

Transcriptomics-Based Investigation of Molecular Mechanisms Underlying Apoptosis Induced by ZnO Nanoparticles in Human Diffuse Large B-Cell Lymphoma

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Introduction: Zinc oxide nanoparticles (ZnO NPs) show anti-cancer activity. Diffuse Large B-cell Lymphoma (DLBCL) is a type of B-cell malignancies with unsatisfying treatment outcomes. This study was set to assess the potential of ZnO NPs to selectively induce apoptosis in human DLBCL cells (OCI-LY3), and to describe possible molecular mechanisms of action.

Methods: The impact of ZnO NPs on DLBCL cells and normal peripheral blood mononuclear cells (PBMCs) was studied using cytotoxicity assay and flow-cytometry. Transcriptomics analysis was conducted to identify ZnO NPs-dependent changes in the transcriptomic profiles of DLBCL cells.

Results: ZnO NPs selectively induced apoptosis in DLBCL cells, and caused changes in their transcriptomes. Differential gene expression (DGE) with fold change (FC) ≥ 3 and $p \leq 0.008$ with corrected $p \leq 0.05$ was identified for 528 genes; 125 genes were over-expressed and 403 genes were under-expressed in ZnO NPs-treated DLBCL cells. The over-expressed genes involved in biological processes and pathways like stress response to metal ion, cellular response to zinc ion, metallothioneins bind metals, oxidative stress, and negative regulation of growth. In contrast, the under-expressed genes were implicated in DNA packaging complex, signaling by NOTCH, negative regulation of gene expression by epigenetic, signaling by WNT, M phase of cell cycle, and telomere maintenance. Setting the FC to ≥ 1.5 with $p \leq 0.05$ and corrected $p \leq 0.1$ showed ZnO NPs to induce over-expression of anti-oxidant genes and under-expression of oncogenes; target B-cell receptor (BCR) signaling pathway and NF- κ B pathway; and promote apoptosis by intrinsic and extrinsic pathways.

Discussion: Overall, ZnO NPs selectively induced apoptosis in DLBCL cells, and possible molecular mechanisms of action were described.

Keywords: transcriptomics, ZnO NPs, DLBCL, apoptosis, mechanism of action

Introduction

A growing body of evidence has shown a therapeutic potential of nanoparticles (NPs) against a wide range of malignant cells.¹ Previous research by us and others have shown a preferential killing of cancer cells, such as acute promyelocytic leukemia cells (HL-60),² cervical carcinoma cells (HeLa),³ hepatocellular carcinoma cells (HepG2),^{4,5} lung cancer cells (A549),^{5,6} and breast cancer cells (MDA-MB-231)⁷ by metal NPs like gold NPs and silver NPs. Furthermore, earlier work on zinc oxide (ZnO) NPs, which are another example of metal NPs, have demonstrated the capability of the ZnO NPs to selectively induce apoptosis in cancer cells like acute promyelocytic leukemia cells (HL60) and chronic myeloid leukemia cells (K562) with almost no impact on the normal peripheral blood mononuclear cells (PBMCs).^{8,9} Similar results were found in breast cancer cells (MCF7),¹⁰ colon cancer cells (Caco-2),¹¹ hepatocellular carcinoma cells (HepG2),¹² lung adenocarcinoma (A549),¹² ovarian cancer cells (SKOV3)¹³ and cervical carcinoma cells (HeLa).¹⁴ Despite the extensive research on the therapeutic potential of ZnO NPs against cancer cells, the molecular mechanism of apoptosis induction by

ZnO NPs has not yet been fully illustrated. Nevertheless, the cytotoxicity of ZnO NPs has been related to the disequilibrium of zinc-dependent protein activity and to overproduction of reactive oxygen species (ROS) in the treated cells.^{15–18}

Transcriptomics is a high-throughput approach that enables a global gene expression profiling of all human coding genes in a single assay.¹⁹ Therefore, transcriptomics is a promising approach to provide insights into the molecular basis of malignant tumors and putative mechanism of NPs cytotoxicity against cancer cells.^{20–23} In this context, different investigators sought alterations induced by ZnO NPs in the transcriptome of cancer cells, such as lung cancer cells (A549),²⁴ colon carcinoma cells (RKO),²⁵ colorectal adenocarcinoma cells (Caco-2),²⁵ melanoma cells (SK Mel-28),²⁵ acute monocytic leukemia cells (THP-1)²⁶ and acute T-cell leukemia cells (Jurkat A3).²⁷ Nevertheless, as indicated by the search engines of “PubMed” and “Google Scholar” using the following key words “ZnO NPs gene expression and diffuse large B-cell lymphoma cells” or “ZnO NPs transcriptomics and diffuse large B-cell lymphoma cells”, and to the best of our knowledge, no transcriptomics work on diffuse large B-cell lymphoma cells (DLBCL) treated with ZnO NPs was reported before.

DLBCL is the most prevalent type of lymphomas seen in adults.²⁸ Despite the improvements that have been made in the new modalities of DLBCL therapy, the disease still follows an aggressive clinical course with unsatisfactory treatment outcomes, as large proportion of DLBCL patients shows recurrence or refractory disease.^{29,30} Achieving full remission in recurrence or refractory patients have been shown to be challenging, putting patients at high-risk of death.³¹ In fact, the five years overall survival of DLBCL patients was reported to be 50%, especially in patients diagnosed with the subtype known as activated B-cell like (ABC), in which relapse and refractory are frequently reported.³² Therefore, there has been a great and urgent need for a new modality of therapy to improve the treatment outcomes of the disease.

Given the selective cytotoxicity of ZnO NPs against a wide range of cancer cells as reported above, the present work was set to investigate the potential of ZnO NPs to induce apoptosis in human DLBCL cell line (OCI-LY3), which has been established from a relapsed DLBCL (subtype ABC) patient with advanced stage of the disease, and has been known for its resistance to standard chemotherapies.³³ Next, molecular mechanisms of action of ZnO NPs cytotoxicity on DLBCL cells were studied using transcriptomics. The expected findings of the current work would possibly advance our understanding about the therapeutic potential of ZnO NPs against the ABC subtype of DLBCL, and may form the fundamentals on which ZnO NPs-based clinical applications could be developed to overcome the refractory feature of this disease.

Materials and Methods

Cell Culture

Human DLBCL cells (OCI-LY3 cell line) were obtained from German Collection of Microorganisms and Cell Cultures GmbH (DSMZ). OCI-LY3 cells were cultivated in RPMI 1649 medium with 10% fetal bovine serum (FBS), 40 µg/mL gentamycin, 100 µg/mL streptomycin sulphate, 100 U/mL penicillin, 4.5 mg/mL glucose and 2 mg/mL sodium bicarbonate under humidified condition containing 5% CO₂ at 37 °C. Blood samples of healthy donors were collected in accordance with the granted ethical approval from the Ethical Committee of Majmaah University in Saudi Arabia (MUREC-April.01/COM-206) and informed consents were obtained from all the participants. This work also adhered to the principles of the Declaration of Helsinki. Density-gradient centrifugation with lymphoprep were utilized for the isolation of peripheral blood mononuclear cells (PBMCs) from blood of healthy subjects. PBMCs were stimulated in RPMI 1640 medium supplemented with 10% FBS, 5 µg/mL phytohemagglutinin, 100 U/mL penicillin, 40 µg/mL gentamycin, 100 µg/mL streptomycin sulphate, 4.5 mg/mL glucose and 2 mg/mL sodium bicarbonate for 3 days at 37 °C under an atmosphere of humidified air containing 5% CO₂. Next, the PBMCs were transferred to a medium as the above but which lacked phytohemagglutinin and contained 5 ng/mL of interleukin 2 and were incubated at 37 °C in a CO₂ incubator for subsequent analysis.

Cytotoxicity Assay

The preparation and characterization of the ZnO NPs (Sigma-Aldrich #721077) used in the present study were previously reported by Triboulet et al.³⁴ To assess the impact of ZnO NPs against DLBCL cells MTT assay was used. Suspension of DLBCL cells (3×10^5 cells/mL) in RPMI 1640 medium supplemented with 10% FBS were seeded into 96-well culture plate (200 µL/ well)

with different concentration of ZnO NPs (0 µg/mL, 10 µg/mL, 20 µg/mL, 30 µg/mL, 40 µg/mL, 50 µg/mL, 60 µg/mL, 70 µg/mL and 80 µg/mL). The 96-well culture plate was then incubated for five days (120 hours) at 37 °C in a CO₂ incubator with no change of the culture medium.

The toxicity of the ZnO NPs on normal PBMCs compared with DLBCL cells was also studied using MTT assay. Suspensions of PBMCs (3×10^5 cells/mL) in culture medium (RPMI 1640 with 10% FBS and 5 ng/mL interleukin 2) and suspensions of DLBCL cells (3×10^5 cells/mL) in culture medium (RPMI 1640 supplemented with 10% FBS) were separately seeded into a 96-well culture plate (200 µL/well) containing different concentrations of ZnO NPs (0 µg/mL, 10 µg/mL, 20 µg/mL, and 30 µg/mL). Next, the cells were continuously incubated for 5 days (120 hours) at 37 °C with the presence of CO₂ with no change of the culture medium.

Post incubation, MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenylt×10 trazolium bromide] assay was utilized to evaluate the viability of the DLBCL cells and PBMCs. Briefly, the medium was carefully removed, and a new medium (RPMI 1640) with FBS (10%) and MTT dye (final concentration 0.5 mg/mL) was added to the 96-well culture plate that was incubated for 3 hours at 37 °C. Next, the medium was removed and the formazan crystals were dissolved in DMSO (200 µL). Absorbance microplate reader was then used to measure the reduced levels of MTT at wavelength = 570 nm.³⁵ All experiments were conducted three times independently.

Identification of the Type of Cell Death

To determine the type of killing of the DLBCL cells induced by the ZnO NPs, the DLBCL cells in RPMI 1640 medium with 10% FBS (3×10^5 cells/mL) were incubate in a 6-well plate with and without ZnO NPs (30 µg/mL) for 24 hours at 37 °C in CO₂ incubator. The type of cell death was investigated using FACS CantoII flow cytometer (Becton Dickinson) with an Annexin V-FITC Kit (Trevigen, Inc.). Briefly, a washing step of the malignant cells (3×10^5 cells/mL) with cold 1X phosphate buffered saline (1X PBS) followed by centrifugation for 5 minutes at 300×g was done. Next, a dye solution (fluorescein-labeled Annexin V, propidium iodide and 1X binding buffer) was used to stain the cells for 15 minutes at room temperature in dark. Next, the stained cells were transferred into a FACS tube and 1X binding buffer was added to them. Finally, the cells were injected into FACS CantoII flow cytometer (Becton Dickinson) and analyzed using FACSDiva software (Becton Dickinson). The experiments were performed three times independently.

Determination of ROS Levels

ROS levels in the DLBCL cells with and without ZnO NPs treatment was measured using DCFH-DA (2',7'-dichlorodihydrofluorescein diacetate). Suspensions of DLBCL cells (3×10^5 cells/mL) in culture medium (RPMI 1640 supplemented with 10% FBS) were cultivated in a 96-well culture plate (200 µL/well) containing different concentrations of ZnO NPs (0 µg/mL, 30 µg/mL, 40 µg/mL, 50 µg/mL) for 24 hours at 37 °C with the presence of CO₂. Post incubation, the cells were washed twice with PBS and were treated with the DCFH-DA (20 µM) and incubated as before for 1 hour. Next, the dichlorofluorescein (DCF) intensity was measured using microplate reader (Synergy-4) with an excitation wavelength = 338nm and emission wavelength = 500nm. The calculation of DCF intensity in the ZnO NPs treated cells was relative to that of the control cells (ZnO NPs = 0 µg/mL); and the final results were expressed in percentage. The experiments were conducted three times independently.

RNA Extraction and Quality Control

The reagents and the manufacturer's protocol of RNeasy Mini Kit (QIAGEN) were used to extract total RNA from the DLBCL cells. NanoDrop ND-1000 spectro-photometer (Thermo Fisher Scientific, Inc.) was then utilized to measure the purity and quantity of the isolated RNA; use of RNA samples was limited to those with high purity (260 nm/280 nm ≥ 2). Similarly, the RNA Nano 6000 chips with the Agilent 2100 Bioanalyzer were used evaluate the integrity of the RNA samples according to the manufacturer's instructions (Agilent Technologies, Inc.) RNA samples used for subsequent experiments had good integrity (RNA Integrity Number (RIN) ≥ 8).

Transcriptomics Profiling

The Agilent protocol of “one-color microarray-based gene expression analysis” was followed to study the transcriptomic profile of the DLBCL cells. Briefly, 100 ng of the isolated RNA from the DLBCL cells along with the RNA spikes from RNA Spike-In Kit, One-Color (Agilent) were converted to cDNA. Next, low input quick amp labeling (one-color) kit (Agilent) was used to transcribe and amplify the cDNA to fluorescent-labeled cRNA that was purified using the materials and protocol of RNeasy Mini Kits (QIAGEN). To assess the quality and quantity of the labeled cRNA, nanoDrop ND-1000 spectrophotometer (Thermo Fisher Scientific, Inc.) was utilized. All samples proceeded to the next step showed: 260 nm/280 nm ≥ 2 , cRNA yield ≥ 825 ng and specific activity (pmol Cy3/ μ g cRNA) ≥ 6 . To prepare the hybridization samples, 600 ng of the labeled cRNA was mixed with 5 μ L of 10X Gene Expression Blocking Agent (Agilent) and 1 μ L of 25X Fragmentation Buffer (Agilent), making a total volume of 25 μ L. The labeled cRNA in the hybridization samples was then fragmented by incubation at 60°C for 30 min. To stop the fragmentation, 25 μ L of 2X Hi-RPM hybridization Buffer (Agilent) was added to the pre-cooled hybridization samples (total volume = 50 μ L). Next, 40 μ L from each hybridization sample was added onto a gasket that was assembled to SurePrint G3 Human Gene Expression 8 \times 60K Microarray slide (Agilent) and incubated for 17 hours at 65°C. Post incubation, a washing step using the Gene Expression Washing Buffer 1 and 2, to which 0.005% Triton X-102 was added (Agilent), was performed for the microarray slides. After that, the Agilent SureScan Microarray Scanner using the AgilentG3_GX_1color protocol was employed to scan the microarray slides. Next, Agilent Feature Extraction Software (version 11.0.1.1; Agilent Technologies, Inc.) was used to extract the quantitative data of probe features from the scanned microarray slides. The quantitative data were then saved for further analysis.

Data Analysis

The quantitative data of probe features that were saved in previous section were loaded into the GeneSpring GX version 12.1 software package (Agilent Technologies, Inc.). The log₂ transformation and normalization (percentile shift algorithm with percentile target = 75) were applied to the quantitative data in order to generate normalized signal value of the probes. The analysis was limited to probe quantitative data with the flag labels: “detected” or “not detected”. Two conditions were made; treated condition (ZnO NPs-treated samples; n= 3) and untreated condition (untreated samples (control samples); n= 3). Probes were filtered according to their flag label; only probes with flag “detected” in all samples of either condition (treated condition or untreated condition) were used for the further analysis. Next, fold change (FC) was used to identify differentially expressed genes (DEGs) in the two groups of samples (the two conditions). To identify the most evident DEGs in the two conditions, FC was set to 3-fold or higher with $p \leq 0.008$ and corrected $p \leq 0.05$. Next, the FC cut-off was reduced to ≥ 1.5 -fold with $p \leq 0.05$ and corrected $p \leq 0.1$ in attempt to broadly investigate ZnO NPs-induced transcriptomics alteration. Cluster analysis and heatmap presentations of DEGs were conducted using hierarchical algorithm and were based on probes (normalized signal value of probes) and conditions with Euclidean method for similarity measure and Wards for linkage rule. Gene ontology (GO) enrichment analyses were conducted using the GeneSpring GX version 12.1 software package (Agilent Technologies, Inc.) and only over-represented GO terms with corrected $p \leq 0.05$ were reported. Functional profiling the DEGs was also performed using the gProfiler³⁶ (<https://biit.cs.ut.ee/gprofiler/gost>) against GO and biological pathways (KEGG pathway database, Reactome pathways database and WikiPathways database); the option “only annotated genes” was selected for statistical domain scope and corrected p-value cut-off was set at ≤ 0.05 .

Statistical Analysis

The significance values (p values) were determined using paired Student’s *t*-test; and corrected p values were calculated on the basis of Benjamini–Hochberg false discovery rate (FDR). GeneSpring GX version 12.1 software package (Agilent Technologies, Inc.) was used for the generation of the p values and corrected p values of DEGs. Excel software was used to calculate the p values of the viability assay and the analyses titled identification of the type of cell death. Excel software was also utilized to calculate the means and standard deviations; and to construct the column graphs.

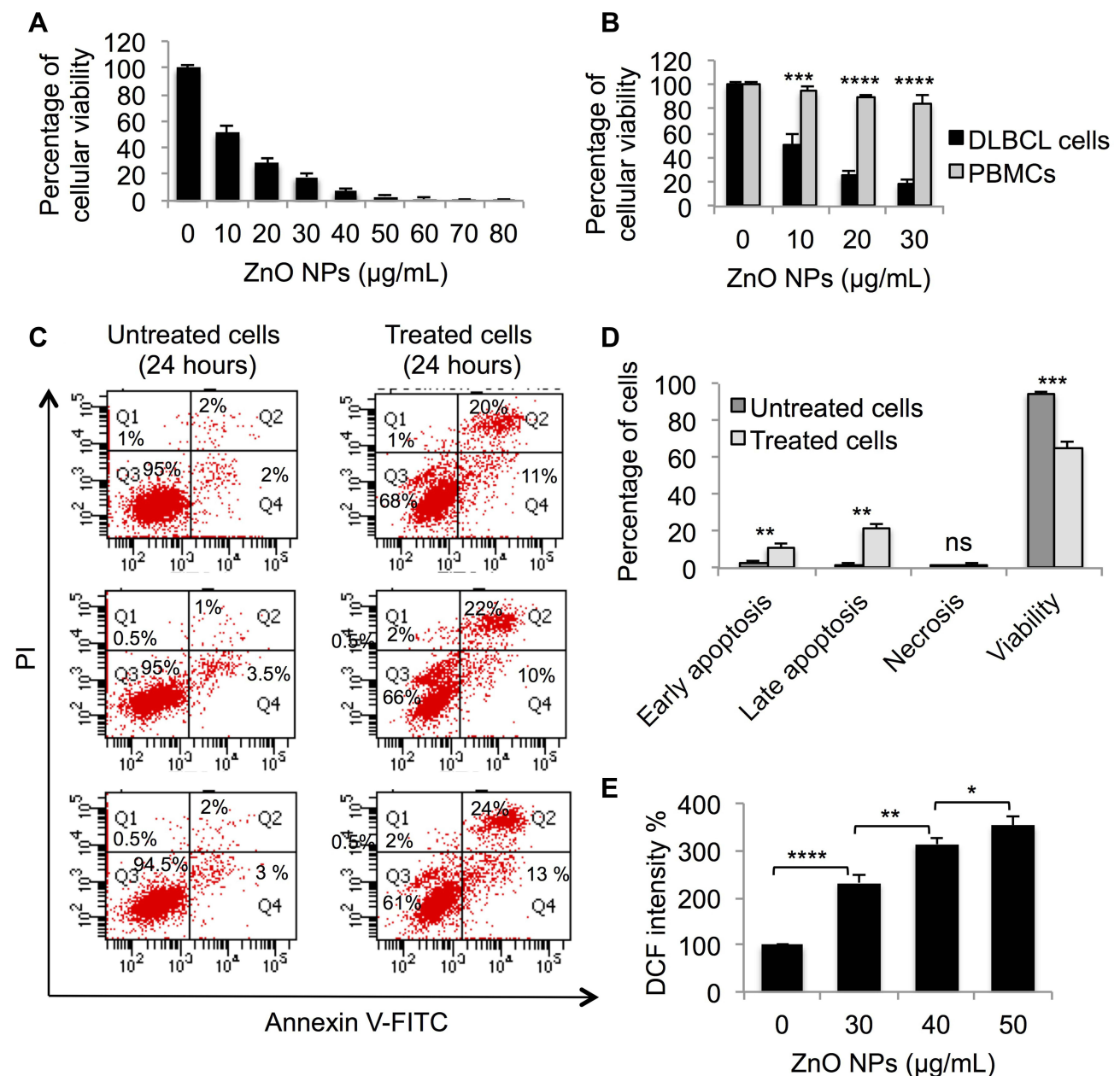


Figure 1 Evaluation of the cytotoxicity of ZnO NPs against DLBCL cells and normal cells. The viability of DLBCL cells (A) and PBMcs (B) following treatment with different concentrations of ZnO NPs for 120 hours was determined using MTT assay. The type of cellular death caused by ZnO NPs (30 µg/mL) in DLBCL cells (C) was identified using annexin V-FITC and propidium iodide (PI) with flow cytometer. The data of flow cytometer were presented using column graph (D). ZnO NPs-induced production of ROS was measured in DLBCL cells using DCFH-DA (E). All experiments were performed in triplicate. Asterisk symbol was used to define p values; $p \leq 0.05$ is * $p \leq 0.01$ is ** $p \leq 0.001$ is *** and $p \leq 0.0001$ is ****.

Abbreviation: PBMcs, peripheral blood mononuclear cells.

Results

Cytotoxicity of ZnO NPs and Identification of the Kind of Cell Death

To determine the impact of ZnO NPs on the viability of DLBCL cells, MTT assay was performed on DLBCL cells that were incubated with different concentrations of ZnO NPs (from 0 to 80 µg/mL) for 120 hours. ZnO NPs were found to confer cytotoxicity on DLBCL cells; the mean of 50% cytotoxic concentration (CC_{50}) was 10 µg/mL (Figure 1A).

To investigate whether the toxicity of ZnO NPs is selective against DLBCL cells but not normal cells, MTT assay was conducted on PBMcs and DLBCL cells treated with four concentrations of ZnO NPs (0 µg/mL, 10 µg/mL, 20 µg/mL,

and 30 $\mu\text{g/mL}$) for 120 hours. **Figure 1B** showed ZnO NPs to selectively target the viability of DLBCL cells; at concentration = 30 $\mu\text{g/mL}$ of ZnO NPs the average viability of DLBCL cells dropped to 18% (range = 14% to 22%) compared with 85% (range = 80% to 92%) for normal PBMCs ($p < 0.0001$).

To identify the type of cellular death induced by ZnO NPs in DLBCL cells, flow-cytometric analysis post staining with Annexin V-FITC and PI was performed. The analysis was done on DLBCL cells treated with ZnO NPs (30 $\mu\text{g/mL}$) for 24 hours and on control cells (untreated DLBCL cells). ZnO NPs targeted the viability of DLBCL cell by inducing apoptosis. The average early apoptotic population (singly positive cells for annexin V-FITC) was 11.3% (range = 10% to 13%) for the treated cells compared with 2.8% (range = 2% to 3.5%) in the control cells ($p = 0.01$; **Figure 1C and D**). Similarly, the average late apoptotic population (doubly positive cells for annexin V-FITC and PI) was 22% (range = 20% to 24%) for the treated cells as opposed to 1.7% (range = 1% to 2%) for the control cells ($p = 0.003$; **Figure 1C and D**). The average viable population (doubly negative cells for annexin V-FITC and PI) was reduced from 94.8% (range = 94.5% to 95%) in the control cells to 65% (range = 61% to 68%) in the treated cells ($p = 0.0004$; **Figure 1C and D**). The treated DLBCL cells with ZnO NPs (30 $\mu\text{g/mL}$) for 24 hours and control cells (untreated DLBCL cells) were stored for subsequent analyses.

ZnO NPs-Induced Production of ROS

To determine whether the ZnO NPs-induced apoptosis in DLBCL cells was associated with changes in the levels of ROS, experiments were conducted to measure ROS levels in DLBCL cells treated with different concentration of ZnO NPs (0 $\mu\text{g/mL}$, 30 $\mu\text{g/mL}$, 40 $\mu\text{g/mL}$, 50 $\mu\text{g/mL}$) for 24 hours. **Figure 1E** shows an increase in the level of ROS in the treated cells compared with the control cells; and that the production of ROS in response to ZnO NPs appeared to be dose-dependent. Compared with the DCF intensity (100%) of the control cells, the DCF intensity of the ZnO NPs-treated cells was 232% for 30 $\mu\text{g/mL}$ NPs, 313% for 40 $\mu\text{g/mL}$ NPs and 353% for the 50 $\mu\text{g/mL}$ NPs.

ZnO NPs-Dependent Transcriptomics Changes in DLBCL Cells

In attempt to illustrate the transcriptomics-related mechanisms that underpin the apoptotic impact of ZnO NPs on DLBCL cells, the transcriptomics profiles of DLBCL cells (three independent samples with independent experiment for each sample) that were treated with ZnO NPs (30 $\mu\text{g/mL}$) for 24 hours were compared with that of untreated DLBCL cells (three independent control samples with independent experiment for each control). The cellular viability of the treated cells was 65% with apoptotic population (early and late apoptosis) = 33.3%. In contrast, the viability of the control cells was 94.8% with apoptotic population (early and late apoptosis) = 4.5%. The transcriptomics comparison included 26,950 probe IDs that had flag label “detected” in all samples of at least one of the two conditions (treated and untreated). The following criteria (FC ≥ 3 -fold with $p \leq 0.008$ and corrected $p \leq 0.05$) were applied to the transcriptomics profiles of the two groups of samples to identify DEGs. ZnO NPs were found to induce differential expression of 528 genes (probe IDs) in DLBCL cells. Of these genes, 125 were over-expressed and 403 were under-expressed in ZnO NPs-treated DLBCL cells compared with untreated DLBCL cells (**Supplementary Table 1**). Volcano plot (**Figure 2A**) shows the impact of the ZnO NPs on the transcriptomes of DLBCL cells; and hierarchical cluster analysis coupled with heatmap presentation (**Figure 2B**) demonstrates the DEGs (FC ≥ 3 -fold with $p \leq 0.008$ and corrected $p \leq 0.05$).

To shed some light onto the functional roles of the DEGs, GO and biological pathway enrichment analysis using KEGG pathway database, Reactome pathways database and WikiPathways database was conducted through the GeneSpring GX version 12.1 software package (Agilent Technologies, Inc.) and the gProfiler (**Table 1**). The over-expressed genes enriched for biological processes and pathways that have been implicated in cellular response to chemical insult. For example, significant over-representation was identified for stress response to metal ion (corrected p value = 4.94×10^{19}), cellular zinc ion homeostasis (corrected p value = 3.51×10^{15}), cellular response to zinc ion (corrected p value = 4.94×10^{14}), metallothioneins bind metals (corrected p value = 1.36×10^9), negative regulation of growth (corrected p value = 3.38×10^6), attenuation phase (corrected p value 3.91×10^5) and oxidative stress (corrected p value = 0.0006). **Figure 3** shows heatmap presentations of the genes that over-represented cellular response to zinc ions, response to chemicals, and negative regulator of growth. In contrast, the under-expressed genes were found to enrich for biological processes and pathways implicated in DNA modifications, gene expression and cell cycle. For instance, significant over-presentation by the under-expressed genes was found for DNA packaging complex (corrected p value 1.74×10^{12}), chromatin assembly (corrected p value 1.02×10^7), signaling by NOTCH (corrected p value

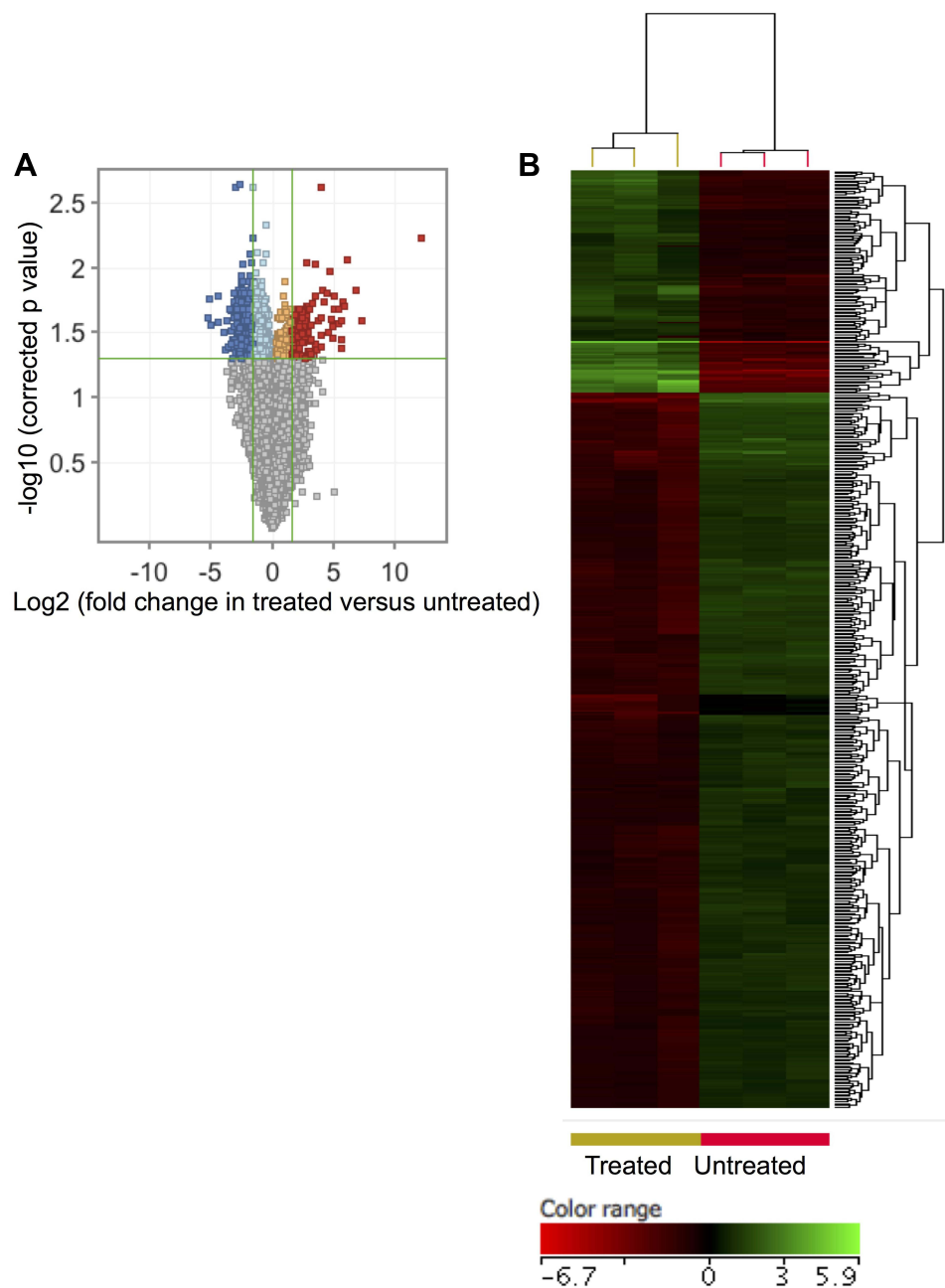


Figure 2 ZnO NPs induced significant changes in the transcriptomic profile of DLBCL cells. Volcano plot demonstrates differentially expressed genes (DEGs: $FC \geq 3$; $p \leq 0.008$; corrected $p \leq 0.05$) in the ZnO NPs-treated DLBCL cells compared with untreated DLBCL cells (**A**). Hierarchical cluster analysis with heatmap presentation were done for the DEGs (**B**). The color range represents the normalized signal value of probes (\log_2 transformation and 75 percentile shift normalization).

1.68×10^7), negative regulation of gene expression by epigenetic (corrected p value 6.69×10^7), signaling by WNT (corrected p value 1.49×10^6), M phase of cell cycle (corrected p value 4.27×10^5), telomere maintenance (corrected p value = 0.005), and negative regulation of transcription (corrected p value 0.02). **Figure 4** shows heatmap presentations of genes that over-represented negative regulation of transcription, chromatin assembly, telomere maintenance and signaling by NOTCH.

Oxidative Stress Responsive Genes

ZnO NPs have been shown to increase the production of reactive oxygen species (ROS) leading to oxidative stress-induced cell death.¹⁵ Among the DEGs ($FC \geq 3$ -fold with $p \leq 0.008$ and corrected $p \leq 0.05$), six oxidative responsive genes,

Table 1 Biological Processes and Pathways Enrichment Analysis of the Differentially Expressed Genes (DEGs: FC \geq 3; P \leq 0.008; Corrected P \leq 0.05) in the ZnO NPs-Treated DLBCL Cells Compared with the Untreated DLBCL Cells

GO or Pathway Accession No	GO or Pathway Term	Reg	Corr. p value	Count in Selection	Count in Total
GO:0097501	Stress response to metal ion	Up	4.94 \times 10 ¹⁹	10	16
GO:0061687	Detoxification of inorganic compound	Up	4.94 \times 10 ¹⁹	10	16
GO:0006882	Cellular zinc ion homeostasis	Up	3.51 \times 10 ¹⁵	10	35
GO:0071294	Cellular response to zinc ion	Up	8.02 \times 10 ¹⁴	8	12
GO:0071241	Cellular response to inorganic substance	Up	1.83 \times 10 ¹²	14	219
GO:0071248	Cellular response to metal ion	Up	6.38 \times 10 ¹¹	11	93
R-HSA-5661231	Metallothioneins bind metals	Up	1.36 \times 10 ⁹	8	16
GO:0010035	Response to inorganic substance	Up	3.36 \times 10 ⁷	13	360
GO:0006873	Cellular ion homeostasis	Up	2.84 \times 10 ⁶	14	684
GO:0045926	Negative regulation of growth	Up	3.38 \times 10 ⁶	10	210
GO:0006950	Response to stress	Up	4.34 \times 10 ⁶	33	4085
GO:0048878	Chemical homeostasis	Up	1.75 \times 10 ⁵	17	1213
R-HSA-3371568	Attenuation phase	Up	3.91 \times 10 ⁵	7	47
R-HSA-3371571	HSF1-dependent transactivation	Up	1.28 \times 10 ⁴	7	59
GO:0070887	Cellular response to chemical stimulus	Up	5.28 \times 10 ⁴	23	2220
GO:0042221	Response to chemical	Up	0.008	28	3716
GO:0034599	Cellular response to oxidative stress	Up	0.02	6	302
GO:0044815	DNA packaging complex	Down	1.74 \times 10 ¹²	17	102
GO:0006305	DNA alkylation	Down	1.66 \times 10 ¹¹	14	68
GO:0006306	DNA methylation	Down	1.66 \times 10 ¹¹	14	68
GO:0006335	DNA replication-dependent nucleosome assembly	Down	6.86 \times 10 ¹⁰	10	31
GO:0031497	Chromatin assembly	Down	1.02 \times 10 ⁷	14	134
REAC:R-HSA-157118	Signaling by NOTCH	Down	1.68 \times 10 ⁷	18	232
REAC:R-HSA-68875	Mitotic Prophase	Down	3.16 \times 10 ⁷	11	135
GO:0045814	Negative regulation of gene expression by epigenetic	Down	6.69 \times 10 ⁷	12	106
GO:0070076	Histone lysine demethylation	Down	1.15 \times 10 ⁶	8	34
REAC:R-HSA-195721	Signaling by WNT	Down	1.49 \times 10 ⁶	15	327
GO:0016577	Histone demethylation	Down	1.83 \times 10 ⁶	8	36
REAC:R-HSA-171306	Packaging Of Telomere Ends	Down	3.19 \times 10 ⁶	7	52
GO:0032451	Demethylase activity	Down	5.02 \times 10 ⁶	8	41
REAC:R-HSA-195258	RHO GTPase Effectors	Down	5.75 \times 10 ⁶	14	319

(Continued)

Table 1 (Continued).

GO or Pathway Accession No	GO or Pathway Term	Reg	Corr. p value	Count in Selection	Count in Total
REAC:R-HSA-68886	M Phase of cell cycle	Down	4.27×10^5	14	385
REAC:R-HSA-194315	Signaling by Rho GTPases	Down	0.0002	14	449
REAC:R-HSA-69278	Cell Cycle, Mitotic	Down	0.001	14	527
GO:0000723	Telomere maintenance	Down	0.005	7	75
GO:0045892	Negative regulation of transcription	Down	0.02	24	983
GO:1902679	Negative regulation of RNA biosynthetic process	Down	0.03	24	1016
GO:0030097	Hemopoiesis	Down	0.05	18	905

Abbreviations: GO, gene ontology; Reg, expression regulation of the genes that enriched GO or pathway terms; Corr p value, corrected p value that was calculated on the basis of Benjamini–Hochberg false discovery rate (FDR); Count in selection, number of DEGs that were assigned to a GO or pathway term; Count in total, number of all genes that have been known to function in a GO or pathway term.

namely heat shock protein family A (Hsp70) member 1A (*HSPA1A*), murine osteosarcoma viral oncogene homolog (*FOS*), matrix metalloproteinase 9 (*MMP9*), heme oxygenase (*HMOX1*), dehydrogenase/reductase (SDR family) member 2 (*DHRS2*) and NAD(P)H dehydrogenase [quinone 1] (*NQO1*), were over-expressed in DLBCL cells upon treatment with ZnO NPs ([Supplementary Table 1](#)). To widen the search for DEGs implicated in cellular response to oxidative stress, the FC cutoff was set to ≥ 1.5 with $p \leq 0.05$ and corrected $p \leq 0.1$. As a result, 11 additional oxidative response genes were identified with over-expression in ZnO NPs- treated DLBCL cells ([Figure 5](#)). Among these genes were oxidation resistance 1 (*OXRI*), dual specificity phosphatase 1 (*DUSP1*), thyroid peroxidase (*TPO*), and thioredoxin reductase 1 and 2 (*TXNRD1* and *TXNRD2*).

Apoptotic Genes

As shown by flowcytometric analysis ([Figure 1](#)), ZnO NPs induced apoptosis in DLBCL cells. In line with this cellular finding, ZnO NPs caused an over-expression of genes (FC ≥ 3 -fold with $p \leq 0.008$ and corrected $p \leq 0.05$) that inhibit the cellular growth (corrected p value = 3.38×10^6 ; [Table 1](#)) such as different types of metallothioneins (*MT1F*, *MT1H*, *MT1G*, *MT1X*, *MT1B*, *MT1E*, *MT1A* and *MT1M*). In contrast, ZnO NPs induced under-expression of genes (FC ≥ 3 -fold with $p \leq 0.008$ and corrected $p \leq 0.05$) implicated in cell cycle (corrected p value = 0.001; [Table 1](#)); such as *MDM4*, *NINLI*, *PSMD10*, *TUBB3* and 10 types of histones. Nevertheless, the DEGs (FC ≥ 3 -fold with $p \leq 0.008$ and corrected $p \leq 0.05$) did not include other core apoptotic genes, such as those that belong to tumor necrosis receptor superfamily and their ligands, BCL2 family or caspases. Consequently, the FC cutoff was set to ≥ 1.5 with $p \leq 0.05$ and corrected $p \leq 0.1$ in order to broaden the search for apoptotic genes. [Figure 6A](#) shows the expression profile of anti-apoptotic genes and [Figure 6B](#) demonstrates the expression profile of pro-apoptotic genes. Interestingly anti-apoptotic genes such as BCL2-like 1 (*BCL2L1*), BCL2-associated athanogene 4 (*BAG4*), CASP8 and FADD-like apoptosis regulator (*CFLAR*), TNF receptor-associated factor 2 (*TRAF2*) were under-expressed in ZnO NPs-treated DLBCL cells. In contrast, pro-apoptotic genes like programmed cell death 5 (*PDCD5*), caspase 1 apoptosis-related cysteine peptidase (*CASP1*), cell death-inducing DFFA-like effector c (*CIDEc*), BCL2-associated agonist of cell death (*BAD*), apoptosis-inducing factor (*SIVA1*) and BCL2-antagonist/killer 1 (*BAK1*) were over-expressed in ZnO NPs-treated DLBCL cells.

Oncogenes

Oncogenes play important roles in the survival and proliferation of malignant cells.³⁷ Setting the FC cutoff to ≥ 1.5 with $p \leq 0.05$ and corrected $p \leq 0.1$ allowed the identification ZnO NPs-induced altered expression of oncogenes in DLBCL cells. Forty oncogenes were differentially expressed in response to treatment with ZnO NPs; 32 genes were under-

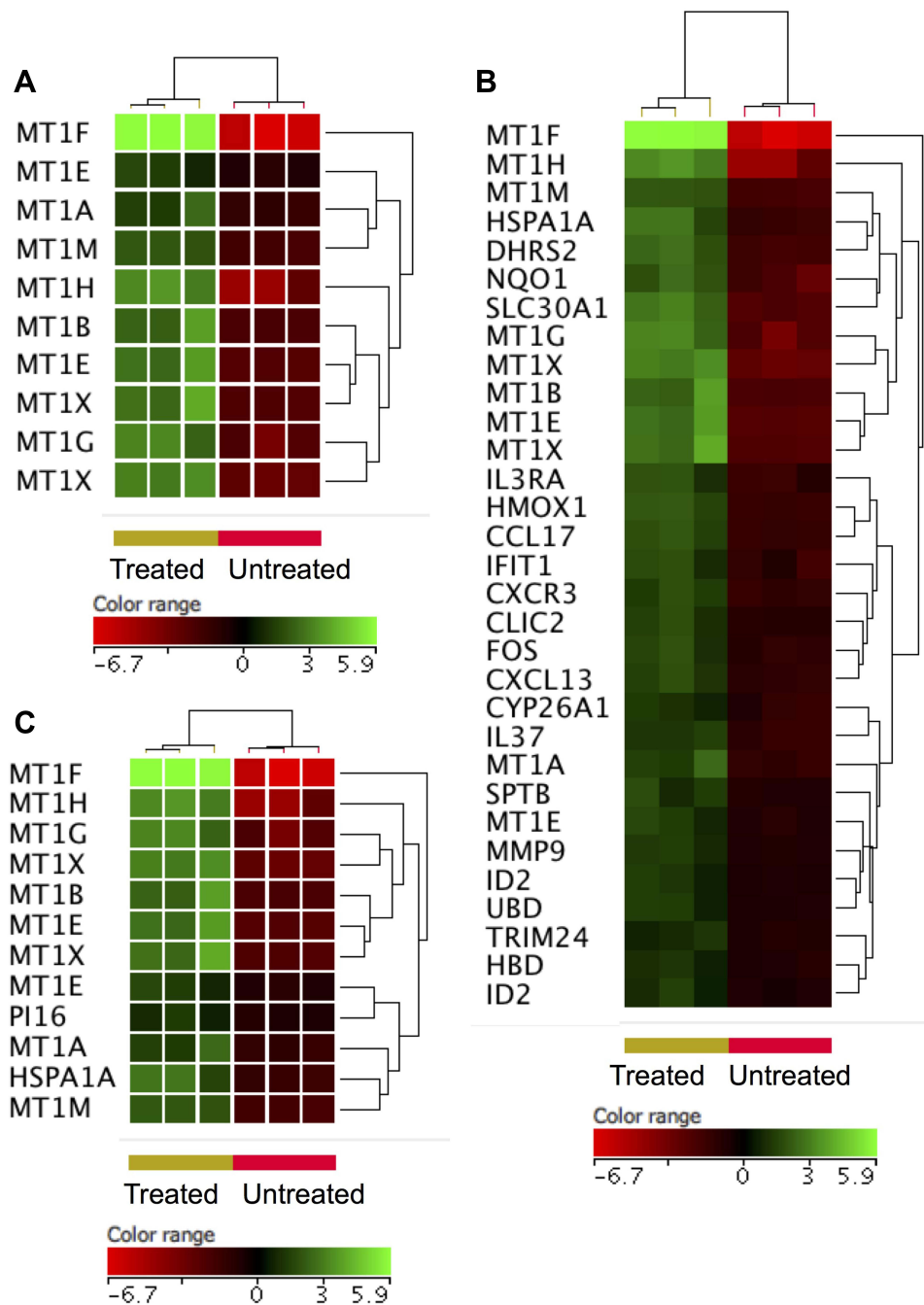


Figure 3 Genes identified with over-expression in the ZnO NPs-treated DLBCL cells compared with the untreated DLBCL cells. Hierarchical cluster analysis with heatmap presentation was conducted on the over-expressed genes ($FC \geq 3$; $p \leq 0.008$; corrected $p \leq 0.05$) that significantly over-represented the biological processes and pathways: cellular response to zinc ions (**A**), response to chemicals (**B**) and negative regulator of growth (**C**). The color range represents the normalized signal value of probes (\log_2 transformation and 75 percentile shift normalization).

expressed and 8 genes were over-expressed in the in the ZnO NPs-treated DLBCL cells. Of interest, ABL proto-oncogene 2 non-receptor tyrosine kinase (*ABL2*), SET nuclear proto-oncogene (*SET*), T-cell leukemia/lymphoma 1A (*TCL1A*), LCK proto-oncogene Src family tyrosine kinase (*LCK*), AKT serine/threonine kinase 2 and 3 (*AKT2* and *AKT3*), lymphoblastic leukemia associated hematopoiesis regulator 1 (*LYL1*), fibroblast growth factor receptor 4 (*FGFR4*), and v-erb-b2 avian erythroblastic leukemia viral oncogene homolog 3 (*ERBB3*) were under-expressed in ZnO NPs-treated DLBCL cells (Figure 7).

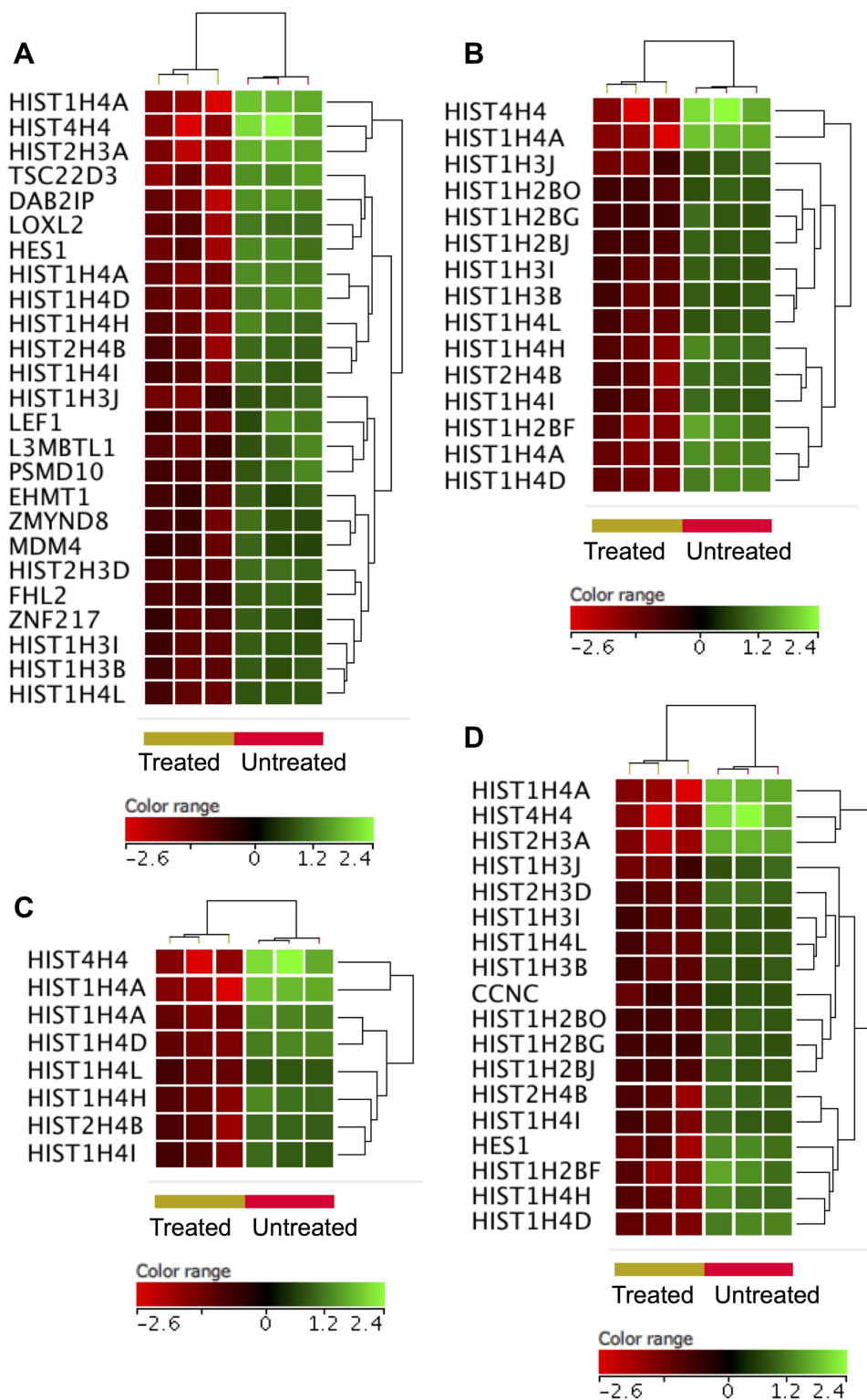


Figure 4 Genes that were under-expressed in the ZnO NPs-treated DLBCL cells compared with the untreated DLBCL cells. Hierarchical cluster analysis with heatmap presentation was conducted on the under-expressed genes ($FC \geq 3$; $p \leq 0.008$; corrected $p \leq 0.05$) that significantly over-represented the biological processes and pathways: negative regulation of transcription (**A**), chromatin assembly (**B**), telomere maintenance (**C**) and NOTCH signaling (**D**). The color range represents the normalized signal value of probes (log2 transformation and 75 percentile shift normalization).

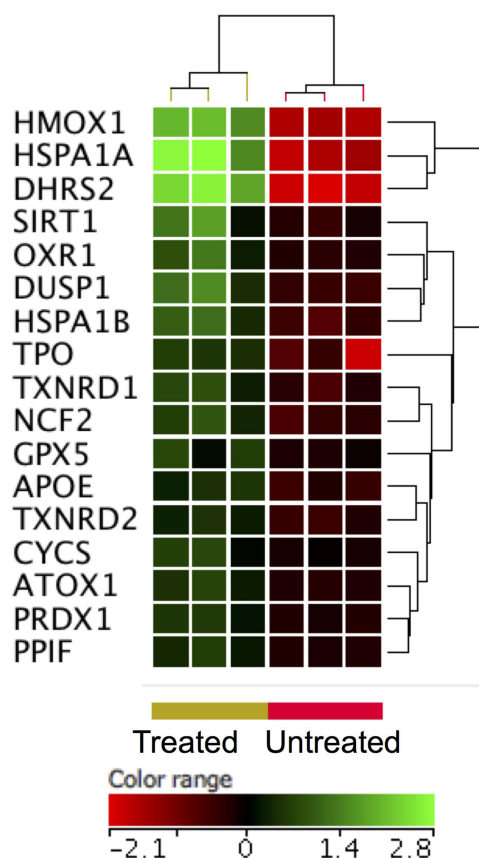


Figure 5 Expression profile of oxidative stress responsive gene. Hierarchical cluster analysis with heatmap presentation was done on oxidative stress responsive gene (FC ≥ 1.5 with $p \leq 0.05$ and corrected $p \leq 0.1$) in ZnO NPs treated DLBCL cells versus untreated DLBCL cells. The color range represents the normalized signal value of probes (log₂ transformation and 75 percentile shift normalization).

B-Cell Receptor Signaling Genes

Signaling through B-cells receptor (BCR) is a key for activation, survival and proliferation of normal and malignant B-cells.³⁸ The transcriptomic profiling showed under-expression of BCR signaling genes (FC cutoff ≥ 1.5 , $p \leq 0.05$, corrected $p \leq 0.1$) in response to the treatment with ZnO NPs. For example, Bruton tyrosine kinase (*BTK*), *AKT1*, *AKT2*, tyrosine-protein kinase Blk (*BLK*), receptor-type tyrosine-protein phosphatase C (*PTPRC*), B-cell lymphoma/leukemia 10 (*BCL10*), hematopoietic progenitor kinase (*HPK*), tyrosine-protein kinase Tec (*TEC*) and linker for activation of B-cells (*LAT2*) showed decreased expression in the ZnO NPs-treated cells (Figure 8).

Nuclear Factor Kappa B Genes

Nuclear factor kappa B (NF κ B) pathway has been heavily implicated in the survival and proliferation of different malignancies including DLBCL.³⁹ The treatment of DLBCL cells with ZnO NPs decreased the expression of NF κ B pathway genes (FC cutoff ≥ 1.5 , $p \leq 0.05$, corrected $p \leq 0.1$). For example I-kappa-B-kinase beta (IKK β), nuclear factor NF-kappa-B p100 (*NFKB2*), nuclear factor NF-kappa-B p110 subunit (*REL*) and transcription factor RelB (*RELB*) where found to be under-expressed in ZnO NPs-treated DLBCL cells (Figure 9).

Discussion

ZnO NPs have been reported by many studies to induced apoptosis in cancer cells.¹⁵ DLBCL (subtype ABC) is an aggressive blood cancer with increased rate of refractory and relapse.³³ The present work demonstrated the capability of ZnO NPs to selectively induce apoptosis in DLBCL cells (subtype ABC), which have been characterized by their

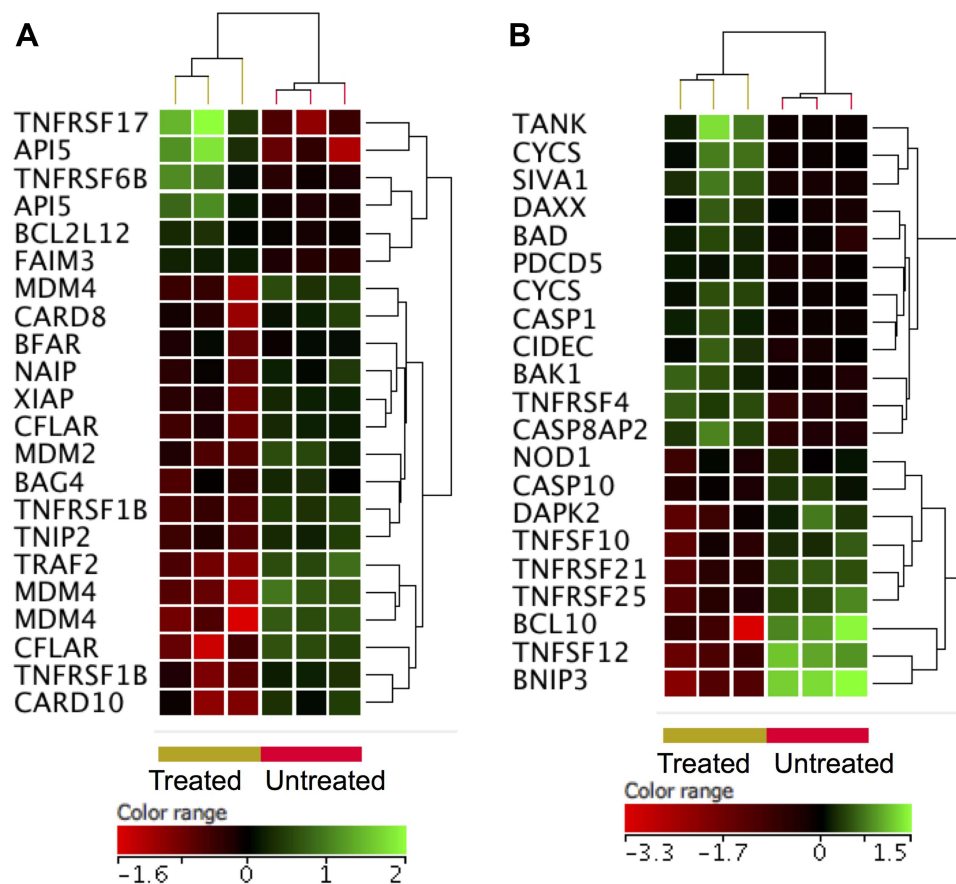


Figure 6 Expression profile of apoptotic gene. Hierarchical cluster analysis with heatmap presentation were constructed for anti-apoptotic genes (**A**) and pro-apoptotic genes (**B**) ($FC \geq 1.5$ with $p \leq 0.05$ and corrected $p \leq 0.1$) in ZnO NPs-treated DLBCL cells compared with untreated DLBCL cells. The color range represents the normalized signal value of probes (log2 transformation and 75 percentile shift normalization).

resistance to standard chemotherapy. In attempt to identify transcriptomics-related mechanisms that underlay the apoptotic effect of the ZnO NPs on DLBCL cells, transcriptomics profiling was conducted on DLBCL cells that were treated with ZnO NPs and control cells (untreated DLBCL cells). Conducting the transcriptomic experiments on ZnO NPs-treated samples with percentage of apoptotic cells = 33.3% was necessary to identify changes in the transcriptomic profile that preceded the ZnO NPs-induced apoptosis (because not all the treated cells were dead; in fact, approximately 65% of the treated cells had not yet undergone apoptosis). Therefore, the identified alterations in the transcriptomic profile of the treated cells are likely to underpin the cytotoxic impact of ZnO NPs.

The molecular findings (transcriptomic profiles) supported and to a large extent explained the cellular findings (induction of apoptosis in DLBCL cells by ZnO NPs). For instance, the treated cells over-expressed genes that have been implicated in response to metal ion, cellular zinc ion homeostasis, cellular response to zinc ion, metallothioneins bind metals and negative regulation of growth. Of these genes, eight types of *MT1* that have been known to play roles in intracellular homeostasis of zinc ions, detoxification of heavy metal ions and suppression of tumor growth.³⁶ The expression of *MT1* genes is down-regulated in different types of cancer perhaps because they target the survival of cancer cells as *MT1* genes inhibits tumor growth.⁴¹⁻⁴⁴ As tumor suppressors, the over-expression of *MT1* genes might have promoted apoptosis in ZnO NPs-treated cells. Agreeing with these data, the treated cells also over-expressed *FOS*, which antagonizes proliferation and promotes apoptosis.⁴⁵ As a self defense mechanism against ZnO NPs the treated cells over-expressed *HSPA1A*, which protects cells from chemical insult,⁴⁶ *SLC30A1*, which exports intracellular Zn ions out of cells to maintain Zn homeostasis,⁴⁷ and cytoskeleton gene *SPTP* perhaps to overcome the previously reported destructive impact of ZnO NPs on cytoskeleton proteins.⁴⁷

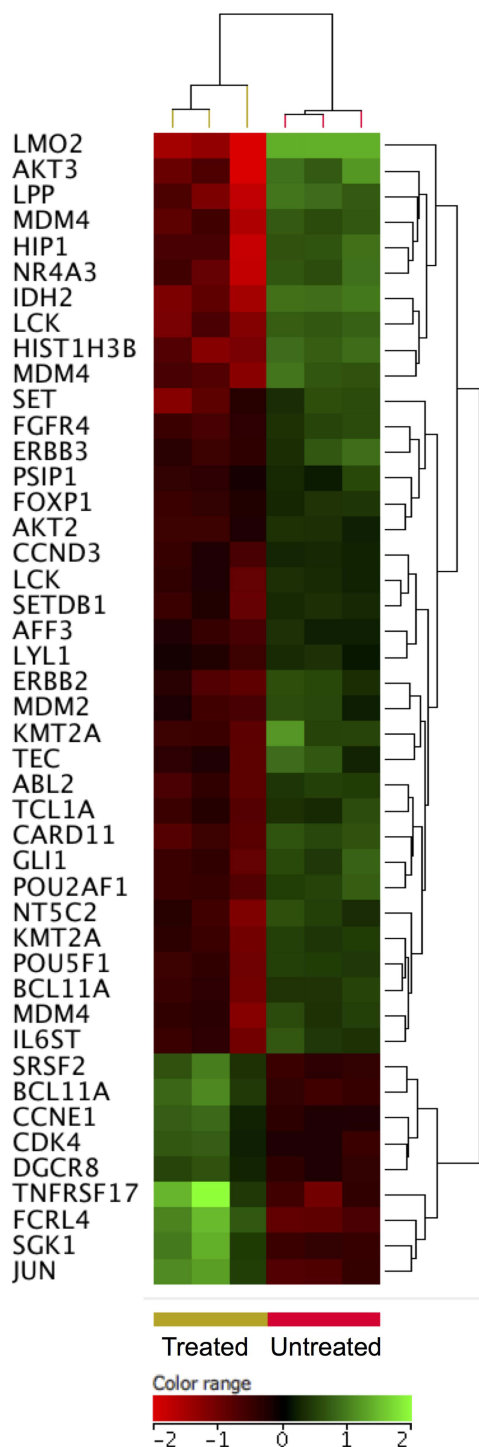


Figure 7 Expression profile of oncogenes. Hierarchical cluster analysis with heatmap presentation were constructed for oncogenes ($FC \geq 1.5$ with $p \leq 0.05$ and corrected $p \leq 0.1$) in ZnO NPs treated DLBCL cells compared with untreated DLBCL cells. The color range represents the normalized signal value of probes (log2 transformation and 75 percentile shift normalization).

The ZnO NPs-treated cells under-expressed genes that play roles in DNA packaging complex, chromatin assembly, signaling by NOTCH, negative regulation of gene expression by epigenetic, signaling by WNT, M phase of cell cycle and telomere maintenance. For example, 14 types of *HIST1* and 3 types of *HIST2*, which are core components of nucleosome that compact DNA into chromatin (DNA packaging and chromatin assembly) restricting access to DNA for transcription,^{48,49} were under-expressed in ZnO NPs-treated cells. Similarly, the transcriptional repressors *L3MBTL1*⁵⁰

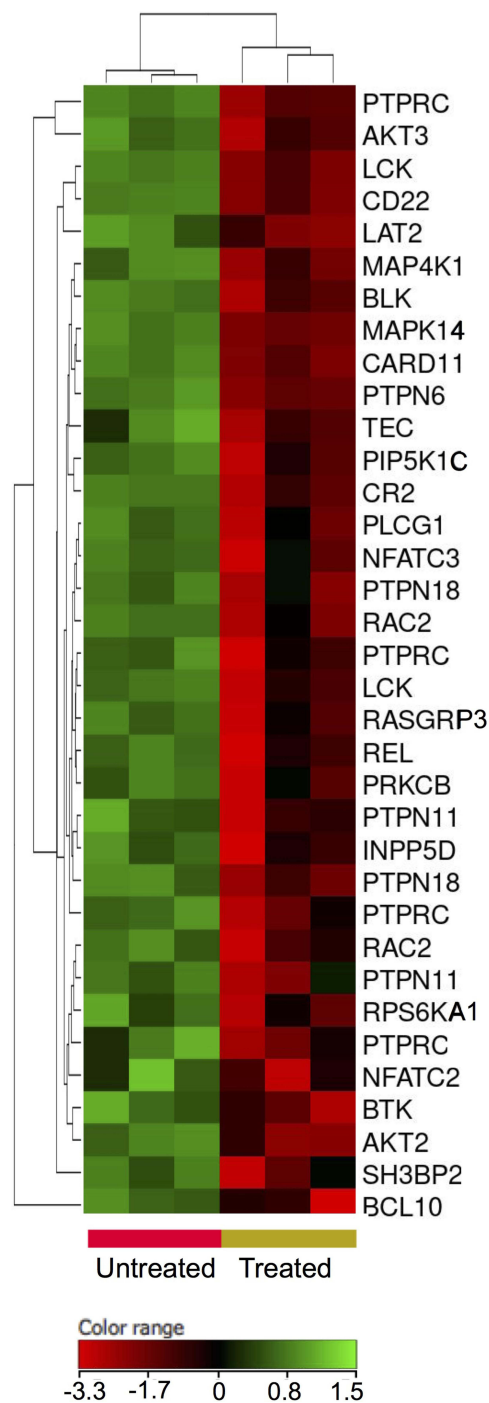


Figure 8 Expression profile of B-cell receptor signaling genes. Hierarchical cluster analysis with heatmap presentation were constructed for the genes ($FC \geq 1.5$ with $p \leq 0.05$ and corrected $p \leq 0.1$) in ZnO NPs-treated DLBCL cells compared with untreated DLBCL cells. The color range represents the normalized signal value of probes (\log_2 transformation and 75 percentile shift normalization).

*ZMYND8*⁵¹ and *ZNF217*⁵² were also under-expressed in the treated cells. Likewise, *EHMT1* which mono and dimethylates lysine residue number 9 of histone 3 leading to repression of DNA transcription,⁵³ was also under-expressed in treated cells. These data suggested that as a defense mechanism, the treated cells attempted to enhance transcription machinery by decreasing the expression of histones (to make DNA more accessible for transcription) and by reducing the expression of transcriptional repressors.

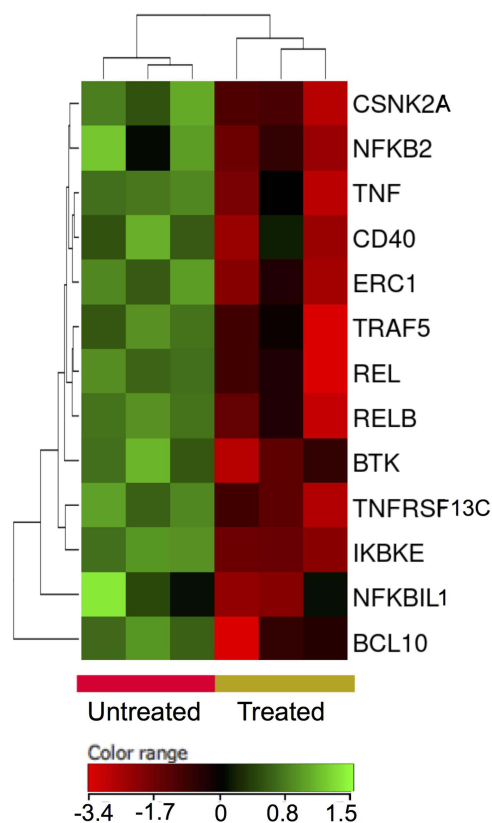


Figure 9 Expression profile of nuclear factor kappa B signaling genes. Hierarchical cluster analysis with heatmap presentation were constructed for the genes ($FC \geq 1.5$ with $p \leq 0.05$ and corrected $p \leq 0.1$) in ZnO NPs-treated DLBCL cells compared with untreated DLBCL cells. The color range represents the normalized signal value of probes (\log_2 transformation and 75 percentile shift normalization).

NOTCH signaling has been implicated in the growth and survival B-cell malignancies including DLBCL⁵⁴ Interestingly, ZnO NPs appeared to target NOTCH signaling as indicated by the decreased expression of the NOTCH target gene *HES1*,⁵⁵ *NOTCH2* ($FC=2$, $p=0.003$, corrected $p=0.05$), which is a receptor engagement of which with its ligands (Jagged-1 and 2) activates NOTCH signaling,⁵⁶ *NOTCH2NL* ($FC=2.4$, $p=0.003$, corrected $p=0.05$), which enhances the signaling of NOTCH pathway by direct interaction with *NOTCH2*,⁵⁷ and increased expression of *NRARP* ($FC=1.88$, $p=0.02$, corrected $p=0.1$), which blocks NOTCH signaling and inhibits the activation of NOTCH target genes.⁵⁸ It appeared from these data that targeting NOTCH signaling is a mechanism by which ZnO NPs target the survival of DLBCL cells.

The treated cells under-expressed variants of *HSIT1*, *HIST2* and *HIST4*. Global expression of histones has been implicated in chromosomal integrity and telomere maintenance.⁵⁹ Cells at replicative-senescence show low expression of histones compared with actively replicative cells. Furthermore, cells with short telomere appeared to express low levels of histones compared with those harboring long telomere.⁶⁰ Furthermore, active telomere maintenance and lengthening by telomerase require increased expression of histones.⁵⁹ Taken together, these findings indicate the importance of histones in telomere maintenance pathway. Agreeing with this view, the treated cells also reduced the expression of genes that constitute human telomerase complex,⁶¹ such as *TEP1* ($FC=1.8$, $p=0.03$, corrected $p=0.1$), and *TERC* ($FC=2.3$, $p=0.001$, corrected $p=0.03$). Moreover, the treated cells also decreased the expression of *PCK* ($FC=1.7$, $p=0.03$, corrected $p=0.1$), *AKT2* ($FC=1.7$, $p=0.003$, corrected $p=0.05$) and *AKT3* ($FC=4.4$, $p=0.007$, corrected $p=0.07$), which phosphorylate *TEP1* and *TRET* contributing to the activation human telomerase complex.⁶¹ Collectively, these data indicated that targeting telomere maintenance is a mechanism by which ZnO NPs exerted their cytotoxicity on DLBCL cells.

A growing body of evidence attributed the cytotoxicity of ZnO NPs on cancer cells to ZnO NPs-dependent production of cellular ROS that promotes oxidative stress-induced cellular death.^{15,62} Agreeing with this, studies have

shown increased expression of cellular oxidative stress responsive genes (anti-oxidant genes); such as SOD, catalase, PRDX2 and GSH, in response to treatment with ZnO NPs.^{63–65} Both the ROS measurement analysis and transcriptomics data of the present study support this notion, where ZnO NPs were found to increase the production of ROS and cause an over-expression of a group of anti-oxidant genes in DLBCL cells. DHRS2, which has NADPH-dependent dicarbonyl reductase activity and reduces ROS,⁶⁶ ORX-1, which increases the expression of genes needed for detoxification hydrogen peroxide,⁶⁷ PRDX1 and GPX5, which detoxify different forms of peroxide,^{68,69} and *TXNRD2*, which keeps thioredoxin in the reduced state,⁷⁰ were all over-expressed upon treatment with ZnO NPs.

As indicated by the flow-cytometric findings, ZnO NPs killed DLBCL cells by apoptosis. The transcriptomic profile of apoptotic genes showed that apoptosis might have occurred in the treated DLBCL cells through the intrinsic (mitochondrial-dependent) and extrinsic (death receptor-dependent) pathways. For example, genes that play anti-apoptotic roles in the intrinsic pathway; such as *MDM4* (*MDMX*), which suppresses the activation of p53,⁷¹ *MDM2*, which induces p53 proteasomal degradation and negatively regulates p53 function⁷² *SIVA1*, which binds to and inhibits the anti-apoptotic gene *BCL-XL*,⁷³ were under-expressed in the treated cells. In line with this, pro-apoptotic genes of the intrinsic pathway like *BAD*, which binds and inactivates Bcl-2 and Bcl-xL,⁷⁴ *PDCD5* (*TRAF19*) and *BAK1*, which decrease the mitochondrial membrane potential and promote the release of cytochrome c,^{74,75} were over-expressed in the treated cells. For the extrinsic pathway, anti-apoptotic genes including *TRAF2*, which blocks the pro-apoptotic signal of TNFSF10 (TRAIL) and CD95L (FASL),⁷⁶ *BAG4* (*SODD*), which blocks TNFRSF1A (TNF-R1) signaling,⁷⁷ *FAIM3* (*TOSO*), which inhibits FAS-induced apoptosis⁷⁸ were under-expressed in the treated cells. In contrast pro-apoptotic genes in the extrinsic pathway like *DAXX*, which bind with Fas death domain and enhances Fas-mediated apoptosis⁷⁹ *CASP8AP2*, which is required for the activation of CAPS8 and FASL-mediated apoptosis⁸⁰ were over-expressed in the treated cells. *XIAP* (*BIRC4*), *NAIP* (*BIRC1*) and *BCL2L12*, which inhibit the activation of executioner caspases (*CASP3* and *CASP7*: common for both pathways)^{81–84} were under-expressed in the treated cells.

The apoptotic impact of ZnO NPs appeared to be conferred, at least partially, through altering the expression of oncogenes in DLBCL cells. For instance, *ABL2*, which is a non-receptor tyrosine-protein kinase that plays roles in cell growth and survival and was implicated in pathogenesis of acute myeloid leukemia and B-cell lymphomas,^{85–87} was under-expressed in the treated cells. Similarly, *TCLIA*, which plays important roles in the progression of chronic lymphocytic leukemia and B-cell lymphomas,^{88,89} showed reduced expression in the treated cells. *AKT2* and *AKT3*, which regulate apoptosis, cell survival, and proliferation and were implicated progression of DLBCL,^{90,91} were also down-regulated in the treated cells. Overall, targeting the expression of oncogenes appeared to be implicated for the exertion of ZnO NPs toxicity on DLBCL cells.

BCR signaling has been heavily involved in the pathogenesis and the aggressive prognosis of DLBCL.^{92,93} Interestingly ZnO NPs targeted the expression (reduced the expression) of *AKT2*, *AKT3*, *BTK*, *BLK*, *BCL10*, *TEC* and *LAT2*, all of which are transducer of BCR signal from the plasma membrane to nucleus.⁹⁴ Targeting BCR signaling has been an attractive approach for the therapy of for many B-cell malignancies including DLBCL.⁹⁵ Of the downstream events of BCR signaling is the activation of NF- κ B pathways, which favors the progression and treatment resistant in many type of cancer including DLBCL.^{96,97} Interestingly, targeting BCR signaling by the ZnO NPs was also associated with reduced expression of *IKBKB*, which is the main activator of NF- κ B pathway,³⁹ and the subunits of NF- κ B (*NFKB2*, *REL* and *RELB*).³⁹ Taken together, these data indicated that ZnO NPs targeted BCR signaling and NF- κ B pathway to induce apoptosis in DLBCL cells.

Collectively, the findings reported here proposed a possible mechanism through which ZnO NPs-induced apoptosis in DLBCL cells happened. The mechanism involved increased production of ROS; and inhibition of proliferation and survival of DLBCL cells by targeting BCR, NF- κ B, NOTCH and telomere maintenance pathways; and activation of extrinsic and intrinsic apoptotic pathways. As self defense mechanism, DLBCL cells responded to ZnO NPs by overexpressing Zn homeostasis, stress responsive and oxidative responsive genes. However, the cellular protection machinery appeared to be defeated by the cytotoxicity of ZnO NPs leading to the induction of apoptosis in DLBCL cells. Interestingly, some of the changes in the transcriptomics profiles of the treated cells were related the nature of the DLBCL subtype ABC. The ZnO NPs targeted DLBCL subtype ABC-dependent proliferation and survival pathways like BCR, NF- κ B and NOTCH; and oncogenes *TCL1A*, *AKT2* and *AKT3*. The alterations in the transcriptomics profiles of the treated cells may also provide

clues about how ZnO toxicity could be measured. The over-expressed genes like *MTIs* (eight genes), *HSPA1A*, *SLC30A1* and *SPTP* are perhaps good biomarker candidates for measuring/monitoring ZnO toxicity.

Conclusion

Overall, the present study reported ZnO NPs to selectively induce apoptosis in DLBCL subtype ABC cells, which have been known to be resistant to chemotherapy. The transcriptomics profiling in the present study revealed ZnO NPs-induced changes in the transcriptomic profile of DLBCL cells. The transcriptomic alterations were consistent with the apoptotic effect of the ZnO NPs on DLBCL cells, and described possible molecular mechanisms through which ZnO NPs killed DLBCL cells. These mechanisms appeared to involve increased production of ROS and down-regulation of DLBCL subtype ABC-dependent proliferation and survival pathways like BCR, NF- κ B and NOTCH, leading to induction of cell death through the intrinsic and extrinsic apoptosis pathways. The ZnO NPs-induced changes in the transcriptomics of DLBCL cells provided a list of genes that could be further studied as biomarker candidates for assessing/monitoring ZnO toxicity.

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Disclosure

The author declares that there are no conflicts of interest.

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