



Published in final edited form as:

J Invest Dermatol. 2013 October ; 133(10): 2489–2492. doi:10.1038/jid.2013.166.

Over-Expression of the Gastrin-Releasing Peptide in Cutaneous Nerve Fibers and its Receptor in Spinal Cord in Primates with Chronic Itch

Leigh A. Nattkemper^{1,2}, Zhong-Qiu Zhao⁵, Anna J. Nichols⁶, Alexandru D.P. Papoiu¹, Carol A. Shively⁴, Zhou-Feng Chen⁵, and Gil Yosipovitch^{1,2,3,*}

¹Department of Dermatology, Wake Forest University School of Medicine, Winston Salem, NC, USA

²Department of Neurobiology & Anatomy, University Wake Forest School of Medicine, Winston Salem, NC, USA

³Department of Regenerative Medicine, Wake Forest University School of Medicine, Winston Salem, NC, USA

⁴Department of Comparative Medicine, Wake Forest University School of Medicine, Winston Salem, NC, USA

⁵Center for the Study of Itch, Washington University School of Medicine, St. Louis, MO, USA

⁶Department of Neuroscience and Physiology, SUNY Upstate Medical University, Syracuse, NY, USA

To the Editor

Chronic pruritus affects millions of patients worldwide and has a significant impact on quality of life similar to that of chronic pain (Kini *et al.*, 2011; Stander *et al.*, 2007). Significant advances have been made in the last five years to elucidate the molecular pathways of acute itch (Liu *et al.*, 2009; Sun and Chen, 2007). However, experimental approaches investigating the pathogenesis of pruritus and the ability to test novel therapeutic agents are largely limited to rodent models. Although these models offer some advantages, their translational potential to human disease remains to be established (Seok *et al.*, 2013; Jeffry *et al.*, 2011). Furthermore, most animal models focus on acute itch, which displays significant pathophysiological differences in comparison with chronic itch (Yosipovitch *et al.*, 2007). Therefore, there is an unmet need for better animal models for chronic itch research.

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

*Correspondence to: Gil Yosipovitch, Department of Dermatology, Neurobiology & Anatomy, and Regenerative Medicine, Wake Forest University Health Sciences, Winston-Salem, NC 27157, Phone: (336) 716-2901, Fax: (336) 716-7732, gyosipov@wakehealth.edu.

Conflict of Interests:

The authors state no conflict of interests.

Recently, gastrin-releasing peptide (GRP) and its receptor (GRPR) were discovered to play a key role in itch transmission, but not in nociception (Sun and Chen, 2007). GRP was found in a subset of unmyelinated dorsal root ganglion (DRG) neurons, while GRPR was expressed in lamina I of the dorsal horn of the spinal cord in mice. In the skin, GRP is present in primary afferent nerve fibers and found to be increased in mice with chronic dermatitis (Tominaga *et al.*, 2009). The GRP/GRPR signaling pathway is considered to be the first molecular pathway specific to itch transmission, but there is no evidence that this pathway has an analogous role in primates or humans.

We identified a subgroup of adult female *Cynomolgus* macaques (*Macaca fascicularis*) suffering from idiopathic chronic itch and observed their scratching behaviors twice every week for 10 minutes for four years using a focal observation technique (Altmann, 1974). The population pattern of GRP and GRPR expression in the skin and spinal cord were assessed in a blinded manner, respectively. The expression of GRP and GRPR was quantified by immunohistochemistry and analyzed with a 1-way ANOVA and Bonferroni post hoc test, while an Unpaired t-test was used to compare the amount of PGP9.5⁺ nerve fibers between groups. The scratching behavior of each animal was then analyzed for potential correlations with GRP and GRPR expression levels using a 2-tailed Spearman correlation test and linear regression.

The frequency (number of scratching episodes per hour) and duration (percent time of scratching during focal observation) of scratching were recorded over four years, totaling 68 hours of observation per animal. The animals were euthanized and histological sections from six randomly selected animals with different itch severities were collected. For each primate, the total count of nerve fibers expressing GRP in lichenified (chronically damaged skin lesion) and non-lichenified skin from thoracic dermatomes were correlated with itch severity. Skin cryosections (20 μ m thickness) were double stained with antibodies for GRP (Immunostar, Hudson, WI) and the neuronal marker Product Gene Protein 9.5 (PGP9.5; Neuromics, Edina, MN) to examine the amount of co-localization at the dermal-epidermal junction. Although fibers were found in both the epidermis and dermis, they were mainly present in the dermal-epidermal junction so all quantification was focused on this area. The population of GRPR expressing cells in the dorsal horn of the thoracic spinal cord of the same animals was also analyzed for a potential correlation to itch intensity by staining the spinal cord cryosections with the GRPR antibody (MBL International, Woburn, MA). For a detailed methods section, see supplementary information.

Primates that exhibited a comparatively higher severity of itching consistently displayed an increased percentage of GRP expressing nerve fibers at the dermal-epidermal junction of lichenified skin ($p=0.03$; Figure 1a & e). The mean percentage of GRP⁺/PGP9.5⁺ fibers was 4.8 fold higher in primates exhibiting the highest itch intensity than in primates with the lowest intensity. The frequency (Figure 1b) and duration (Figure 1c) of scratching were significantly correlated with the percentage of GRP in lichenified skin ($r=0.94$, $p=0.02$; for both frequency and duration), but not in non-lichenified skin ($r=0.77$, $p=0.10$; for both frequency and duration). The innervation pattern of PGP9.5⁺ fibers in the skin show no significant difference of fibers in animals with severe itch compared to those with mild/moderate itch (Figure 1d).

Additionally, an increased amount of GRPR expressing cells was observed in the dorsal horn of the spinal cord of primates exhibiting a higher itch severity compared to animals with lower itch severity ($p=0.002$; Figure 2a & d). The frequency (Figure 2b) and duration (Figure 2c) of scratching was also found to be significantly correlated with the expression of GRPR in the superficial lamina I & II ($r=0.94$, $p=0.02$; for both frequency and duration), but only trended toward significance in the deep lamina III–V ($r=0.83$, $p=0.06$; for both frequency and duration) of the dorsal horn of the thoracic spinal cord. The number of GRP⁺ fibers in lichenified skin also significantly correlated to the amount of staining of GRPR in the superficial ($r=0.94$, $p=0.01$) and deep ($r=0.89$, $p=0.03$) lamina of the dorsal horn of each primate (Fig. 2e).

GRP and its ligand GRP are highly expressed in the spinal cord and skin of primates with chronic itch, which to our knowledge is previously unreported. It is consistent with previous finding in non-chronic itch mice models (Andoh *et al.*, 2011; Sun and Chen, 2007; Sun *et al.*, 2009). Therefore, the GRP/GRPR system is a promising drug target for the treatment of chronic pruritus in humans. Furthermore, this subset of female *Cynomolgus* macaques offers a novel model of itch that could better represent chronic itch in humans. This would allow for long term follow up studies assessing other mediators involved in chronic itch.

Acknowledgments

The authors wish to thank Dr. Carol Milligan and members of her lab for providing invaluable assistance. Dr. Chen is funded by NIH NIAMS grant 1R01AR056318. Dr. Shively is funded by NIH grant R01 HL08713. Dr. Yosipovitch is funded by NIH NIAMS grant R01 55902-4.

Abbreviations

GRP	gastrin-releasing peptide
GRPR	gastrin-releasing peptide receptor
PGP9.5	product gene protein 9.5

References

1. Altmann J. Observational study of behavior: sampling methods. *Behaviour*. 1974; 48:227–67. [PubMed: 4597405]
2. Andoh T, Kuwazono T, Lee JB, et al. Gastrin-releasing peptide induces itch-related responses through mast cell degranulation in mice. *Peptides*. 2011; 32(10):2098–103. [PubMed: 21933692]
3. Jeffrey J, Kim S, Chen ZF. Itch signaling in the nervous system. *Physiology (Bethesda)*. 2011; 26(4): 286–92. [PubMed: 21841076]
4. Kini SP, DeLong LK, Veledar E, et al. The impact of pruritus on quality of life: the skin equivalent of pain. *Arch Dermatol*. 2011; 147(10):1153–6. [PubMed: 21680760]
5. Liu Q, Tang Z, Surdenikova L, et al. Sensory neuron-specific GPCR Mrgprs are itch receptors mediating chloroquine-induced pruritus. *Cell*. 2009; 139(7):1353–65. [PubMed: 20004959]
6. Seok J, Warren HS, Cuenca AG, et al. Genomic responses in mouse models poorly mimic human inflammatory diseases. *PNAS*. 2013; 110(9):3507–3512. [PubMed: 23401516]
7. Ständer S, Weisshaar E, Mettang T, et al. Clinical classification of itch: a position paper of the International Forum of the Study of Itch. *Acta Derm Venereol*. 2007; 87(4):291–4. [PubMed: 17598029]

8. Sun YG, Chen ZF. A gastrin-releasing peptide receptor mediates the itch sensation in the spinal cord. *Nature*. 2007; 448(7154):700–3. [PubMed: 17653196]
9. Sun YG, Zhao ZQ, Meng XL, et al. Cellular basis of itch sensation. *Science*. 2009; 325(5947): 1531–4. [PubMed: 19661382]
10. Tominaga M, Ogawa H, Takamori K. Histological characterization of cutaneous nerve fibers containing gastrin-releasing peptide in NC/Nga mice: an atopic dermatitis model. *J Invest Dermatol*. 2009; 129:2901–5. [PubMed: 19571818]
11. Yosipovitch G, Carstens E, McGlone F. Chronic itch and chronic pain: Analogous mechanisms. *Pain*. 2007; 131(1–2):4–7. [PubMed: 17524558]

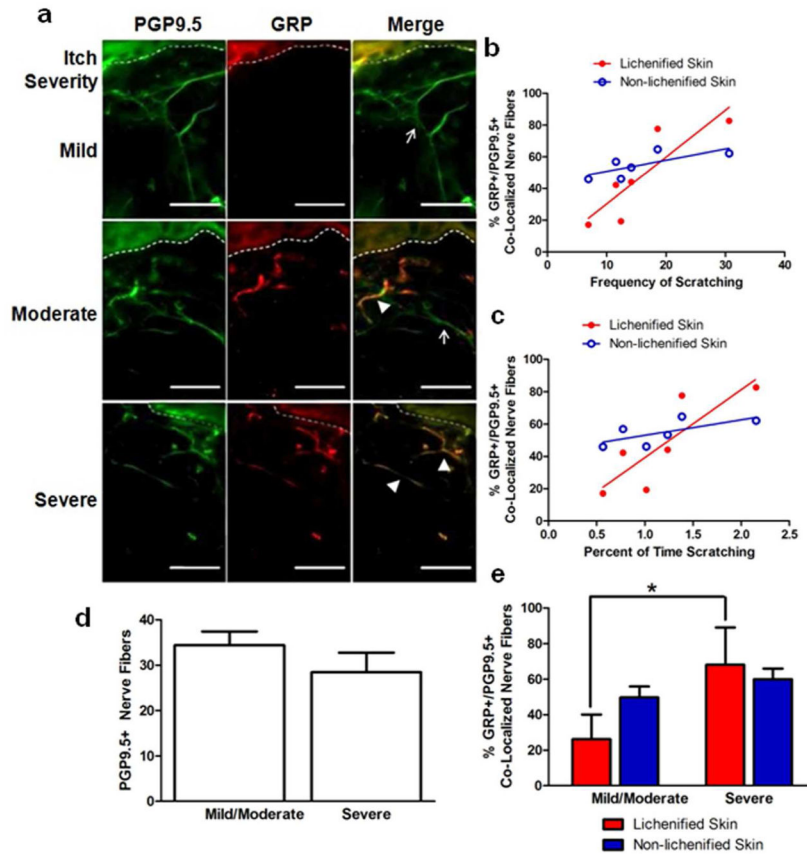


Figure 1. Immunohistochemical staining of GRP⁺ nerve fibers in the skin of primates with chronic itch

(a) Double-labeling of PGP9.5 (green) and GRP (red) in lichenified skin of primates representing mild, moderate, and severe itch intensities. Primates with higher scratching severity showed an increase of co-localization (yellow; arrowhead) of GRP in PGP9.5⁺ fibers at the dermal-epidermal junction compared to primates with lower scratching severity (not co-localized; arrow). The border between the epidermis and dermis is indicated with a dashed line (- -). Scale bars = 56 μ m. The mean percent of double labeled GRP⁺/PGP9.5⁺ fibers at the dermal-epidermal junction in lichenified skin but not in non-lichenified skin significantly correlate with the frequency of scratching (b) and the percent time spent scratching (c). The amount of PGP9.5⁺ nerve fibers do not change between itch severities (d), while the expression of GRP significantly increases with high itch severity when compared to mild/moderate severity in lichenified skin (e) (n=3 per group; *p<0.05).

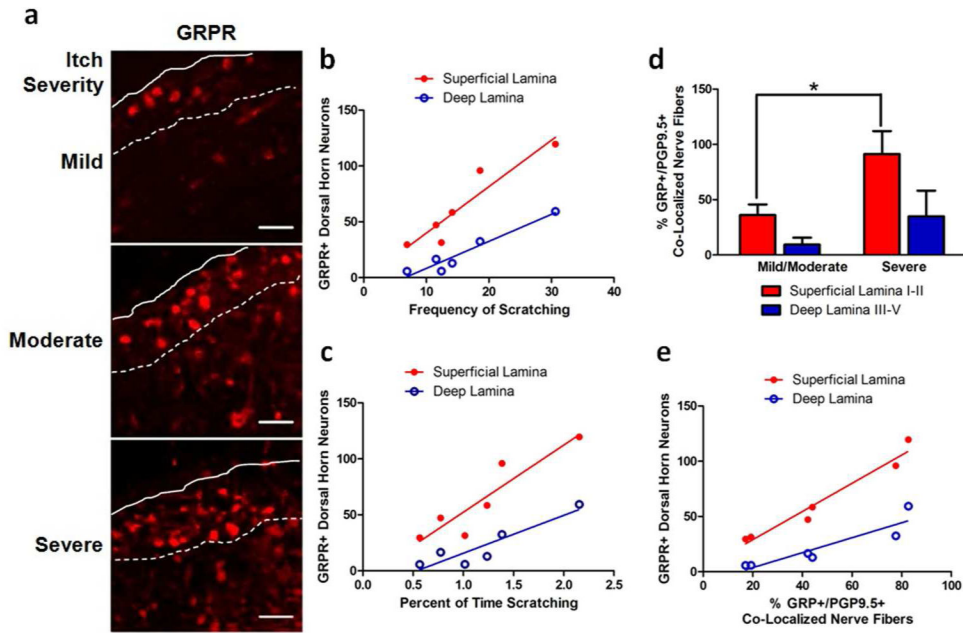


Figure 2. Immunohistochemical staining of GRPR⁺ cells in the dorsal horn of primates with chronic itch

(a) Labeling of GRPR⁺ (red) cells in the dorsal horn of the spinal cord of primates representing mild, moderate, and severe itch severities. A significant increase of GRPR⁺ cells was found in the superficial lamina but not in the deep lamina of primates exhibiting severe scratching. Superficial and deep lamina are separated with a dashed line (- -) and the dorsal horn surface is indicated with a solid line (-). Scale bars = 56µm. The number of labeled GRPR⁺ cells in the superficial lamina of the dorsal horn correlate with the frequency of scratching (b) and the percent time spent scratching (c). (d) The number of GRPR⁺ neurons significantly increases with severe itch intensity compared to mild/moderate intensity in superficial lamina (n=3 per group; *p<0.05). (e) The percentage of GRP⁺ fibers in lichenified skin significantly correlated to the amount of GRPR⁺ cells in the dorsal horn for each primate.