

REVIEW ARTICLES

e-ISSN 1643-3750 © Med Sci Monit, 2019; 25: 3329-3335 DOI: 10.12659/MSM.914027

Received:2018.11.07Accepted:2019.01.28Published:2019.05.06

Authors' Contribution: Study Design A Data Collection B Statistical Analysis C Data Interpretation D Manuscript Preparation E Literature Search F Funds Collection G

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Source of support:

Exosomes in Pathogenesis, Diagnosis, and Treatment of Alzheimer's Disease

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Alzheimer's disease (AD) is a neurodegenerative disorder characterized by the accumulation of β -amyloid peptide 1-42 and phosphorylation of tau protein in the brain. Thus far, the transfer mechanism of these cytotoxic proteins between nerve cells remains unclear. Recent studies have shown that nanoscale extracellular vesicles (exosomes) originating from cells may play important roles in this transfer process. In addition, several genetic materials and proteins are also involved in intercellular communication by the secretion of the exosomes. That proposes novel avenues for early diagnosis and biological treatment in AD, based on exosome detection and intervention. In this review, exosome-related pathways of cytotoxic protein intercellular transfer in AD, and the effect of membrane proteins on exosomes targeting cells are first introduced. The advances in exosomerelated biomarker detection in AD are summarized. Finally, the advantages and challenges of reducing cytotoxic protein accumulation via exosomal intervention for AD treatment are discussed. It is envisaged that future research in exosomes may well provide new insights into the pathogenesis, diagnosis, and treatment of AD.

MeSH Keywords: Alzheimer Disease • Biological Markers • Exosomes

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Background

Alzheimer's disease (AD) is a late-onset neurodegenerative disorder involving memory and other cognitive impairments. The initiating mechanism of its onset is related to poisoning, metabolism, genetic factors, and so on. Recent evidences have demonstrated that vascular, inflammatory and degenerative pathways critically interact and contribute to its pathology [1-3]. However, no hypothesis has yet managed to account for all symptoms and pathogenic mechanism in AD, it is commonly accepted that the main pathological features of AD involve amyloid β -peptide 1–42 (A β 1–42) protein accumulation and phosphorylated tau protein (p-tau)-induced apoptosis in nerve cells. AB1-42 toxic protein accumulation may not be easily degraded and eliminated by ubiquitin or the autophagy pathway [4]. Moreover, abnormal p-tau tends to lose its function of maintaining microtubule stability [5,6]. The consequent microtubule assembly leads to microtubule associated axonal transport barriers, which interfere with the formation of lysosomes and contribute to the aggregation of toxic proteins in axons [7].

In another hand, AD familial studies have suggested that toxic proteins are associated with multiple specific gene mutations, such as the amyloid precursor protein gene (*APP*) and presenilin (*PS-1, PS-2*), which appear to facilitate A β 1-42 assembly by promoting A β production [8–10]. However, AD genetic studies have indicated that several genes, such as *ApoE4* and *ApoJ*, affect the degradation of A β 1–42 protein [11–13]. These results have provided the theoretical basis for diagnostic genetic factor-based biomarker discovery and gene therapy for AD.

In recent studies, the presence of $A\beta1-42$ protein and p-tau in the cerebrospinal fluid (CSF) of patients with AD was verified [14–16], suggesting that toxic proteins may penetrate into the extracellular fluid through nerve cell secretion. In addition, adding toxic proteins to cell culture medium or CSF induced pathological and behavioral changes both in nerve cells and animal models analogous to AD [17–19]. These results suggest that the transmission of toxic proteins between cells may be an important pathway for AD progression after the initiation of AD pathogenesis. However, the transfer mechanism of these toxic proteins between the cells and extracellular fluid remains unclear.

Exosomes, a kind of nanoscale vesicle widely present in various cells and the extracellular fluid, may carry small molecular genetic material and proteins involved in information transfer between cells [20,21]. This vesicle transport may relate to the toxic protein generation, transportation, and degradation process in AD [22–24]. This might provide a new line of research for revealing the disease's progressive pathogenesis and improving the current poor treatment status of AD. In this review, we focus on discussing the roles of exosomes in toxic protein

production, transportation, and degradation, with a mind to providing new viewpoints to the pathogenesis, model establishment, and biological therapy of AD.

Biological Characteristics of Exosomes

Exosomes are a kind of single lipid membrane vesicles secreted by cells, with diameters ranging from 40 to 100 nm, and are widely distributed in body fluids [25]. Many kinds of cells such as neurons, glial, stem, and tumor cells may release exosomes to the extracellular fluid through direct fusion of the multivesicular bodies with the plasma membrane [26]. However, it was only in the last decade that exosomes were considered to play an important role in intercellular communication based on genetic content transport. Exosomes in the extracellular fluid can bind to target cells through membrane receptors, directly fuse with the plasma membrane, or be endocytosed to release their contents [20]. This process was verified by co-culture of R18-labeled exosomes and PKH-67-labeled cells; the membrane fusion process was observed under the fluorescence microscope [27]. Moreover, exosome microRNA transfer to target cells using fluorescent labeling has also been reported [28]. These findings confirmed the extracellular transport mechanism through exosomes.

A database of exosomes content was roughly established by large-scale analysis of various tissues, and there were11261 protein entries, 2375 mRNA entries and 764 miRNA entries in exosomes [29]. In addition, modified exosomes successfully delivered exogenous siRNA into the brain tissue of mice after intravenous administration, indicating that modified exosomes may cross biological barriers, such as the blood-brain barrier (BBB) [30]. The technology of exosomes content detection and exogenous genetic material modify may deliver exogenous hereditary materials to acting sites on nerve cells and create new avenues for disease biomarkers detection and gene therapy for neurological diseases such as AD.

Furthermore, the issue of how exosomes function on target cells with high selectivity remains a challenge for targeted treatment. In several studies, the rabies virus glycoprotein (RVG) peptide was bound to the extra-exosomal N terminus of murine Lamp2b [30,31], a protein found abundantly in exosomal membranes [32], in order to selectively act on neural cells. It is noted that modified exosomes have successfully delivered exogenous interfering RNA exclusively into neural cells *in vivo* [30]. At the same time, the exosomes of homologous cells have immunogenicity, which can effectively avoid inducing immune response. In addition, through the fluorescent labeling of exosomes and animal imaging technology, the acting sites of exosomes can be dynamically tracked, which makes it possible to provide effective technical support for accurate gene therapy.

Exosomes in the Pathogenesis of AD

Exosomes are believed to be involved in the spread of many neurodegenerative diseases, including Parkinson's disease (PD) and AD [33]. After the occurrence of AD initiating factors, Aβ may be secreted from cells in association with exosomes and be released to the extracellular space. However, most tau proteins released into extracellular fluids are cut-off mid-region tau [34,35], lacking tail ends that cause tau aggregation. Therefore, Full-length tau in exosomes via endocytosis or axonal transmission may be the main vector of abnormal tau transmission [36]. Polanco et al. established a simple model of neural circuit with hippocampal neurons. They observed the exosomes spreading in the interconnected neurons and found the exosomes can spread the $A\beta$ and tau protein by endosomal pathway and axonal transport [37]. Moreover, in the plaques of patients with AD, enriched accumulation was found in A β and specific exosomal proteins, such as Alix, indicating the A β in the plagues may constitute the contents of the exosomes secreted from neural cells in AD [38]. In addition, research on tau protein has reported that the concentration of abnormal p-tau in exosomes isolated from the CSF of patients with AD at the mild/severe stage (Braak stages 3-6) was significantly higher than that from patients with AD at the early stage (Braak stages 0-2). This phenomenon indicated that exosome-mediated secretion of p-tau might play a significant role in the abnormal processing of tau and in the increase of CSF tau during early AD [39]. Furthermore, hypophosphorylated tau protein-rich exosomes extracted from CSF of AD patients can promote the aggregation of tau protein in neurons and microglia [40]. Exosomes play a similar role in other neurodegenerative diseases. For instance, α -synuclein was released from exosomes to the extracellular fluid, based on a calcium dependent mechanism, which aggravates and propagates PD-related pathological features [41]. Moreover, exosomes may wrap prion proteins and be secreted from the infected cells to the extracellular fluid, and deliver prion proteins to the target cells by membrane fusion [42]. This is similar to the pathological mechanism in AD and PD.

In addition, exosomes are also involved nerve cell injury in AD. The mutation of the *PS* gene could downregulate cystatin C in exosomes, a protein targeting the classical secretory pathway by its signal peptide sequence and providing a neuroprotective function in AD [43], which induced the decrease of soluble APP and increase of A β 1–42 [44]. In contrast, lysosomes play an important role in neurodegenerative diseases by degrading unnecessary proteins [45]. Lysosomal dysfunction promotes exosome secretion from nerve cells in neurodegenerative diseases and aggravates extracellular toxic protein accumulation [41].

Not only the exosomes of neurons aggravate the progress of AD, but also the exosomes in astrocytic apoptosis involved in AD pathogenesis. In the central nervous system (CNS), astrocytes are indispensable for the support of neurons. However, astrocytes may also activate inflammatory and proapoptotic signaling pathways if they are triggered to migrate and proliferate [46-48]. Astrocytic reactivation has been observed in many neurodegenerative diseases, including AD [49,50]. Glial apoptosis has been reported to be correlated with the number of senile plaques, and caspases activation has been suggested to contribute to astrocytic damage [51,52]. Bieberich et al. found that expression of prostate apoptosis response 4 (PAR-4) and the simultaneous elevation of ceramide can induce apoptosis in neural progenitor cells [53,54], and they verified that $A\beta$ could induce apoptosis in astrocytes in vitro through a PAR-4dependent mechanism. They also suggested that the mechanism of astrocytic apoptosis in AD may be associated with the secretion of PAR-4/ceramide-containing exosomes, which could induce cell death [55]. Furthermore, they found neutral sphingomyelinase 2 (nSMase2) deficient astrocytes were protected from AB induced apoptosis, and nSMase2 could modulate the secretion of exosomes [24].

Although exosomes are involved in nerve cell injury in AD, in the nervous system they are active messengers, protecting neurons from oxidative stress [56]. Similar neuroprotective effects are found in neurodegenerative diseases. In AD research, it was found that exosomes both played a role in toxic protein spread and acted on A β to reduce injury in the nervous system. In the extracellular fluid, exosomes derived from N2a cells may abrogate the synaptic-plasticity-disrupting activity of both synthetic and AD brain-derived AB, and rescue long-term potentiation from Aβ-mediated impairment in vivo. These effects are mainly due to the sequestration of A β oligomers via exosomal surface proteins, such as the p75 neurotrophin receptor (PrPc), which has high affinity to A β oligomers [57]. Furthermore, in the CNS, microglia around the neurons can remove damaged structures, including exosomes-delivered AB. Enhancement of exosome secretion could efficiently reduce extracellular levels of A β by the glycosphingolipids (GSLs) on its surface [24].

Exosomes as Biomarkers in the Diagnosis of AD

Biomarkers are necessary for improving diagnostic sensitivity and specificity and for monitoring the biological activities of AD, prior to apparent clinical symptom manifestation. Toxic proteins in exosomes can be detected in the early stage of the disease, and exosomes are carriers of specific toxic proteins between cells and the extracellular fluid in a variety of neurodegenerative diseases. Therefore, the detection of specific toxic proteins in peripheral exosomes from body fluids is expected to become a biomarker-based method in the diagnosis of early stage AD and other neurodegenerative diseases. Recently, the combination of CSF p-tau and CSF A_{β1}-42 has widely been studied as a diagnostic biomarker capable of distinguishing AD from other dementia in the early stages. Comparing the AD, dementia with Lewy bodies, and vascular dementia, concentration of CSF p-tau, it was highest in patients with AD [14,15,58]. Although CSF p-tau detection can help distinguish AD from other types of dementia, comparing p-tau concentrations in exosomes is more helpful for identifying the degree and stage of AD by combining symptoms based on the positive correlation between the amount of p-tau in CSF exosomes and the severity of AD. Moreover, as p-tau can be found in CSF exosomes in the early stage, it is helpful for the early diagnosis of AD [39]. Recent study has found that the full-length tau to mid-region tau in CSF and plasma exosomes of AD patients was much higher than free solution, while the CSF and plasma exosomes of healthy people did not contain full-length tau [36]. Therefore, CSF exosomes may be used as a biomarker in the diagnosis of AD. But at present, p-tau can also be identified in a small amount of CSF [59]. Detection of specific tau in CSF exosomes for diagnosis of AD does not show better application prospects.

There have been several studies on the detection of AB1-42 in the CSF and plasma of patients with AD at the early stage [60-63]. Current investigations suggest that Aβ accumulation could bring forward the diagnosis of AD by more than 15 years [64]. However, the detection rate of A β 1–42 in the CSF in early stage AD was only 40%-50%, and the concentrations remained stable over a period of 12 months [65]. Furthermore, the accumulation of A β 1–42 in the plasma was dissimilar to that in the CSF [66], and there was no variation in plasma A β over time in patients with AD [67]. Therefore, plasma A β cannot be utilized as an AD biomarker in previous viewpoint. As CSF A β is mainly derived from the exosomes secreted by lesion cells, detection of A β 1–42 in CSF exosomes is prospected to improve the diagnostic sensitivity of AD. In the study of combined detection of CSF p-tau and Aβ1-42, sensitivity and specificity were higher than 86% [68]. Other studies have reported that the combined detection of CSF A β 1–42 and p-tau might help assist early diagnosis for AD 10 years prior to clinical onset and differentiate early stage AD from frontotemporal lobar degeneration [69,70]. Therefore, the combined detection of these two potential biomarkers in CSF exosomes may be more valuable for the early diagnosis of AD.

In recent years, advantages of miRNAs in disease diagnosis have attracted research attention to miRNA detection. miRNA could be a biomarker for AD diagnosis. As miRNA secreted by the host cells is rich in exosomes, there have been several studies in which exosomes were extracted from different types of body fluids for miRNA micro-quantitative determination. In a case study, five miRNAs were found to be significantly different in CSF exosomes from the normal control group (two upregulated, three downregulated) [71]. More miRNAs were found to be abnormal in AD plasma exosomes (19 upregulated, 33 downregulated) [72,73]. In the downregulated miRNAs, mir-342-3p was also significantly reduced in the blood of patients with AD [74]. Other microRNAs, such as miR-9-5p and miR-598, could be detected rich in the exosomes delivered from CSF of AD patients [75]. The comparison of these results could provide accurate early diagnosis for AD.

Exosomes in the Treatment of AD

In the nervous system, exosomes are active messengers, protecting neurons from oxidative stress [56]. As mentioned before, exogenous exosomes can assist in the degradation of A β 1–42 in AD and other neurodegenerative diseases. Hao et al. co-cultured injured cortical neurons with human adipose-derived mesenchymal stem cells (ADSCs) using a semi-porous membrane, and the results demonstrated that AMSCs-conditioned medium, enriched with exosomes, mediates direct neuroprotection by inhibiting neuronal cell apoptosis, promoting nerve regeneration and repair, and restoring bioenergy following energy depletion caused by glutamate excitotoxicity [76]. In another study, exosomes were extracted from mesenchymal stromal cell (MSC)-conditioned medium and injected into a rat stroke model; it was found that exosomes could reduce nerve cell injury [77]. These results suggest that exosomes originating from ADSCs, and other exogenous stem cells, could be used for the treatment of nervous system diseases. In the exosomes isolated from ADSCs, the content of Neprilysin (NEP) was significantly higher than that of nerve cells [78], and this neutral endopeptidase was related to the degradation of A β [79,80]. The exosomes isolated from rat primary neurons contained cystatin C, which is also involved in the degradation of A β [81–83]. The physiological high concentration of cystatin C in the CSF and its proliferative effect on neural rat stem cells suggested it could exert a trophic function in the brain [84]. Therefore, exogenous exosomes are considered to be potential agents for the treatment of AD.

Owing to their RNA transport capacity, stable presence in body fluids, and capability of crossing the BBB, exosomes can be used as carriers to deliver nucleic acid fragments, such as miRNA and siRNA, for the treatment of AD. In 2011, a study first reported successful treatment in AD mice by using exosomes carrying siRNA [30]. Purified RVG-targeted exosomes were loaded with exogenous siRNA by electroporation. Significantly decreased protein expression and A β deposition in the AD mouse brain were confirmed. This provided evidence for the validity of exosome therapy for neurodegenerative diseases through siRNA delivery. In another study, delivery of miR-124a through exosomes enhanced the expression of excitatory amino acid transporter 2 (GLT1) on the surface of astrocytes, for modulating

synapse activity and improving glutamate uptake [85]. This method is expected to alleviate neuronal apoptosis in AD.

Compared to traditional candidate vectors for gene therapy, such as viruses, polyethylenimine nanoparticles, and liposomes, exosomes show greater advantages in therapeutic effect, targeting ability, low immune response, and safety [86]. Exosomes are naturally secreted components from body cells, and are widely present in the extracellular fluid [25]. Exosomes for therapeutic treatment can be obtained from the culture medium of MSCs or self-derived dendritic cells with reduced cell immunogenicity [30,87,88]. Exosomes can prevent the activation of the immune system and phagocytosis by mononuclear macrophages due to the expression of CD55 and CD59 [89]. Therefore, exosomes may be safely applied in AD therapy.

Conclusions

Exosomes have been confirmed as an important agent in AD pathogenesis progression. Elucidating the underlying mechanisms involved will promote AD model establishment, early stage diagnosis, and gene therapy. It may also provide insights into potential therapies for other neurodegenerative diseases.

As small genetic fragments and toxic proteins could be carried by exosomes and transported between cells and extracellular fluids, which might also be the mechanism of the slow progress of neurodegenerative diseases, it can be assumed that in the co-culture system of nerve cells and isolated exosomes from plasma or CSF of patients with AD, nerve cells may be induced into AD-like injury cells. Furthermore, applying specific amounts

References:

- Lattanzi S, Brigo F, Vernieri F, Silvestrini M: Visit-to-visit variability in blood pressure and Alzheimer's disease. J Clin Hypertens (Greenwich), 2018; 20: 918–24
- Lattanzi S, Carbonari L, Pagliariccio G et al: Neurocognitive functioning and cerebrovascular reactivity after carotid endarterectomy. Neurology, 2018; 90: e307–15
- McKenzie JA, Spielman LJ, Pointer CB et al: Neuroinflammation as a common mechanism associated with the modifiable risk factors for Alzheimer's and Parkinson's diseases. Curr Aging Sci, 2017; 10: 158–76
- 4. Ling D, Salvaterra PM: A central role for autophagy in Alzheimer-type neurodegeneration. Autophagy, 2009; 5: 738–40
- Alonso Adel C, Mederlyova A, Novak M et al: Promotion of hyperphosphorylation by frontotemporal dementia tau mutations. J Biol Chem, 2004; 279: 34873–81
- Li B, Chohan MO, Grundke-Iqbal I, Iqbal K: Disruption of microtubule network by Alzheimer abnormally hyperphosphorylated tau. Acta Neuropathol, 2007; 113: 501–11
- 7. Dixit R, Ross JL, Goldman YE, Holzbaur EL: Differential regulation of dynein and kinesin motor proteins by tau. Science, 2008; 319: 1086–89
- Hardy J, Selkoe DJ: The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. Science, 2002; 297: 353–56
- 9. Tanzi RE: The genetics of Alzheimer disease. Cold Spring Harb Perspect Med, 2012; 2: pii: a006296

of exosomes from AD to the CSF may induce AD symptoms in animal models. Intracerebral aggregation of alpha-synuclein can be induced by injecting exosomes extracted from brain tissues of dementia with Lewy bodies patients in animal model brains [90]; in another study, exosomes containing pathogenic proteins from the conditioned medium of HEK293-APP Swe/Ind cells showed high neurotoxicity to the hippocampal dentate gyrus region *in vivo* [91]. These results would promote a new approach for establishing AD and other neurodegenerative disease models. Compared to the existing AD models, exosome-based cell and animal models more closely resemble the actual pathogenesis.

In the early stage of AD, the combined detection of p-tau and A β 1–42 can effectively enhance diagnostic sensitivity and specificity, as these two potential biomarkers are more likely delivered to extracellular fluids by exosomes. With the development of clinical laboratory technology, toxic proteins can be detected in a small amount of CSF, which offsets the advantages of CSF exosomes. Detection of toxic proteins and specific microRNA in plasma exosomes will improve the convenience of early diagnosis of AD.

Finally, exosomes will become a new hot spot in molecular treatment of AD due to their security, selectivity to target cells, and their capacity for small molecule drug delivery. In addition, exosome diffusion is not restricted by biological barriers, providing more options for exosome administration.

Conflicts of interest

None.

- Tysoe C, Whittaker J, Xuereb J et al: A presenilin-1 truncating mutation is present in two cases with autopsy-confirmed early-onset Alzheimer disease. Am J Hum Genet, 1998; 62: 70–76
- 11. Evans DA, Beckett LA, Field TS et al: Apolipoprotein E epsilon4 and incidence of Alzheimer disease in a community population of older persons. JAMA, 1997; 277: 822–24
- 12. Egensperger R, Kosel S, von Eitzen U, Graeber MB: Microglial activation in Alzheimer disease: Association with APOE genotype. Brain Pathol, 1998; 8: 439–47
- Harold D, Abraham R, Hollingworth P et al: Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. Nat Genet, 2009; 41: 1088–93
- Arai H, Satoh-Nakagawa T, Higuchi M et al: No increase in cerebrospinal fluid tau protein levels in patients with vascular dementia. Neurosci Lett, 1998; 256: 174–76
- 15. Parnetti L, Lanari A, Amici S et al: Phospho-Tau International Study Group: CSF phosphorylated tau is a possible marker for discriminating Alzheimer's disease from dementia with Lewy bodies. Phospho-Tau International Study Group. Neurol Sci, 2001; 22: 77–78
- Maddalena AS, Papassotiropoulos A, Gonzalez-Agosti C et al: Cerebrospinal fluid profile of amyloid beta peptides in patients with Alzheimer's disease determined by protein biochip technology. Neurodegener Dis, 2004; 1: 231–35

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- 17. Tian X, Wang J, Dai J et al: Hyperbaric oxygen and *Ginkgo Biloba* extract inhibit Abeta25–35-induced toxicity and oxidative stress *in vivo*: A potential role in Alzheimer's disease. Int J Neurosci, 2012; 122: 563–69
- Gimenez-Llort L, Blazquez G, Canete T et al: Modeling behavioral and neuronal symptoms of Alzheimer's disease in mice: A role for intraneuronal amyloid. Neurosci Biobehav Rev, 2007; 31: 125–47
- 19. Lord A, Kalimo H, Eckman C et al: The Arctic Alzheimer mutation facilitates early intraneuronal Abeta aggregation and senile plaque formation in transgenic mice. Neurobiol Aging, 2006; 27: 67–77
- Simons M, Raposo G: Exosomes vesicular carriers for intercellular communication. Curr Opin Cell Biol, 2009; 21: 575–81
- 21. Faure J, Lachenal G, Court M et al: Exosomes are released by cultured cortical neurones. Mol Cell Neurosci, 2006; 31: 642–48
- 22. Sharples RA, Vella LJ, Nisbet RM et al: Inhibition of gamma-secretase causes increased secretion of amyloid precursor protein C-terminal fragments in association with exosomes. FASEB J, 2008; 22: 1469–78
- Bulloj A, Leal MC, Xu H et al: Insulin-degrading enzyme sorting in exosomes: A secretory pathway for a key brain amyloid-beta degrading protease. J Alzheimers Dis, 2010; 19: 79–95
- 24. Yuyama K, Sun H, Mitsutake S, Igarashi Y: Sphingolipid-modulated exosome secretion promotes clearance of amyloid-beta by microglia. J Biol Chem, 2012; 287: 10977–89
- 25. Lasser C, Alikhani VS, Ekstrom K et al: Human saliva, plasma and breast milk exosomes contain RNA: Uptake by macrophages. J Transl Med, 2011; 9: 9
- 26. van Niel G, Porto-Carreiro I, Simoes S, Raposo G: Exosomes: A common pathway for a specialized function. J Biochem, 2006; 140: 13–21
- Parolini I, Federici C, Raggi C et al: Microenvironmental pH is a key factor for exosome traffic in tumor cells. J Biol Chem, 2009; 284: 34211–22
- Montecalvo A, Larregina AT, Shufesky WJ et al: Mechanism of transfer of functional microRNAs between mouse dendritic cells via exosomes. Blood, 2012; 119: 756–66
- 29. Mathivanan S, Fahner CJ, Reid GE, Simpson RJ: ExoCarta 2012: Database of exosomal proteins, RNA and lipids. Nucleic Acids Res, 2012; 40: D1241–44
- Alvarez-Erviti L, Seow Y, Yin H et al: Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. Nat Biotechnol, 2011; 29: 341–45
- Cooper JM, Wiklander PB, Nordin JZ et al: Systemic exosomal siRNA delivery reduced alpha-synuclein aggregates in brains of transgenic mice. Mov Disord, 2014; 29: 1476–85
- Simhadri VR, Reiners KS, Hansen HP et al: Dendritic cells release HLA-Bassociated transcript-3 positive exosomes to regulate natural killer function. PLoS One, 2008; 3: e3377
- Emmanouilidou E, Melachroinou K, Roumeliotis T et al: Cell-produced alpha-synuclein is secreted in a calcium-dependent manner by exosomes and impacts neuronal survival. J Neurosci, 2010; 30: 6838–51
- 34. Kanmert D, Cantlon A, Muratore CR et al: C-terminally truncated forms of Tau, but not full-length Tau or its C-terminal fragments, are released from neurons independently of cell death. J Neurosci, 2015; 35: 10851–65
- 35. Wagshal D, Sankaranarayanan S, Guss V et al: Divergent CSF tau alterations in two common tauopathies: Alzheimer's disease and progressive supranuclear palsy. J Neurol Neurosurg Psychiatry, 2015; 86: 244–50
- 36. Guix FX, Corbett GT, Cha DJ et al: Detection of aggregation-competent Tau in neuron-derived extracellular vesicles. Int J Mol Sci, 2018; 19: pii: E663
- 37. Polanco JC, Li C, Durisic N et al: Exosomes taken up by neurons hijack the endosomal pathway to spread to interconnected neurons. Acta Neuropathol Commun, 2018; 6: 10
- Rajendran L, Honsho M, Zahn TR et al: Alzheimer's disease beta-amyloid peptides are released in association with exosomes. Proc Natl Acad Sci USA, 2006; 103: 11172–77
- Saman S, Kim W, Raya M et al: Exosome-associated tau is secreted in tauopathy models and is selectively phosphorylated in cerebrospinal fluid in early Alzheimer disease. J Biol Chem, 2012; 287: 3842–49
- 40. Wang Y, Balaji V, Kaniyappan S et al: The release and trans-synaptic transmission of Tau via exosomes. Mol Neurodegener, 2017; 12: 5
- 41. Alvarez-Erviti L, Seow Y, Schapira AH et al: Lysosomal dysfunction increases exosome-mediated alpha-synuclein release and transmission. Neurobiol Dis, 2011; 42: 360–67
- 42. Fevrier B, Vilette D, Archer F et al: Cells release prions in association with exosomes. Proc Natl Acad Sci USA, 2004; 101: 9683–88

- Sundelof J, Arnlov J, Ingelsson E et al: Serum cystatin C and the risk of Alzheimer disease in elderly men. Neurology, 2008; 71: 1072–79
- 44. Ghidoni R, Paterlini A, Albertini V et al: Cystatin C is released in association with exosomes: A new tool of neuronal communication which is unbalanced in Alzheimer's disease. Neurobiol Aging, 2011; 32: 1435–42
- Paxinou E, Chen Q, Weisse M et al: Induction of alpha-synuclein aggregation by intracellular nitrative insult. J Neurosci, 2001; 21: 8053–61
- Agostinho P, Cunha RA, Oliveira C: Neuroinflammation, oxidative stress and the pathogenesis of Alzheimer's disease. Curr Pharm Des, 2010; 16: 2766–78
- Li C, Zhao R, Gao K et al: Astrocytes: Implications for neuroinflammatory pathogenesis of Alzheimer's disease. Curr Alzheimer Res, 2011; 8: 67–80
- Fuller S, Steele M, Munch G: Activated astroglia during chronic inflammation in Alzheimer's disease – do they neglect their neurosupportive roles? Mutat Res, 2010; 690: 40–49
- Meda L, Baron P, Scarlato G: Glial activation in Alzheimer's disease: The role of Abeta and its associated proteins. Neurobiol Aging, 2001; 22: 885–93
- Croisier E, Graeber MB: Glial degeneration and reactive gliosis in alphasynucleinopathies: The emerging concept of primary gliodegeneration. Acta Neuropathol, 2006; 112: 517–30
- 51. Kobayashi K, Hayashi M, Nakano H et al: Apoptosis of astrocytes with enhanced lysosomal activity and oligodendrocytes in white matter lesions in Alzheimer's disease. Neuropathol Appl Neurobiol, 2002; 28: 238–51
- Mouser PE, Head E, Ha KH, Rohn TT: Caspase-mediated cleavage of glial fibrillary acidic protein within degenerating astrocytes of the Alzheimer's disease brain. Am J Pathol, 2006; 168: 936–46
- Bieberich E, MacKinnon S, Silva J et al: Regulation of cell death in mitotic neural progenitor cells by asymmetric distribution of prostate apoptosis response 4 (PAR-4) and simultaneous elevation of endogenous ceramide. J Cell Biol, 2003; 162: 469–79
- Bieberich E, Silva J, Wang G et al: Selective apoptosis of pluripotent mouse and human stem cells by novel ceramide analogues prevents teratoma formation and enriches for neural precursors in ES cell-derived neural transplants. J Cell Biol, 2004; 167: 723–34
- 55. Wang G, Dinkins M, He Q et al: Astrocytes secrete exosomes enriched with proapoptotic ceramide and prostate apoptosis response 4 (PAR-4): Potential mechanism of apoptosis induction in Alzheimer disease (AD). J Biol Chem, 2012; 287: 21384–95
- 56. Wang S, Cesca F, Loers G et al: Synapsin I is an oligomannose-carrying glycoprotein, acts as an oligomannose-binding lectin, and promotes neurite outgrowth and neuronal survival when released via glia-derived exosomes. J Neurosci, 2011; 31: 7275–90
- An K, Klyubin I, Kim Y et al: Exosomes neutralize synaptic-plasticity-disrupting activity of Abeta assemblies *in vivo*. Mol Brain, 2013; 6: 47
- Vanderstichele H, De Vreese K, Blennow K et al: Analytical performance and clinical utility of the INNOTEST PHOSPHO-TAU181P assay for discrimination between Alzheimer's disease and dementia with Lewy bodies. Clin Chem Lab Med, 2006; 44: 1472–80
- Olsson B, Lautner R, Andreasson U et al: CSF and blood biomarkers for the diagnosis of Alzheimer's disease: A systematic review and meta-analysis. Lancet Neurol, 2016; 15: 673–84
- Lewczuk P, Kornhuber J, Vanmechelen E et al: Amyloid beta peptides in plasma in early diagnosis of Alzheimer's disease: A multicenter study with multiplexing. Exp Neurol, 2010; 223: 366–70
- 61. Lewczuk P, Esselmann H, Groemer TW et al: Amyloid beta peptides in cerebrospinal fluid as profiled with surface enhanced laser desorption/ionization time-of-flight mass spectrometry: Evidence of novel biomarkers in Alzheimer's disease. Biol Psychiatry, 2004; 55: 524–30
- 62. Takeda S, Sato N, Rakugi H, Morishita R: Plasma beta-amyloid as potential biomarker of Alzheimer disease: Possibility of diagnostic tool for Alzheimer disease. Mol Biosyst, 2010; 6: 1760–66
- Perneczky R, Tsolakidou A, Arnold A et al: CSF soluble amyloid precursor proteins in the diagnosis of incipient Alzheimer disease. Neurology, 2011; 77: 35–38
- 64. Rowe CC, Ellis KA, Rimajova M et al: Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging. Neurobiol Aging, 2010; 31: 1275–83
- 65. Andreasen N, Hesse C, Davidsson P et al: Cerebrospinal fluid beta-amyloid(1-42) in Alzheimer disease: differences between early- and late-onset Alzheimer disease and stability during the course of disease. Arch Neurol, 1999; 56: 673–80

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- 66. Mehta PD, Pirttila T, Patrick BA et al: Amyloid beta protein 1–40 and 1–42 levels in matched cerebrospinal fluid and plasma from patients with Alzheimer disease. Neurosci Lett, 2001; 304: 102–6
- 67. Mayeux R, Honig LS, Tang MX et al: Plasma A[beta]40 and A[beta]42 and Alzheimer's disease: Relation to age, mortality, and risk. Neurology, 2003; 61: 1185–90
- Clark CM, Xie S, Chittams J et al: Cerebrospinal fluid tau and beta-amyloid: how well do these biomarkers reflect autopsy-confirmed dementia diagnoses? Arch Neurol, 2003; 60: 1696–702
- 69. Schoonenboom NS, Pijnenburg YA, Mulder C et al: Amyloid beta(1–42) and phosphorylated tau in CSF as markers for early-onset Alzheimer disease. Neurology, 2004; 62: 1580–84
- Fiandaca MS, Kapogiannis D, Mapstone M et al: Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: A case-control study. Alzheimers Dement, 2015; 11: 600–7e1
- Lugli G, Cohen AM, Bennett DA et al: Plasma exosomal miRNAs in persons with and without Alzheimer disease: Altered expression and prospects for biomarkers. PLoS One, 2015; 10: e0139233
- 72. Cheng L, Doecke JD, Sharples RA et al., Australian Imaging, Biomarkers and Lifestyle (AIBL) Research Group: Prognostic serum miRNA biomarkers associated with Alzheimer's disease shows concordance with neuropsychological and neuroimaging assessment. Mol Psychiatry, 2015; 20: 1188–96
- Gui Y, Liu H, Zhang L et al: Altered microRNA profiles in cerebrospinal fluid exosome in Parkinson disease and Alzheimer disease. Oncotarget, 2015; 6: 37043–53
- 74. Tan L, Yu JT, Tan MS et al: Genome-wide serum microRNA expression profiling identifies serum biomarkers for Alzheimer's disease. J Alzheimers Dis, 2014; 40: 1017–27
- 75. Riancho J, Vazquez-Higuera JL, Pozueta A et al: MicroRNA profile in patients with Alzheimer's disease: Analysis of miR-9-5p and miR-598 in raw and exosome enriched cerebrospinal fluid samples. J Alzheimers Dis, 2017; 57: 483–91
- 76. Hao P, Liang Z, Piao H et al: Conditioned medium of human adipose-derived mesenchymal stem cells mediates protection in neurons following glutamate excitotoxicity by regulating energy metabolism and GAP-43 expression. Metab Brain Dis, 2014; 29: 193–205
- 77. Xin H, Li Y, Cui Y et al: Systemic administration of exosomes released from mesenchymal stromal cells promote functional recovery and neurovascular plasticity after stroke in rats. J Cereb Blood Flow Metab, 2013; 33: 1711–15

- Katsuda T, Tsuchiya R, Kosaka N et al: Human adipose tissue-derived mesenchymal stem cells secrete functional neprilysin-bound exosomes. Sci Rep, 2013; 3: 1197
- 79. Iwata N, Tsubuki S, Takaki Y et al: Metabolic regulation of brain Abeta by neprilysin. Science, 2001; 292: 1550–52
- Yasojima K, Akiyama H, McGeer EG, McGeer PL: Reduced neprilysin in high plaque areas of Alzheimer brain: A possible relationship to deficient degradation of beta-amyloid peptide. Neurosci Lett, 2001; 297: 97–100
- 81. Kaeser SA, Herzig MC, Coomaraswamy J et al: Cystatin C modulates cerebral beta-amyloidosis. Nat Genet, 2007; 39: 1437–39
- Mi W, Pawlik M, Sastre M et al: Cystatin C inhibits amyloid-beta deposition in Alzheimer's disease mouse models. Nat Genet, 2007; 39: 1440–42
- Sastre M, Calero M, Pawlik M et al: Binding of cystatin C to Alzheimer's amyloid beta inhibits in vitro amyloid fibril formation. Neurobiol Aging, 2004; 25: 1033–43
- Taupin P, Ray J, Fischer WH et al: FGF-2-responsive neural stem cell proliferation requires CCg, a novel autocrine/paracrine cofactor. Neuron, 2000; 28: 385–97
- Morel L, Regan M, Higashimori H et al: Neuronal exosomal miRNA-dependent translational regulation of astroglial glutamate transporter GLT1. J Biol Chem, 2013; 288: 7105–16
- Liu R, Liu J, Ji X, Liu Y: Synthetic nucleic acids delivered by exosomes: A potential therapeutic for gene-related metabolic brain diseases. Metab Brain Dis, 2013; 28: 551–62
- Di Nicola M, Carlo-Stella C, Magni M et al: Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. Blood, 2002; 99: 3838–43
- Krampera M, Glennie S, Dyson J et al: Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. Blood, 2003; 101: 3722–29
- Clayton A, Harris CL, Court J et al: Antigen-presenting cell exosomes are protected from complement-mediated lysis by expression of CD55 and CD59. Eur J Immunol, 2003; 33: 522–31
- 90. Ngolab J, Trinh I, Rockenstein E et al: Brain-derived exosomes from dementia with Lewy bodies propagate alpha-synuclein pathology. Acta Neuropathol Commun, 2017; 5: 46
- 91. Zheng T, Pu J, Chen Y et al: Exosomes secreted from HEK293-APP Swe/Ind cells impair the hippocampal neurogenesis. Neurotox Res, 2017; 32: 82–93

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