular plant species were recorded and relative abundance was estimated by percentage cover in five randomly placed quadrats (2 m×2 m) at each site. Canopy height was measured to the nearest mm at the centre of each of the four 1 m² quadrants comprising the 2 m×2 m quadrat, and averaged. 5–10 g of *Calluna* shoots were collected into paper bags and air dried for later chemical analyses. Three soil cores (15 cm depth ×4 cm diameter) were extracted from the centre of the fifth quadrat for chemical analyses; these were stored in an ice-filled cool-box before transfer to controlled temperature rooms upon return to the laboratory, typically within 1–3 days. Five samples of fresh *Calluna* litter (5–10g) were also collected from each quadrat for enzyme assays. The litter was air dried and stored in paper bags.

Chemical analyses

To determine extractable soil ammonium and nitrate concentrations, 40 g of fresh soil was mixed with 150 ml of 1M KCl and shaken for one hour on a rotary shaker before filtering. The filtrate was measured for inorganic N (NH $_4^+$ and NO $_3^-$) using a SKALAR san flow $^{++}$ segmented flow analyser (Skalar UK (Ltd), York, UK). Litter phosphomonoesterase (PME) activity was assayed using the artificial substrate p-nitrophenyl phosphate (p-

NPP), in accordance with the method outlined by Johnson *et al.* [36]. Samples were centrifuged for 5 minutes at 3000 rpm and the resulting liquid was analysed colourimetrically (using a Perkin Elmer Lambda 3 UV spectrophotometer, Perkin Elmer, USA) at a wavelength of 460 nm. Paired non- reactive controls were prepared for each of the samples by adding the PNP and toluene solution immediately after the stopping chemicals to prevent chemical reaction. The readings were compared with pigment densities from a calibration curve obtained using six standards of p-nitrophenol prepared within the absorbance range of the samples.

Total N and carbon (C) concentrations were determined using a FlashEA 1112 CN analyser (Thermo Scientific, UK). *Calluna* shoot and litter samples were dried at 80° C for 24 hours before being ground into a fine powder with a ball mill for two minutes. 15–20 µg of milled material was weighed into tin capsules and combusted. Standard samples were prepared using certified reference materials. Soil pH $_{\rm (H2O)}$ was measured by mixing 10g field moist soil with 25ml deionised water, then allowing the mixture to stand for 30 minutes; pH was then recorded using a CORNING 220 pH meter (Corning Inc., USA).

Litter phenol oxidase activity was measured by vortex-mixing 1 g of ground *Calluna* litter with 9 ml deionised water. 3 ml of this solution was then added to 4.5 ml water and 7.5 ml of 1 M dihydroxyphenylalanine (DOPA) and then shaken for 9 minutes. After shaking, the mixture was centrifuged at 6000 rpm for 5

Table 2. Summary of optimal models for vegetation responses in relation to N deposition and climate variables.

	Model 1	(tota	l N d	deposition	+ er	viro	nmental v	arial	Model 2 (oxidized/reduced N + environmental variables)										
Response variable	All sites	d.f.	Р	Uplands	d.f.	. Р	Lowlands	d.f.	Р	All sites	d.f.	Р	Uplands	d.f.	. Р	Lowlands	d.f.	. Р	
Graminoid cover (%)	(+) N	50	***	(+) N	23	**	(+) N	22	**	(+) NH _x	50	***	(+) NH _x	23	**	(+) NH _x	25	*	
							gdd x rain		**										
Bryophyte cover (%)	(-) gdd	50	***	(-) gdd	23	*	(-) gdd	25	**	(-) gdd	50	***	(-) gdd	25	*	(-) gdd	25	**	
Lichen cover (%)	(-) N	47	**	(-) N	20	*	(-) N	25	*	(-) NH _x	47	**	(-) NO _y	19	*	(-) NH _x	25	**	
	(+) rain		*	(+) rain		**				rain x ht		***	(+) rain		***				
	rain x ht		**	rain x ht		***							NO _y x ht		*				
													rain x ht		***				
Canopy height (cm)	(+) N	49	***	(+) N	22	***	(+) gdd	25	*	(+) NH _x	49	***	(+) NH _x	22	***	(+) gdd	25	*	
	(+) gdd		***	(+) gdd		***				(+) gdd		**	(+) gdd		***				
Pleurozium (%)	(-) N	48	***	(+) rain	22	**	(-) N	24	***	(-) NO _y	47	**	(+) rain	22	**	(-) NH _x	24	**	
	(+) rain		***	N x rain		***	(-) gdd		**	(-) NH _x		***	NH _x x rain		***	(-) gdd		**	
	N x rain		***							(+) rain		***							
										NH _x x rain		***							
Brachythecium (%)	(+) N	50	×	-	-	-	-	-	-	(+) NH _x	50	*	-	-	-	-	-	-	
Hylocomnium (%)	(-) N	49	**	(-) gdd	23	*	(-) gdd	25	*	(-) NH _x	48	**	(-) gdd	23	*	(-) NO _y	23	×	
	(-) gdd		**							(-) gdd		**				(-) gdd		*	
Deschampsia (%)	(+) N	50	***	(+) N	23	**	-	-	-	(+) NH _x	49	***	-	-	-	(+) NH _x	25	*	
										(-) gdd		*							

(+/-) = direction of response; x = interaction; P values = * <0.05, **<0.001, ***<0.001; gdd = growing degree days; ht = *Calluna* canopy height. doi:10.1371/journal.pone.0059031.t002

minutes. A sub-sample was then taken, filtered (0.2 μ m) and absorbance measured at 460 nm using a Perkin Elmer Lambda 3 UV spectrophotometer, Perkin Elmer, USA. Soil moisture content was determined gravimetrically by weighing a 10g sample of fresh soil, drying this at 105 °C for 24 hours, then re-weighing.

Data analyses

Relationships between response and explanatory variables (N deposition, rainfall and temperature) were analysed using the R statistical package, version 2.12.2 (R development core team, 2011). The climate data used were based upon UK 5 km² gridded data sets from the UK Meteorological Office. Variables representing total annual precipitation and temperature were used, the latter represented by data on growing degree days (GDD, sum of degree days above 5°C in a given year). Both precipitation and GDD were averaged over the period 1997–2006. Since total deposition is the product of oxidised and reduced N deposition these three explanatory variables could not be included in the same statistical model. In order to separate the combined effects of total N deposition from the individual influences of oxidised and reduced N forms, two separate models were, therefore, run for each response variable. Model 1 = total N deposition, rainfall and

temperature; Model 2 = reduced N, oxidised N, rainfall and temperature. Percentage cover data for vegetation were arc-sine transformed prior to analysis. Log or square root transformations were used to transform other non-normally distributed data prior to modelling. Relationships were tested using multiple regression analysis or, for count data (e.g. species richness), generalised linear models (GLM) with poisson errors. Relationships with species richness were investigated firstly for all species recorded at a site and subsequently for the different plant functional groups present (lichens, mosses, grasses and forbs). Where curvature in the data was evident, models were fitted with a quadratic function. The most complicated models were fitted (to include interactions between explanatory variables, plus quadratic terms to test for non-linearity of responses) and then simplified using likelihood ratio deletion tests until the minimum adequate model was attained. Evidence of overdispersal was corrected by re-fitting the model with quasi-empirical structures, followed by F tests as outlined by Crawley [37]. Three analyses were performed for each parameter to encompass (a) all sites, (b) upland sites only and (c) lowland sites only.

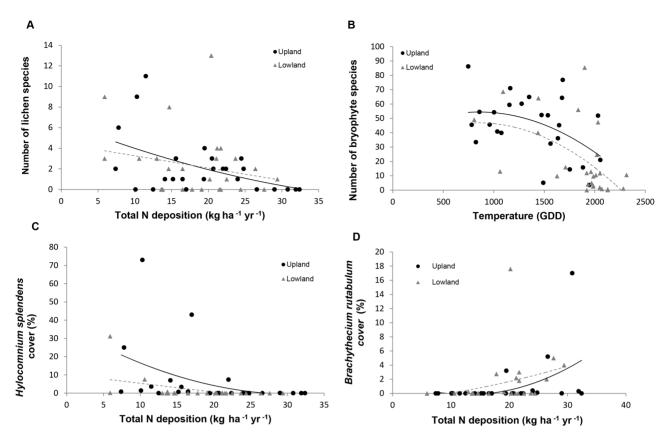


Figure 3. Relationships between N deposition and lower plant species richness and abundance. Declines in lichen species richness are related to increasing N deposition (A) whereas bryophyte richness per se is principally influenced by temperature (B). At a species level, however, significant species-specific relationships between N deposition and bryophytes were evident (C, D). doi:10.1371/journal.pone.0059031.q003

Results

Higher plant responses

A significant decline in overall species richness was observed across all sites in relation to increasing levels of total N deposition $(F_{1,50} = 27.5, P < 0.001, Figure 2a)$. The biggest losses were seen at the lower end of the N deposition gradient, with an average of 13 species lost per site between 5–10 kg N ha⁻¹ yr⁻¹, compared to an average of only 3 species lost as N increased from 10 to 20 kg N ha⁻¹ yr⁻¹. Beyond this, declines in species richness plateaued, indicating a reduction in sensitivity as N loading increased. Temperature was also a significant factor across all sites, both on its own and in interaction with N deposition ($F_{1.50} = 4.4$, P<0.05, Table 1: model 1). Species richness was seen to decline by 50 % across all sites around a temperature threshold of 1000 growing degree days (GDD) and total N inputs of 15 kg N ha⁻¹ yr⁻¹, and declined by a further 50 % when GDD exceeded 2000; this relationship was significant in the lowland-only dataset and as a not quite significant trend for the uplands.

The number of forb species also decreased with increasing total N deposition ($F_{1,50} = 21.2$, P<0.001, Table 1: model 1; Figure 2b), with the steepest rate of decline seen with modest increases of N at the lower end of the deposition gradient (4 species lost on average per site between 5–10 kg N ha⁻¹ yr⁻¹ with negligible losses after this). Detailed analysis of the role of N forms indicates that, for forbs across all sites, stronger relationships were found with oxidised N ($F_{1,50} = 16.0$, P<0.001, Table 1: model 2). Significant interactions between N deposition and rainfall indicate that forb diversity decline is exacerbated in wetter areas receiving higher

levels of N deposition. In the uplands, the relationship was significant with total N deposition ($F_{1,19} = 6.9$, P < 0.05, Table 1: model 1), whilst in the lowlands, NO_y and NH_x both had significant interactions with rain ($F_{1,20} = 4.6$, P < 0.05, $F_{1,19} = 6.2$, P < 0.05 respectively) Table 1: model 2).

Graminoid species richness declined significantly across all sites with increasing total N inputs ($F_{1,50} = 9.\overline{3}$, P<0.01, Figure 2c), as well as with increasing temperature ($F_{1.50} = 10.9$, P<0.01). Losses associated with increasing N were highest at the lower end of the deposition range (an average of 4 species lost per site between 5-10 kg ha⁻¹ yr⁻¹) with a slower rate of loss as inputs increased further. Additionally, each incremental temperature increase of 500 growing degree days corresponded to an average loss of 1 species per site. Across all sites, both forms of N (oxidised and reduced) were associated with graminoid species diversity decline. However, when upland and lowland datasets were considered independently, stronger relationships with NO_v were found in the lowland sites, and with NH_x in the upland sites. Graminoid abundance, by contrast, increased with increasing N ($F_{1.50} = 15.8$, P<0.001), indicating a shift towards more grass dominated systems with greater N deposition (Figure 2d); reduced N was the most significant driver of this response ($F_{1,50} = 19.5$, P<0.001, Table 2: model 2). Increases in grass abundance were, however, found to be chiefly characterised by a few species, specifically Molinia caerulea and Deschampsia flexuosa, the latter being significantly more abundant at sites receiving greater reduced N inputs ($F_{2,49} = 9.2$, P<0.01, Table 2: model 1).

Across both models and all sites, significant positive relationships were observed between Calluna canopy height and temper-

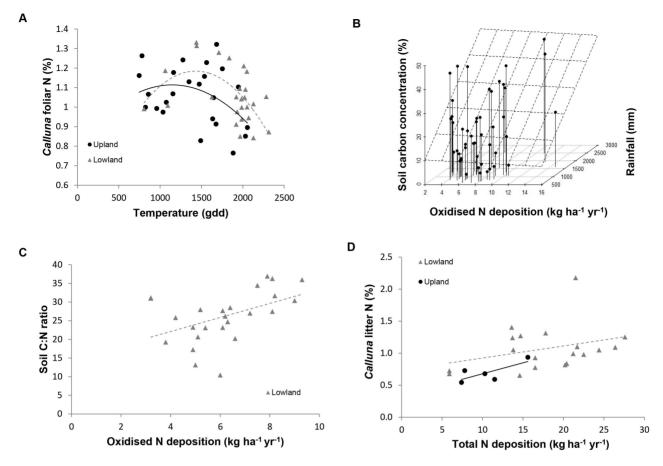


Figure 4. The influence of N deposition and climate on heathland plant and soil chemistry. *Calluna* foliar N concentrations were principally related to changes in temperature (A). However, positive relationships were seen between *Calluna* litter N concentrations and total N deposition (B). Soil carbon concentrations were higher in areas receiving greater oxidised N inputs and rainfall (C), and soil C:N ratios were also positively related to levels of oxidised N deposition (D). doi:10.1371/journal.pone.0059031.g004

ature ($F_{2,49} = 12.6$, P<0.001, Table 1: model 1). Whilst height increased in a linear fashion in relation to total N deposition across all sites ($F_{2,49} = 15.3$, P<0.001, Table 2: model 1), this relationship was largely driven by NHy in the uplands ($F_{2,22} = 24.8$, P<0.001, Table 2: model 2).

Lower plant responses

Significant negative relationships between lichen species richness and total N deposition were found across the combined (all sites) dataset ($F_{1,50} = 7.8$, P<0.01, Table 1: model 1, Figure 3a), driven largely by upland sites. An average of 5 lichen species per site was recorded in areas receiving less than 10 kg N ha⁻¹ yr⁻¹. This declined to an average of 1.4 species at input rates of 20 kg ha⁻¹ yr⁻¹, indicating that even modest rates of N loading are associated with a considerable loss of lichen species. Significant interactions indicate that the negative effects of N deposition on lichen richness were greater at sites with high temperature ($F_{1,48} = 4.8$, P<0.05, Table 1: model 1), particularly in the uplands ($F_{1,20} = 8.9$, P<0.01, Table 1: model 1).

Considered independently, patterns in lichen abundance at upland and lowland sites were best explained by the different forms of N. The relationship with oxidised N was statistically stronger in upland sites ($F_{5,19} = 4.5$, P < 0.05, Table 2: model 2), whilst the relationship was stronger with reduced N in the lowlands ($F_{1,25} = 8.9$, P < 0.01, Table 2: model 2). Climatic

conditions were also important (particularly in the wetter, upland areas), with lichen abundance generally greater in areas receiving more rainfall ($F_{4,47} = 4.4$, P < 0.05, Table 2: model 1). Lichen cover was also generally lower in wetter areas with taller *Calluna* canopies ($F_{4,47} = 9.1$, P < 0.001, Table 2: model 1).

Temperature was the main driver influencing bryophyte species richness and abundance across all sites ($F_{1.50} = 21.3$, P<0.001, Table 1: model 1, Figure 3b), with less species recorded at the warmer end of the temperature gradient (<1000 GDD = 10 species (on average) per site; > 2000 GDD = 5 species (on average) per site). This overall relationship was driven by the strong decline at lowland sites experiencing the warmest conditions, with a non-significant relationship in the uplands dataset. Whilst a general downward trend in bryophyte diversity was apparent in relation to N deposition across all sites, the only significant N signal was found in the lowland sites in the form of oxidised N ($F_{1,26} = 11.2$, P<0.01, Table 1: model 2). Stronger relationships with N were, however, found at the species level: the characteristic heathland mosses Hylocomium splendens (Figure 3c) and Pleurozium schreberii were seen to severely decline with increasing N (Table 2) whilst, conversely, the nitrophilous species [38] Brachythecium rutabulum (Figure 3d) was more abundant at sites receiving higher N inputs $(F_{1.50} = 6.1, <0.05)$.

Table 3. Summary of optimal models for plant and soil biogeochemical responses in relation to N deposition and climatic variables.

	Model 1 ((Tota	al N d	eposition -	⊦ env	/ironr	nental vari	able	s)	Model 2 (oxidized/reduced N + environmental variables)										
Response variable	All sites	d.f.	Р	Uplands	d.f.	Р	Lowlands	d.f.	Р	All sites	d.f.	Р	Uplands	d.f.	Р	Lowlands	d.f.	P		
·	(-) N	49	**	(-) N	20	***	-	-	-	(-) NO _y	47	***	(-) NO _y	23	***	-	-	-		
	(+) gdd		**	gdd x rain		*				(+) gdd		*								
				N x rain		*				NH_{x} x rain		*								
Extractable NO ₃	gdd x moist	38	**	(+) moist	12	***	(+) moist	24	**	(+) moist	36	***	(+) NH _x	10	*	(+) moist		*		
										gdd x moist		*	(+) moist		*					
Extractable NH _x	(+) gdd	44	**	(+) N	11	**	-	-	-	(+) gdd	44	**	(+) NO _y	10	**	-	-	-		
				(+) moist		*							(+) gdd		*					
Total soil N	(+) rain	45	***	N x rain	19	**	-	-	-	NO _y x rain	44	***	(+) rain	18	*	(-) NO _y	23	*		
Total soil C	(+) rain	45	***	(-) rain	17	*	-	-	-	NO_y x rain	44	***	NO_y x rain	19	**	-	-	-		
				N x rain		*														
				gdd x rain		*														
Soil C:N ratio	(-) rain	43	*	-	-	-	-	-	-	(-) rain	43	*	-	-	-	(+) NO _y	22	**		
Litter Ph-Oxidase	(-) gdd	44	*	-	-	-	gdd x rain	19	**	(-) gdd	44	*	(+) NH _x	19	**	NH _x x gdd	19	**		
	gdd x rain		*							gdd x rain		*	NO_y x rain		**					
Litter PME	(-) rain	47	***	(+) N	21	*	-	-	-	(+) NH _x	46	*	(-) rain	21	*	(+) NO _y	23	*		
	rain x N		**	(-) rain		*				(-) rain		***								
										NH_{x} x rain		**								
Calluna litter N	(+) N	23	*	(+) N	2	*	-	-	-	-	-	-	(+) NH _x	2	*	-	-	-		
				(-) gdd		*							(-) gdd		*					
Calluna litter C:N	(-) N	23	**	-	-	-	-	-	-	(-) NH _x	23	**	-	-	-	(+) NH _x	18	*		
Calluna foliar N	(-) gdd	48	*	-	-	-	(+) gdd	20	*	(+) gdd	48	*	-	-	-	NH _x x gdd	22	×		
							N x rain		*											

(+/-) = direction of response; x = interaction; P values = * <0.05, **<0.001, ***<0.001; gdd = growing degree days; moist = soil moisture; PME = phosphomonoesterase.

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Calluna tissue chemistry

The main factor affecting *Calluna* tissue N concentrations across all sites was temperature (Figure 4a). A significant quadratic relationship was found between (increasing) temperature and (higher) tissue N concentrations up to a threshold equivalent to 1500 GDD ($F_{1,48} = 5.4$, P < 0.05, Table 3: Model 1). When upland and lowland datasets were considered separately, this relationship was only significant for the lowlands ($F_{2,23} = 5.5$, P < 0.05).

Soil pH

Soil pH ranged from 3.5 to 5.4, decreasing significantly in relation to total N deposition ($F_{2,49} = 12.0$, P<0.01, Table 3: model 1) and increasing at sites with higher temperatures ($F_{2,49} = 7.5$, P<0.01, Table 3: model 1) across all sites. The relationship with N appears to be driven by oxidised N ($F_{4,47} = 13.7$, P<0.001, Table 3: model 2), particularly in the upland sites.

Extractable soil nutrients

A positive relationship between reduced N deposition and extractable soil nitrate (NO_3^-) concentrations was found for upland sites ($F_{3,10} = 9.6$, P<0.05, Table 3: model 2). Across the combined, all sites dataset, however, interactions between temperature and soil moisture ($F_{2,38} = 6.9$, P<0.01) suggest that

extractable ${\rm NO_3}^-$ concentrations are greater in areas with element climatic conditions (i.e. warm and wet).

Extractable soil ammonium (NH₄⁺) concentrations also increased with increasing total N in the upland sites (F_{2,11} = 11.9, P<0.01, Table 3: model 1), although this pattern was not significant in the lowlands. As with NO₃⁻, significant relationships with climate indicate that extractable NH₄⁺ concentrations are higher in warmer areas (F_{4,9} = 8.6, P<0.05, Table 3: model 2); this relationship appears to be driven by the uplands dataset, where greater soil NH₄⁺ availability is also associated with higher rainfall (F_{4,9} = 18.1, P<0.01, Table 3: model 2).

Total soil nitrogen and carbon concentrations

Significant interactions between total N deposition and rainfall were found for total soil N ($F_{2,19} = 10.7$, P < 0.001, Table 3: model 1) and C ($F_{4,17} = 6.5$, P < 0.05, Table 3: model 1) in upland sites, with the highest concentrations occurring at wetter sites receiving high N inputs. Model 2 analyses show that oxidised N is driving this relationship (N: $F_{2,44} = 23.9$, P < 0.001; C: $F_{2,44} = 9.3$, P < 0.001, Table 3, Figure 4b). At lowland sites, a negative relationship was seen between oxidised N and total soil N concentrations, although this was only significant at the 5% level ($F_{1,23} = 6.3$, P < 0.05, Table 3: model 2). Climate is clearly also an important driver of soil C and N, with significantly higher concentrations at sites receiving more rainfall (N, $F_{1,45} = 15.1$,

P<0.001; C: $F_{1,45}$ = 13.3, P<0.001, Table 3: model 1). Indeed, at upland sites, an interaction between temperature and precipitation indicates that soil N and C concentrations are highest where rainfall and temperature are both high (N: $F_{3,18}$ = 9.7, P<0.01, model 2; C: $F_{4,17}$ = 5.8, P<0.05, model 1). At lowland sites, soil C:N ratios were highest at sites receiving greater oxidised N inputs ($F_{2,22}$ = 9.0, P<0.01, Table 3: model 2, Figure 4 ϵ).

Litter chemistry, enzyme activity and nutrient turnover

Across all sites, litter N concentrations were significantly higher in areas of higher N deposition ($F_{1,23} = 5.3$, P < 0.05, Table 3: model 1, Figure 4*d*). This relationship was driven by data from upland sites, with the strongest patterns seen in relation to reduced N ($F_{2,2} = ,34.0$, P < 0.05, Table 3: model 2). Litter C:N ratios were lower at sites with the highest N deposition ($F_{1,23} = 8.6$, P < 0.01, Table 3: model 1); this relationship was strongest in the lowlands for reduced N ($F_{1,18} = 5.5$, P < 0.05, Table 3: model 2), with a similar (non-significant) trend apparent for the uplands.

Overall, litter phenol oxidase activity was more strongly related to climate than to levels of N deposition. Across the full (all sites) data set, enzyme activities were lower at warmer sites ($F_{2,44} = 6.1$, P < 0.05, Table 3: model 1); a significant interaction was also apparent between temperature and rainfall, with the lowest activities recorded from areas where higher temperatures combined with greater quantities of rainfall ($F_{2,44} = 6.7$, P < 0.05, Table 3: model 1). In upland areas, phenol-oxidase activity was, however, higher at sites receiving greater inputs of reduced N ($F_{3,19} = 8.3$, P < 0.01, Table 3: model 2, Figure 5a), as well as those with high levels of both rainfall and oxidised N deposition ($F_{3,19} = 12.2$, P < 0.01, Table 3: model 2).

As for phenol-oxidase, litter PME activity across all sites was significantly related to climate, with activity decreasing as precipitation levels increased ($F_{2,47} = 15.0$, P < 0.001, Table 3: model 1). A positive relationship between total N deposition and litter PME activity was, however, apparent for upland sites ($F_{2,21} = 4.3$, P < 0.05, Table 3: model 1). Analysis of N forms revealed that this was driven by reduced N and that this relationship was also significant for the all sites dataset ($F_{3,46} = 4.3$, P < 0.05, model 2, Figure 5*b*).

Discussion

Above-ground responses

A decline in plant species diversity in relation to increasing atmospheric N deposition has been observed in a variety of ecosystems ranging from acid and calcareous grasslands [39,40] to boreal and temperate forests [41,42] and even arctic tundra [43]. Measurement of plant species richness can provide a reliable indication of habitat quality [44,45] and is considered to be a fundamental tool in assessing community and wider-scale diversity [46]. Where species losses are countered by increases in a few, dominant and/or invasive species, the risk to habitats are homogenisation with declines in genetic-, ecosystem- and landscape-scale diversity. The findings from this survey provide clear evidence that elevated N deposition significantly reduces the floristic diversity of heathland ecosystems, either solely or in combination with climatic drivers - particularly increasing temperature - a major driver of global biodiversity loss in both terrestrial and non-terrestrial ecosystems [4,47]. The nature of the relationship between total N deposition and species richness suggests similar patterns in both upland and lowland systems, namely that the greatest proportional reduction of species richness in response to increasing N inputs is seen at sites currently experiencing the lowest deposition loads. It appears that both oxidised and reduced forms of N are contributing to heathland biodiversity loss, although, at the functional group level there were more significant relationships between species richness and reduced N, compared to oxidised N, as hypothesised. This is in line with findings from similar surveys elsewhere for grassland [48) and heathland [19], providing further support for an additional impact associated with the acidifying properties of reduced N and/ or direct toxicity of ammonium ions, over and above that of Nenrichment per se. Given the broad cover of spatial, climatic and pollution indices, the magnitude of diversity loss that has been observed amongst forb, graminoid, bryophyte and lichen functional groups in this study not only validates experimental findings from many N manipulation studies [39,49] but, more profoundly, provides compelling evidence that N deposition is driving biodiversity loss amongst UK heathlands at a national scale.

In N-limited environments, increased N availability frequently leads to increased plant growth and/or expansion of nitrophilous species, potentially leading to competitive exclusion of slower growing habitat-specialist species adapted to low levels of N

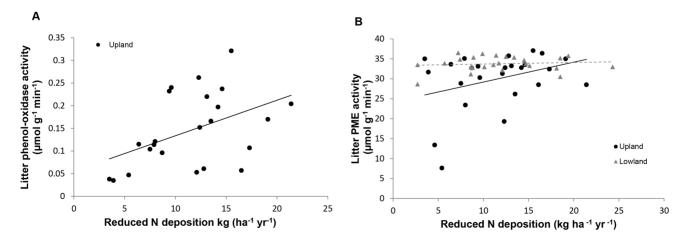


Figure 5. Relationships between N deposition and litter enzyme activity. Increasing reduced N deposition was positively correlated with increased litter phenol-oxidase activity in upland sites (A) and litter phosphomonoesterase (PME) activity across all sites (B). doi:10.1371/journal.pone.0059031.g005

availability [40]. Effects of N on soil pH, particularly in its reduced form [50], can also result in the decline of sensitive plant species, particularly those with narrow pH niches or low tolerance of high NH₄⁺ ion concentrations [51,52]. In the current study, Calluna plants were taller and soil pH lower in areas receiving high N inputs, suggesting that reduced understorey light availability and/ or unfavourable soil pH may be mechanisms contributing to biodiversity loss in British heathlands. Whilst bryophyte cover was not related per se to N deposition, shifts within bryophyte communities, notable at the species level, were significantly related to N inputs. For example, the characteristic heathland moss species Hylocomnium splendens and Pleurozium schreberii were seen to decline as N deposition increased, whilst Brachythecium rutabulum, a nitrophilous species more commonly associated with base rich, eutrophicated habitats [53] was more abundant at sites with higher N deposition. Similar species shifts have been seen in boreal forest systems [41,54] with H. splendens declining, and being replaced by B. rutabulum, at relatively modest input rates of 10 kg N $ha^{-1} yr^{-}$

Although graminoid abundance increased significantly with increasing N deposition, a concurrent decrease in graminoid diversity, coupled with sharp declines in the abundance of typical heathland grass species (e.g. *Festuca ovina, Danthonia decumbens*) indicates that grass invasion at higher N deposition loads is dominated by only a small number of species (specifically *Deschampsia flexuousa* or *Molinia caerulea*). These findings support earlier observations of *D. flexuousa* and *M. caerulea* increases in high N deposition areas in the Netherlands [55] and an associated conversion of 35 % of Dutch heathlands to grasslands in the 1980s [56]. These species have also been found to increase in Danish bog systems exposed to N levels of 10–15 kg ha⁻¹ yr⁻¹, to the detriment of characteristic ombrotrophic vegetation [57].

N deposition is associated with negative impacts on lichen communities in many ecosystems [58-60], a phenomenon often linked to increased competition for light as a result of N-driven stimulation in higher plant growth [9,61]. However, direct toxicity of NO₂⁻, NO₃⁻ and NH₄⁺ is also likely to be a contributing factor in the decline of non-vascular plants [16,62,63]. Whilst lichen abundance was negatively related to both reduced and oxidised forms of N in this study, declines in both the cover and species richness of lichens were statistically stronger with reduced N. As with most higher plants, lichens can effectively assimilate NH₄⁺ at modest levels [64] rapidly converting it into amino acids to mitigate its cytotoxicity. However, for acidophytic species (such as those found in heathlands) this assimilation capacity is limited and toxic levels of unassimilated NH₄⁺ can accumulate in the lichen thallus [65]. It seems likely, therefore, that both competitive exclusion and potentially also direct toxicity effects are contributing to the decline in abundance and diversity of lichens observed with increasing N deposition in the current study.

Below-ground responses

At all sites in this study, litter N concentrations increased, and C:N ratios decreased, with increasing N inputs suggesting greater availability of organic N for soil microorganisms and thus faster rates of nutrient cycling. Microbial enzyme activity provides a useful proxy for decomposition rates and has been frequently used to monitor the functional responses of microbial populations to global change perturbations [10,66]. Our results show that overall the activity of phenol-oxidase — an enzyme responsible for breakdown of the structural components of plant material [67] — in litter was greater at sites receiving higher inputs of reduced N. This relationship was seen in both upland and lowland systems, the latter, however, stronger in areas of greater rainfall. Significant

interactions between oxidised N and rainfall in upland systems were also apparent, possibly reflecting the wider span of oxidised N inputs (3–15 kg ha⁻¹ yr⁻¹) compared to lowland sites (3–9 kg ha⁻¹ yr⁻¹). The pattern of increased litter/soil enzyme activity with increasing N deposition supports results from a recent survey of biochemical indicators in lowland heathlands [19]. However, contrasting results have been reported previously for moorlands [18] and temperate forests [66,67].

The positive relationship between N deposition – particularly reduced N - and litter PME activity provides the first field scale evidence of an increase in P demand across both upland and lowland heathland sites receiving high rates of N deposition. These findings are in line with those from manipulation experiments across a range of ecosystems [68,69], and with survey evidence of increased plant uptake of P across transects of increasing N deposition in lowland heathland [15,70].

The main factors regulating decomposition are moisture and temperature [6,71]; this is also apparent in our survey results with enzyme activities highest in warmer, wetter areas. Climate change predictions for warmer, drier summers and cooler, wetter winters will, therefore, have implications for the activities of enzymes involved in the decomposition process, with likely consequences for nutrient availabilities and uptake. Concentrations of soil extractable NO₃ and NH₄ were significantly higher with increasing N deposition in the upland sites, although no such relationship was evident in the lowlands. The strong influence of climate on rates of nutrient turnover and availability [72,73], and generally lower annual rainfall in lowland (600–1300 mm) compared to upland (800-2500 mm) sites, may explain the lack of relationship with N in lowlands and the generally higher abundance of litter in upland sites. This may be particularly the case in this survey since sampling was carried out in the summer when soil moisture levels are at their lowest and plant and microbial immobilisation of N are at their highest [74]. Caution must be exercised when making assumptions based on 'snapshot' measurements of mineral pools (particularly across broad temporal and spatial gradients) that are inherently subject to large interseasonal and inter-annual variations [75]. Furthermore, recent Nfocused surveys carried out by Jones & Power [19] and Stevens et al. [76] found that soil type, underlying geology, climatic and altitudinal variation were important drivers of soil and plant responses, influencing the nature of relationships with N deposition.

Higher soil C:N ratios at sites with higher N inputs, specifically in the lowlands, indicate a shift towards immobilisation with increasing N deposition. Lower litter C:N ratios at high N sites contrast with this pattern, although may be explained by the previously mentioned N-driven stimulation at the early stage of decomposition where labile, energy rich substrates are abundant, followed by a slowdown in later stages as the relative proportions of recalcitrant compounds (such as lignin) progressively increase [77]. Whilst many studies of N deposition impacts have found that total soil N content increased with elevated N inputs [69,78,79], no evidence of N or C accumulation in relation to elevated N as a sole driver was found in this survey. This may reflect the depth (0– 15cm) of the soil core sampled, which not only results in high levels of between site variability in depth of organic layer included in the sample (from <1cm to 15 cm), but may also be too deep to capture nutrient signals from recent decades that are present in the upper horizons.

Across all sites, the most significant driver influencing soil total N and C content was precipitation. Since climatic factors are known to strongly influence the production of plant material (and, subsequently, plant residues) [80], the dominant influence of

rainfall is not surprising. Indeed, our results suggest that climatic variables are important drivers of many ecosystem processes and, therefore, are likely to influence responses to N deposition. Given that climate models predict altered precipitation and temperature patterns for the northern hemisphere [81], and that widespread exceedance of critical N loads is predicted across Europe for the foreseeable future [2,82], the implications of global change for the biodiversity, structure and function of these internationally important heathland ecosystems are very considerable.

Conclusion

The findings from this survey provide novel support to a growing body of evidence demonstrating a strong link between N deposition and an increase in nitrophilous plant species across a wide range of habitats [17,22]. An associated decline in the diversity of both higher and lower plants across broad spatial and climatic gradients - in both upland and lowland environments highlights the ubiquity of relationships with N, providing clear field evidence of widespread N-driven changes in the composition, structure and diversity of British heathlands. There is also strong evidence to suggest that current rates of N deposition are altering many biogeochemical processes, particularly rates of nutrient

References

- 1. Vitousek PD, Aber JD, Howarth PW, Likens GE, Matson PA, et al. (1997) Human alteration of the global nitrogen cycle: sources and consequences. Ecological applications 7: 737-750.
- 2. Galloway JN, Dentener FJ, Capone DG, Boyer EW, Howarth RW (2004) Nitrogen cycles: past, present and future. Biogeochemistry 70: 153-226.
- EMEP (European Monitoring and Evaluation Programme). Available: http:// www.emep.int. Accessed 15 August 2012.
- 4. Sala OE, Chapin FS, Armesto JL (2000) Global biodiversity scenarios for the year 2100. Science 287: 1770-1774.
- 5. Phoenix GK, Hicks WK, Cinderby S, Kuylenstierna JCI, Stock WD, et al. (2006) Atmospheric nitrogen deposition in world biodiversity hotspots: the need for a greater global perspective in assessing N deposition impacts. Global Change Biology 12: 470-476.
- Kleijn D, Bekker RM, Bobbink R, de Graaf MC, Roelofs JGM (2008) In search for key biogeochemical factors affecting plant species persistence in heathlands and acidic grasslands: a comparison of common and rare species. Journal of Applied Ecology 45: 680–687
- Pîlkington MG, Caporn SJM, Carroll JA, Cresswell N, Lee JL, et al. (2007) Phosphorus supply influences heathland responses to atmospheric nitrogen deposition. Environmental Pollution 148: 191-200.
- Power SA, Ashmore MR, Cousins DA, Sheppard LJ (1998) Effects of nitrogen addition on the stress sensitivity of Calluna vulgaris. New Phytologist 138: 663-
- Bobbink RK, Hicks TK, Galloway JN, Spranger T, Alkemade T, et al. (2010) Global assessment of nitrogen deposition effects on terrestrial plant diversity, a synthesis. Ecological Applications 20: 30-59.
- Schimel JP, Bennett J, Fierer N (2005) Microbial community composition and soil nitrogen cycling: is there really a connection? In: Bardgett RD, et al. (editors). Biological diversity and function in soils. Cambridge University Press, Cambridge, UK. pp. 171-188.
- 11. Power SA, Barker CG, Allchin EA, Ashmore MR, Bell JNB (2001) Habitat management: a tool to modify ecosystem impacts of N deposition? The Scientific
- 12. Green ER, Ellis RJ, Gadsdon SC, Milcu A, Power SA (2013) How does N deposition affect belowground heathland recovery following wildfire? Soil Biology and Biochemistry 57: 775-783.
- 13. Phoenix GK, Leake JR, Read DJ, Grime JP, Lee JA (2004) Accumulation of pollutant N in calcareous and acidic grasslands: evidence from N flux and 15N tracer studies. Water, Air and Soil Pollution, Focus 4: 159-167.
- 14. Sutton MA, Pitcairn CER, Whitfield CP, Leith ID, Sheppard LJ, et al. (2004) Bioindicator and biomonitoring methods for assessing the effects of atmospheric nitrogen on statutory nature conservation sites. JNCC Report. JNCC, Peterborough, UK. pp. 230-356.
- 15. Pitcairn CER, Fowler D, Leith I, Sheppard LJ, Tang S, et al. (2006) Diagnostic indicators of nitrogen deposition. Environmental Pollution 144: 941-950.
- 16. Remke E, Brouwer E, Kooijman A (2009) Even low to medium nitrogen deposition impacts vegetation of dry, coastal dunes around the Baltic Sea. Environmental Pollution 157: 792-800.
- 17. Maskell LC, Smart SM, Bullock JM, Thompson K, Stevens CJ (2010) Nitrogen deposition causes widespread loss of species richness in British habitats. Global Change Biology 16: 671-679.

cycling and availability, and that these relationships are influenced by climatic conditions. Litter N concentrations, litter enzyme activity (phenol-oxidase and PME) and, for the uplands, soil extractable N concentrations, all appear to have consistent relationships with total N deposition and may, therefore, be useful biochemical indicators of, and responses to, N deposition. Given future predictions for climate change and continued wide scale exceedance of critical N loads across Europe, further effects of these global change phenomena on heathland diversity and associated ecosystem services can be expected.

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Author Contributions

Conceived and designed the experiments: SP SC AB. Performed the experiments: GS CF AB. Analyzed the data: GS SP. Contributed reagents/materials/analysis tools: GS CF SP . Wrote the paper: GS SP CF SC AB.

- 18. Edmondson JL, Carroll JA, Price EA (2010) Bio-indicators of nitrogen pollution in heather moorland. Science of the Total Environment 408: 6202-6209.
- 19. Jones AG, Power SA (2012) Field scale evaluation of effects of nitrogen deposition on the functioning of heathland ecosystems. Journal of Ecology 100:
- 20. Power SA, Collins CM (2010) Biochemical signals of nitrogen deposition across an urban-rural gradient. Atmospheric Environment 44: 1772-1780.
- 21. Stevens CJ, Dise NB, Gowing DJG, Mountford JO (2006) Loss of forb diversity in relation to nitrogen deposition in the UK: regional trends and potential controls. Global Change Biology 12: 1823-1833.
- Stevens CJ, Dupre C, Dorland E, Gaudnik C, Gowing DJG (2010) Nitrogen deposition threatens species richness of grasslands across Europe. Environmental Pollution 158: 2940-2945.
- Phoenix GK, Emmett BA, Britton AJ, Caporn SJM, Dise NB, et al. (2012) Impacts of atmospheric nitrogen deposition: responses of multiple plant and soil parameters across contrasting ecosystems in long-term field experiments. Global Change Biology 18: 1197–2215.
- 24. Aerts R (1990) Competition in heathland along an experimental gradient of nutrient availability. Oikos 57: 310-318.
- 25. Power SA, Green ER, Barker CG, Bell JNB, Ashmore MR (2006) Ecosystem recovery: heathland response to a reduction in N deposition. Global Change Biology 12: 1241-1252.
- Barker CG, Power SA, Bell JNB (2004) Effects of habitat management on heathland response to atmospheric nitrogen deposition. Biological Conservation 120: 41-52
- 27. RoTAP (2012) Review of Transboundary Air Pollution: Acidification, Eutrophication, Ground Level Ozone and Heavy Metals in the UK. Contract Report to the Department for Environment, Food and Rural Affairs. Centre for Ecology & Hydrology, UK
- 28. Sheppard LJ, Crossley A, Leith ID, Hargreaves KJ, Carfrae JA, et al. (2009) An automated wet deposition system to compare the effects of reduced and oxidised N on ombrotrophic bog species: practical considerations. Water, Air and Soil Pollution Focus 4: 197-205.
- Smart SM, Clarke RT, van de Poll HM, Robertson EJ, Shield ER, et al. (2003) National-scale vegetation change across Britain; an analysis of sample-based surveillance data from the Countryside Surveys of 1990 and 1998. Journal of Environmental Management 67: 239-254.
- 30. Stevens CJ, Dise NB, Gowing DJG, Mountford JO (2006) Loss of forb diversity in relation to N deposition in the UK: regional trends and potential controls. Global Change Biology 12: 1823-1833.
- 31. JNCC (Joint Nature Conservancy Committee) (2004) Common standards monitoring guidance for lowland heathland. Report ISSN 1743-8160. Available: http://jncc.defra.gov.uk/pdf/CSM_lowland_heathland.pdf Accessed 20 April 2012.
- Gimingham CH (1972) Ecology of Heathlands. Chapman & Hall. London. Watson RT, Zinyowera MC, Moss RH (editors.) (1995) Climate Change 1995: Impacts, adaptations and mitigations of climate change: Scientific technical analyses. Cambridge University Press, Cambridge UK. 878 pp.
- 34. Carroll JA, Caporn SJM, Cawley L, Read DJ, Lee JA (1999) The effect of increased deposition of atmospheric nitrogen on Calluna vulgaris in upland Britain. New Phytologist 141: 423-431.

- Southon GE, Green ER, Jones AG, Barker CG, Power SA (2012) Long-term nitrogen additions increase likelihood of climate stress and affect recovery from wildfire in a lowland heath. Global Change Biology 18: 2824–2837.
- Johnson D, Leake JR, Lee JA, Campbell CD (1998) Changes in soil microbial biomass and microbial activities in response to 7 years simulated pollutant nitrogen deposition on a heathland and two grasslands. Environmental Pollution 103: 239–250.
- 37. Crawley MJ (2007) The R Book. Wiley Blackwell. London, UK.
- Siebel HN (2005) Standaardlijst van de Nederlandse blad-, lever- en hauwmossen [Checklist of Dutch Bryophytes and Liverworts]. Buxbaumiella 73: 26–64.
- Clark CM, Tilman D (2008) Loss of plant species after chronic low-level nitrogen deposition to prairie grasslands. Nature 451: 712–715.
- Stevens CJ, Dise NB, Moutford JO (2004) Impact of nitrogen deposition on the species richness of grasslands. Science 303: 1876–1879.
- Strengbom JA, Nordin A, Nasholm T, Ericson L (2001) Slow recovery of a boreal forest ecosystem following decreased nitrogen input. Functional Ecology 15: 454–457.
- 42. Gilliam FS, Roberts ME (2003) The herbaceous layer in forests of eastern North America. Oxford University Press, New York, USA.
- Gordon C, Wynn JM, Woodin SJ (2001) Impacts of increased nitrogen supply on high Arctic heath: the importance of bryophytes and phosphorus availability. New Phytologist 149: 461–471.
- Godefroid S, Koedam N (2003) Distribution pattern of the flora in a peri-urban forest: an effect of the city-forest ecotone. Landscape and Urban Planning 65: 169–185.
- Dupre C (2001) Regional and local variation in plant species richness. Acta Universaitatis Upsaliensis. Comprehensive summaries of Uppsala dissertations from the Faculty of Science and Technology. Uppsala. ISBN 91-554-5064-4.
- Gotelli NJ, Colwell RK (2001) Quantifying biodiversity: procedures and pitfalls in the measurements and comparison of species richness. Ecology Letters 4: 379–391.
- Van Vuuren DP, Sala OE, Pereira HM (2006) The future of vascular plant diversity under four global scenarios. Ecology and Society 11. Available: www. ecologyandsociety.org vol11/iss2/art25/[Accessed 20-8-12].
- Stevens C, Maskell LC, Smart SM, Caporn SJM, Dise NB, et al. (2009) Identifying indicators of atmospheric nitrogen deposition in calcifugous grasslands. Biological Conservation 142: 2069–2075.
- Čarroll JA, Caporn SJM, Johnson D, Morecroft MD, Lee JA (2003) The interactions between plant growth, vegetation structure and soil processes in semi-natural acidic and calcareous grasslands receiving long-term inputs of simulated pollutant nitrogen deposition. Environmental Pollution 121: 363–376.
- Sheppard L, Wallander H (2004) In: Amancio S, Stulen I, (editors). Nitrogen acquisition and assimilation in higher plants. Kluwer Academic Publishers, Netherlands. pp. 65–98.
- Nordin A, Strengbom J, Forsum A (2009) Complex biotic interactions drive long-term vegetation change in a nitrogen enriched boreal forest. Ecosystems 12: 1204–1211.
- Stevens C, Dise NB, Gowing DJ (2009) Regional trends in soil acidification and exchangeable metal concentrations in relation to acid deposition rates. Environmental Pollution 157: 313–319.
- Hill MO, Preston CD, Bosanquet SDS, Roy DB (2007) BRYOATT: attributes
 of British and Irish mosses, liverworts and hornworts. Centre for Ecology and
 Hydrology, Cambridge, UK.
- Hallingback T (1992) The effect of air pollution on mosses in southern Sweden. Biological Conservation 59: 163–170.
- 55. Heil GW, Aerts R (1993) Synthesis: perspectives for heathlands. In: Heil GW, Aerts R (editors). Heathlands, patterns and processes in a changing environment. Kluwer Academic Publishers. pp. 201–217.
- Bobbink R, Heil G (1993) Atmospheric deposition of sulphur and nitrogen in heathland ecosystems. In: Aerts R, Heil GW (editors). Heathlands, Patterns and Processes in a Changing Environment . Kluwer Academic Publishers, The Netherlands. pp. 25–50.
- 57. Aaby B (1994) Monitoring Danish raised bogs. In: Gruenig, A (editor). Mires and man: Mire conservation in a densely populated country: The Swiss experience. The Swiss Federal Institute for forests, snow and landscape. Birmensdorf, Switzerland. pp 284–300, 391–40.
- Baddeley JA, Thompson DBE, Lee JA (1994) Regional and historical variation in the nitrogen-content of *Racomitrium lanuginosum* in Britain in relation to atmospheric nitrogen deposition. Environmental Pollution 84: 189–196.
- Britton AJ, Fisher JM (2007) Interactive effects of nitrogen deposition, fire and grazing on diversity and composition of low-alpine prostrate *Calluna vulgaris* heathland. Journal of Applied Ecology 44: 125–135.

- Fenn ME, Jovan S, Yuan F (2008) Empirical and simulated critical loads for nitrogen deposition in Californian mixed conifer forests. Environmental Pollution 155: 492–511.
- Barker C (2001) The impact of management on heathland response to increased nitrogen deposition. PhD Thesis. Imperial College London, UK.
- Hogan E, Minnullina G, Sheppard LJ, Leith ID, Crittenden PD (2010) Effects of nitrogen enrichment on phosphatase activity and nitrogen:phosphorus relationships in *Cladonia portentosa*. New Phytologist 186: 911–925.
- Hyvarinen M, Crittenden PD (1998) Relationships between atmospheric nitrogen inputs and the vertical nitrogen and phosphorus concentration gradients in the lichen *Cladonia portentosa*. New Phytologist 140: 519–530.
- Hauck M (2010) Ammonium and nitrate tolerance in lichens. Environmental Pollution 158: 1127–1133.
- Dahlman L, Persson J, Nasholm T (2003) Carbon and nitrogen distribution in the green algal lichens *Hypogymnia physodes* and *Platismatia glauca* in relation to nutrient supply. Planta 217: 41–48.
- Carreiro MM, Sinsabaugh RL, Repert DA (2000) Microbial enzyme shifts explain litter decay responses to simulated nitrogen deposition. Ecology 81: 2359–2365.
- Sinsabaugh RL, Carreiro MM, Alvarez S (2002) In:Burns RG, Dick RP (editors). Enzymes in the environment: activity, ecology and applications. Marcel Dekker Inc., New York, USA. pp. 249–265.
- Phoenix GK, Booth RE, Leake JR, Read DJ, Grime P, et al. (2003) Effects of enhanced N deposition and phosphorus limitation on N budgets of semi-natural grasslands. Global Change Biology 9: 130–132.
- Saiya-Cork KR, Sinsabaugh RL, Zak DR (2002) The effects of long term nitrogen deposition on extracellular enzyme activity in an *Acer saccharum* forest soil. Soil Biology and Biochemistry 34: 1309–1315.
- Rowe EC, Smart SM, Kennedy VH, Emmett BA, Evans CD (2008) N deposition increases the acquisition of phosphorus and potassium by heather Calluna vulgaris. Environmental Pollution 155: 201–207.
- Giller KE (2005) Soil biodiversity: scaling down and scaling up. In: Bardgett RD, et al. (editors). Biological diversity and function in soils. Cambridge University Press. Cambridge, UK. pp. 295–319.
- Criquet S, Ferre E, Farnet AM (2004) Annual dynamics of phosphatase activities in an evergreen oak litter: influence of biotic and abiotic factors. Soil Biology and Biochemistry 36: 1111–1118.
- Jamieson N, Barraclough S, Unkovich M (1998) Soil N dynamics in a natural calcareous grassland under a changing climate. Biology and Fertility of Soils 27: 267–273.
- Morecroft MD, Marrs RH, Woodward FI (1992) Altitudinal and seasonal trends in soil nitrogen mineralisation rate in the Scottish highlands. Journal of Ecology 80: 49–56.
- O'Sullivan O, Horswill P, Phoenix G (2011) Recovery of soil nitrogen pools In species-rich grasslands after 12 years of simulated pollutant nitrogen deposition: 16 year experimental analysis. Global Change Biology 17: 2615–2628.
- Stevens CJ, Manning P, van den Berg LJL, de Graaf MCC, Wamelink GW, et al. (2011) Ecosystem responses to reduced and oxidised nitrogen inputs in European terrestrial habitats. Environmental Pollution 159: 665–676.
- Kirk TK, Farrell RL (1987) Enzymatic combustion. The microbial degradation of lignin. Annual Review of Microbiology 41: 465–505.
- Power SA, Ashmore MR Cousins DA (1998) Impacts and fate of experimentally enhanced nitrogen deposition on a British lowland heath. Environmental Pollution 102: 27–30.
- Pilkington MG, Caporn SJM, Carroll JA, Lee JA, Reynolds B, et al. (2005) Effects of increased deposition of atmospheric nitrogen on an upland moor, nitrogen budgets and nutrient accumulation. Environmental Pollution 138: 473

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- Wells ED, Warren WG (1997) Height growth, needle mass and needle nutrient concentration of N, P, K, Ca, Mg and Cu in a 6 year old black spruce peatland plantation in Newfoundland, Canada. Scandinavian Journal of Forest Research 12: 138–148.
- 81. IPCC (2007) Climate Change Synthesis Report. Contribution of Working Groups I, II and III to the Fourth Assessment. In: Pachauri, RK, Reisinger A, editors. Report of the Intergovernmental Panel on Climate Change. IPCC, Geneva, Switzerland. pp.104.
- Matejko M, Dore AJ, Hall J, Dore CJ, Blas M, et al. (2009) The influence of long term trends in pollutant emissions on deposition of sulphur and nitrogen and exceedance of critical loads in the United Kingdom. Environmental Science and Policy 12: 882–896.