

Article

Effects and Impacts of Different Oxidative Digestion Treatments on Virgin and Aged Microplastic Particles

Ilaria Savino ^{1,2}, Claudia Campanale ^{1,*} , Pasquale Trotti ³ , Carmine Massarelli ¹ , Giuseppe Corriero ² and Vito Felice Uricchio ¹ 

- ¹ Italian National Council of Research, Water Research Institute, 70132 Bari, Italy; ilaria.savino@ba.irsa.cnr.it (I.S.); carmine.massarelli@ba.irsa.cnr.it (C.M.); vito.uricchio@ba.irsa.cnr.it (V.F.U.)
- ² Department of Biology, University of Bari Aldo Moro, 70121 Bari, Italy; giuseppe.corriero@uniba.it
- ³ Sezione di Entomologia e Zoologia Agraria, Dipartimento di Scienze del Suolo, della Pianta e degli Alimenti, University of Bari Aldo Moro, 70121 Bari, Italy; pasquale.trotti@uniba.it
- * Correspondence: claudia.campanale@ba.irsa.cnr.it

Abstract: Although several sample preparation methods for analyzing microplastics (MPs) in environmental matrices have been implemented in recent years, important uncertainties and criticalities in the approaches adopted still persist. Preliminary purification of samples, based on oxidative digestion, is an important phase to isolate microplastics from the environmental matrix; it should guarantee both efficacy and minimal damage to the particles. In this context, our study aims to evaluate Fenton's reaction digestion pre-treatment used to isolate and extract microplastics from environmental matrices. We evaluated the particle recovery efficiency and the impact of the oxidation method on the integrity of the MPs subjected to digestion considering different particles' polymeric composition, size, and morphology. For this purpose, two laboratory experiments were set up: the first one to evaluate the efficacy of various digestion protocols in the MPs extraction from a complex matrix, and the second one to assess the possible harm of different treatments, differing in temperatures and volume reagents used, on virgin and aged MPs. Morphological, physicochemical, and dimensional changes were verified by Scanning Electron Microscope (SEM) and Fourier Transformed Infrared (FTIR) spectroscopy. The findings of the first experiment showed the greatest difference in recovery rates especially for polyvinyl chloride and polyethylene terephthalate particles, indicating the role of temperature and the kind of polymer as the major factors influencing MPs extraction. In the second experiment, the SEM analysis revealed morphological and particle size alterations of various entities, in particular for the particles treated at 75 °C and with major evident alterations of aged MPs to virgin ones. In conclusion, this study highlights how several factors, including temperature and polymer, influence the integrity of the particles altering the quality of the final data.

Keywords: microplastics; oxidative digestion; Fenton's reagent; virgin; aged; weathering; SEM; FTIR



Citation: Savino, I.; Campanale, C.; Trotti, P.; Massarelli, C.; Corriero, G.; Uricchio, V.F. Effects and Impacts of Different Oxidative Digestion Treatments on Virgin and Aged Microplastic Particles. *Polymers* **2022**, *14*, 1958. <https://doi.org/10.3390/polym14101958>

Academic Editor: Jacopo La Nasa

Received: 1 April 2022

Accepted: 9 May 2022

Published: 11 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Microplastics (MPs) are “synthetic solid particle or polymeric matrix, with regular or irregular shape from 1 µm to 5 mm size, of either primary or secondary manufacturing origin” [1]. Their presence has been reported in all environmental matrices, becoming an emerging problem worldwide [2–4]. Due to their small size, high volume surface ratio, and their ability to adsorb or release pollutants [5], MPs' threats mainly concern their effects on organisms and human health [6–9]. Therefore, MPs monitoring is important to understand their presence in the environment. Microplastic studies require several methodological approaches to isolate, identify and quantify particles spread in environmental matrices [10–12].

However, when environmental matrices are rich in organic matter, a chemical digestion treatment is necessary to remove it and release particles. Organic residues have a

density similar to that of polymers, and they may float together MPs during the density separation phase, hindering extraction and quantitative analysis of particles [13]. Organic digestion treatments may be based on oxidizing agents, acids, basics, or enzymes [10,14–16]. However, not all procedures remove organic matter without damaging polymers [17–19].

Applying strong acids, such as nitric acid (HNO₃), produced efficient digestion of biota but they are toxic, corrosive, and cause polymers degradation such as polystyrene, polyamide, and polyethylene [10,20,21]. Alternatively, studies have used alkaline solutions and enzymes for biota digestion, but these require much time, may damage some polymers, and are very expensive [22–24].

Oxidizing agents are increasingly used for water, soil, and sediment because the type of organic matter is more difficult to digest (leaves, woody debris, algae, etc.) [13,25,26]. However, at high concentrations and temperatures, agents such as hydrogen peroxide (H₂O₂) could destroy polyamide particles, reduce their size and alter the colour of polypropylene particles [27]. Digestion protocols should have minimal impact on the morphology, colour, and weight of MPs [25,28].

Several studies analyzed the effects of digestion treatments on MPs, testing different reaction times, temperatures, and reagent volumes [19,27,29–33]. However, most methodological studies tested treatments on virgin MP rather than aged, neglecting their effect on fragile and damaged particles, more representative of reality [28–30]. Despite the recent development of biodegradable plastics, less impactful on the environment [34], many biotic and abiotic factors act on plastics and MPs, leading to changes in polymer properties through different degradation mechanisms [35–39]. Light and temperature, for example, involve free radical formation, chain scission, and subsequent reduction of molecular weight. This, together with mechanical and biotic stress, makes the polymers fragile and more susceptible to fragmentation. The formation of superficial cracks becomes, then, a site of other degradation reactions, leading to the disintegration of material [35,40].

In this context, the present study aims to assess the goodness of the most popular protocol of oxidative digestion used as a preparative step, to purify samples isolating and extracting MPs from complex environmental matrices. The method has been evaluated in terms of efficiency of extraction and recovery of MPs from the environmental matrix and, the impact and aggressiveness of the chemical digestion on the integrity of particles.

Moreover, we tested different experimental digestion conditions on virgin and aged MPs of various morphology, polymer, and size to assess if a different reaction to the chemical digestion and, an eventual alteration of items, occur based on MPs properties. We hypothesize that the rapid oxidation and the stringent exothermic reaction could destroy some polymer particles, especially the most aged ones. These particles are already fragile due to weathering caused by the time of permanence in the environment. The final objective is to advise a less impactful digestion protocol for the extraction of MPs from environmental matrices.

2. Materials and Methods

2.1. Experimental Design

Two different laboratory experiments were set up to assess the efficiency of the most used digestion protocol [14] and its impact on MPs integrity. For this purpose, the oxidative treatment, based on the Fenton reaction ($\text{Fe}^{2+} + \text{H}_2\text{O}_2 \rightarrow \text{Fe}^{3+} + \text{OH} + \text{OH}^-$) [25], was tested using different temperature ranges and reagent volumes. Moreover, to reproduce the difficulties linked to the MPs extraction from complex environmental samples, virgin and aged MPs standards of different sizes and compositions were added to unpolluted soil samples. The integrity of particles and the level of alteration before and after the different treatments were evaluated through Scanning Electron Microscopy (SEM), Fourier Transformed Infrared (FTIR) spectroscopy.

In Table 1 the experimental set-up of the two trials is shown.

Table 1. Indication of the experimental conditions followed for each trial.

Experiment One	
AIM	Evaluate the efficiency of extraction of the most commonly used chemical digestion protocol (based on Wet Peroxide Oxidation [14]) on the recovery of virgin MPs standards from a complex matrix
Particle selection	Virgin MPs
Matrix selected	Soil
Starting digestion condition	Reagents volume: 20 mL of 30% H ₂ O ₂ solution add to 20 mL of 0.05 M iron sulphate heptahydrate (FeSO ₄ ·7H ₂ O) every 30' until complete sample digestion. The temperature of reaction: 75 °C.
Density separation	NaI (1.8 g cm ³)
Qualitative analysis	Stereomicroscope
Experiment Two	
AIM	Evaluate the impact of the most commonly used chemical digestion protocol on the integrity of virgin and aged MPs standards
Particle selection	Virgin and aged MPs
Matrix selected	Soil
Starting digestion condition	Reagents volume: 20 mL of 30% H ₂ O ₂ solution add to 20 mL of 0.05 M iron sulphate heptahydrate (FeSO ₄ ·7H ₂ O) every 30' until complete sample digestion. The temperature of reaction: 75 °C.
Density separation	NaI (1.8 g cm ³)
Qualitative analysis	FTIR—SEM

2.2. Microplastic Standards Selection

Virgin MPs of different shapes and polymers were selected by common plastic items (Table 2). Particle colour was chosen to facilitate detection and counting during the extraction phase. They were cut and smoothed in the laboratory, and particles were passed through sieves with mesh sizes from 5 mm to 1 mm, from 1 mm to 500 µm, and from 500 µm to 100 µm, obtaining MPs of three size ranges. Even, 5 mm size PE, and PP pre-production pellets were added to evaluate the impact of the most commonly used chemical digestion protocol [14] on the integrity of MPs standards (Figure 1).

Table 2. Polymer, density, source, colour, and shape of MPs selected as standards for the experiments. (*): image reworked from source [10].

Polymers	Density (g cm ³) (*)	Source	Colour	Shape
Polystyrene (PS)	0.01–1.06	Food box	White	Fragment
Polypropylene (PP)	0.85–0.92	Disposable glass	Red	Fragment
Polyethylene (PE)	0.89–0.98	Mulching films	Black	Fragment
Polyamide (PA)	1.12–1.15	Textile	Black	Fibre
Polyvinyl chloride (PVC)	1.38–1.41	Building material	Black	Fragment
Polyethylene terephthalate (PET)	1.38–1.41	Plastics bottle	Green	Fragment

2.3. Experiment One: Evaluating the Efficiency of MPs Digestion Treatment through Recovery Tests

Digestion Treatment Conditions

Virgin MPs, 30 particles for each polymer (PE, PP, PET, PVC, PS), underwent six oxidative digestion treatments at three different temperatures (75 °C, 50 °C, and 30 °C), and reagent volumes (100 or 60 mL of H₂O₂ + 20 mL of FeSO₄·7H₂O) (Table 3). The ferrous

ion (Fe^{2+}) of the iron sulphate heptahydrate initiates and catalyses the reaction leading to the generation of hydroxyl and hydroperoxyl radicals, powerful oxidants that degrade organic compounds [41].

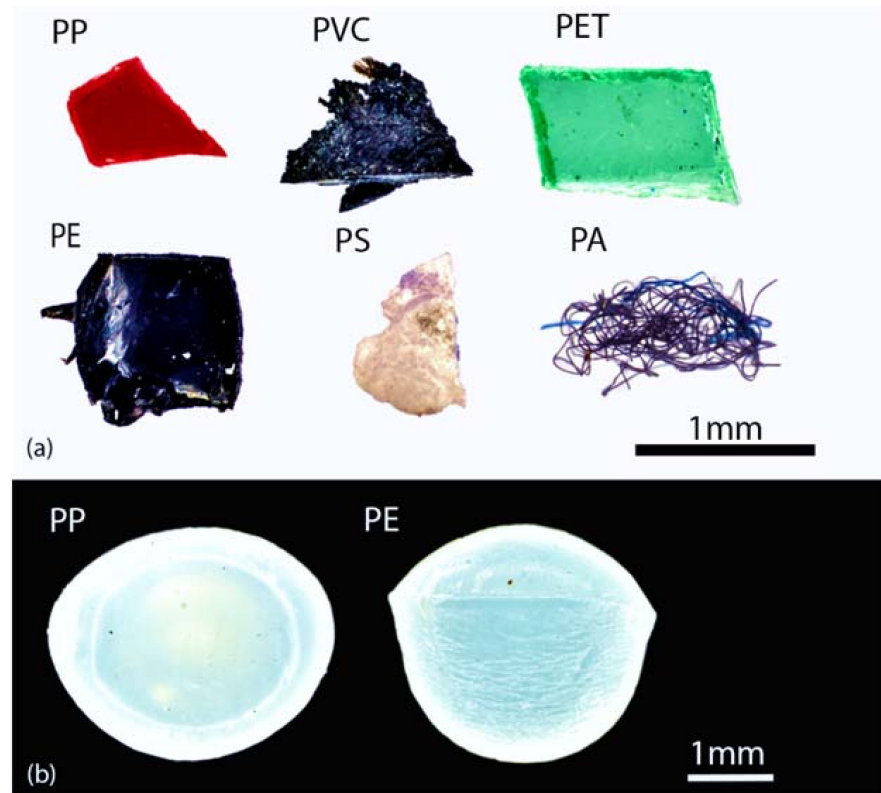


Figure 1. (a) Virgin MPs products by cutting common plastic items. (b) Pre-production pellets added in experiment two. Images produced by Carl Zeiss Tessovar Microscope.

Table 3. Summary of the different oxidative digestion conditions used in experiment one. “-” Treatment made in the absence of the matrix on particle sizes from 500 to 100 μm .

Treatment	Reagent Volumes	Temperature ($^{\circ}\text{C}$)	Polymers	Size	Soil Matrix (g)
1	100 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	75 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm	50
				1 mm–500 μm	-
2	60 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	75 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm	50
				1 mm–500 μm	-
3	100 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	50 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5 mm–1 mm	50
				1 mm–500 μm	-
4	60 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	50 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm	50
				1 mm–500 μm	-
5	100 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	30 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm	50
				1 mm–500 μm	-
6	60 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	30 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm	50
				1 mm–500 μm	-

Three size ranges of particles (5–1 mm; 1 mm–500 μm ; 500–100 μm) were assessed for recoveries. The biggest particles (5–1 mm; 1 mm–500 μm) were added to 50 g of soil to simulate the extraction from a complex matrix while the smallest ones (500–100 μm) were added just of digestion reagents to exclude the influence of matrix on the recovery of particles and evaluate just the effect of the digestion protocol.

A solution of NaI (1.8 g cm^3) was prepared by dissolving the salt in distilled water, to extract MPs from the environmental matrix. After the digestion treatments, the solution was added to the sample, it was shaken for about 10 s and decanted for 1 h. The supernatant was filtrated by a vacuum filtration unit (Sartorius, Goettingen, Germany) using a nitrocellulose filter (Whatman nitrocellulose membrane filters diam. 47 mm, pore size 0.45 μm) and particles were observed under a stereomicroscope (Motic SMZ – 171, Hong Kong, China).

The polymer recovery rate was calculated as the number of extracted particles on the number of added particles. The final value was expressed as a percentage.

2.4. Experiment Two: Evaluating the Impact of Digestion on Virgin vs. Aged MPs Integrity

2.4.1. Ageing of Microplastics

Some virgin microplastics were artificially weathered in a climate room equipped with UVA lamps, calibrated at 340 nm, and programmed at a temperature of 22 $^{\circ}\text{C}$, an irradiance of 12 h, and humidity at 60%, for a total of 20 days. Afterwards, samples were subjected to thermally ageing at 45 $^{\circ}\text{C}$ in an air-circulated oven, for another 20 days (Figure 2).

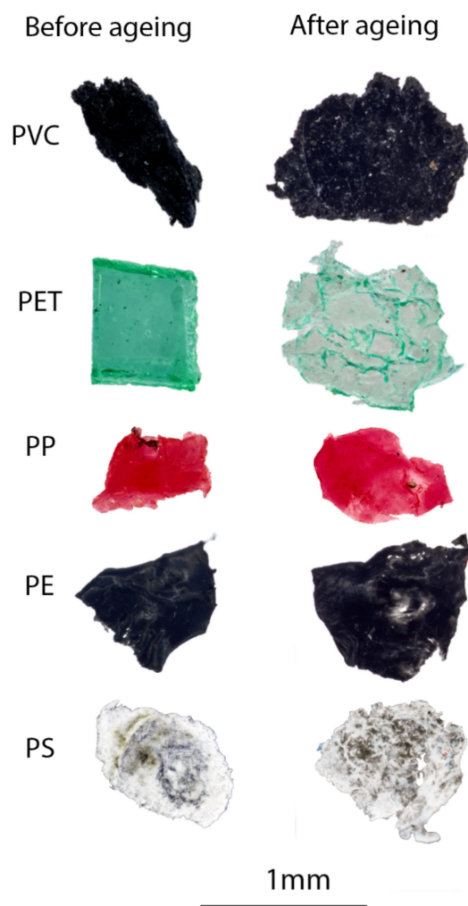


Figure 2. The morphological aspect of some polymers (PVC, PET, PP, PE, PS) before and after the ageing process. These polymers, together with PA fibre and pellets (PP, PE), were exposed to UVA (photo-oxidation) in the climatic chamber for 20 days and then at a temperature of 45 $^{\circ}\text{C}$ for a further 20 days in dry conditions.

2.4.2. Digestion Treatment Conditions

Three oxidative digestion treatments, with different experimental conditions, were tested on PVC, PE, PP, PS, PET fragment; PP and PE pellets, and PA fibre to evaluate the impact on virgin vs. aged MPs by SEM analysis. For each polymer, were selected particles of size from 5 mm to 500 μm and added to 13 g of soil, with the exception of PA to exclude possible contamination from fibre present in the environmental matrix (Table 4). After digestion, particles were separated by the matrix using a saturated NaI solution, and their integrity was observed by SEM, before and after treatments. Virgin and aged fibre were analyzed as tangles before treatments for the difficulty of obtaining single filaments and handling especially those being aged and fragile.

Table 4. Summary of different oxidative digestion conditions used in experiment two. “-” Treatment made in the absence of the matrix on particle sizes from 5 to 1 mm.

Treatment	Reagent Volumes	Temperature ($^{\circ}\text{C}$)	Polymers	Size	Soil Matrix (g)
a	100 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	75 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm 1 mm–500 μm	13
			PA	5–1 mm	-
b	60 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	50 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm 1 mm–500 μm	13
			PA	5–1 mm	-
c	60 mL H_2O_2 + 20 mL $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	30 $^{\circ}\text{C}$	PE, PP, PET, PVC, PS	5–1 mm 1 mm–500 μm	13
			PA	5–1 mm	-

2.5. Fourier Transform Infrared Spectroscopy (FTIR) Acquisition

Aged particles were analyzed, before and after weathering by Fourier Transform Infrared spectroscopy using a Thermo Scientific NICOLET Summit FTIR Spectrometer (Waltham, MA, USA) equipped with an Everest ATR with a diamond Crystal plate and a DTGS KBr detector. The FTIR spectra were recorded in the region of 4000–400 cm^{-1} with 32 scans at a resolution of 4 cm^{-1} .

2.6. Scanning Electron Microscopy (SEM) Acquisition

Scanning Electron Microscopy (HITACHI TM 3000 Tabletop, Tokyo, Japan) was used to observe morphology polymers before and after oxidative digestion treatments. Particles were fixed on carbon adhesive and coated with a thin layer of gold and palladium for 2 min and 10 mA to avoid charging during electron microscopy. Larger particles, such as pellets, were measured operating at 5 kV, while for other particles it was operated at 15 kV. The size of some particles was measured before and after treatments by SEM image software (Hitachi TM 3000, ver. 02-03-02, Tokyo, Japan).

2.7. Quality Control

A cotton coat was worn during the laboratory procedures, preventing any contamination from synthetic clothing. All glass instruments were washed three times with Milli-Q water and covered with aluminium foil. The NaI solution was filtered through a nitrocellulose filter before its use. All analytical steps were performed in a laminar flow cabinet to avoid laboratory airborne contamination.

3. Results

3.1. Results of Experiment One: Evaluating the Efficiency of MPs Digestion Treatment through Recovery Tests

As an overall result, the experimental tests performed in different temperature and peroxide volume conditions showed a recovery efficiency of about 100% for most of the MP materials used (Figure 3). The major criticalities in the extraction efficiency emerged above all for PVC and PET items.

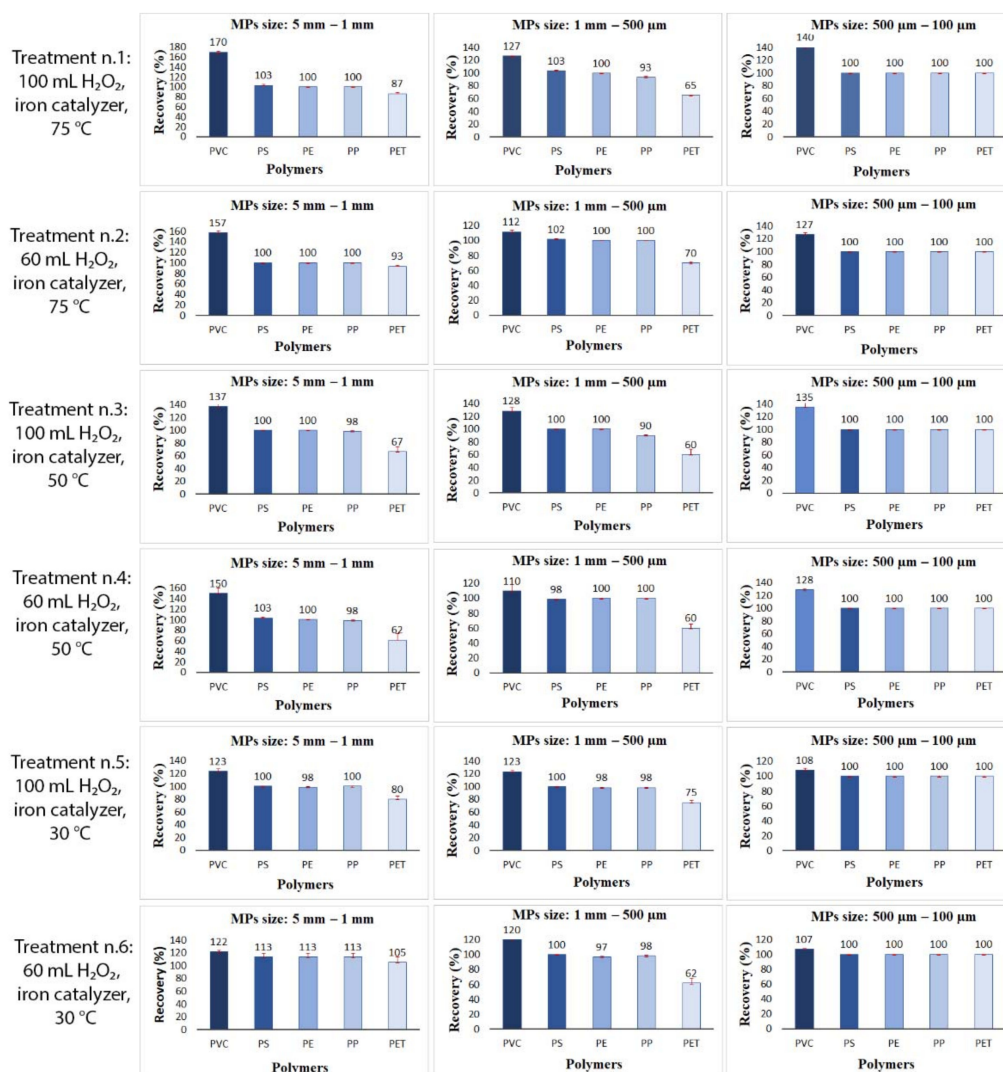


Figure 3. Recovery rates of PVC, PS, PE, PP, and PET, of three-dimensional sizes, after six different treatments varying for H₂O₂ volumes and temperatures. Values are expressed as a percentage mean value of two replicates \pm the standard deviation.

The extraction of 1–5 mm PVC particles testing the treatment n. 1 (75 °C, 100 mL H₂O₂) showed the highest rate of recovery above the 100% (170 \pm 1.4%) followed by treatment n. 2 (75 °C, 60 mL) with a recovery rate of 157 \pm 3.5%, treatment n. 4 (50 °C, 60 mL) with 150 \pm 10%, treatment n. 3 (50 °C, 100 mL) 137 \pm 4%, treatment n. 5 (30 °C, 100 mL) 123 \pm 4% and treatment n. 6 (30 °C, 60 mL) 122 \pm 2%.

This enhancement of observed particles with respect to the initial number of added items is due to the aggressiveness of the digestion treatment on PVC, which led to its fragmentation in smaller particles observed and identified both in suspension and in the soil matrix used (Figure 4).

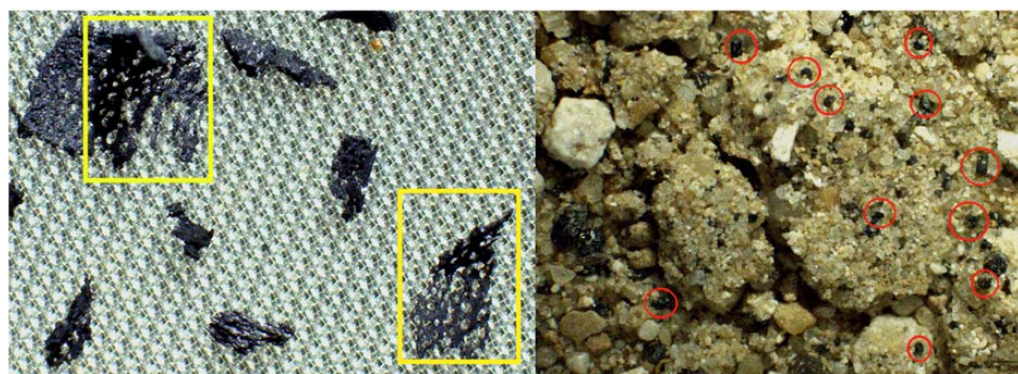


Figure 4. Particulars of PVC fractures (yellow boxes on the left) due to the abrupt oxidation reaction at 75 °C that led to the fragmentation of PVC in tiny particles trapped in the soil matrix (red circles on the right).

Likewise, the smallest PVC fragments (1 mm–500 µm; 500–100 µm) showed the same behaviour with all recoveries above 100%, confirming a fragmentation of this polymer. In each size range, the treatment n.6 (30 °C, 60 mL) resulted in having a lower impact on particles with recoveries close to 100% ($116 \pm 2\%$).

Many other small and tiny PVC particles lower than 100 µm were also observed especially in treatments 1 and 2, probably generated from the fragmentation of bigger particles. As shown in Figure 4 the surface of bigger PVC particles appears greatly modified with holes and cracks.

Differently from the PVC behaviour, all the treatments tested in the different ranges of temperature and reagents volume (treatments n. 1, 2, 3, 4, 5, and 6) demonstrated a good recovery efficiency for PS, PE and PP polymers in all the three size ranges evaluated (5–1 mm, 1 mm–500 µm and 500–100 µm). Indeed, the recoveries obtained were $101 \pm 1\%$, $100.3 \pm 0.5\%$, $99.3 \pm 0.9\%$, for PS, PE, and PP, respectively, showing good resistance to the oxidation reaction and an equally satisfying recovery efficiency from the matrix.

Otherwise, the recoveries of 5–1 mm and 1 mm–500 µm PET fragments showed the lowest recovery rates among all polymers, ranging from 24 and 93% of recovered particles. However, this particle loss is mostly attributable to an effect of the soil matrix used which made difficult the recovery of MPs trapping them in the bottom. Indeed, the tests on the smallest size range of PET MPs (500–100 µm), set up without the soil matrix, showed recovery rates equal to 100% for all the six treatments evaluated.

3.2. Results of Experiment Two: Evaluating the Impact of Digestion Treatment on Virgin and Aged MPs through Qualitative Evaluations

3.2.1. Ageing of Microplastics: FTIR Acquisition

The FTIR acquisitions of MP standards of different polymer compositions made before (black lines, Figure 5) and after the ageing of particles (red lines, Figure 5), show that new absorption peaks were formed consecutively to weathering suggesting strong differences with the pristine materials probably due to their degradation (red lines, Figure 5). In the spectra of each artificially weathered particle, is evident the presence of broad peaks in the region from 3100 to 3700 cm^{-1} (OH stretching).

The ageing process produces new bands at 3423 cm^{-1} in the IR spectra of PET, PA, and PP (pellets and fragments) (Figure 5a).

In the spectra of aged PET particles, forty days after the artificial weathering, a new peak at 1614 cm^{-1} , non-existent in the same particles before the ageing process, appears. (Figure 5c).

Regarding the PE particles, compared to the unaltered pristine MPs acquired by FTIR at the time zero before ageing, the weathered fragment spectra show the presence of new intense peaks at 3414 and 1577 cm^{-1} and others less intense at 873 and 777 cm^{-1} . Similarly, the aged PE pellet spectra show new weathering bands at 3458 and 1618 cm^{-1} .

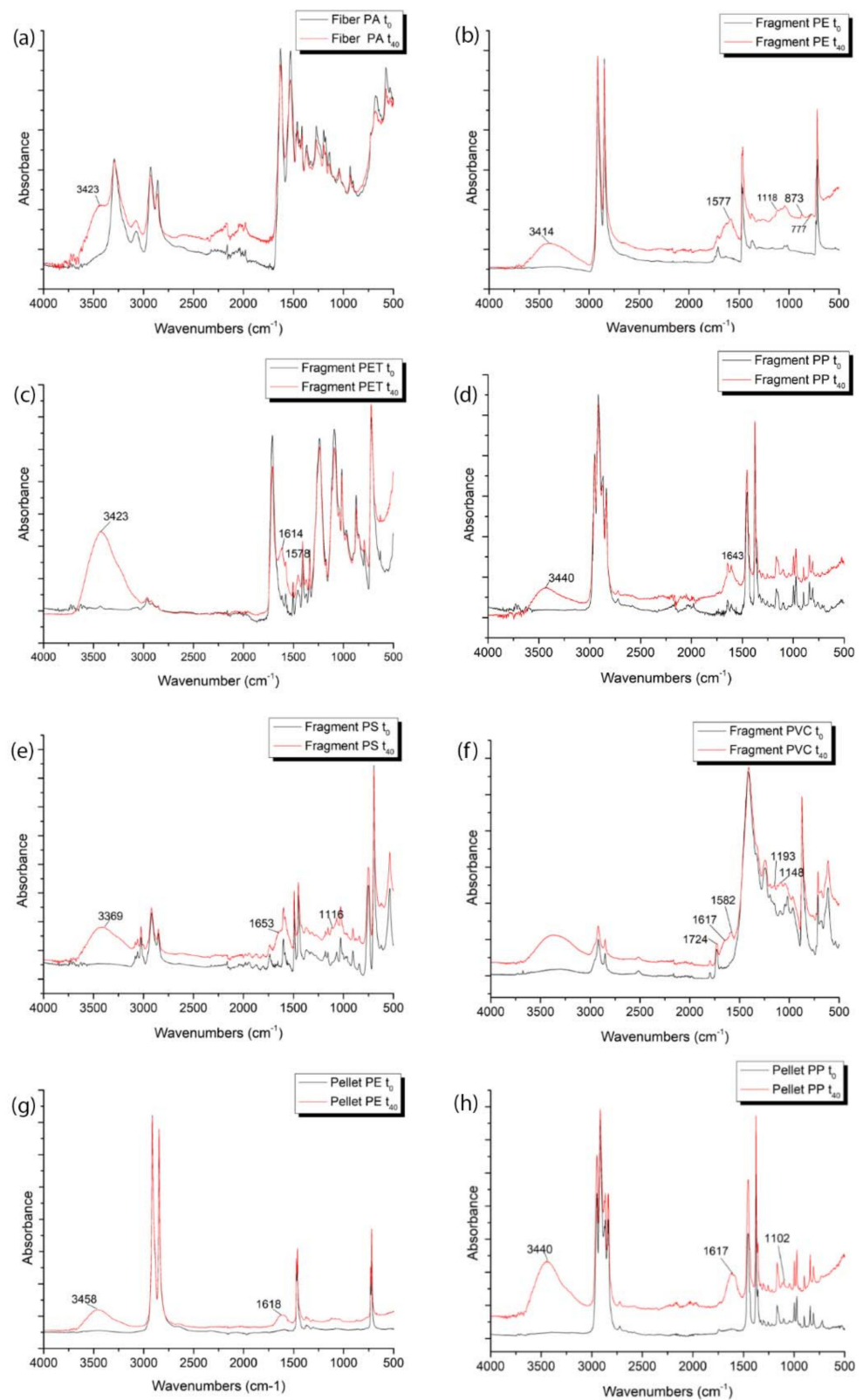


Figure 5. Comparison between the IR spectra of different polymers before (black lines, t_0) and after 40 days of ageing (red lines, t_{40}). New peaks formed after ageing are indicated in the spectra of each polymer. Absorption areas related to ageing from 3100 to 3700 cm^{-1} (hydroxyl groups) are evident in all polymers. (a) PA fiber; (b) PE fragment (c) PET fragment; (d) PP fragment; (e) PS fragment; (f) PVC fragment; (g) PE pellet; (h) PP pellet.

Otherwise, the following peaks appear in the IR spectrum of PP fragments and pellets after ageing: 3404 , 3440 cm^{-1} , and 1643 cm^{-1} (OH bending) (Figure 5d–h). Moreover, the PP pellets show a new band at 1102 cm^{-1} . The IR spectra of PS show changes corresponding to the formation of new bands at 3369 cm^{-1} , 1653 cm^{-1} , and 1116 cm^{-1} (Figure 5e). Regarding the analysis of the IR spectrum of weathered PVC, a broad peak of moderate intensity can be detected in the region $3000\text{--}3500\text{ cm}^{-1}$ and new peaks, with respect to virgin materials, are evident at 1617 cm^{-1} , 1582 cm^{-1} , 1193 cm^{-1} , and 1148 cm^{-1} (Figure 5f).

3.2.2. Scanning Electron Microscopy (SEM) Acquisition

Scanning Electron Microscopy acquisition shows the physical effects of three different oxidative treatments on the integrity of both virgin and aged particles.

Virgin MPs appear compact and solid, with a three-dimensional structure and smooth surfaces. The treatment at $30\text{ }^{\circ}\text{C}$ (treatment c) generates a dimensional reduction of PET MPs associated with margins corrosion (Figure 6a). An expansion of the PVC (Figure 6b), showing its surface damaged by small holes, is also visible together with the PP and PS particles abrasion (Figure S1).

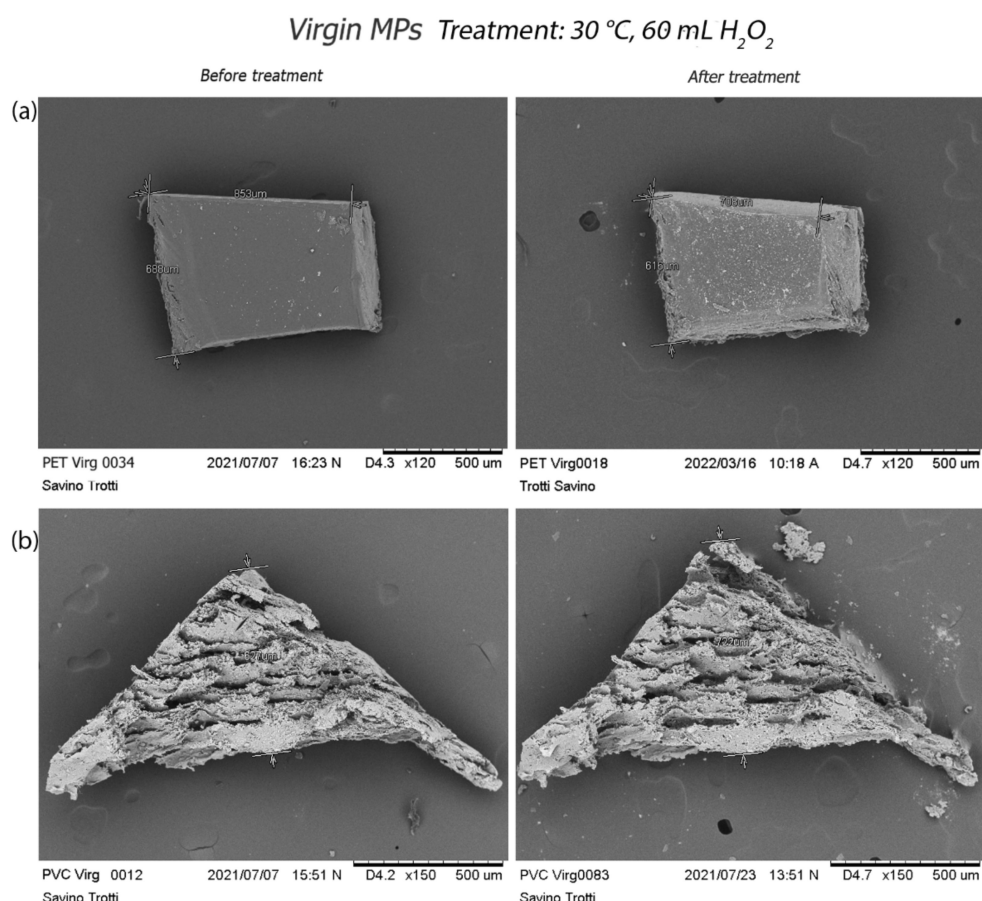


Figure 6. Alterations caused by treatment at $30\text{ }^{\circ}\text{C}$ on virgin PET and PVC: (a) size reduction and corrosion of virgin PET margins, (from 853 to $708\text{ }\mu\text{m}$); (b) PVC dimensional expansion from $627\text{ }\mu\text{m}$ to $722\text{ }\mu\text{m}$.

The treatment at $50\text{ }^{\circ}\text{C}$ (treatment b) affects PVC and PS particles by forming holes, material loss, and corrosion (Figure 7). Even in this case, a slight reduction in the size of PET and PP fragments is visible (Figure S2).

Virgin MPs Treatment: 50 °C, 60 mL H₂O₂

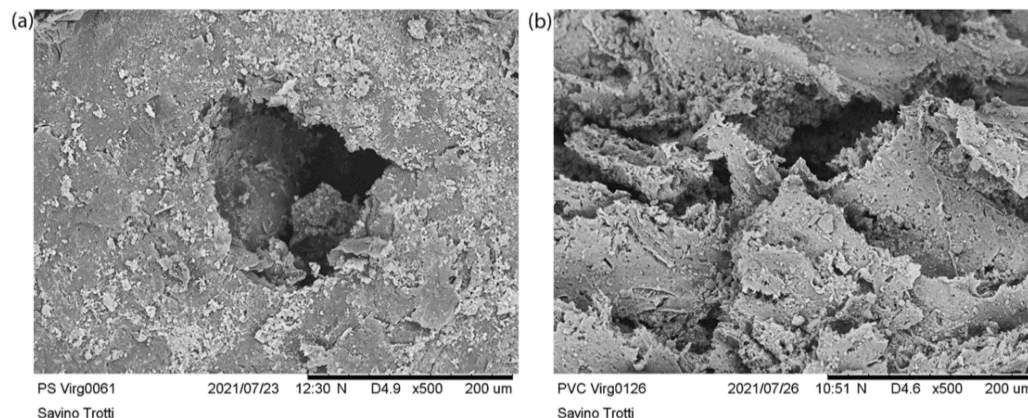


Figure 7. Effects of treatment at 50 °C on virgin PS and PVC: (a) formation of a hole on the surface of the PS; (b) PVC particle surface with small holes.

However, the greatest surface changes occur after treatment at 75 °C (treatment a) (Figure S3). On the one hand, PVC particles manifest wide holes (Figure 8) and a lost material of PS fragment. On the other hand, PP and PET fragments show corroded margins (Figure 9). In each treatment, virgin PE and PP pellets (Figure 10) and PE fragments highlight high resistance to oxidative digestion (Figure S4).

Virgin MPs Treatment: 75 °C, 100 mL H₂O₂

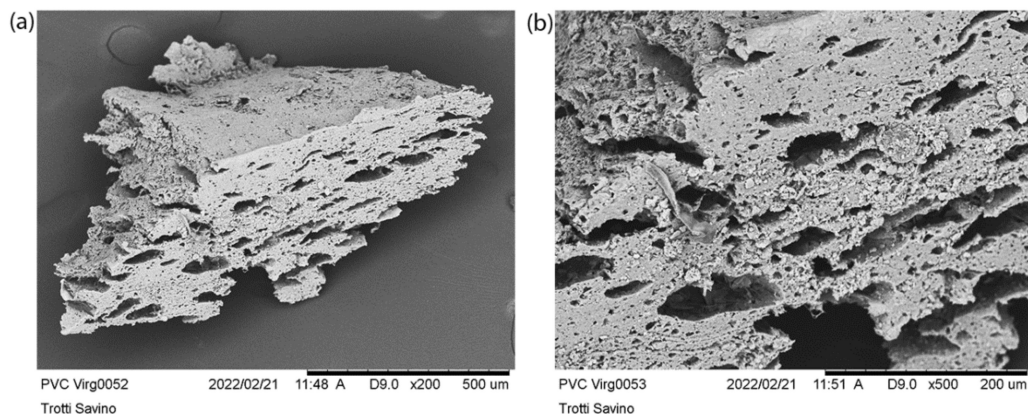


Figure 8. Effects of oxidative digestion treatment at 75 °C on the virgin PVC: (a) morphological acquisition of the entire PVC particle after treatment; (b) detail of large holes inside the PVC particle.

Compared to virgin standards, aged MPs appear, before the digestion treatments, flattened, brittle, with undefined shapes with some cracks on their surface.

As well as for virgin MPs, the milder and intermediate treatments do not produce strong changes in aged particles. In all treatments, a fraying of the fibre from the initial tangle is evident.

The treatment at 30 °C generates new cracks in PVC and PS fragments, and accentuates those already present, due to weathering, in PE and PET (Figure S5).

The treatment at 50 °C produces curling of PE fragments, a fraying of the fibre, and many cracks in PET ones. Even in this case, PVC particles show a surface full of small holes (Figure 11).

Virgin MPs Treatment: 75 °C, 100 mL H₂O₂

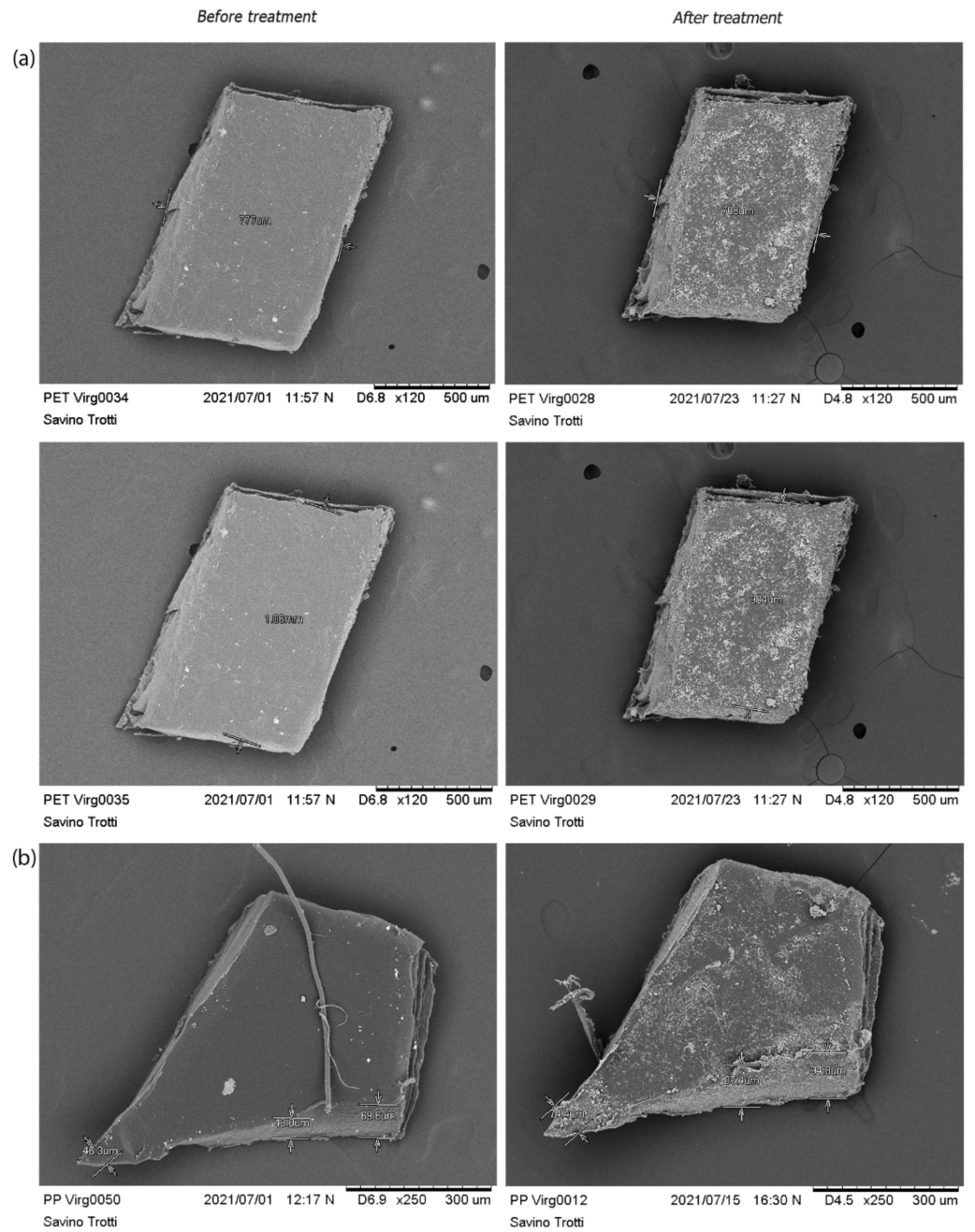


Figure 9. Corrosive treatment effects on PET and PP particles: comparison of size measurements, before and after treatment, emphasizes the corrosion of virgin PET (a) and PP (b).

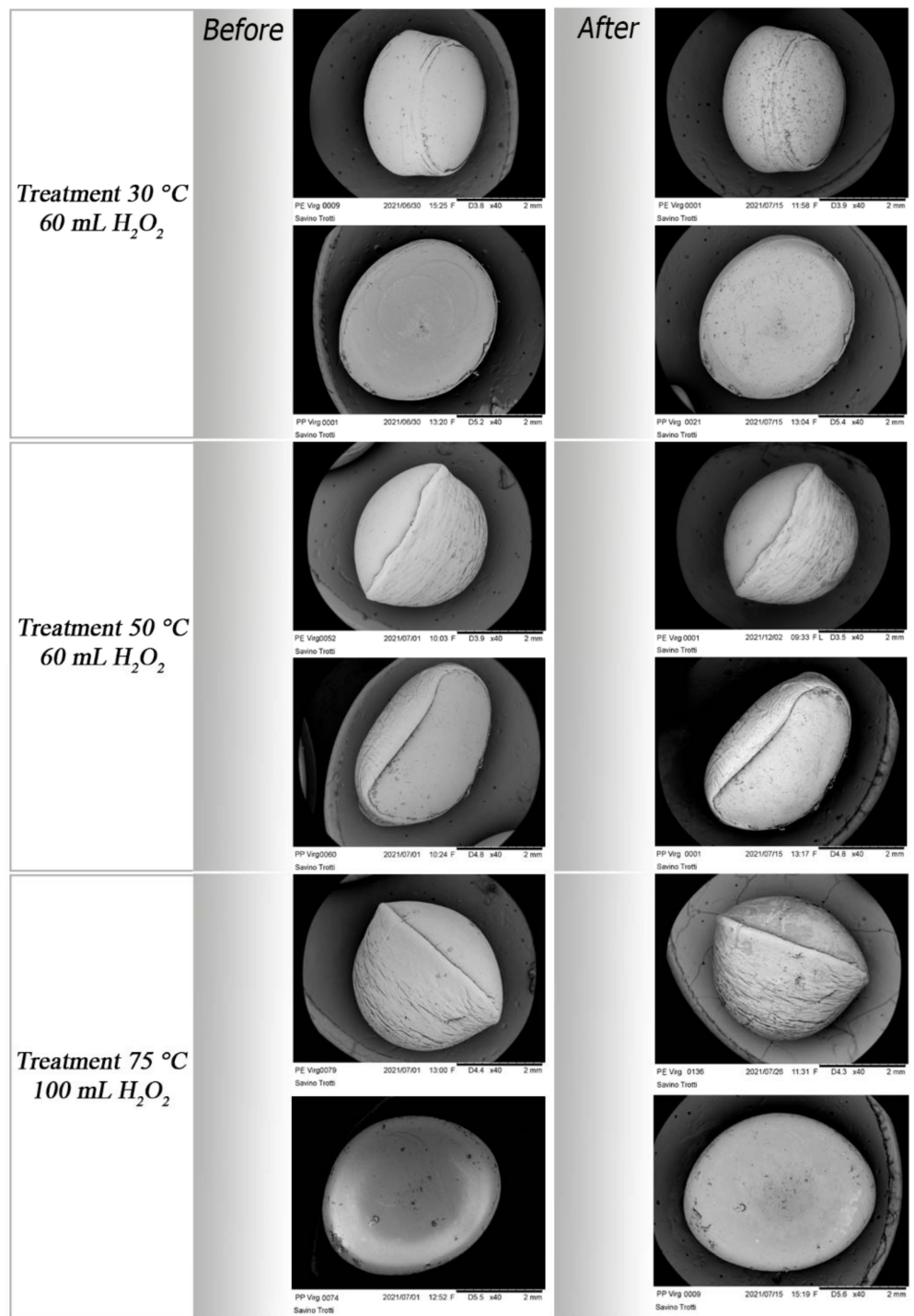


Figure 10. Morphological aspects of pellets before and after different treatments. Virgin PE and PP pellets highlight high resistance to oxidative digestion.

Aged MPs Treatment: 50 °C, 60 mL H₂O₂

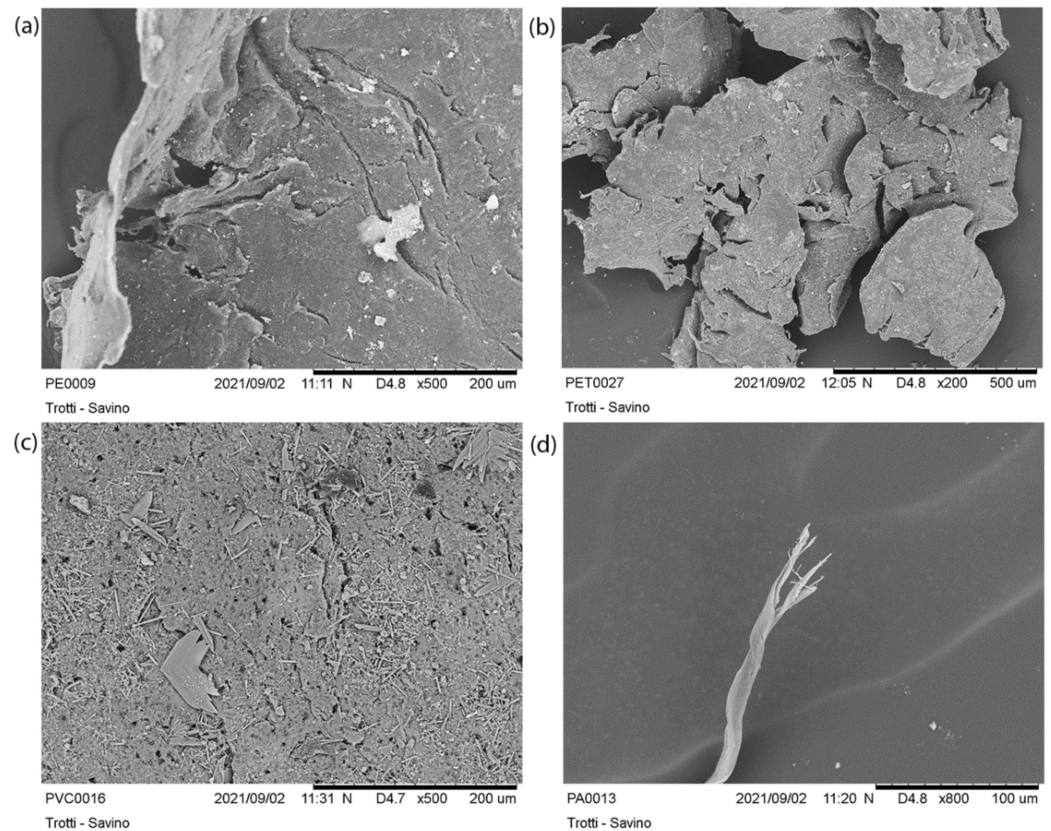


Figure 11. Focus on the effects of treatment at 50 °C on aged particles: (a) detail of PE curling; (b) more cracks in the PET; (c) small holes on the PVC surface; (d) fraying of PA fibre.

The digestion treatment at 75 °C, applied on aged MPs, causes a radical alteration of most particles (Figure S6) with evident changes such as the loss of material of PVC, PS, and PP particles, cracks expansion of PET, PE corrosion, and a fraying of PA (Figure 12). Aged pellets show several abrasions on their surface, especially after the most aggressive treatment at 75 °C (Figure 13).

Aged MPs Treatment: 75 °C, 100 mL H₂O₂

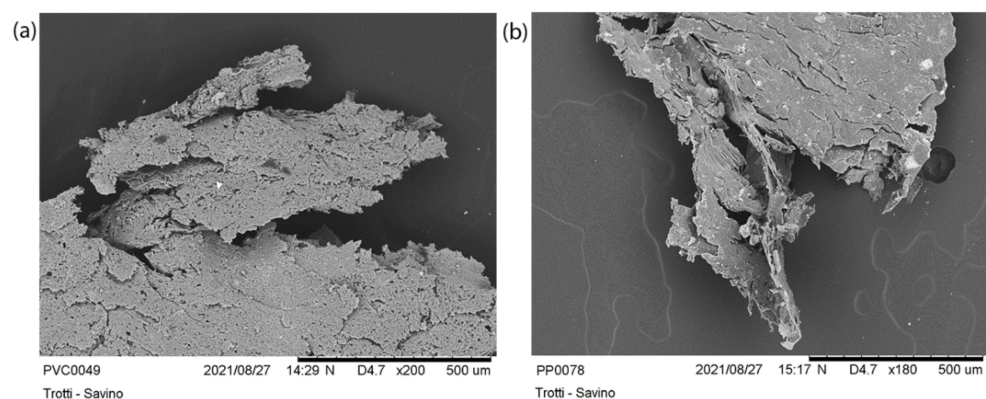


Figure 12. *Cont.*

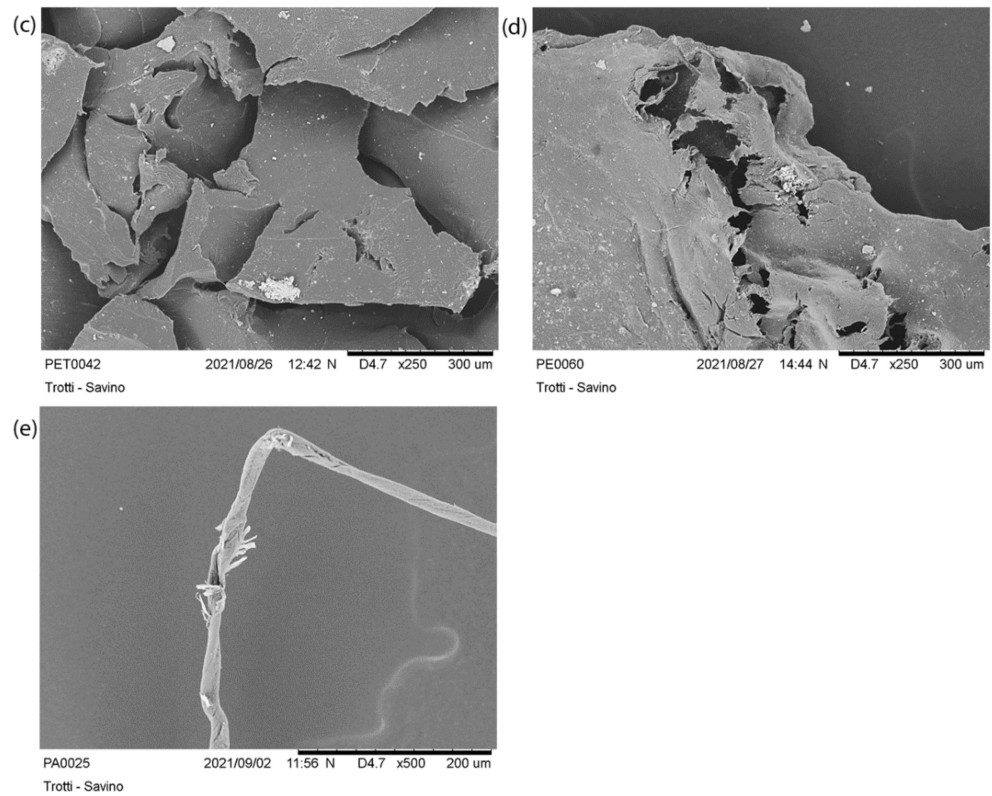


Figure 12. Focus on the effects of treatment at 75 °C on aged particles: (a) small holes on the PVC surface and loss of the polymer material; (b) corrosion and loss of PP polymer material; (c) formation of large cracks in PET; (d) corrosion and loss of PE polymer material; (e) breaking and fraying of the fibre.

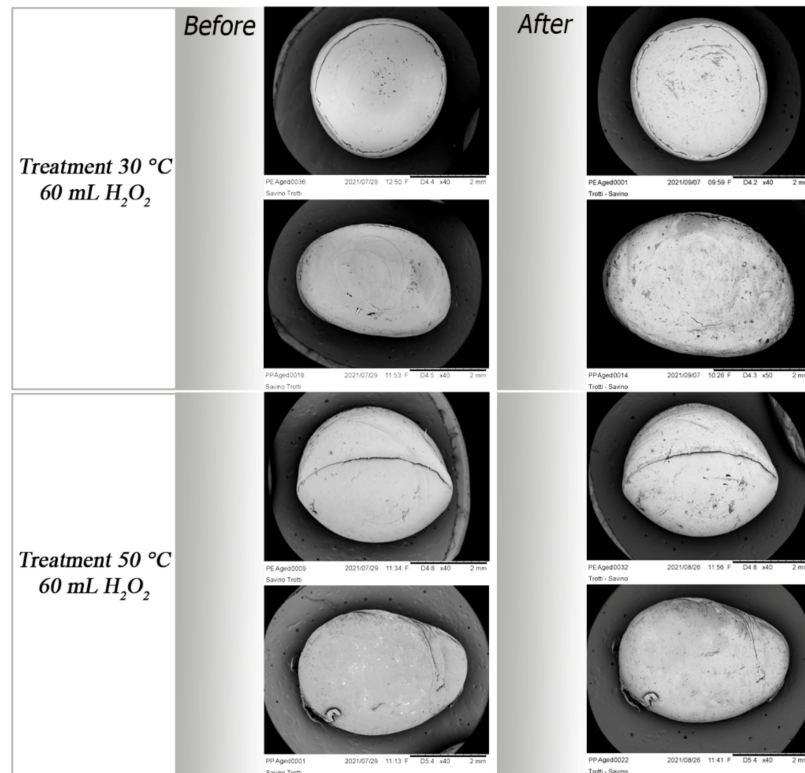


Figure 13. Cont.

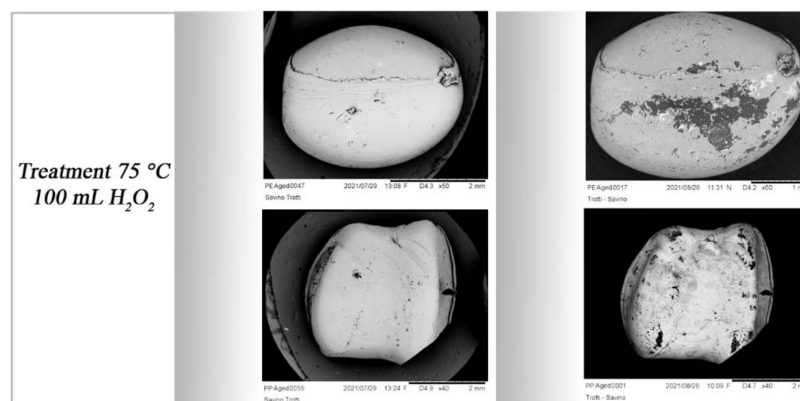


Figure 13. Morphological aspects of aged pellets before and after different treatments: abraded areas are highlighted as the temperature increases.

4. Discussion

Chemical digestion treatment is a crucial step of MPs analysis, especially when environmental samples are rich in organic matter; indeed, removing the natural debris promotes the subsequent extraction of MPs from the matrix. However, the choice of the protocol must consider its digestion effectiveness and, at the same time, the integrity of polymers, so as not to compromise the identification and quantification of particles.

In this regard in the present study, attention has been paid to the impact that one of the most common digestion protocols, based on Fenton's reaction, could have on MPs in terms of recoveries and integrity of particles.

In experiment number one, we hypothesized that the recovery of virgin MPs may be subjected to the type of sample in which the particles are dispersed. Although the density of the NaI solution, adopted for MPs extraction, is higher than high-density polymer standards (e.g., PVC and PET), it was not always possible to fully recover all particles.

Most low-density polymers were recovered almost always at 100%, instead the PET of dimensions from 5 mm to 500 microns, and the PVC of dimensions between 1 mm and 500 microns, were recovered with low efficiency because they remained inside the matrix. A similar result was also observed in a previous study [42], where small PVC fragments were trapped in the sediment after the density separation, but the protocol used does not correspond to this study. Nevertheless, the recoveries of these two polymers were also demonstrated to be low in other studies [43,44]. This behaviour may depend on the effect of the oxidation of the polymer surface, which leads to increased hydrophilicity, reducing the number of possible air bubbles and the buoyancy of the material [43,45]. Previous studies have already reported a relationship between size and recovery rate highlighting that separation of particles with diameters lower than one mm is more difficult than large MPs [27,46,47]. Electrostatic interactions between MPs and other particles depend on the morphology, composition, size, and surface charge of MPs [48]. Therefore, the presence of the matrix can interfere with the MPs extraction phase from the soil and our study suggests this with the recovery of 100% of PET, in the absence of soil.

Our results also highlight the presence of more PVC particles in the suspension and in the soil than initially added. It is appended both in the presence of the matrix and in the absence, after all treatments, especially at 75 °C using 100 mL of H₂O₂, suggesting the influence of temperature in polymer fragmentation. Indeed, an increase in the temperature of the digestion treatment caused the loss of some types of MPs also in previous studies [27,28]. The temperature is an important factor in the extraction procedure as it can affect the characteristics of a polymer based on its glass transition temperature (T_g), which is the critical temperature at which a material changes its features from "glassy material" to "rubbery material" [49].

In experiment number two, we obtained a visual demonstration of how Fenton's reagent impacts MPs. In this case, both virgin and aged particles were analyzed with the

addition of PE and PP pellets, and PA fibres. Various previous studies tested digestion protocols on polymers but pellets are the most used type followed by fragments and fibres [27–29,50,51].

The greater number of small PVC particles counted could be explained by observing the formation of large pores with a slight expansion of the size of the PVC polymer. These pores, favoured by an increase in temperature, may generate a release of tiny fragments of PVC such as those found in the soil. The presence of larger cavities has been reported in [52] for some polymers after the Fenton oxidation process.

SEM analysis also shows a size reduction and margin corrosion of PET and PP, after treatment. Even in [53] particles of PE and PP show a size reduction of about 10%, after long treatment with H₂O₂ but in our case, virgin PE showed high resistance to treatment along with pellets. The corrosion of PET margins and the formation of roughness may have favoured the creation of active sites on the polymer increasing the adhesion of soil particles on the MPs surface. In addition, a particle size reduction could affect those results that include the evaluation of MPs size ranges because particles of a lower size range may not be detected because they are strongly aggravated by the treatment.

As for the virgin PS, particles were damaged at 50 °C and 75 °C as reported by [27]. We have not observed any major changes in this polymer despite the known damage caused by a temperature from 70 °C to 100 °C on the PA [29,54]. However, our data cannot be fully compared with these studies because we show a morphological alteration of the particles and not an assessment of weight.

The results observed so far suggest that the oxidative treatment generates an impact on virgin particles caused by high temperatures. However, environmental MPs are particles already altered and damaged due to biotic and abiotic degradation [35,55,56].

Indeed, the FTIR acquisitions highlighted the formation of new peaks in the polymer spectra consecutively to the ageing of virgin polymers in artificial conditions. As previously observed by other authors [57–59], the regions reflecting ageing-related changes (hydroxyl groups, peaks from 3100 to 3700 cm⁻¹, carbon double bonds, 1600 and 1680 cm⁻¹ and carbonyl groups, 1690 and 1810 cm⁻¹) appeared greatly modified compared to the pristine materials in each type of MPs.

Several previous works conducted under simulated environmental conditions and on field-collected samples showed that photo and thermal oxidation together with humidity could alter the physicochemical structure of MPs leading to the introduction of oxygen into the polymer chain with the formation of carbonyl (CO) and hydroxyl (OH) functional groups [60–63].

As a matter of fact, in the spectra of our degraded MPs, exposed to UVA followed by a period of incubation at 45 °C in dry conditions, new broad hydroxyl peaks (centred at 3300–3400 cm⁻¹), appeared in the aged particles and were the most readily identified. The forced weathering, resulted, in the concomitant occurrence of oxidation reactions, chain degradation, and the formation of surface cracks and fractures. This alteration could allow an easier and deeper infiltration of water and oxygen from the atmosphere into the sample leading, in time, to an increased effect of ageing [64]. However, our weathered samples have been dried immediately before FTIR acquisition, therefore, the OH bands origin could be minimally related to the presence of adsorbed humidity from the atmosphere and more likely to the polymeric alcohols, used as a lubricant in plastic, or other by-products easily released during the degradation process [5].

Moreover, the entire region between 1550 and 1810 cm⁻¹ usually referred to as the “carbonyl groups” is indicative of oxidized carbon in the plastic hydrocarbon chain. The presence of carbonyls, in almost all our weathered polymers, even if less distinctive compared to hydroxyl groups, suggests that oxygen has bonded with the hydrocarbon chain [60,65,66].

Indeed, the use of a “carbonyl index” (CI) is frequently used to measure the light-induced photo-oxidation, since it increases with increasing exposure time of plastic into the environment or progressive ageing of MPs [57,59,67].

Moreover, in our experiments, new peaks are present almost all in the fingerprint region ($1500\text{--}500\text{ cm}^{-1}$).

However, the spectral changes during these processes are not yet fully understood and it is complex to monitor and predict these changes.

The increased peaks around 1760 and 1690 in PVC, PP, and PE fragment spectra, indicates the presence of carbonyl groups formed by photo-oxidation in the climate chamber [57,67].

Therefore, the assessment of methodological protocols for MPs studies should consider the intrinsic physicochemical difference between virgin and aged starting materials preferring the use of weathered particles to simulate the real environmental conditions. To our knowledge, only one study used aged particles for protocol testing [18].

Compared to virgin particles, PA fibres and PE fragments, already damaged by the ageing process, showed signs of breakage and fraying. The impact of the treatment temperature was strongly evident on the aged particles because it caused the loss of polymeric material and a strong alteration of all particles. Even aged pellets showed a rough surface already after the milder treatments until to show the most evident abrasions after the digestion at $75\text{ }^{\circ}\text{C}$.

5. Conclusions

This study evaluated the impact of the most common oxidative digestion protocol used to extract MPs from environmental samples. We have provided a visual demonstration of particle alterations by SEM analysis and we find that this technique can help in the evaluation of protocols. Wet peroxide oxidation is an effective organic digestion method for different environmental matrices but our results showed morphological changes in polymers not observed in other studies. Both virgin and aged MPs were damaged, especially after treatment at $75\text{ }^{\circ}\text{C}$. Several factors must be considered in the assessment of experimental conditions. Furthermore, methods recommending temperatures below $50\text{ }^{\circ}\text{C}$ should be preferred at the expense of longer digestion times. We also suggest evaluating the type of matrix, particle size, and shape and exploring a broad type of polymer in methodological analysis, to ensure a comprehensive assessment of the impact on MPs. Moreover, the weathering of particles in simulated environmental conditions, showed great alterations of FTIR spectra influencing the correct polymer characterization of real environmental samples and making difficult the interpretation of spectra. The degree of degradation is probably connected to the time of exposure of particles to environmental weather conditions.

Nonetheless, the evaluation of methodological approaches on aged MPs rather than on pristine materials is essential to ensure a more realistic vision of obtained results and a better quality of the final data.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/polym14101958/s1>, Figure S1. Detail on the effects of treatment at $30\text{ }^{\circ}\text{C}$ on polymers: (a) slight damage to the PS; (b) formation of small holes on the surface of the PVC; (c) corrosion of PET margins; (d) abrasion and corrosion of PP margins. Figure S2. Slight size alteration caused by treatment at $50\text{ }^{\circ}\text{C}$ on virgin PP (a) and PET (b). Figure S3. Overview of virgin polymer morphology before and after treatment at $75\text{ }^{\circ}\text{C}$: loss of polymer material in PS; formation of large holes in the inside of PVC; corrosion of PP and PET margins, high resistance to PE and PA treatment. Figure S4. Visual demonstration of the high resistance of virgin PE after treatment at $30\text{ }^{\circ}\text{C}$ (a), treatment at $50\text{ }^{\circ}\text{C}$ (b), and $75\text{ }^{\circ}\text{C}$ (c). Figure S5. Focus on damage caused by treatment at $30\text{ }^{\circ}\text{C}$ on aged MPs: formation of cracks and breaks on PVC (a); PS (b); PET (c); and PE (d). Figure S6. Overview of aged polymer morphology before and after treatment at $75\text{ }^{\circ}\text{C}$: loss of material of PVC, PS, and PP particles, cracks expansion of PET, PE corrosion, and a fraying of PA.

Author Contributions: Conceptualization, C.C. and I.S.; methodology, C.C., I.S. and P.T.; formal analysis, I.S. and P.T.; investigation, I.S.; resources, V.F.U. and C.M.; data curation, I.S. and C.C.; writing—original draft preparation, I.S. and C.C.; writing—review and editing, C.C.; visualization,

I.S. and P.T.; supervision, C.C. and G.C.; project administration, C.M.; funding acquisition, C.M. and V.F.U. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors gratefully acknowledge Domenico Bellifemine for his precious support in technical improvements in laboratory equipment and Francesco Porcelli for his support and availability in SEM acquisitions.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Frias, J.P.G.L.; Nash, R. Microplastics: Finding a consensus on the definition. *Mar. Pollut. Bull.* **2019**, *138*, 145–147. [[CrossRef](#)] [[PubMed](#)]
2. Gasperi, J.; Wright, S.L.; Dris, R.; Collard, F.; Mandin, C.; Guerrouache, M.; Langlois, V.; Kelly, F.J.; Tassin, B. Microplastics in air: Are we breathing it in? *Curr. Opin. Environ. Sci. Health* **2018**, *1*, 1–5. [[CrossRef](#)]
3. Kanhai, L.D.K.; Gårdfeldt, K.; Lyashevskaya, O.; Hassellöv, M.; Thompson, R.C.; O'Connor, I. Microplastics in sub-surface waters of the Arctic Central Basin. *Mar. Pollut. Bull.* **2018**, *130*, 8–18. [[CrossRef](#)] [[PubMed](#)]
4. Campanale, C.; Galafassi, S.; Savino, I.; Massarelli, C.; Ancona, V.; Volta, P.; Uricchio, V.F. Microplastics pollution in the terrestrial environments: Poorly known diffuse sources and implications for plants. *Sci. Total Environ.* **2022**, *805*, 150431. [[CrossRef](#)]
5. Campanale, C.; Dierkes, G.; Massarelli, C.; Bagnuolo, G.; Uricchio, V.F. A Relevant Screening of Organic Contaminants Present on Freshwater and Pre-Production Microplastics. *Toxics* **2020**, *8*, 100. [[CrossRef](#)]
6. Batel, A.; Borchert, F.; Reinwald, H.; Erdinger, L.; Braunbeck, T. Microplastic accumulation patterns and transfer of benzo[a]pyrene to adult zebrafish (*Danio rerio*) gills and zebrafish embryos. *Environ. Pollut.* **2018**, *235*, 918–930. [[CrossRef](#)]
7. Barboza, L.G.A.; Lopes, C.; Oliveira, P.; Bessa, F.; Otero, V.; Henriques, B.; Raimundo, J.; Caetano, M.; Vale, C.; Guilhermino, L. Microplastics in wild fish from North East Atlantic Ocean and its potential for causing neurotoxic effects, lipid oxidative damage, and human health risks associated with ingestion exposure. *Sci. Total Environ.* **2020**, *717*, 134625. [[CrossRef](#)]
8. Campanale, C.; Massarelli, C.; Savino, I.; Locaputo, V.; Uricchio, V.F. A Detailed Review Study on Potential Effects of Microplastics and Additives of Concern on Human Health. *Int. J. Environ. Res. Public Health* **2020**, *17*, 1212. [[CrossRef](#)]
9. Cormier, B.; Gambardella, C.; Tato, T.; Perdriat, Q.; Costa, E.; Veclin, C.; Le Bihanic, F.; Grassl, B.; Dubocq, F.; Kärrman, A.; et al. Chemicals sorbed to environmental microplastics are toxic to early life stages of aquatic organisms. *Ecotoxicol. Environ. Saf.* **2021**, *208*, 111665. [[CrossRef](#)]
10. Campanale, C.; Savino, I.; Pojar, I.; Massarelli, C.; Uricchio, V.F. A Practical Overview of Methodologies for Sampling and Analysis of Microplastics in Riverine Environments. *Sustainability* **2020**, *12*, 6755. [[CrossRef](#)]
11. Massarelli, C.; Campanale, C.; Uricchio, V.F. A handy open-source application based on computer vision and machine learning algorithms to count and classify microplastics. *Water* **2021**, *13*, 2104. [[CrossRef](#)]
12. Lusher, A.L.; Munno, K.; Hermabessiere, L.; Carr, S. Isolation and Extraction of Microplastics from Environmental Samples: An Evaluation of Practical Approaches and Recommendations for Further Harmonization. *Appl. Spectrosc.* **2020**, *74*, 1049–1065. [[CrossRef](#)] [[PubMed](#)]
13. Li, Q.; Wu, J.; Zhao, X.; Gu, X.; Ji, R. Separation and identification of microplastics from soil and sewage sludge. *Environ. Pollut.* **2019**, *254*, 113076. [[CrossRef](#)]
14. Masura, J.; Baker, J.; Foster, G.; Arthur, C.; Laboratory Methods for the Analysis of Microplastics in the Marine Environment: Recommendations for Quantifying Synthetic Particles in Waters and Sediments. NOAA Technical Memorandum NOS-OR&R-48. 2015. Available online: <https://repository.library.noaa.gov/view/noaa/10296> (accessed on 1 January 2022).
15. von Friesen, L.W.; Granberg, M.E.; Hassellöv, M.; Gabrielsen, G.W.; Magnusson, K. An efficient and gentle enzymatic digestion protocol for the extraction of microplastics from bivalve tissue. *Mar. Pollut. Bull.* **2019**, *142*, 129–134. [[CrossRef](#)] [[PubMed](#)]
16. Bretas Alvim, C.; Bes-Piá, M.A.; Mendoza-Roca, J.A. Separation and identification of microplastics from primary and secondary effluents and activated sludge from wastewater treatment plants. *Chem. Eng. J.* **2020**, *402*, 126293. [[CrossRef](#)]
17. Thiele, C.J.; Hudson, M.D.; Russell, A.E. Evaluation of existing methods to extract microplastics from bivalve tissue: Adapted KOH digestion protocol improves filtration at single-digit pore size. *Mar. Pollut. Bull.* **2019**, *142*, 384–393. [[CrossRef](#)]
18. Prata, J.C.; da Costa, J.P.; Girão, A.V.; Lopes, I.; Duarte, A.C.; Rocha-Santos, T. Identifying a quick and efficient method of removing organic matter without damaging microplastic samples. *Sci. Total Environ.* **2019**, *686*, 131–139. [[CrossRef](#)]
19. Pfohl, P.; Roth, C.; Meyer, L.; Heinemeyer, U.; Gruending, T.; Lang, C.; Nestle, N.; Hofmann, T.; Wohlleben, W.; Jessl, S. Microplastic extraction protocols can impact the polymer structure. *Microplast. Nanoplast.* **2021**, *1*, 8. [[CrossRef](#)]
20. Naidoo, T.; Goordiyal, K.; Glassom, D. Are Nitric Acid (HNO₃) Digestions Efficient in Isolating Microplastics from Juvenile Fish? *Water Air Soil Pollut.* **2017**, *228*, 470. [[CrossRef](#)]

21. Stock, F.; Kochleus, C.; Bänsch-Baltruschat, B.; Brennholt, N.; Reifferscheid, G. Sampling techniques and preparation methods for microplastic analyses in the aquatic environment—A review. *TrAC Trends Anal. Chem.* **2019**, *113*, 84–92. [[CrossRef](#)]
22. Löder, M.G.J.; Imhof, H.K.; Ladehoff, M.; Löschel, L.A.; Lorenz, C.; Mintenig, S.; Piehl, S.; Primpke, S.; Schrank, I.; Laforsch, C.; et al. Enzymatic Purification of Microplastics in Environmental Samples. *Environ. Sci. Technol.* **2017**, *51*, 14283–14292. [[CrossRef](#)] [[PubMed](#)]
23. Kühn, S.; van Werven, B.; van Oyen, A.; Meijboom, A.; Bravo Rebolledo, E.L.; van Franeker, J.A. The use of potassium hydroxide (KOH) solution as a suitable approach to isolate plastics ingested by marine organisms. *Mar. Pollut. Bull.* **2017**, *115*, 86–90. [[CrossRef](#)] [[PubMed](#)]
24. Mbachu, O.; Jenkins, G.; Pratt, C.; Kaparaju, P. Enzymatic purification of microplastics in soil. *MethodsX* **2021**, *8*, 101254. [[CrossRef](#)] [[PubMed](#)]
25. Tagg, A.S.; Harrison, J.P.; Ju-Nam, Y.; Sapp, M.; Bradley, E.L.; Sinclair, C.J.; Ojeda, J.J. Fenton's reagent for the rapid and efficient isolation of microplastics from wastewater. *Chem. Commun.* **2017**, *53*, 372–375. [[CrossRef](#)]
26. Zobkov, M.; Belkina, N.; Kovalevski, V.; Zobkova, M.; Efremova, T.; Galakhina, N. Microplastic abundance and accumulation behavior in Lake Onego sediments: A journey from the river mouth to pelagic waters of the large boreal lake. *J. Environ. Chem. Eng.* **2020**, *8*, 104367. [[CrossRef](#)]
27. Hurley, R.R.; Lusher, A.L.; Olsen, M.; Nizzetto, L. Validation of a Method for Extracting Microplastics from Complex, Organic-Rich, Environmental Matrices. *Environ. Sci. Technol.* **2018**, *52*, 7409–7417. [[CrossRef](#)]
28. Munno, K.; Helm, P.A.; Jackson, D.A.; Rochman, C.; Sims, A. Impacts of temperature and selected chemical digestion methods on microplastic particles. *Environ. Toxicol. Chem.* **2018**, *37*, 91–98. [[CrossRef](#)]
29. Pfeiffer, F.; Fischer, E.K. Various Digestion Protocols Within Microplastic Sample Processing—Evaluating the Resistance of Different Synthetic Polymers and the Efficiency of Biogenic Organic Matter Destruction. *Front. Environ. Sci.* **2020**, *8*, 263. [[CrossRef](#)]
30. Enders, K.; Lenz, R.; Beer, S.; Stedmon, C.A. Extraction of microplastic from biota: Recommended acidic digestion destroys common plastic polymers. *ICES J. Mar. Sci.* **2017**, *74*, 326–331. [[CrossRef](#)]
31. Gulizia, A.M.; Brodie, E.; Daumuller, R.; Bloom, S.B.; Corbett, T.; Santana, M.M.F.; Motti, C.A.; Vamvounis, G. Evaluating the Effect of Chemical Digestion Treatments on Polystyrene Microplastics: Recommended Updates to Chemical Digestion Protocols. *Macromol. Chem. Phys.* **2022**, 2100485. [[CrossRef](#)]
32. Alfonso, M.B.; Takashima, K.; Yamaguchi, S.; Tanaka, M.; Isobe, A. Microplastics on plankton samples: Multiple digestion techniques assessment based on weight, size, and FTIR spectroscopy analyses. *Mar. Pollut. Bull.* **2021**, *173*, 113027. [[CrossRef](#)] [[PubMed](#)]
33. Kallenbach, E.M.F.; Hurley, R.R.; Lusher, A.; Friberg, N. Chitinase digestion for the analysis of microplastics in chitinous organisms using the terrestrial isopod *Oniscus asellus* L. as a model organism. *Sci. Total Environ.* **2021**, *786*, 147455. [[CrossRef](#)] [[PubMed](#)]
34. Dordevic, D.; Necasova, L.; Antonic, B.; Jancikova, S.; Tremlová, B. Plastic cutlery alternative: Case study with biodegradable spoons. *Foods* **2021**, *10*, 1612. [[CrossRef](#)] [[PubMed](#)]
35. Zhang, K.; Hamidian, A.H.; Tubić, A.; Zhang, Y.; Fang, J.K.H.; Wu, C.; Lam, P.K.S. Understanding plastic degradation and microplastic formation in the environment: A review. *Environ. Pollut.* **2021**, *274*, 116554. [[CrossRef](#)]
36. Lang, K.; Bhattacharya, S.; Ning, Z.; Sánchez-Leija, R.J.; Bramson, M.T.K.; Centore, R.; Corr, D.T.; Linhardt, R.J.; Gross, R.A. Enzymatic Polymerization of Poly(glycerol-1,8-octanediol-sebacate): Versatile Poly(glycerol sebacate) Analogues that Form Monocomponent Biodegradable Fiber Scaffolds. *Biomacromolecules* **2020**, *21*, 3197–3206. [[CrossRef](#)]
37. Chamas, A.; Moon, H.; Zheng, J.; Qiu, Y.; Tabassum, T.; Jang, J.H.; Abu-Omar, M.; Scott, S.L.; Suh, S. Degradation Rates of Plastics in the Environment. *ACS Sustain. Chem. Eng.* **2020**, *8*, 3494–3511. [[CrossRef](#)]
38. Lear, G.; Maday, S.D.M.; Gambarini, V.; Northcott, G.; Abbel, R.; Kingsbury, J.M.; Weaver, L.; Wallbank, J.A.; Pantos, O. Microbial abilities to degrade global environmental plastic polymer waste are overstated. *Environ. Res. Lett.* **2022**, *17*, 043002. [[CrossRef](#)]
39. Glaser, J.A. Biological Degradation of Polymers in the Environment. In *Plastics in the Environment*; IntechOpen: London, UK, 2019.
40. Fairbrother, A.; Hsueh, H.-C.; Kim, J.H.; Jacobs, D.; Perry, L.; Goodwin, D.; White, C.; Watson, S.; Sung, L.-P. Temperature and light intensity effects on photodegradation of high-density polyethylene. *Polym. Degrad. Stab.* **2019**, *165*, 153–160. [[CrossRef](#)]
41. Lyngsie, G.; Krumina, L.; Tunlid, A.; Persson, P. Generation of hydroxyl radicals from reactions between a dimethoxyhydroquinone and iron oxide nanoparticles. *Sci. Rep.* **2018**, *8*, 10834. [[CrossRef](#)]
42. Monteiro, S.S.; Rocha-Santos, T.; Prata, J.C.; Duarte, A.C.; Girão, A.V.; Lopes, P.; Cristovão, T.; da Costa, J.P. A straightforward method for microplastic extraction from organic-rich freshwater samples. *Sci. Total Environ.* **2022**, *815*, 152941. [[CrossRef](#)]
43. Grause, G.; Kuniyasu, Y.; Chien, M.-F.; Inoue, C. Separation of microplastic from soil by centrifugation and its application to agricultural soil. *Chemosphere* **2022**, *288*, 132654. [[CrossRef](#)] [[PubMed](#)]
44. Duong, T.T.; Le, P.T.; Nguyen, T.N.H.; Hoang, T.Q.; Ngo, H.M.; Doan, T.O.; Le, T.P.Q.; Bui, H.T.; Bui, M.H.; Trinh, V.T.; et al. Selection of a density separation solution to study microplastics in tropical riverine sediment. *Environ. Monit. Assess.* **2022**, *194*, 65. [[CrossRef](#)] [[PubMed](#)]
45. Pongstabodee, S.; Kunachitpimol, N.; Damronglerd, S. Combination of three-stage sink–float method and selective flotation technique for separation of mixed post-consumer plastic waste. *Waste Manag.* **2008**, *28*, 475–483. [[CrossRef](#)] [[PubMed](#)]

46. Cashman, M.A.; Ho, K.T.; Boving, T.B.; Russo, S.; Robinson, S.; Burgess, R.M. Comparison of microplastic isolation and extraction procedures from marine sediments. *Mar. Pollut. Bull.* **2020**, *159*, 111507. [[CrossRef](#)]
47. Nguyen, B.; Claveau-Mallet, D.; Hernandez, L.M.; Xu, E.G.; Farner, J.M.; Tufenkji, N. Separation and Analysis of Microplastics and Nanoplastics in Complex Environmental Samples. *Acc. Chem. Res.* **2019**, *52*, 858–866. [[CrossRef](#)]
48. Brewer, A.; Dror, I.; Berkowitz, B. The Mobility of Plastic Nanoparticles in Aqueous and Soil Environments: A Critical Review. *ACS ES&T Water* **2021**, *1*, 48–57. [[CrossRef](#)]
49. He, J.; Liu, W.; Huang, Y.-X. Simultaneous Determination of Glass Transition Temperatures of Several Polymers. *PLoS ONE* **2016**, *11*, e0151454. [[CrossRef](#)]
50. Catarino, A.I.; Thompson, R.; Sanderson, W.; Henry, T.B. Development and optimization of a standard method for extraction of microplastics in mussels by enzyme digestion of soft tissues. *Environ. Toxicol. Chem.* **2017**, *36*, 947–951. [[CrossRef](#)]
51. Treilles, R.; Cayla, A.; Gaspéri, J.; Strich, B.; Ausset, P.; Tassin, B. Impacts of organic matter digestion protocols on synthetic, artificial and natural raw fibers. *Sci. Total Environ.* **2020**, *748*, 141230. [[CrossRef](#)]
52. Hu, K.; Zhou, P.; Yang, Y.; Hall, T.; Nie, G.; Yao, Y.; Duan, X.; Wang, S. Degradation of Microplastics by a Thermal Fenton Reaction. *ACS ES&T Eng.* **2021**, *2*, 110–120. [[CrossRef](#)]
53. Nuelle, M.-T.; Dekiff, J.H.; Remy, D.; Fries, E. A new analytical approach for monitoring microplastics in marine sediments. *Environ. Pollut.* **2014**, *184*, 161–169. [[CrossRef](#)] [[PubMed](#)]
54. Duan, J.; Han, J.; Zhou, H.; Lau, Y.L.; An, W.; Wei, P.; Cheung, S.G.; Yang, Y.; Tam, N.F. Development of a digestion method for determining microplastic pollution in vegetal-rich clayey mangrove sediments. *Sci. Total Environ.* **2020**, *707*, 136030. [[CrossRef](#)] [[PubMed](#)]
55. Fotopoulou, K.N.; Karapanagioti, H.K. Degradation of Various Plastics in the Environment. In *Hazardous Chemicals Associated with Plastics in the Marine Environment*; Barcelò, D., Kostianoy, A.G., Eds.; The Handbook of Environmental Chemistry; Springer: Cham, Switzerland; New York, NY, USA, 2017; Volume 78, pp. 71–92. [[CrossRef](#)]
56. McGivney, E.; Cederholm, L.; Barth, A.; Hakkarainen, M.; Hamacher-Barth, E.; Ogonowski, M.; Gorokhova, E. Rapid Physico-chemical Changes in Microplastic Induced by Biofilm Formation. *Front. Bioeng. Biotechnol.* **2020**, *8*, 205. [[CrossRef](#)] [[PubMed](#)]
57. Fernández-González, V.; Andrade-Garda, J.M.; López-Mahía, P.; Muniategui-Lorenzo, S. Impact of weathering on the chemical identification of microplastics from usual packaging polymers in the marine environment. *Anal. Chim. Acta* **2021**, *1142*, 179–188. [[CrossRef](#)] [[PubMed](#)]
58. Prata, J.C.; Reis, V.; Paço, A.; Martins, P.; Cruz, A.; da Costa, J.P.; Duarte, A.C.; Rocha-Santos, T. Effects of spatial and seasonal factors on the characteristics and carbonyl index of (micro)plastics in a sandy beach in Aveiro, Portugal. *Sci. Total Environ.* **2020**, *709*, 135892. [[CrossRef](#)]
59. Rodrigues, M.O.; Abrantes, N.; Gonçalves, F.J.M.; Nogueira, H.; Marques, J.C.; Gonçalves, A.M.M. Spatial and temporal distribution of microplastics in water and sediments of a freshwater system (Antuã River, Portugal). *Sci. Total Environ.* **2018**, *633*, 1549–1559. [[CrossRef](#)]
60. Brandon, J.; Goldstein, M.; Ohman, M.D. Long-term aging and degradation of microplastic particles: Comparing in situ oceanic and experimental weathering patterns. *Mar. Pollut. Bull.* **2016**, *110*, 299–308. [[CrossRef](#)]
61. Miranda, M.N.; Sampaio, M.J.; Tavares, P.B.; Silva, A.M.T.; Pereira, M.F.R. Aging assessment of microplastics (LDPE, PET and uPVC) under urban environment stressors. *Sci. Total Environ.* **2021**, *796*, 148914. [[CrossRef](#)]
62. Xu, J.L.; Thomas, K.V.; Luo, Z.; Gowen, A.A. FTIR and Raman imaging for microplastics analysis: State of the art, challenges and prospects. *TrAC Trends Anal. Chem.* **2019**, *119*, 115629. [[CrossRef](#)]
63. Liu, P.; Qian, L.; Wang, H.; Zhan, X.; Lu, K.; Gu, C.; Gao, S. New Insights into the Aging Behavior of Microplastics Accelerated by Advanced Oxidation Processes. *Environ. Sci. Technol.* **2019**, *53*, 3579–3588. [[CrossRef](#)]
64. Gulmine, J.V.; Akcelrud, L. FTIR characterization of aged XLPE. *Polym. Test.* **2006**, *25*, 932–942. [[CrossRef](#)]
65. Lacoste, J.; Carlsson, D.J. Gamma-, photo-, and thermally-initiated oxidation of linear low density polyethylene: A quantitative comparison of oxidation products. *J. Polym. Sci. Part A Polym. Chem.* **1992**, *30*, 493–500. [[CrossRef](#)]
66. Socrates, G. *Infrared and Raman Characteristic Group Frequencies: Tables and Charts—George Socrates—Google Libri*, 3rd ed.; Chichester, E., Ed.; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2004.
67. Veerasingam, S.; Ranjani, M.; Venkatachalapathy, R.; Bagaev, A.; Mukhanov, V.; Litvinyuk, D.; Mugilarasan, M.; Gurumoorthi, K.; Gunganathan, L.; Aboobacker, V.M.; et al. Contributions of Fourier transform infrared spectroscopy in microplastic pollution research: A review. *Crit. Rev. Environ. Sci. Technol.* **2020**, *51*, 2681–2743. [[CrossRef](#)]