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## The role of leptin in osteoarthritis

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### Abstract

Background: The pathogenesis of osteoarthritis (OA) is not clear; leptin may be related to its pathogenesis.

**Methods:** We reviewed articles on leptin in OA, chondrocytes, and in vitro experiments. It is concluded that leptin may lead to OA via some signaling pathways. At the same time, the concentration of leptin in vitro experiments and OA/rheumatoid arthritis (RA) patients was summarized.

**Results:** Leptin levels in serum and synovial fluid of OA/RA patients were higher than normal person. In the condition of infection and immunity, serum leptin levels in the peripheral blood significantly increase. Because of the close relationship between obesity, leptin, and OA, it is crucial to study the effects of weight loss and exercise intervention on serum leptin levels to improve the symptoms of OA patients.

**Conclusion:** Treatment for leptin-increased obesity may be a treatment for OA. The role of leptin in OA cannot be ignored and needs to be further studied.

**Abbreviations:** OA = osteoarthritis, RA = rheumatoid arthritis.

Keywords: cartilage, chronic disease, leptin, osteoarthritis, signaling pathway

### 1. Introduction

Osteoarthritis (OA) is a common degenerative disease of articular cartilage which mainly occurs in the older population.<sup>[1]</sup> There are a lot of risk factors for OA, such as obesity, age, trauma, sex, and so on. And this disease will eventually lead to severe pain and joint movement disorders.<sup>[2]</sup> The pathogenesis of OA has not been clear, whereas a recently found adipocyte factor called leptin was involved in the body's metabolism and the immune adjustment and its expression was significantly increased in OA patients. Leptin was considered as an important participant in the development of OA. This article will focus on the role of leptin in OA development.

### 2. Results

## 2.1. The structure and function of leptin and its relationship with OA

Leptin is a peptide hormone which was first reported in 1994 mainly comes from fat tissue. This 16kD hormone was produced

Medicine (2018) 97:14(e0257)

Received: 25 November 2017 / Received in final form: 4 March 2018 / Accepted: 6 March 2018

http://dx.doi.org/10.1097/MD.000000000010257

by the ob/ob gene and belonged to the type 1 cytokine superfamily.<sup>[3,4]</sup> Its receptor was encoded by db/db gene and belonged to the type 1 cytokine receptor superfamily.<sup>[5]</sup> There are lots of homology of leptin receptors, such as obRa, obRb, obRc, obRd, obRe, and obRf,<sup>[6]</sup> in which the only long receptor obRb is the most widely expression and functional receptor mainly through JAK/STAT pathway.<sup>[7]</sup>Figure 1 shows how JAK/STAT signaling regulates leptin expression. Like adiponectin and visfatin,<sup>[8,9]</sup> leptin was also known as a adipokine,<sup>[10]</sup> It was earliest found to play an important role in energy metabolism<sup>[11]</sup> because it could lead to a loss of appetite and an increased energy consumption.<sup>[12]</sup> Leptin levels in obesity, in turn, were significantly elevated in the human body.<sup>[13]</sup> Due to its high serum levels in high weight individuals and the relieving joint symptoms by losing weight in OA patients,<sup>[14,15]</sup> we hypothesized that leptin has some connections with OA caused by obesity. Afterward, leptin proved to participate in the inflammatory response which further shows that leptin may play an important role in the development of OA.<sup>[16]</sup> Griffin et al proved that a lack of leptin does not cause spontaneous OA with the experiment in mouse model indicating that losing of leptin signaling pathways may protect body from the development of OA.<sup>[17]</sup> There is a genetic correlation of leptin and OA.<sup>[18]</sup> First, the leptin gene was increasingly expressed in OA cartilage chondrocyte. And it is also showed that leptin gene and its receptor gene are associated with OA with single nucleotide polymorphism analysis.<sup>[19,20]</sup>

## 2.2. The expression of leptin in serum and synovial fluid of OA patients

Normal leptin levels in human blood were related to sex and age. Argente et al found that leptin levels in female were significantly higher than male with the same age especially after the age of 12 because leptin expression decreased in male and rose in female after the age of 12.<sup>[21]</sup> Leptin levels in peripheral blood of OA and rheumatoid arthritis (RA) patients were higher than normal person.<sup>[22]</sup> A high aggregation phenomenon of leptin in synovial fluid of patients with RA was detected. However, its concentration was lower when compared with the serum leptin levels.<sup>[23]</sup> A

Editor: Antonino Bianco.

MY and JZ equally contributed to this work.

This study was approved by the ethics review committee of The First Affiliated Hospital of Soochow University, and written informed consent was obtained from all participants of the study.

The authors have no funding and conflicts of interest to disclose.

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Figure 1. Leptin binds to the 2 CK domains and causes receptor dimerization. The dimerization receptor activates JAK2, which then phosphorylates STAT3. STAT3 forms dimers and exposes nuclear signals, then enters the nucleus to regulate gene expression.

similar situation was found in osteoarthritic synovial fluid because the severity of OA and the level of synovial fluid leptin were positively correlated.<sup>[24]</sup> Leptin levels in serum and synovial fluid of normal adults and OA/RA patients were shown in Tables 1 and 2. The expression of leptin's short receptor was detected in articular cartilage in normal people and patients with early OA, whereas the long receptor which actually involved in signaling pathways was poor expressed. In joints of patients with severe OA, the expression of leptin and its long receptor were significantly increased, and the expression of leptin and its receptor and the severity of OA were positively correlated. This

### Table 1

Leptin levels in serum and synovial fluid of normal adults and OA patients<sup>[26,27]</sup>.

First author	Control	OA	Р
de Boer			
Ν	132	172	
Serum, ng/mL	18 (13) <sup>*</sup>	73 (61) <sup>*</sup>	<.001
Ku			
Ν	10	42	
Synovial fluid, ng/mL	2.05 (1.0-4.6) <sup>†</sup>	4.40 (0.5–15.8) <sup>†</sup>	<.001

OA = osteoarthritis.

Data shown are means (SD).

<sup>+</sup> Values are median (range).

suggests that the expression of leptin and its receptor increased with the development of  $OA.^{[24,25]}$ 

Cartilage wear and degradation is the key step in the development of OA. The high concentration of leptin in synovial fluid and the expression of leptin receptor on cartilage cell surface indicate that leptin may act a certain role in cartilage degeneration.<sup>[27]</sup> The expression of inflammation factor in normal articular cavity was scanty, whereas synovial fibroblasts presented a dose-dependent rising in the expression of IL6 when treated with leptin.<sup>[28]</sup> IL6 broadly existed in serum and synovial fluid of OA patients as a kind of inflammatory cytokine,<sup>[29]</sup> which can lead to degradation of proteoglycan<sup>[30]</sup> and suppress the formation of cartilage by reducing cartilage proteoglycan synthesis through NOTCH pathway and increasing the expression the expression of cartilage by reducing the expression of the expressin the expression of the expression of the expressi

# Table 2 Leptin levels in serum and synovial fluid of normal adults and RA patients<sup>[23]</sup>.

	Control (n=15)	RA (n=20)
.eptin, ng/mL		
Serum	$11.01 \pm 7.08$	$18.35 \pm 9.85^{*}$
Synovial fluid	$7.69 \pm 5.22$	$14.31 \pm 6.84^{*}$

RA = rheumatoid arthritis.

<sup>\*</sup> P<.05.

sion of decomposition factor MMP13.<sup>[31]</sup> The expression of IL6 decreases obviously in synovial fibroblasts in patients with OA when leptin receptor blockers were used. This suggested that leptin can induce the release of inflammatory cytokines and promote the progress of OA through cartilage damage mediated by IL6.<sup>[28]</sup> Yang et al found that IL-1 expression in cartilage cells greatly increased on the seventh day when treated with leptin. In addition to IL1, content of cartilage collagen degradationpromoting factors MMP9 and MMP13 was also increased.<sup>[25,32]</sup> Leptin and IL1 together could result in the abnormal expression of NOS<sub>2</sub> which can promote cartilage cell apoptosis by p53 signal pathway and induce the production of matrix metalloproteinase and prostaglandin.<sup>[28,33]</sup> These evidences suggested that leptin promotes the progress of OA by enhancing the NOS<sub>2</sub> system with the help of IL1. In addition to the above several inflammatory cytokines, leptin also promoted the expression of other cartilage decomposition factors, such as IL8, MMP2, cathepsinD, and so on.<sup>[34]</sup> What is more, leptin mediated a dose-dependent expression of VCAM in the synovium in RA and OA cartilage cells and mice ATDC-5 cells. VCAM correlated with severe OA and hand OA, and block of Leptin pathway can significantly reduce the production of VCAM.<sup>[35,36]</sup>

Leptin itself did not damage joints and cartilage, and its effects on cartilage were two-tier. In addition to promoting the progress of OA, leptin can also promote the synthesis of cartilage proteoglycan. Leptin expression in osteoblast was around 5 times in patients within OA patients than normal people.<sup>[34,37,38]</sup> A similar phenomenon can be observed in articular cartilage cells. Osteoblast and articular cartilage cells produce increased TGFbeta and IGF-1 under leptin treatment. The 2 kinds of growth factors can promote the synthesis of proteoglycan. Leptin in low concentration can promote proteoglycan and type 2 collagen formation, whereas high concentration of leptin can induce the proliferation of articular cartilage cells.<sup>[39]</sup>

### 2.3. The relationship between leptin and immunemediated OA

Leptin also participated in the immune regulating system. The expression of leptin increases under different acute inflammatory stimulation of irritants.<sup>[40]</sup> In the condition of infection and immunity, serum leptin levels in the peripheral blood significantly increase.<sup>[23]</sup> Interactions between Inflammatory factors will also promote the expression of leptin. For example, the level of leptin will change as the concentration of IL and TNF changes. Leptin is reflected in many autoimmune diseases, such as RA, diabetes, and multiple sclerosis.<sup>[41]</sup> Pathogenesis of OA is still unclear. Recently, Wang et al found that immune responses played an important role in the onset of OA with abnormal expression and activation of complement in the synovium and synovial fluid,<sup>[42]</sup> suggesting that complement was involved in the pathogenesis of OA. In addition to complement, inflammatory markers such as IL-1 beta in osteoarthritic synovial tissues were also detected,<sup>[43]</sup> IL-1 beta is not only involved in the immune inflammation in the joints, but also can inhibit the type 2 collagen synthesis.<sup>[44]</sup> B cells and CD4 + T cells and macrophages' infiltration in osteoarthritic synovial tissue were also reported.<sup>[45,46]</sup> All the evidence suggests the immune system was involved in the occurrence of OA. In addition to regulate metabolism, leptin also has certain effects in immune system, particularly in the T-cell proliferation and differentiation,<sup>[47]</sup> and the regulation of immune function.<sup>[48]</sup> Decreased T lymphocytes and immune system function were found in leptin receptor lacking mouse model.<sup>[49]</sup> Leptin levels in

serum and synovial fluid were increased significantly in patients with OA.<sup>[50]</sup> Inflammation was weakened in leptin-lacking mouse model where symptoms of RA were also relieved. Exogenous leptin abdominal cavity injection in ob/ob mice can lead to different levels of oxidative stress and increased inflammation-related gene expression,<sup>[51]</sup> indicating that leptin might change the course of the arthritis by regulating immune response in the whole body or the articular cavity.<sup>[17]</sup> Synovitis is a very important process in early OA and there are now experiments targeting synovial angiogenesis early treatment to reduce symptoms of OA. Synovitis is often associated with mononuclear cell infiltrates. Leptin receptor was expressed on the surface of mononuclear cells, but whether leptin has a role in monocyte infiltration process is still not clear.<sup>[52]</sup>

### 2.4. The relationship between leptin, obesity, and OA

Obesity affects the body's material circulation and energy metabolism. Obesity is closely related to many diseases, such as diabetes, hypertension, and cardiovascular disease.<sup>[53,54]</sup> OA is no exception. On the one hand, joints of obese individuals bear more pressure than the normal population, which would lead to greater joint wear.<sup>[55]</sup> On the other hand, leptin levels in obese people were significantly increased may be due to leptin resistance. The increased leptin accelerated the process of inflammation of the joints.<sup>[56]</sup> Recent studies have found that obese mice caused by leptin knockout were not associated with a higher risk of OA, suggesting that leptin is an essential link in obese-mediated OA.<sup>[17]</sup> The possible role of leptin in the pathogenesis of OA was elaborated in Figure 2.

Leptin was mainly produced by adipose tissue in other organs, such as teeth, periodontal tissue,<sup>[57]</sup> stomach,<sup>[58]</sup> placenta,<sup>[59]</sup> osteoblast,<sup>[31]</sup> and joint cartilage.<sup>[25]</sup> In addition to the synovial membrane,<sup>[60]</sup> synthesis of leptin was also identified in osteoarthritic joint adipose tissue. Patellar fat pad is also a source of leptin.<sup>[61]</sup> Obese individuals often present with high leptin levels because of too much fat.<sup>[62]</sup> On the contrary, leptin resistance was not unusual in obese people,<sup>[63]</sup> thus leading to a negative feedback between leptin sensitivity and leptin concentration and then a continuously reduction in sensitivity of leptin and an increase in leptin secretion which might explain the phenomenon that older people often present with higher OA occurrence and lower leptin sensitivity.<sup>[64]</sup>

Because there is such a close link between obesity, leptin, and OA, it is crucial to study the effects of weight loss and exercise intervention on serum leptin levels to improve the symptoms of OA patients. Decreases in serum leptin may be one mechanism by which weight loss improves physical function and symptoms in OA patients.<sup>[65]</sup> Recent findings suggest that high-fat diet (HFD)-induced obesity can lead to the development of OA, whereas resveratrol may relieve OA pathology by reducing systemic inflammation and/or inhibiting TLR4 signaling in cartilage.<sup>[66]</sup> Therefore, resveratrol may be a promising treatment for obesity-related OA. For dietary structure, dietary fatty acid content plays an important role in the pathogenesis of OA after joint injury and supports the need for further study of dietary fatty acid supplements as a potential treatment for OA.<sup>[67]</sup>

### 3. Conclusion

OA is a common chronic disease characterized by local inflammation, cartilage damage, great pain, and joint movement disorder. Abnormal expression of leptin was shown in patients



Figure 2. The possible role of leptin in the pathogenesis of osteoarthritis.

with OA . Leptin was involved in obesity and various inflammatory processes. Its role in OA cannot be ignored. Yet the mechanism of leptin in the development of OA is still not clear. The interaction between leptin signaling pathway and other adipokines or inflammation factors and the potential immuno-logical function of Leptin in OA still remains to be elucidated.

#### Author contributions

Conceptualization: J. Zhang. Supervision: H. Yang. Validation: J. Zhang, Y. Sun. Writing – original draft: M. Yan. Writing – review & editing: Y. Sun.

### References

- Musumeci G, Castrogiovanni P, Trovato FM, et al. Physical activity ameliorates cartilage degeneration in a rat model of aging: a study on lubricin expression. Scand J Med Sci Sports 2015;25:222–30.
- [2] Musumeci G, Trovato FM, Pichler K, et al. Extra-virgin olive oil diet and mild physical activity prevent cartilage degeneration in an osteoarthritis model: an in vivo and in vitro study on lubricin expression. J Nutr Biochem 2013;24:2064.
- [3] Zhang YY, Proenca R, Maffei M, et al. Positional cloning of the mouse obese gene and its human homolog (vol. 372, Pg 425, 1994). Nature 1995;374:479–1479.

- [4] Friedman JM, Halaas JL. Leptin and the regulation of body weight in mammals. Nature 1998;395:763–70.
- [5] Carter S, Caron A, Richard D, et al. Role of leptin resistance in the development of obesity in older patients. Clin Interv Aging 2013;8:829–44.
- [6] Lee GH, Proenca R, Montez JM, et al. Abnormal splicing of the leptin receptor in diabetic mice. Nature 1996;379:632–5.
- [7] Heshka JT, Jones PJH. A role for dietary fat in leptin receptor, OB-Rb, function. Life Sci 2001;69:987–1003.
- [8] de Luis DA, Sagrado MG, Conde R, et al. Relation of visfatin to cardiovascular risk factors and adipocytokines in patients with impaired fasting glucose. Nutrition 2013;29:1300–3.
- [9] Golbidi S, Laher I. Exercise induced adipokine changes and the metabolic syndrome. J Diabetes Res 2014;2014:726861.
- [10] Hamrick MW, Herberg S, Arounleut P, et al. The adipokine leptin increases skeletal muscle mass and significantly alters skeletal muscle miRNA expression profile in aged mice. Biochem Biophys Res Commun 2010;400:379–83.
- [11] Trayhurn P. The biology of obesity. Proc Nutr Soc 2005;64:31-8.
- [12] Ahima RS, Prabakaran D, Mantzoros C, et al. Role of leptin in the neuroendocrine response to fasting. Nature 1996;382:250–2.
- [13] Vuolteenaho K, Koskinen A, Moilanen E. Leptin—a link between obesity and osteoarthritis. Applications for prevention and treatment. Basic Clin Pharmacol Toxicol 2014;114:103–8.
- [14] Miller GD, Nicklas BJ, Davis CC, et al. Is serum leptin related to physical function and is it modifiable through weight loss and exercise in older adults with knee osteoarthritis? Int J Obes Relat Metab Disord 2004;28:1383–90.
- [15] Vuolteenaho K, Koskinen A, Moilanen T, et al. Leptin levels are increased and its negative regulators, SOCS-3 and sOb-R are decreased in obese patients with osteoarthritis: a link between obesity and osteoarthritis. Ann Rheum Dis 2012;71:1912–3.

- [16] Scotece M, Conde J, Lopez V, et al. Adiponectin and leptin: new targets in inflammation. Basic Clin Pharmacol Toxicol 2014;114:97–102.
- [17] Griffin TM, Huebner JL, Kraus VB, et al. Extreme obesity due to impaired leptin signaling in mice does not cause knee osteoarthritis. Arthritis Rheum 2009;60:2935–44.
- [18] Iliopoulos D, Malizos KN, Tsezou A. Epigenetic regulation of leptin affects MMP-13 expression in osteoarthritic chondrocytes: possible molecular target for osteoarthritis therapeutic intervention. Ann Rheum Dis 2007;66:1616–21.
- [19] Qin J, Shi D, Dai J, et al. Association of the leptin gene with knee osteoarthritis susceptibility in a Han Chinese population: a case-control study. J Hum Genet 2010;55:704–6.
- [20] Ma X, Guo H, Hao S, et al. Association of single nucleotide polymorphisms (SNPs) in leptin receptor gene with knee osteoarthritis in the Ningxia Hui population. Hereditas (Beijing) 2013;35:359–64.
- [21] Argente J, Barrios V, Chowen JA, et al. Leptin plasma levels in healthy Spanish children and adolescents, children with obesity, and adolescents with anorexia nervosa and bulimia nervosa. J Pediatr 1997;131:833–8.
- [22] Otero M, Lago R, Gomez R, et al. Changes in plasma levels of fat-derived hormones adiponectin, leptin, resistin and visfatin in patients with rheumatoid arthritis. Ann Rheum Dis 2006;65:1198–201.
- [23] Seven A, Guzel S, Aslan M, et al. Serum and synovial fluid leptin levels and markers of inflammation in rheumatoid arthritis. Rheumatol Int 2009;29:743–7.
- [24] Ku JH, Lee CK, Joo BS, et al. Correlation of synovial fluid leptin concentrations with the severity of osteoarthritis. Clin Rheumatol 2009;28:1431–5.
- [25] Simopoulou T, Malizos KN, Iliopoulos D, et al. Differential expression of leptin and leptin's receptor isoform (Ob-Rb) mRNA between advanced and minimally affected osteoarthritic cartilage; effect on cartilage metabolism. Osteoarthritis Cartilage 2007;15:872–83.
- [26] de Boer TN, Spil WEV, Huisman AM, et al. Serum adipokines in osteoarthritis; comparison with controls and relationship with local parameters of synovial inflammation and cartilage damage. Osteoarthritis Cartilage 2012;20:846.
- [27] Steppan CM, Crawford DT, Chidsey-Frink KL, et al. Leptin is a potent stimulator of bone growth in ob/ob mice. Regul Pept 2000;92:73–8.
- [28] Yang W-H, Liu S-C, Tsai C-H, et al. Leptin induces IL-6 expression through OBRI receptor signaling pathway in human synovial fibroblasts. PLoS One 2013;8:
- [29] Kaneko S, Satoh T, Chiba J, et al. Interleukin-6 and interleukin-8 levels in serum and synovial fluid of patients with osteoarthritis. Cytokines Cell Mol Ther 2000;6:71–9.
- [30] Sui Y, Lee JH, DiMicco MA, et al. Mechanical injury potentiates proteoglycan catabolism induced by interleukin-6 with soluble interleukin-6 receptor and tumor necrosis factor alpha in immature bovine and adult human articular cartilage. Arthritis Rheum 2009; 60:2985–96.
- [31] Zanotti S, Canalis E. Interleukin 6 mediates selected effects of Notch in chondrocytes. Osteoarthritis Cartilage 2013;21:1766–73.
- [32] Troeberg L, Nagase H. Proteases involved in cartilage matrix degradation in osteoarthritis. Osteoarthritis Cartilage 2012;1824: 133–45.
- [33] Sasaki K, Hattori T, Fujisawa T, et al. Nitric oxide mediates interleukin-1-induced gene expression of matrix metalloproteinases and basic fibroblast growth factor in cultured rabbit articular chondrocytes. J Biochem 1998;123:431–9.
- [34] Bao JP, Chen WP, Feng J, et al. Leptin plays a catabolic role on articular cartilage. Mol Biol Rep 2010;37:3265–72.
- [35] Conde J, Scotece M, Lopez V, et al. Adiponectin and leptin induce VCAM-1 expression in human and murine chondrocytes. PLoS One 2012;7:
- [36] Schett G, Kiechl S, Bonora E, et al. Vascular cell adhesion molecule 1 as a predictor of severe osteoarthritis of the hip and knee joints. Arthritis Rheumatism 2009;60:2381–9.
- [37] Dumond H, Presle N, Terlain B, et al. Evidence for a key role of leptin in osteoarthritis. Arthritis Rheumatism 2003;48:3118–29.
- [38] Mutabaruka M-S, Aissa MA, Delalandre A, et al. Local leptin production in osteoarthritis subchondral osteoblasts may be responsible for their abnormal phenotypic expression. Arthritis Res Ther 2010;12:R20.
- [39] Loeser RF. Systemic and local regulation of articular cartilage metabolism: where does leptin fit in the puzzle? Arthritis Rheumatism 2003;48:3009–12.
- [40] Otero M, Lago R, Gomez R, et al. Towards a pro-inflammatory and immunomodulatory emerging role of leptin. Rheumatology 2006;45: 944–50.

- [41] Otero M, Lago R, Lago F, et al. Leptin, from fat to inflammation: old questions and new insights. FEBS Lett 2005;579:295–301.
- [42] Wang Q, Rozelle AL, Lepus CM, et al. Identification of a central role for complement in osteoarthritis. Nat Med 2011;17:1674–9.
- [43] Smith MD, Triantafillou S, Parker A, et al. Synovial membrane inflammation and cytokine production in patients with early osteoarthritis. J Rheumatol 1997;24:365–71.
- [44] Goldring MB, Birkhead J, Sandell LJ, et al. Interleukin-1 suppresses expression of cartilage-specific type-II and type-IX collagens and increases type-I and type-III collagens in human chondrocytes. J Clin Invest 1988;82:2026–37.
- [45] Benito MJ, Veale DJ, Fitzgerald O, et al. Synovial tissue inflammation in early and late osteoarthritis. Ann Rheum Dis 2005;64:1263–7.
- [46] Shiokawa S, Matsumoto N, Nishimura J. Clonal analysis of B cells in the osteoarthritis synovium. Ann Rheum Dis 2001;60:802–5.
- [47] Lord GM, Matarese G, Howard LK, et al. Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. Nature 1998;394:897–901.
- [48] Matarese G, Moschos S, Mantzoros CS. Leptin in immunology. J Immunol 2005;174:3137–42.
- [49] Kimura M, Tanaka S, Isoda F, et al. T lymphopenia in obese diabetic (db/ db) mice is non-selective and thymus independent. Life Sci 1998; 62:1243–50.
- [50] Busso N, So A, Chobaz-Peclat V, et al. Leptin signaling deficiency impairs humoral and cellular immune responses and attenuates experimental arthritis. J Immunol 2002;168:875–82.
- [51] Sainz N, Rodriguez A, Catalan V, et al. Leptin administration downregulates the increased expression levels of genes related to oxidative stress and inflammation in the skeletal muscle of ob/ob mice. Mediators Inflamm 2010;2010:784343.
- [52] Zarkesh-Esfahani H, Pockley G, Metcalfe RA, et al. High-dose leptin activates human leukocytes via receptor expression on monocytes. J Immunol 2001;167:4593–9.
- [53] Hui E, Xu A, Yang HB, et al. Obesity as the common soil of nonalcoholic fatty liver disease and diabetes: role of adipokines. J Diabetes Investig 2013;4:413–25.
- [54] Boden G, Salehi S. Why does obesity increase the risk for cardiovascular disease? Curr Pharm Des 2013;19:5678–83.
- [55] Terlain B, Presle N, Pottie P, et al. Leptin: a link between obesity and osteoarthritis? Bull Acad Natl Med 2006;190:1421–35.
- [56] Flier JS. Hormone resistance in diabetes and obesity: insulin, leptin, and FGF21. Yale J Biol Med 2012;85:405–14.
- [57] Li W, Zhu W, Hou J, et al. Leptin and its receptor expression in dental and periodontal tissues of primates. Cell Tissue Res 2014;355: 181–8.
- [58] Cammisotto PG, Bendayan M. Leptin secretion by white adipose tissue and gastric mucosa. Histol Histopathol 2007;22:199–210.
- [59] Hoggard N, Hunter L, Duncan JS, et al. Leptin and leptin receptor mRNA and protein expression in the murine fetus and placenta. Proc Natl Acad Sci USA 1997;94:11073–8.
- [60] Conde J, Scotece M, Lopez V, et al. Differential expression of adipokines in infrapatellar fat pad (IPFP) and synovium of osteoarthritis patients and healthy individuals. Ann Rheum Dis 2014;73:631–3.
- [61] Clockaerts S, Bastiaansen-Jenniskens YM, Feijt C, et al. Cytokine production by infrapatellar fat pad can be stimulated by interleukin 1 beta and inhibited by peroxisome proliferator activated receptor alpha agonist. Ann Rheum Dis 2012;71:1012–8.
- [62] Lonnqvist F, Arner P, Nordfors L, et al. Overexpression of the obese (ob) gene in adipose-tissue of human obese subjects. Nat Med 1995;1: 950–3.
- [63] Berbari NF, Pasek RC, Malarkey EB, et al. Leptin resistance is a secondary consequence of the obesity in ciliopathy mutant mice. Proc Natl Acad Sci USA 2013;110:7796–801.
- [64] Sanchez-Rodriguez M, Garcia-Sanchez A, Retana-Ugalde R, et al. Serum leptin levels and blood pressure in the overweight elderly. Arch Med Res 2000;31:425–8.
- [65] Miller GD, Jenks MZ, Vendela M, et al. Influence of weight loss, body composition and lifestyle behaviors on plasma adipokines: a randomized weight loss trial in older men and women with symptomatic knee osteoarthritis. J Obes 2012;2012:708505.
- [66] Jiang M, Li X, Yu X, et al. Oral administration of resveratrol alleviates osteoarthritis pathology in C57BL/6J mice model induced by a high-fat diet. Mediators of Inflamm 2017;2017:7659023.
- [67] Wu CL, Jain D, Mcneill JN, et al. Dietary fatty acid content regulates wound repair and the pathogenesis of osteoarthritis following joint injury. Ann Rheum Dis 2015;74:2076–83.