



Association between inflammatory biomarkers and acute respiratory distress syndrome or acute lung injury risk

A systematic review and meta-analysis

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Summary

Background The relationship between acute respiratory distress syndrome (ARDS)/acute lung injury (ALI) and levels of certain inflammatory factors remains controversial. The purpose of this meta-analysis was to summarize the available studies evaluating the association between levels of inflammatory factors and ARDS/ALI incidence.

Methods We searched the PubMed, EmBase, and Cochrane databases for studies published up to July 2017. For each inflammatory factor, a random effects model was employed to pool results from different studies.

Results We identified 63 studies that included 6243 patients in our meta-analysis. Overall, the results indi-

cated that the levels of angiotensin II (ANG)-2 (standard mean difference, SMD: 1.34; $P < 0.001$), interleukin (IL)-1 β (SMD: 0.92; $P = 0.012$), IL-6 (SMD: 0.66; $P = 0.005$), and tumor necrosis factor (TNF)- α (SMD: 0.98; $P = 0.001$) were significantly higher in patients with ARDS/ALI than in unaffected individuals. No significant differences were observed between patients with ARDS/ALI and unaffected individuals in terms of the levels of IL-8 (SMD: 0.61; $P = 0.159$), IL-10 (SMD: 1.10; $P = 0.231$), and plasminogen activator inhibitor (PAI)-1 (SMD: 0.70; $P = 0.060$).

Conclusions ARDS/ALI is associated with a significantly elevated levels of ANG-2, IL-1 β , IL-6, and TNF- α , but not with IL-8, IL-10, and PAI-1 levels.

Keywords Inflammation · Acute lung injury · Acute respiratory distress syndrome · Systematic review · Meta-analysis

Availability of data and material Not applicable

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Abbreviations

ALI	Acute lung injury
ANG	Angiotensin
ARDS	Acute respiratory distress syndrome
IL	Interleukin
KL	Krebs von den Lungen
LDH	Lactate dehydrogenase
PAI	Plasminogen activator inhibitor
RAGE	Receptor for advanced glycation end products
SMD	Standard mean difference
TNF	Tumor necrosis factor
vWF	von Willebrand factor

Background

Acute respiratory distress syndrome (ARDS) and acute lung injury (ALI) are pulmonary diseases characterized by inflammatory pulmonary edema, acute hy-

poxemia, and accumulation of bilateral pulmonary infiltrates [1]. In the USA the annual number of ARDS cases is more than 140,000 [2] and the reported rates of ARDS and ALI are 59 and 79 cases for every 100,000 individuals, respectively, with reported mortality rates ranging from 22% to 41% [3]. ARDS and ALI are triggered by several factors that are associated with either direct or indirect injury. Direct injuries that trigger ARDS/ALI include serious pulmonary infection, aspiration of foreign bodies, pulmonary contusion, and oxygen poisoning. Indirect injuries associated with ARDS/ALI include severe systemic infection, severe non-pulmonary trauma, acute pancreatitis, major blood transfusion, and disseminated intravascular coagulation [4, 5]. Patients exposed to the abovementioned factors are at high risk for developing ARDS/ALI. Although the mechanisms underlying ARDS/ALI pathogenesis remain unclear, inflammation has been considered as one of the major inducing factors.

The pathology of ARDS is characterized by diffuse pulmonary interstitial and alveolar edema due to capillary endothelial and alveolar epithelial cell injuries that lead to acute respiratory insufficiency or failure [6]. In the early stages of ARDS, edema fluid is concentrated in the alveolar interstitial spaces, and neutrophils adhere to the surface of damaged vascular endothelial cells and migrate to pulmonary interstitial and alveolar cavities. Afterwards, proinflammatory factors, such as inflammatory cytokines and proteases, are released and promote neutrophil-mediated lung injury [7]. In the later stage of ARDS, the injured lung is characterized by severe fibrosis and alveolar destruction and reconstruction [8]. The majority of ARDS/ALI patients show rapid disease progression because of this acute inflammatory process.

Various inflammatory mediators that activate the inflammatory cascade and induce secondary diffuse lung parenchymal injury are the primary causes of ARDS/ALI [9, 10]; however, even low levels of inflammatory factors that do not contribute to the amplification of inflammation are associated with the onset of ARDS/ALI, suggesting their possible role as critical drivers of the disease. Therefore, inflammation-related factors can potentially serve as reliable predictors of ARDS/ALI risk.

A previous study systematically reviewed the relationship between plasma biomarkers and ARDS onset and found that levels of Krebs von den Lungen-6 (KL-6), lactate dehydrogenase (LDH), the soluble receptor for advanced glycation end products (RAGE), and von Willebrand factor (vWF) were significantly correlated with ARDS incidence [11]; however, the study did not investigate the inflammatory factors present in the pulmonary edema fluid, and conclusions were based on a relatively small number of studies for each inflammatory factor. Therefore, further validation of these putative associations is required. This study provided update results by analyzing recently pub-

lished literature that investigated the relationships between specific inflammatory biomarkers and the onset of ARDS/ALI. Furthermore, these relationships were analyzed according to different characteristics.

Methods

We conducted the meta-analysis in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement [12].

Search strategy

We systematically searched the PubMed, EmBase, and Cochrane Central Register of Controlled Trials databases for publications up to July 2017 using the keywords “acute respiratory distress syndrome,” “acute lung injury,” “inflammation,” “C-reactive protein,” “interleukin,” “tumour necrosis factor,” “cytokines,” “interferon,” “transforming growth factor,” and “risk factor.” The search strategy used for the PubMed database is described in Supplementary information (searching strategy in PubMed). We restricted our search to reports published in English. We also included relevant articles cited as references of the studies.

Data selection and extraction

Literature search and selection were independently performed by two researchers, and any inconsistencies were resolved by group discussion. A study was eligible for inclusion if the following criteria were met: (1) the study included patients with ARDS/ALI; (2) participants in the control group were not diagnosed with ARDS/ALI; (3) the primary outcomes of interest included Angiopoietin (ANG)-2, Interleukin (IL)-1 β , IL-6, IL-8, IL-10, Plasminogen activator inhibitor (PAI)-1, and Tumor necrosis factor (TNF)- α , while the secondary outcomes included albumin, ANG-1, Clara cell secretory protein (CC16), C-reactive protein (CRP), endotoxin, granulocyte colony-stimulating factor (G-CSF), intercellular cell adhesion molecule (ICAM), IL-2, IL-4, IL-12, KL-6, Lactate dehydrogenase (LDH), myeloperoxidase (MPO), nuclear factor (NF)- κ β , procalcitonin (PCT), protein C, RAGE, sE-selectin, surfactant protein (SP-D), transforming growth factor (TGF)- β 1, tissue factor (TF), Tumor necrosis factor receptor (TNFR)-1, TNFR-2, vascular endothelial growth factor (VEGF), and vWF. Reviews, editorials, non-human studies, letters, and conference papers were excluded because of insufficient data.

The following parameters were extracted from the articles: first author, country, publication year, study design, sample size, and average age, gender, underlying disease of participants, patient disease status, method of ARDS/ALI diagnosis, specimen source, test time, and follow-up. The Newcastle-Ottawa Scale (NOS) was used to evaluate the methodolog-

ical quality of each study [13]. The NOS is based on the following three subscales: selection of the study group (four categories), comparability of the groups (one category), and outcome assessment (three categories). Data extraction and quality assessment were conducted independently by two authors, and results were examined and adjudicated by an additional author who referred to the original study.

Statistical analysis

In this meta-analysis, the standard mean difference (SMD) and 95% confidence interval (CI) were determined to evaluate the effect of sample size across studies [14]. We pooled the SMDs for each inflammatory factor using a random effects model [15]. The I^2 statistic was used to assess heterogeneity of the SMDs across multiple studies [16]. Means and variances were estimated from medians and ranges as previously described [17]. A sensitivity analysis was performed by sequentially removing each individual study from the meta-analysis [16]. Meta-regression was also conducted for ANG-2, IL-1 β , IL6, IL-8, IL10, PAI-1, and TNF- α based on sample size and mean age. Furthermore, stratified analyses were conducted based on sample size, mean age, patient status, and sample origin. Visual inspection of funnel plots from the Egger's [18] or Begg's test [19] was conducted to evaluate publication bias. All tests were two-tailed, and P -value < 0.05 was considered statistically significant. Data analyses were performed using STATA software (version 10.0; Stata Corporation, College Station, TX, USA).

Results

Literature search

In this study, a total of 302 articles were retrieved from PubMed, 704 from EmBase, and 28 from the Cochrane Library. After removing duplicates, 851 articles passed the inclusion criteria in the meta-analysis. A total of 757 articles were excluded because they were considered irrelevant after scanning the titles and abstracts. Furthermore, articles that were considered to be unrelated based on full-text assessment ($n=5$), a duplicate publication ($n=1$), mortality-related studies ($n=18$), studies with undesired outcomes ($n=4$), and studies with control groups without ARDS/ALI risk ($n=3$) were also excluded. Finally, 63 studies that studied a total of 6243 patients were included in our systematic review (Fig. 1; [20–82]).

Study characteristics

A total of 59 studies implemented a prospective design, while the remaining 4 studies employed a retrospective design. Most studies included patient groups with an average age ranging from 40 to 60 years. All participants enrolled in the included studies were at risk of ARDS/ALI, and the majority of samples were collected less than 1 day after study recruitment. All included studies employed standardized American-European Consensus Conference (AECC) criteria, Berlin definition of ARDS, lung injury score (LIS), and oxygenation index (PaO₂/FiO₂) for ARDS/ALI diagnosis. In earlier studies, there were no general criteria for defining ARDS, so the criteria were defined by the authors of each study. The NOS quality analysis for all studies returned scores ranging from 6 to 8, indicating a good overall quality of the included studies (Table 1).

Inflammatory biomarkers and ARDS/ALI

The relationship between ARDS/ALI and angiopoietin (ANG)-2 levels is presented in Fig. 2. The overall standard mean difference (SMD) from six studies showed that ARDS/ALI patients had higher ANG-2 levels than those of unaffected individuals (SMD: 1.34; 95% CI: 0.59–2.10; $P < 0.001$); however, significant heterogeneity was detected ($I^2 = 97.4\%$; $P < 0.001$). Sensitivity analysis showed that the conclusion did not change after sequential removal of each study (Supplementary information Table S1).

The relationship between ARDS/ALI and interleukin (IL)-1 β levels is presented in Fig. 3. The pooled SMD from 10 studies indicated that ARDS/ALI patients exhibited significantly higher IL-1 β levels than those of the individuals without ARDS/ALI (SMD: 0.92; 95% CI: 0.20–1.64; $P = 0.012$). Although substantial heterogeneity was observed across all studies ($I^2 = 93.1\%$; $P < 0.001$), the conclusion did not change

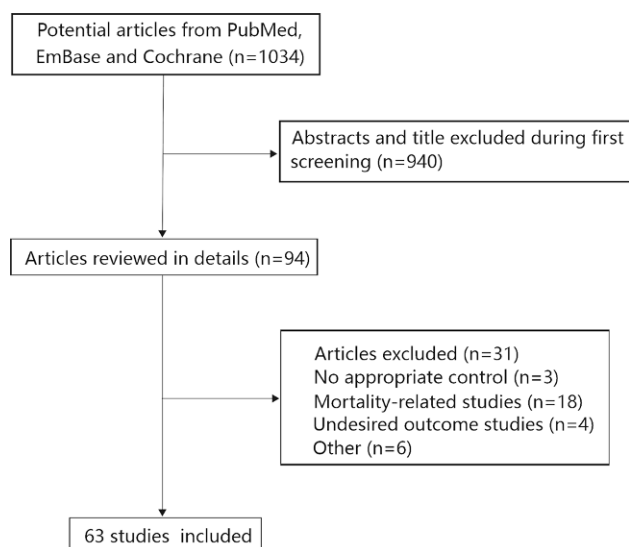


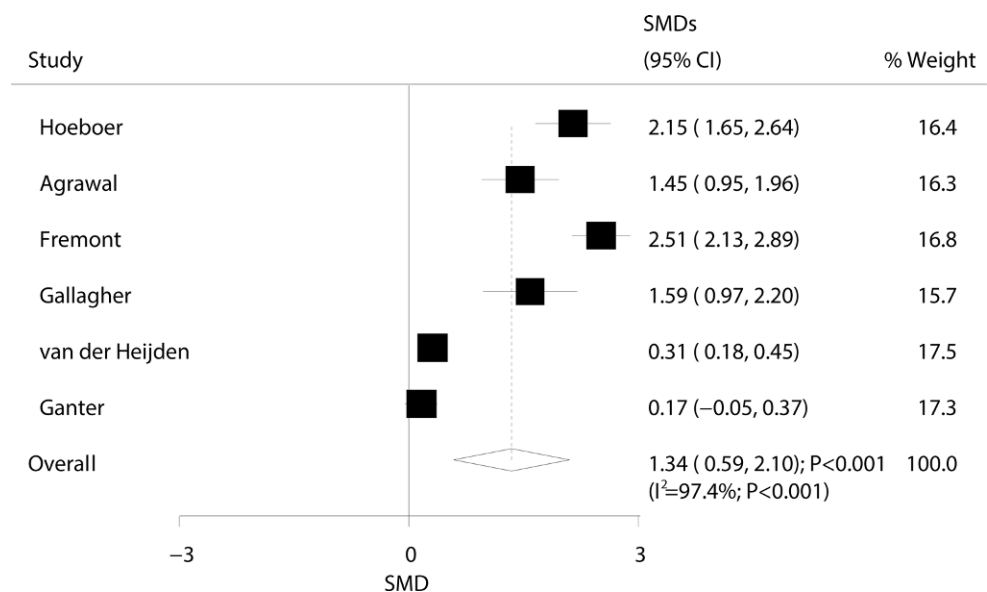
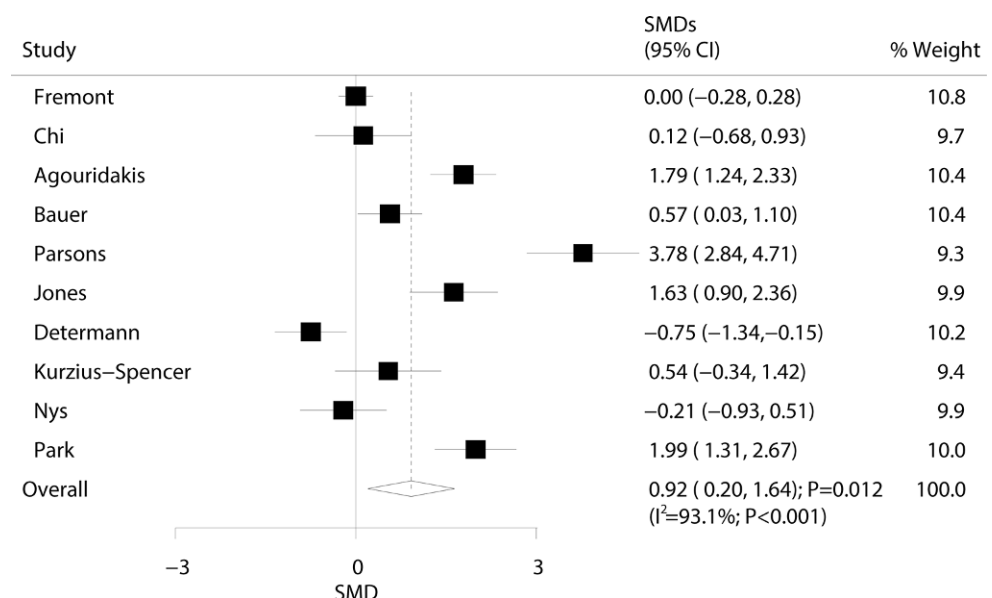
Fig. 1 PRISMA flowchart illustrating the process of study selection in our analysis

Table 1 Characteristics of subjects in eligible studies

Authors	Country	Year	Study design	Sample size	Mean or median age (years)	Gender (m/f)	Underlying disease	Patient status	Judgment method of ARDS/ALI	Specimen source	Test time	Follow-up	NOS score
Hoebner [12]	Netherlands	2015	Prospective	101	64.0	69/32	Fever	ARDS	Berlin definition/LIS	Blood	7 days	7 days	8
Roubinian [13]	U.S.	2015	Prospective	317	58.0	153/164	Pulmonary transfusion reactions hypoxemia	ALI	Defined	Blood	1 day	NA	6
Jones [14]	U.S.	2013	Prospective	43	45.6	40/9	Inhalation and burns	ALI	PaO ₂ /FIO ₂	BALF	3 days	3 days	7
Agrawal [15]	U.S.	2013	Prospective	230	65.0	79/88	Critically ill	ALI	Berlin definition	Blood	1 day	60 days	7
Schultz [16]	Netherlands	2012	Retrospective	20	59.0	13/7	Mechanical ventilation	ALI	LIS	BALF	6 days	6 days	7
Quesnel [17]	France	2012	Retrospective	122	49.0	79/43	Critically ill	ALI/ARDS	AECC	BALF	NA	28 days	6
Osaka [18]	Japan	2011	Prospective	27	50.0	12/15	Pneumonia	ALI	PaO ₂ /FIO ₂	Blood	1 day	10 days	8
Guerivily [19]	France	2011	Prospective	74	58.0	53/21	Critically ill	ALI	AECC	Blood/BALF	1 day	28 days	7
Jabaudon [20]	France	2011	Prospective	64	59.0	41/23	Severe Sepsis	ALI	AECC	Blood	1 day	28 days	7
Anan [21]	Netherlands	2010	Prospective	83	60.0	65/18	Mechanical ventilation	ALI/ARDS	AECC	Blood	1 day	NA	7
Kohno [22]	Japan	2011	Prospective	20	71.0	15/5	Thoracic aortic aneurysm repair	ARDS	PaO ₂ /FIO ₂	Blood	1–4 days	22 days	7
Determann [23]	Netherlands	2010	Prospective	36	58.0	22/14	Mechanical ventilation	ALI/ARDS	LIS	Blood	2 days	2 days	8
Determann [24]	Netherlands	2010	Prospective	150	61.0	99/51	Mechanical ventilation	ALI	LIS	Blood/BALF	1 day	4 days	8
Fremont [25]	U.S.	2010	Retrospective	192	39.0	131/61	Traumatic injuries	ALI	PaO ₂ /FIO ₂	Blood	3 days	6–10 days	7
Determann [26]	Netherlands	2009	Retrospective	22	65.0	17/5	Ventilator-associated pneumonia	ALI/ARDS	AECC	BALF	1 day	8 days	7
Chi [27]	China	2009	Prospective	27	NA	NA	Orthotopic liver transplantation	ALI	PaO ₂ /FIO ₂	Blood	1 day	7 days	6
Kroposki [28]	U.S.	2009	Prospective	32	43.0	15/17	Mechanical ventilation	ARDS	AECC	Blood/BALF	1 day	2–14 days	7
Calfee [29]	U.S.	2009	Prospective	67	51.0	40/27	Hydrostatic pulmonary edema	ALI	AECC	Blood/BALF	4 h	3 days	8
Van der Heijden [30]	Netherlands	2008	Prospective	112	56.0	NA	Critically ill	ALI/ARDS	AECC/LIS	Blood	1 day	NA	7
Kurzus-Spencer [31]	U.S.	2008	Prospective	21	NA	20/1	Smoke inhalation injury	ARDS	AECC/PaO ₂ /FIO ₂	BALF	36 h	72 h	8
Nathani [32]	U.K.	2008	Prospective	42	62.0	24/18	ARDS risk population	ARDS/ALI	AECC	Blood/BALF	1 day	4 days	7
Ganter [33]	U.S.	2008	Prospective	208	41.0	155/53	Traumatic injuries	ALI	AECC	Blood	1 day	28 days	7
Gallagher [34]	U.S.	2008	Prospective	63	67.0	35/28	Critically ill	ALI/ARDS	AECC	Blood	1 day	2 months	7
Calfee [35]	U.S.	2007	Prospective	1451	52.0	609/839	Trauma	ALI	Defined	Blood	1 day	180 days	7
Ware [36]	U.S.	2007	Prospective	878	52.0	514/364	Acute cardiogenic pulmonary edema	ALI/ARDS	Defined	Blood	1 day	3 days	7
Perkins [37]	UK	2007	Prospective	54	NA	NA	ARDS risk population	ALI/ARDS	AECC	BALF	1 day	4 days	6
El Solh [38]	U.S.	2006	Prospective	51	36.6	22/29	Aspiration pneumonitis	ALI	AECC	Blood/BALF	1 day	28 days	8
Parsons [39]	U.S.	2005	Prospective	49	NA	NA	Critically ill	ALI	AECC	Blood	1 day	180 days	8
Bourros [40]	Greece	2004	Prospective	59	51.7	43/16	Critically ill	ALI	AECC	Blood/BALF	1 day	NA	7
Nakae [41]	Japan	2003	Prospective	21	62.0	15/6	Sepsis	ARDS	Defined	Blood	NA	NA	7
Sato [42]	UK	2004	Prospective	37	39.5	32/5	Mechanical ventilation	ARDS	AECC	Blood	1 day	6.5 days	8

Table 1 (Continued)

Authors	Country	Year	Study design	Sample size	Mean or median age (years)	Gender (m/f)	Underlying disease	Patient status	Judgment method of ARDS/ALI	Specimen source	Test time	Follow-up	NOS score
Nys [43]	Belgium	2003	Prospective	67	54.0	43/24	Pneumonia	ARDS	PaO ₂ /FIO ₂	BALF	1–2 days	NA	7
Gessner [44]	Germany	2003	Prospective	35	60.0	16/19	Acute respiratory failure	ARDS	AECC	BALF	1 day	6 months	7
Ishizaka [45]	Japan	2004	Prospective	35	68.0	27/8	Cardiogenic pulmonary edema	ALI	AECC	BALF	1 day	NA	6
Prabhakaran [46]	U.S.	2003	Prospective	51	50.0	29/22	Hydrostatic edema	ARDS	AECC	Blood/BALF	1 day	NA	7
Grissom [47]	U.S.	2003	Prospective	39	51.0	16/17	ARDS risk population	ARDS	AECC	Blood/BALF	96h	42 days	8
Agouridakis [48]	Greece	2002	Prospective	65	44.0	40/25	Mechanical ventilation	ARDS	AECC	Blood/BALF	1 day	15 days	8
Agouridakis [49]	Greece	2002	Prospective	34	49.0	23/11	Mechanical ventilation	ARDS	AECC	Blood/BALF	1 day	6 months	8
Thickett [50]	UK	2002	Prospective	68	65.0	45/23	ARDS risk population	ARDS	AECC	BALF	1 day	4 days	6
Hamacher [51]	France	2002	Prospective	36	43.0	28/8	ARDS risk population	ARDS	AECC	BALF	1 day	21 days	7
Takala [52]	Finland	2002	Prospective	52	54.0	29/19	Critically ill	ARDS	AECC	Blood	1 day	7 days	8
Park [53]	Switzerland	2001	Prospective	69	43.8	41/28	ARDS risk population	ARDS	AECC	BALF	1 day	21 days	7
Hirani [54]	UK	2001	Prospective	56	48.0	NA	Major trauma	ARDS	AECC	BALF	1 day	36 months	8
Geerts [55]	Belgium	2001	Prospective	26	52.0	19/7	ARDS risk population	ARDS	AECC	BALF	1 day	NA	7
Siemiatkowski [56]	Poland	2000	Prospective	36	44.3	27/9	Major trauma	ARDS	LIS	Blood	1 day	10 days	7
Armstrong [57]	UK	2000	Prospective	67	62.0	44/23	Critically ill	ARDS/ALI	AECC	BALF	48h	NA	7
Bauer [58]	Spain	2000	Prospective	66	57.2	NA	Pneumonia	ARDS	AECC	Blood	1 day	NA	8
Gando [59]	Japan	1999	Prospective	58	58.0	37/21	Critically ill	ARDS	Defined	Blood	1 day	4 days	7
Donnelly [60]	Scotland	1999	Prospective	61	NA	NA	Trauma	ARDS	Defined	BALF	NA	NA	6
Parsons [61]	U.S.	1997	Prospective	77	37.5	53/24	ARDS risk population	ARDS	Defined	Blood	1 day	2 days	7
Schutte [62]	Germany	1996	Prospective	56	54.5	45/11	Pneumonia	ARDS	AECC	Blood/BALF	2 days	10 days	7
Chollet-Martin [63]	France	1996	Prospective	14	61.0	NA	Pneumonia	ARDS	LIS	Blood/BALF	3 days	7 days	8
Ricou [64]	U.S.	1996	Prospective	33	48.0	24/9	Critically ill	ARDS	LIS	Blood/BALF	3 days	2 weeks	8
Schwartz [65]	U.S.	1996	Prospective	12	45.0	7/5	Mechanical ventilation	ARDS	LIS	BALF	1 day	NA	7
Jorens [66]	UK	1995	Prospective	35	56.6	31/4	Carotipulmonary bypass	ARDS	Defined	BALF	1 day	3 days	7
Fuchs-buder [67]	Switzerland	1996	Prospective	21	NA	NA	Critically ill	ARDS	LIS	BALF	2 days	10 days	8
Leff [68]	U.S.	1993	Prospective	26	NA	NA	Sepsis	ARDS	Defined	Blood	1 day	2 days	7
Sakamaki [69]	Japan	1995	Prospective	48	49.0	29/19	Sepsis	ARDS	Defined	Blood	1 day	15 days	7
Donnelly [70]	U.S.	1994	Prospective	82	49.5	NA	ARDS risk population	ARDS	LIS	Blood	1–3 days	NA	8
Moaali [71]	U.S.	1989	Prospective	47	66.0	NA	ARDS risk population	ARDS	Defined	Blood	1 day	22 days	7
Rubin [72]	U.S.	1990	Prospective	45	NA	NA	Sepsis	ARDS	LIS	Blood	1 day	3 days	8
Roten [73]	Switzerland	1990	Prospective	50	49.0	31/19	Critically ill	ARDS	Defined	Blood	1 day	5 days	7
Parsons [74]	U.S.	1992	Prospective	103	46.0	77/26	ARDS risk population	ARDS	Defined	Blood	1 day	2 days	7
AECC American-European Consensus Conference, BALF Bronchoalveolar Lavage Fluid, ALI Acute lung injury, ARDS Acute respiratory distress syndrome, NA not available													

Fig. 2 Forest plot comparing ANG-2 levels between ARDS/ALI patients and unaffected individuals**Fig. 3** Forest plot comparing IL-1 β levels between ARDS/ALI patients and unaffected individuals

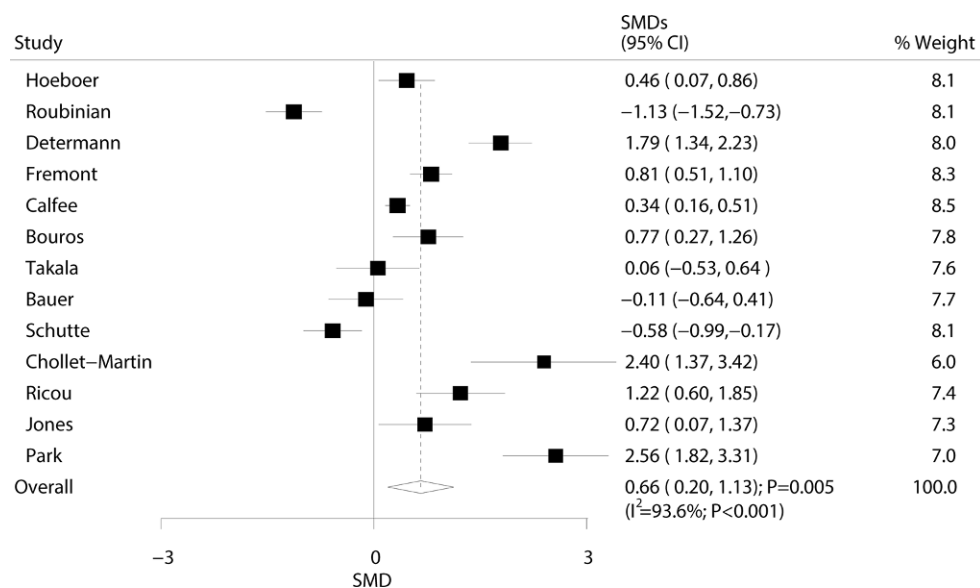
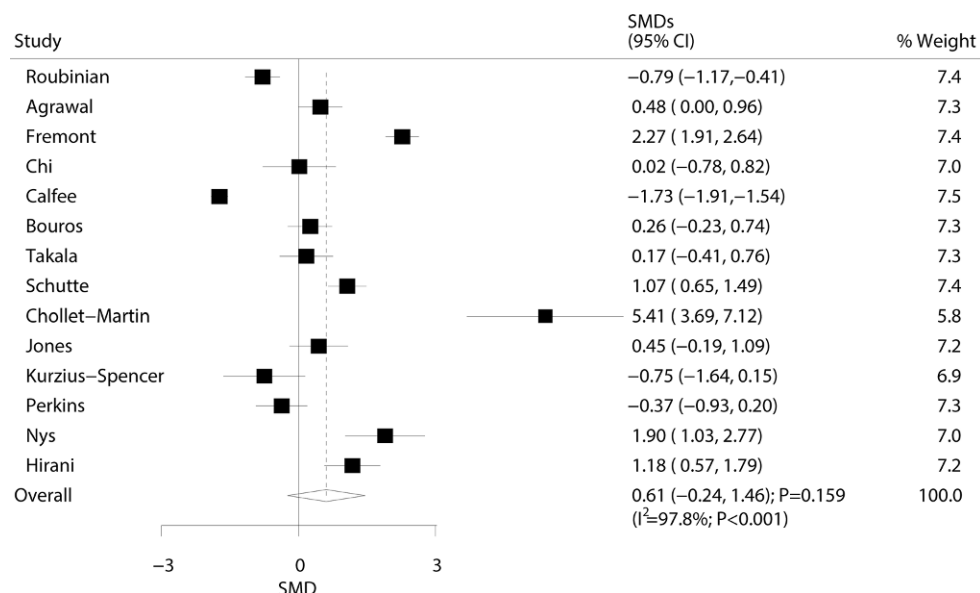
after sequential exclusion of each study (Supplementary information Table S2).

The relationship between ARDS/ALI and IL-6 levels is shown in Fig. 4. Overall results showed that ARDS/ALI patients had higher IL-6 levels than those of individuals in the population without ARDS/ALI (SMD: 0.66; 95% CI: 0.20 to 1.13; $P=0.005$). Heterogeneity was observed at the same degree as the effect across the studies ($I^2=93.6%$; $P<0.001$). Sensitivity analysis showed that the conclusion was not affected by the exclusion of any specific study from the pooled analysis (Supplementary information Table S3).

The relationship between ARDS/ALI and IL-8 levels was analyzed in 14 studies, and results are shown in Fig. 5. No significant differences in IL-8 levels were observed between ARDS/ALI patients and individuals of the population without ARDS/ALI (SMD: 0.61; 95% CI: −0.24–1.46; $P=0.159$). Furthermore, substan-

tial heterogeneity was detected ($I^2=97.8%$; $P<0.001$). Based on sensitivity analysis, we excluded the study conducted by Calfee et al. [57], which specifically included a large sample size of trauma patients. We concluded that ARDS/ALI were associated with higher IL-8 levels (SMD: 0.76; 95% CI: 0.11–1.40; $P=0.021$) (Supplementary information Table S4).

The relationship between ARDS/ALI and IL-10 levels was investigated in seven studies, and results are presented in Fig. 6. We detected no significant differences in IL-10 levels between ARDS/ALI and non-ARDS/ALI patients (SMD: 1.10; 95% CI: −0.70–2.91; $P=0.231$). Substantial heterogeneity was observed ($I^2=98.3%$; $P<0.001$). Sensitivity analysis indicated that ARDS/ALI patients had higher IL-10 levels than those of non-ARDS/ALI patients when the study conducted by Roubinian et al. was excluded (Supplementary information Table S5) [82].

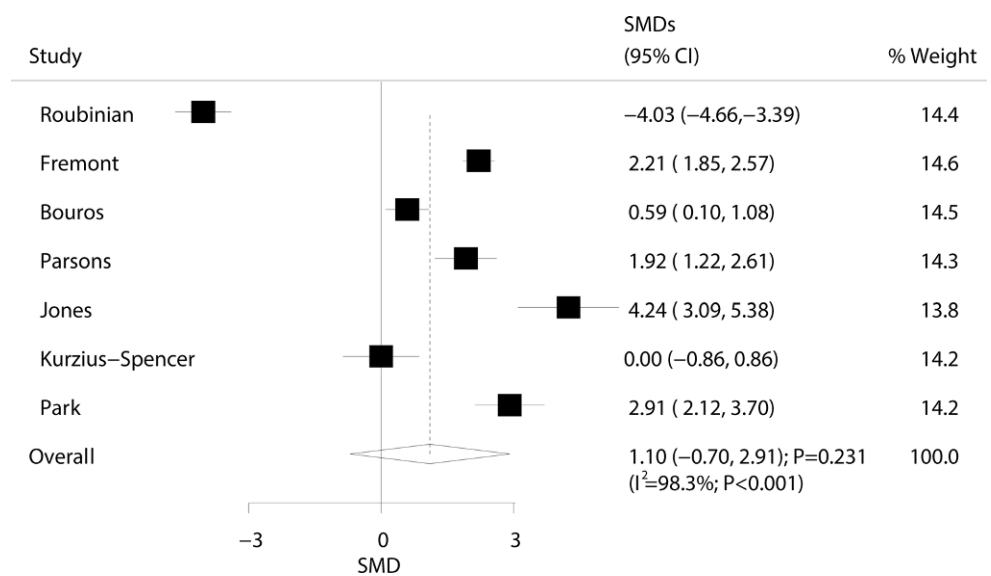
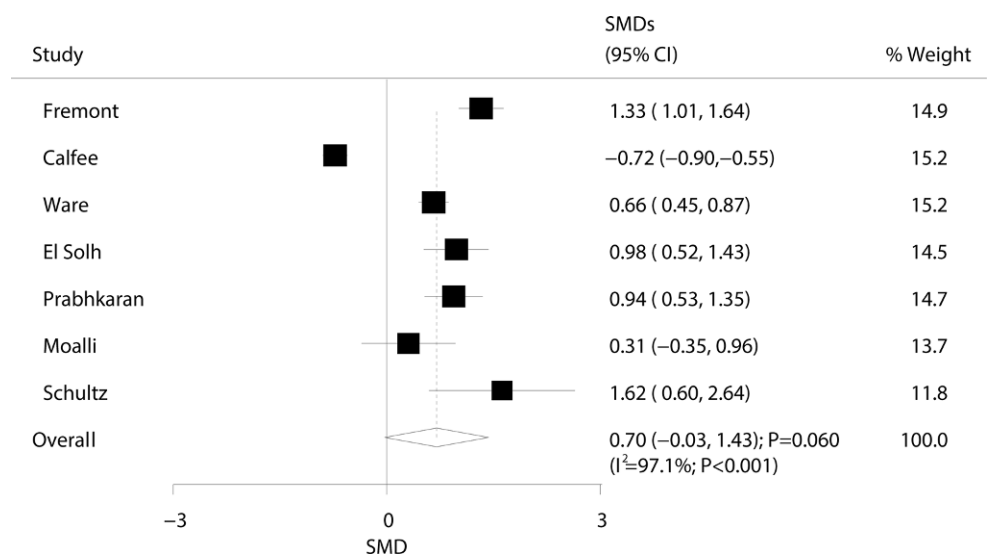
Fig. 4 Forest plot comparing IL-6 levels between ARDS/ALI patients and unaffected individuals**Fig. 5** Forest plot comparing IL-8 levels between ARDS/ALI patients and unaffected individuals

The relationship between ARDS/ALI and plasminogen activator inhibitor-1 (PAI-1) levels was investigated in seven studies, and results are presented in Fig. 7. We detected no significant differences in PAI-1 levels between ARDS/ALI patients and non-ARDS/ALI individuals (SMD: 0.70; 95% CI: -0.03–1.43; $P=0.060$). Substantial heterogeneity was observed ($I^2=97.1%$; $P<0.001$). Sensitivity analysis showed that this result changed after excluding the study conducted by Calfee et al. (Supplementary information Table S6) [57].

The relationship between ARDS/ALI and tumour necrosis factor (TNF)- α levels was investigated in 16 studies, and results are shown in Fig. 8. Pooled results showed that ARDS/ALI patients had significantly higher TNF- α levels than those of individuals without ARDS/ALI (SMD: 0.98; 95% CI: 0.41–1.56; $P=0.001$). Significant heterogeneity was detected across all in-

cluded studies ($I^2=94.0%$; $P<0.001$). These results did not change after sequential exclusion of any specific study (Supplementary information Table S7).

The correlations between ARDS/ALI and other inflammatory factors based on sample origin are summarized in Table 2. Overall, ARDS/ALI patients showed higher levels of albumin (SMD: 2.15; $P=0.010$), ANG-1 (SMD: 4.60; $P<0.001$), KL-6 (SMD: 2.23; $P=0.044$), myeloperoxidase (MPO) (SMD: 1.75; $P<0.001$), transforming growth factor (TGF)- β 1 (SMD: 0.83; $P=0.013$), transfer factor (TF) (SMD: 5.57; $P<0.001$), and TNF receptor-1 (SMD: 5.40; $P<0.001$). Moreover, ARDS/ALI patients had lower levels of IL-12 (SMD: -1.47; $P<0.001$), surfactant protein D (SP-D) (SMD: -1.17; $P=0.012$), and vascular endothelial growth factor (VEGF) (SMD: -4.52; $P<0.001$) in the bronchial alveolar lavage fluid (BALF). In addition, ARDS/ALI patients had higher levels of KL-6 (SMD:

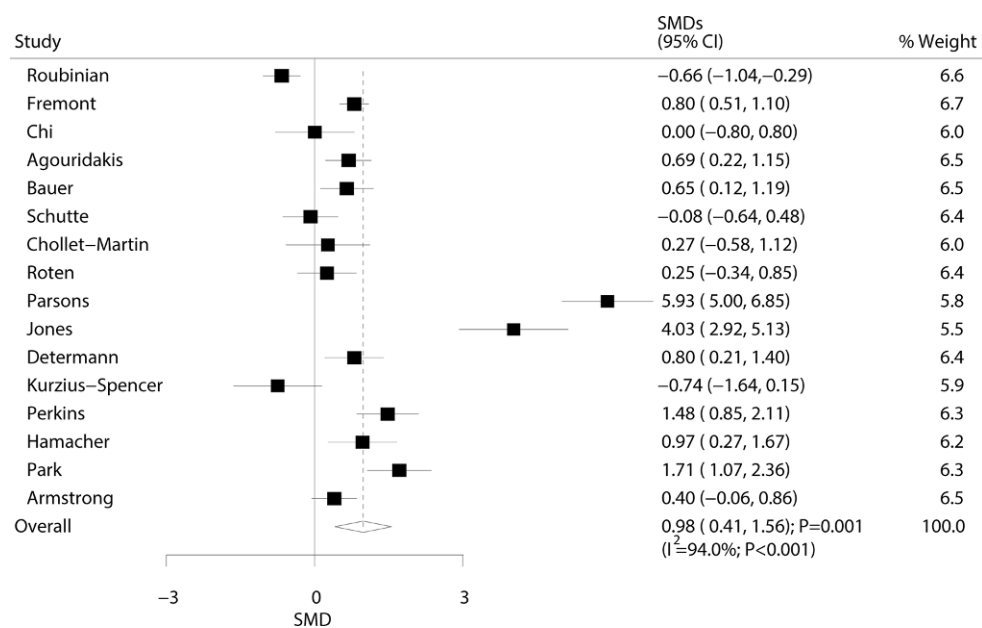
Fig. 6 Forest plot comparing IL-10 levels between ARDS/ALI patients and unaffected individuals**Fig. 7** Forest plot comparing PAI-1 levels between ARDS/ALI patients and unaffected individuals

3.36; $P<0.001$), MPO (SMD: 2.58; $P<0.001$), procalcitonin (PCT) (SMD: 0.41; $P=0.038$), receptor for advanced glycation end products (RAGE) (SMD: 1.64; $P=0.031$), sE-selectin (SMD: 0.55; $P=0.011$), TF (SMD: 3.55; $P<0.001$), and TNF receptor-2 (SMD: 3.82; $P<0.001$) than unaffected individuals. ARDS/ALI was associated with lower IL-12 (SMD: -0.80; $P<0.001$) levels in the blood. No other significant differences were observed between ARDS/ALI and non-ARDS/ALI patients.

Meta-regression and subgroup analyses

A relatively large heterogeneity was observed among the studies included in our meta-analysis. We therefore performed a meta-regression analysis for ANG-2, IL-1 β , IL-6, IL-8, IL-10, PAI-1, and TNF- α ; results are presented in Supplementary information Figures S1–S14. Overall, sample size was determined to influence the association between PAI-1 levels and

ARDS/ALI ($P=0.025$); no other significant associations were observed. Subgroup analyses were also conducted based on sample size, mean age, patient status, and sample source (Table 3). First, ARDS/ALI did not show a significant influence on ANG-2 levels when the mean age was <60.0 years, and patients with ALI. Second, ARDS/ALI was not associated with IL-1 β levels if the study sample size was ≥ 100 , patients with ALI, or samples were collected from the BALF. Third, no significant associations were detected between ARDS/ALI and IL-6 levels when the study sample size was ≥ 100 , the mean age was <60.0 years, patients with ALI, and samples were collected from the blood. Fourth, ARDS/ALI were associated with higher IL-8 levels if the study sample size was <100, patients had ARDS, or samples were collected from BALF. Fifth, ARDS/ALI were significantly associated with higher IL-10 levels when the study sample size was <100, patients with ARDS, and samples were collected from the BALF. Sixth, ARDS/ALI patients showed signifi-

Fig. 8 Forest plot comparing TNF- α levels between ARDS/ALI patients and unaffected individuals

cantly higher PAI-1 levels when the study sample size was < 100, patients with ARDS or ARDS/ALI, and samples were collected from the BALF. Finally, ARDS/ALI were not associated with TNF- α levels if the study sample size was \geq 100, patients with ALI or ARDS/ALI.

Publication bias

Funnel plots of inflammatory factors and ARDS/ALI incidence are presented in Supplementary information Figures S15–S21. No significant publication biases were detected between ARDS/ALI and IL-1 β (P -value for Egger's test (P_{Egger}): 0.148; P -value for Begg's test (P_{Begg}): 0.283), IL-6 (P_{Egger} : 0.330; P_{Begg} : 0.161), IL-10 (P_{Egger} : 0.874; P_{Egger} : 1.000), PAI-1 (P_{Egger} : 0.184; P_{Begg} : 0.548), and TNF- α (P_{Egger} : 0.111; P_{Begg} : 0.224). Although results of the Begg's tests showed no evidence of publication bias for ANG-2 ($P=0.707$) and IL-8 ($P=0.827$), results of Egger's test showed potential publication bias (P -value for ANG-2: 0.048; P -value for IL-8: 0.013). Conclusions did not change after correction using the trim and fill method [83].

Discussion

In our study, ARDS/ALI were found to be associated with higher levels of ANG-2, IL-1 β , IL-6, and TNF- α , whereas no significant associations were detected between ARDS/ALI and IL-8, IL-10, and PAI-1 levels. Furthermore, serum levels of KL-6, MPO, RAGE, sE-selectin, TE, and TNF receptor-2 were significantly higher in ARDS/ALI patients than in unaffected individuals; however, ARDS/ALI patients had lower IL-12 levels. The BALF concentrations of albumin, ANG-1, KL-6, MPO, TGF- β 1, TE, and TNF receptor-1, were significantly higher in ARDS/ALI patients than in individuals without ARDS/ALI. In addition, ARDS/ALI

were associated with lower levels of IL-12 and VEGF; however, heterogeneity among studies was substantial, and the amount of data available was insufficient. Therefore, more research is needed to verify the results of our meta-analysis.

Current treatment for ARDS/ALI consists of respiratory support and immunological treatment. Evidence suggests that the dynamic balance between proinflammatory and anti-inflammatory factors plays a key role in the pathogenesis and prognosis of ARDS/ALI [84]; however, cytokine interactions are highly complex and difficult to study. When proinflammatory and anti-inflammatory factors are unbalanced, excess inflammatory cytokines are released, which in turn damage the lung tissues or even whole body tissues. Therefore, studies that investigate inflammatory factors present during the onset of ARDS/ALI can help elucidate the mechanisms underlying ARDS/ALI pathogenesis and serve as the basis for the development of new treatment approaches for ARDS/ALI.

ANG-2 is a proinflammatory cytokine and a member of the vascular growth factor family. ANG-2 mainly promotes cell apoptosis and disrupts vascularization and can also act in conjunction with VEGF to promote neovascularization [85, 86]. The findings of the present study indicated that serum levels of ANG-2 were significantly higher in ARDS/ALI patients than in unaffected individuals. Similar to our current findings, serum levels of ANG-2 have been associated with other diseases, such as sepsis and pulmonary hypertension [87]. In particular, ANG-2 serum levels were associated with the onset of septic shock, and ANG-2 blood concentrations have been observed to increase during endothelial cell inflammation. Furthermore, elevated ANG-2 levels in the blood are known to promote vascular permeability and leakage; however, only a small number of studies have explored the re-

Table 2 Summary of results of the association of other inflammatory factors with ARDS/ALI based on specimen source

Factors	No. of studies	Groups	SMD	95% CI	P value	Heterogeneity (%)	P for heterogeneity
Albumin	1	Blood	-0.82	-1.79 to 0.14	0.095	-	-
	2	BALF	2.15	0.51 to 3.79	0.010	82.2	0.018
ANG-1	1	Blood	0.80	0.28 to 2.30	0.676	-	-
	1	BALF	4.60	3.09 to 6.12	<0.001	-	-
CC16	3	Blood	-0.31	-2.7 to 2.08	0.799	96.8	<0.001
	4	BALF	-0.44	-3.06 to 2.18	0.742	96.1	<0.001
CRP	3	Blood	1.64	-0.31 to 3.59	0.100	92.2	<0.001
Endotoxin	2	BALF	0.30	-0.15 to 0.75	0.191	0.0	0.887
G-CSF	2	Blood	-0.49	-1.47 to 0.49	0.326	93.9	<0.001
ICAM	4	Blood	-0.14	-2.47 to 2.20	0.909	99.4	<0.001
	2	BALF	0.79	-1.61 to 3.18	0.520	96.3	<0.001
IL-2	2	Blood	0.01	-0.26 to 0.28	0.934	0.0	0.817
	1	BALF	-0.36	-1.16 to 0.44	0.380	-	-
IL-4	2	Blood	0.69	-0.16 to 1.54	0.111	81.1	0.022
	1	BALF	0.30	-0.38 to 0.99	0.387	-	-
IL-12	1	Blood	-0.8	-1.09 to -0.50	<0.001	-	-
	1	BALF	-1.47	-2.18 to -0.75	<0.001	-	-
KL-6	3	Blood	3.36	2.50 to 4.21	<0.001	49.5	0.138
	3	BALF	2.23	0.06 to 4.41	0.044	93.0	<0.001
LDH	2	Blood	1.82	-0.23 to 3.87	0.082	85.9	<0.001
MPO	1	Blood	2.58	2.20 to 2.97	<0.001	-	-
	1	BALF	1.75	0.90 to 2.60	<0.001	-	-
NF- κ β	2	BALF	0.86	-0.45 to 2.17	0.198	67.1	0.081
PCT	2	Blood	0.41	0.02 to 0.80	0.038	23.7	0.252
Protein C	2	Blood	-2.00	-7.15 to 3.16	0.447	99.9	<0.001
RAGE	4	Blood	1.64	0.15 to 3.14	0.031	96.3	<0.001
	1	BALF	0.16	-0.68 to 1.00	0.704	-	-
sE-selectin	3	Blood	0.55	0.13 to 0.97	0.011	15.2	0.307
SP-D	4	Blood	-0.05	-1.65 to 1.55	0.950	98.5	<0.001
	1	BALF	-1.17	-2.08 to -0.25	0.012	-	-
TGF- β 1	4	BALF	0.83	0.17 to 1.49	0.013	81.2	<0.001
TF	1	Blood	3.55	2.71 to 4.39	<0.001	-	-
	1	BALF	5.57	3.55 to 7.60	<0.001	-	-
TNFR-1	2	Blood	1.61	-4.42 to 7.64	0.601	98.9	<0.001
	1	BALF	5.40	4.22 to 6.58	<0.001	-	-
TNFR-2	1	Blood	3.82	2.80 to 4.83	<0.001	-	-
	2	BALF	3.22	-2.62 to 9.05	0.280	98.4	<0.001
VEGF	1	Blood	0.49	-0.23 to 1.04	0.063	-	-
	1	BALF	-4.52	-5.43 to -3.61	<0.001	-	-
vWF	6	Blood	0.81	-0.94 to 2.54	0.365	99.2	<0.001

ANG-1 Angiotensin-1, *CC16* Clara cell secretory protein, *CRP* C-reactive protein, *G-CSF* granulocyte colony-stimulating factor, *ICAM* intercellular cell adhesion molecule, *IL-2* interleukin-2, *LDH* Lactate dehydrogenase, *MPO* myeloperoxidase, *NF- κ β* nuclear factor- κ β , *PCT* procalcitonin, *RAGE* receptor for advanced glycation end products, *SP-D* surfactant protein, *TGF- β 1* transforming growth factor- β 1, *TF* tissue factor, *TNFR-1* Tumor necrosis factor receptor-1, *TNFR-2* Tumor necrosis factor receptor-2, *VEGF* vascular endothelial growth factor, *vWF* von Willebrand factor

relationship between ANG-2 and ARDS/ALI incidence, and the overall results might have been altered by more recent findings from subsequent studies.

Our current results demonstrated that ARDS/ALI patients had significantly higher serum levels of IL-1 β than individuals without ARDS/ALI. IL-1 β is synthesized and released by mononuclear macrophages. It is recognized as the primary proinflammatory cytokine that triggers inflammation and is known to ex-

ert multiple biological functions, such as promoting the activity of natural killer cells, increasing chemotaxis of macrophages and neutrophils, and regulating the immune response as an endogenous heat source [88, 89]. During infection or sepsis, IL-1 β can destroy the blood-brain barrier and increase the risk of patient mortality. IL-1 β has an inherent antagonist in the human body, IL-1ra, which can inhibit IL-1 β activity by competitively binding to its receptor.

Table 3 Subgroup analysis of inflammatory related factors and incidence of ARDS/ALI

Inflam-matory factors	Groups	No. of stud-ies	SMD	95%CI	P value	Hetero-geneity (%)	P for hetero-geneity	
ANG-2	Sample size							
	≥ 100	5	1.30	0.47 to 2.13	0.002	97.8	< 0.001	
	< 100	1	1.59	0.97 to 2.20	< 0.001	–	–	
	Mean age (years)							
	≥ 60.0	3	1.74	1.30 to 2.18	< 0.001	51.1	0.129	
	< 60.0	3	0.98	–0.02 to 1.98	0.055	98.4	< 0.001	
	Patients' status							
	ARDS	1	2.15	1.65 to 2.64	< 0.001	–	–	
	ALI	3	1.38	–0.19 to 2.93	0.085	98.3	< 0.001	
	ARDS/ALI	2	0.91	–0.34 to 2.16	0.152	93.7	< 0.001	
	Specimen source							
	Blood	6	1.34	0.59 to 2.10	< 0.001	97.4	< 0.001	
	BALF	0	–	–	–	–	–	
	IL-1β	Sample size						
≥ 100		2	–0.33	–1.05 to 0.40	0.379	79.7	0.026	
< 100		8	1.26	0.48 to 2.04	0.002	89.9	< 0.001	
Mean age (years)								
≥ 60.0		1	–0.75	–1.34 to –0.15	0.014	–	–	
< 60.0		7	1.33	0.43 to 2.23	0.004	94.4	< 0.001	
Patients' status								
ARDS		6	1.39	0.41 to 2.37	0.005	91.8	< 0.001	
ALI		4	0.22	–0.59 to 1.04	0.590	88.0	< 0.001	
Specimen source								
Blood		5	1.39	0.15 to 2.62	0.028	95.3	< 0.001	
BALF		6	0.73	–0.19 to 1.66	0.121	90.2	< 0.001	
IL-6		Sample size						
		≥ 100	5	0.45	–0.26 to 1.16	0.215	96.1	< 0.001
	< 100	8	0.83	0.11 to 1.55	0.024	91.6	< 0.001	
	Mean age (years)							
	≥ 60.0	3	1.49	0.37 to 2.61	0.009	92.1	< 0.001	
	< 60.0	10	0.43	–0.06 to 0.92	0.088	93.1	< 0.001	
	Patients' status							
	ARDS	7	0.80	0.03 to 1.57	0.043	92.6	< 0.001	
	ALI	6	0.54	–0.12 to 1.20	0.108	95.3	< 0.001	
	Specimen source							
	Blood	11	0.37	–0.04 to 0.77	0.074	88.8	< 0.001	
	BALF	7	1.33	0.24 to 2.42	0.016	93.2	< 0.001	
	IL-8	Sample size						
		≥ 100	4	0.06	–1.84 to 1.95	0.953	99.2	< 0.001
< 100		10	0.74	0.15 to 1.33	0.014	87.7	< 0.001	
Mean age (years)								
≥ 60.0		2	2.87	–1.95 to 7.70	0.243	96.6	< 0.001	
< 60.0		9	0.52	–0.61 to 1.65	0.364	98.5	< 0.001	
Patients' status								
ARDS		6	1.28	0.34 to 2.21	0.007	90.4	< 0.001	
ALI		7	0.13	–1.13 to 1.40	0.834	98.6	< 0.001	
ARDS/ALI		1	–0.37	–0.93 to 0.20	0.201	–	–	
Specimen source								
Blood		9	0.67	–0.50 to 1.85	0.262	98.3	< 0.001	
BALF		8	0.78	0.08 to 1.48	0.029	86.1	< 0.001	

Table 3 (Continued)

Inflam-matory factors	Groups	No. of stud-ies	SMD	95%CI	P value	Hetero-geneity (%)	P for hetero-geneity	
IL-10	Sample size							
	≥ 100	2	–0.90	–7.01 to 5.21	0.772	99.6	< 0.001	
	< 100	5	1.89	0.59 to 3.19	0.004	93.4	< 0.001	
	Mean age (years)							
	≥ 60.0	0	–	–	–	–	–	
	< 60.0	6	1.29	–0.75 to 3.32	0.216	98.6	< 0.001	
	Patients' status							
	ARDS	3	1.62	0.05 to 3.19	0.043	91.7	< 0.001	
	ALI	4	0.73	–2.15 to 3.61	0.618	99.0	< 0.001	
	Specimen source							
	Blood	4	0.18	–2.64 to 3.00	0.899	99.0	< 0.001	
	BALF	4	1.90	0.12 to 3.68	0.037	94.3	< 0.001	
	PAI-1	Sample size						
		≥ 100	3	0.42	–0.78 to 1.61	0.497	98.8	< 0.001
< 100		4	0.89	0.52 to 1.27	< 0.001	42.1	0.159	
Mean age (years)								
≥ 60.0		1	0.31	–0.35 to 0.96	0.363	–	–	
< 60.0		6	0.77	–0.04 to 1.57	0.063	97.6	< 0.001	
Patients' status								
ARDS		2	0.68	0.06 to 1.29	0.030	61.1	0.109	
ALI		4	0.77	–0.54 to 2.08	0.250	98.1	< 0.001	
ARDS/ALI		1	0.66	0.45 to 0.87	< 0.001	–	–	
Specimen source								
Blood		6	0.44	–0.34 to 1.22	0.267	97.2	< 0.001	
BALF		3	1.57	0.77 to 2.36	< 0.001	69.6	0.037	
TNF-α		Sample size						
	≥ 100	4	1.66	–0.10 to 3.43	0.065	98.3	< 0.001	
	< 100	12	0.76	0.27 to 1.24	0.002	85.5	< 0.001	
	Mean age (years)							
	≥ 60.0	3	0.51	0.17 to 0.84	0.003	0.0	0.482	
	< 60.0	10	1.36	0.52 to 2.20	0.002	96.0	< 0.001	
	Patients' status							
	ARDS	9	1.05	0.11 to 1.99	0.028	94.7	< 0.001	
	ALI	5	0.90	–0.14 to 1.95	0.090	95.1	< 0.001	
	ARDS/ALI	2	0.92	–0.14 to 1.97	0.089	86.5	0.007	
	Specimen source							
	Blood	9	1.00	0.12 to 1.89	0.026	95.7	< 0.001	
	BALF	10	0.91	0.26 to 1.57	0.006	87.6	< 0.001	

One study included in this analysis showed that IL-1ra is significantly upregulated in ARDS patients [41]. Anakinra is an IL-1β antagonist approved by the U.S. Food and Drug Administration (FDA) for the treatment of rheumatoid arthritis and other autoimmune diseases to reduce clinical symptoms and suppress joint destruction [90]. Determining whether this antagonistic effect can also be observed in ARDS patients is an interesting topic for future research.

IL-6 is an acute inflammatory mediator that is released by various cell types. IL-6 is not expressed un-

der normal physiological conditions but is secreted upon stimulation by inflammatory factors [91]. Our study showed that IL-6 concentrations in the BALF were significantly higher in ARDS/ALI patients than in unaffected individuals. Consistent with these results, IL-6 is recognized as a reliable and objective indicator of local lung tissue damage.

IL-8 is a member of the C-X-C subfamily of chemokines and is produced by various cell types [92]. IL-8 plays a significant role in neutrophil chemotaxis and also inhibits neutrophil apoptosis. In response to local lung tissue injury, IL-8 specifically binds to its receptor, which in turn induces neutrophil aggregation and triggers the release of proteolytic enzymes that mediate inflammation and severe tissue damage. Our findings suggested that IL-8 concentrations are associated with ARDS/ALI incidence.

IL-10 is an anti-inflammatory cytokine that inhibits the secretion of TNF- α , IL-1, and IL-6 [93]. IL-10 can also suppress NF- κ B activity and regulate the Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathway. Our analysis suggested that IL-10 levels are correlated with ARDS/ALI incidence; however, the elevated IL-10 levels could have been caused by inflammation in the lungs.

PAI-1 is mainly secreted by vascular endothelial cells, and its production is a risk factor for thrombosis and atherosclerosis [94]. During ARDS pathogenesis, the coagulation fibrinolysis system is impaired, leading to disseminated intravascular coagulation. Increased PAI-1 levels may lead to local fibrin deposition in lungs. Our results support the idea that PAI-1 plays an important role in the development of ARDS. In addition, PAI-1 has been demonstrated to promote local formation of diseased connective tissue and has been used as an indicator of prognosis of ARDS patients.

The results of our study suggested a strong correlation between TNF- α concentrations in the BALF and ARDS incidence. TNF- α is considered as one of the most important proinflammatory factors in ARDS/ALI [95]. TNF- α is a multifunctional proinflammatory factor that stimulates the secretion of endothelin and nitric oxide by endothelial cells, promotes the expression of adhesion molecules by endothelial cells and leukocytes, and contributes to the progression of severe microcirculatory disorder. Therefore, TNF- α inhibition can potentially serve as an important approach for ARDS prevention and treatment.

The general objective of this study was to identify inflammatory factors that can serve as drug targets to reduce the incidence of ARDS/ALI. Multiple lines of evidence have suggested that proinflammatory cytokines participate in or trigger the inflammatory response in the lungs; however, currently available clinical data are insufficient to verify the correlations between the levels of proinflammatory factors and ARDS/ALI incidence.

Our current meta-analysis has several limitations. First, results were based on other studies but not at the individual level. Second, the included studies showed significant heterogeneity, making it difficult to eliminate alternative explanations for the results, such as differences in the definition of ARDS/ALI, severity of disease, underlying diseases, sample collection times, and treatment strategies. Third, unpublished articles and articles written in other languages were not searched, which could have skewed the obtained results.

Conclusion

The results of our study indicated that ARDS/ALI are associated with elevated levels of ANG-2, IL-1 β , IL-6, and TNF- α , but do not significantly affect IL-8, IL-10, and PAI-1 levels. Furthermore, ARDS/ALI incidence was also determined to be significantly associated with several other inflammatory factors; however, further studies using large sample sizes are required to verify our conclusions. Future studies should also measure the levels of inflammatory factors over time. Log transformation of the measures of inflammatory factors is recommended to obtain a normally distributed data, especially in studies with small sample sizes.

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Author Contribution Daishun Liu conceived and designed the experiments. Zhenfeng Liu, Guoqi Zhou, Yugang Zou, Haixia Wang, Xiao Li and DeliangZheng performed the experiments. Zhenfeng Liu analyzed the data and wrote the paper. Daishun Liu contributed reagents/materials/analysis tools. All authors have read and approved the final version of this manuscript.

Declarations

Conflict of interest Z. Liu, D. Liu, Z. Wang, Y. Zou, H. Wang, X. Li, D. Zheng and G. Zhou declare that they have no competing interests.

Ethical standards For this article no studies with human participants or animals were performed by any of the authors. All studies cited were in accordance with the ethical standards indicated in each case. No ethical approval was required. Consent for publication: not applicable.

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References

- Gotts JE, Matthay MA. Mesenchymal stem cells and acute lung injury. *Crit Care Clin*. 2011;27:719–33.
- Rubenfeld GD, Caldwell E, Peabody E, Weaver J, Martin DP, Neff M. Incidence and outcomes of acute lung injury. *N Engl J Med*. 2005;353:1685–93.
- Brodie D, Bacchetta M. Extracorporeal membrane oxygenation for ARDS in adults. *N Engl J Med*. 2011;365:1905–14.
- Calfee CS, Janz DR, Bernard GR, May AK, Kangelaris KN, Matthay MA. Distinct molecular phenotypes of direct vs indirect ARDS in single-center and multicenter studies. *Chest*. 2015;147:1539–48.
- Morisawa K, Fujitani S, Taira Y, Kushimoto S, Kitazawa Y, Okuchi K. Difference in pulmonary permeability between indirect and direct acute respiratory distress syndrome assessed by the transpulmonary thermodilution technique: a prospective, observational, multi-institutional study. *J Intensive Care*. 2014;2:24.
- Yuan SM. Postperfusion lung syndrome: respiratory mechanics, respiratory indices and biomarkers. *Ann Thorac Med*. 2015;10:151–7.
- Andreoli SP, McAteer JA. Reactive oxygen molecule-mediated injury in endothelial and renal tubular epithelial cells in vitro. *Kidney Int*. 1990;38:785–94.
- Rockx B, Baas T, Zornetzer GA, Haagmans B, Sheahan T, Frieman M. Early upregulation of acute respiratory distress syndrome-associated cytokines promotes lethal disease in an aged-mouse model of severe acute respiratory syndrome coronavirus infection. *J Virol*. 2009;83:7062–74.
- Groeneveld AB. Vascular pharmacology of acute lung injury and acute respiratory distress syndrome. *Vascul Pharmacol*. 2002;39:247–56.
- Bhargava M, Wendt CH. Biomarkers in acute lung injury. *Transl Res*. 2012;159:205–17.
- Terpstra ML, Aman J, van Nieuw Amerongen GP, Groeneveld AB. Plasma biomarkers for acute respiratory distress syndrome: a systematic review and meta-analysis. *Crit Care Med*. 2014;42:691–700.
- Moher D, Liberati A, Tetzlaff J, Altman DG, Group P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *J Clin Epidemiol*. 2009;62:1006–12.
- Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol*. 2010;25:603–5.
- DiSipio T, Rye S, Newman B, Hayes S. Incidence of unilateral arm lymphoedema after breast cancer: a systematic review and meta-analysis. *Lancet Oncol*. 2013;14:500–15.
- Ades AE, Lu G, Higgins JP. The interpretation of random-effects meta-analysis in decision models. *Med Decis Making*. 2005;25:646–54.
- Deeks JJ, Higgins JPT, Altman DG. Analysing data and undertaking meta-analyses. In: Higgins JPT, Green S, editors. *Cochrane handbook for systematic reviews of interventions*. Version 5.1.0. London: The Cochrane Collaboration; 2008. Part 2: General methods for Cochrane reviews.
- Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. *BMC Med Res Methodol*. 2005;5:13.
- Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315:629–34.
- Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics*. 1994;50:1088–101.
- Moalli R, Doyle JM, Tahhan HR, Hasan FM, Braman SS, Saldeen T. Fibrinolysis in critically ill patients. *Am Rev Respir Dis*. 1989;140:287–93.
- Rubin DB, Wiener-Kronish JP, Murray JF, Green DR, Turner J, Luce JM. Elevated von Willebrand factor antigen is an early plasma predictor of acute lung injury in nonpulmonary sepsis syndrome. *J Clin Invest*. 1990;86:474–80.
- Roten R, Markert M, Feihl F, Schaller MD, Tagan MC, Perret C. Plasma levels of tumor necrosis factor in the adult respiratory distress syndrome. *Am Rev Respir Dis*. 1991;143:590–2.
- Parsons PE, Moore FA, Moore EE, Ikle DN, Henson PM, Worthen GS. Studies on the role of tumor necrosis factor in adult respiratory distress syndrome. *Am Rev Respir Dis*. 1992;146:694–700.
- Leff JA, Parsons PE, Day CE, Taniguchi N, Jochum M, Fritz H. Serum antioxidants as predictors of adult respiratory distress syndrome in patients with sepsis. *Lancet*. 1993;341:777–80.
- Donnelly SC, Haslett C, Dransfield I, Robertson CE, Carter DC, Ross JA. Role of selectins in development of adult respiratory distress syndrome. *Lancet*. 1994;344:215–9.
- Jorens PG, Sibille Y, Goulding NJ, van Overveld FJ, Herman AG, Bossaert L. Potential role of Clara cell protein, an endogenous phospholipase A2 inhibitor, in acute lung injury. *Eur Respir J*. 1995;8:1647–53.
- Sakamaki F, Ishizaka A, Handa M, Fujishima S, Urano T, Sayama K. Soluble form of P-selectin in plasma is elevated in acute lung injury. *Am J Respir Crit Care Med*. 1995;151:1821–6.
- Chollet-Martin S, Jourdain B, Gibert C, Elbim C, Chastre J, Gougerot-Pocidalo MA. Interactions between neutrophils and cytokines in blood and alveolar spaces during ARDS. *Am J Respir Crit Care Med*. 1996;154:594–601.
- Fuchs-Buder T, de Moerloose P, Ricou B, Reber G, Vifian C, Nicod L. Time course of procoagulant activity and D dimer in bronchoalveolar fluid of patients at risk for or with acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 1996;153:163–7.
- Ricou B, Nicod L, Lacraz S, Welgus HG, Suter PM, Dayer JM. Matrix metalloproteinases and TIMP in acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 1996;154:346–52.
- Schütte H, Lohmeyer J, Rosseau S, Ziegler S, Siebert C, Kielisch H. Bronchoalveolar and systemic cytokine profiles in patients with ARDS, severe pneumonia and cardiogenic pulmonary oedema. *Eur Respir J*. 1996;9:1858–67.
- Schwartz MD, Moore EE, Moore FA, Shenkar R, Moine P, Haenel JB. Nuclear factor-kappa B is activated in alveolar macrophages from patients with acute respiratory distress syndrome. *Crit Care Med*. 1996;24:1285–92.
- Parsons PE, Moss M, Vannice JL, Moore EE, Moore FA, Repine JE. Circulating IL-1ra and IL-10 levels are increased but do not predict the development of acute respiratory distress syndrome in at-risk patients. *Am J Respir Crit Care Med*. 1997;155:1469–73.

34. Donnelly SC, Bucala R, Metz CN, Grant IS, Robertson CR, Haslett C. Macrophage migration inhibitory factor and acute lung injury. *Chest*. 1999;116:111S.
35. Gando S, Nanzaki S, Morimoto Y, Kobayashi S, Kemmotsu O. Systemic activation of tissue-factor dependent coagulation pathway in evolving acute respiratory distress syndrome in patients with trauma and sepsis. *J Trauma*. 1999;47:719–23.
36. Armstrong L, Thickett DR, Christie SJ, Kendall H, Millar AB. Increased expression of functionally active membrane-associated tumor necrosis factor in acute respiratory distress syndrome. *Am J Respir Cell Mol Biol*. 2000;22:68–74.
37. Bauer TT, Monton C, Torres A, Cabello H, Fillela X, Maldonado A. Comparison of systemic cytokine levels in patients with acute respiratory distress syndrome, severe pneumonia, and controls. *Thorax*. 2000;55:46–52.
38. Siemiatkowski A, Kloczko J, Galar M, Czaban S. von Willebrand factor antigen as a prognostic marker in posttraumatic acute lung injury. *Haemostasis*. 2000;30:189–95.
39. Geerts L, Jorens PG, Willems J, De Ley M, Slegers H. Natural inhibitors of neutrophil function in acute respiratory distress syndrome. *Crit Care Med*. 2001;29:1920–4.
40. Hirani N, Antonicelli F, Strieter RM, Wiesener MS, Ratcliffe PJ, Haslett C. The regulation of interleukin-8 by hypoxia in human macrophages—a potential role in the pathogenesis of the acute respiratory distress syndrome (ARDS). *Mol Med*. 2001;7:685–97.
41. Park WY, Goodman RB, Steinberg KP, Ruzinski JT, Radella F 2nd, Park DR. Cytokine balance in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2001;164:1896–903.
42. Agouridakis P, Kyriakou D, Alexandrakis MG, Perisinakis K, Karkavitsas N, Bourros D. Association between increased levels of IL-2 and IL-15 and outcome in patients with early acute respiratory distress syndrome. *Eur J Clin Invest*. 2002;32:862–7.
43. Agouridakis P, Kyriakou D, Alexandrakis MG, Prekates A, Perisinakis K, Karkavitsas N. The predictive role of serum and bronchoalveolar lavage cytokines and adhesion molecules for acute respiratory distress syndrome development and outcome. *Respir Res*. 2002;3:25.
44. Endo S, Sato N, Nakae H, Yamada Y, Makabe H, Abe H. Surfactant protein A and D (SP-A, AP-D) levels in patients with septic ARDS. *Res Commun Mol Pathol Pharmacol*. 2002;111:245–51.
45. Hamacher J, Lucas R, Lijnen HR, Buschke S, Dunant Y, Wendel A. Tumor necrosis factor-alpha and angiostatin are mediators of endothelial cytotoxicity in bronchoalveolar lavages of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2002;166:651–6.
46. Takala A, Jousela I, Takkunen O, Kautiainen H, Jansson SE, Orpana A. A prospective study of inflammation markers in patients at risk of indirect acute lung injury. *Shock*. 2002;17:252–7.
47. Thickett DR, Armstrong L, Millar AB. A role for vascular endothelial growth factor in acute and resolving lung injury. *Am J Respir Crit Care Med*. 2002;166:1332–7.
48. Gessner C, Hammerschmidt S, Kuhn H, Lange T, Engelmann L, Schauer J. Exhaled breath condensate nitrite and its relation to tidal volume in acute lung injury. *Chest*. 2003;124:1046–52.
49. Grissom CK, Orme JF Jr, Richer LD, McIntyre TM, Zimmerman GA, Elstad MR. Platelet-activating factor acetylhydrolase is increased in lung lavage fluid from patients with acute respiratory distress syndrome. *Crit Care Med*. 2003;31:770–5.
50. Nys M, Deby-Dupont G, Habraken Y, Legrand-Poels S, Kohonen S, Ledoux D. Bronchoalveolar lavage fluids of ventilated patients with acute lung injury activate NF-kappaB in alveolar epithelial cell line: role of reactive oxygen/nitrogen species and cytokines. *Nitric Oxide*. 2003;9:33–43.
51. Prabhakaran P, Ware LB, White KE, Cross MT, Matthay MA, Olman MA. Elevated levels of plasminogen activator inhibitor-1 in pulmonary edema fluid are associated with mortality in acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2003;285:L20–L8.
52. Bouros D, Alexandrakis MG, Antoniou KM, Agouridakis P, Pneumatikos I, Anevlavis S. The clinical significance of serum and bronchoalveolar lavage inflammatory cytokines in patients at risk for acute respiratory distress syndrome. *BMC Pulm Med*. 2004;4:6.
53. Ishizaka A, Matsuda T, Albertine KH, Koh H, Tasaka S, Hasegawa N. Elevation of KL-6, a lung epithelial cell marker, in plasma and epithelial lining fluid in acute respiratory distress syndrome. *Am J Physiol Lung Cell Mol Physiol*. 2004;286:L1088–L94.
54. Sato H, Callister ME, Mumby S, Quinlan GJ, Welsh KL, duBois RM. KL-6 levels are elevated in plasma from patients with acute respiratory distress syndrome. *Eur Respir J*. 2004;23:142–5.
55. Parsons PE, Matthay MA, Ware LB, Eisner MD, National Heart, Lung, Blood Institute Acute Respiratory Distress Syndrome Clinical Trials Network. Elevated plasma levels of soluble TNF receptors are associated with morbidity and mortality in patients with acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2005;288(3):L426–L31.
56. El Solh AA, Bhora M, Pineda L, Aquilina A, Abbetessa L, Berbari E. Alveolar plasminogen activator inhibitor-1 predicts ARDS in aspiration pneumonitis. *Intensive Care Med*. 2006;32:110–5.
57. Calfee CS, Eisner MD, Ware LB, Thompson BT, Parsons PE, Wheeler AP. Trauma-associated lung injury differs clinically and biologically from acute lung injury due to other clinical disorders. *Crit Care Med*. 2007;35:2243–50.
58. Perkins GD, Nathani N, McAuley DF, Gao F, Thickett DR. In vitro and in vivo effects of salbutamol on neutrophil function in acute lung injury. *Thorax*. 2007;62:36–42.
59. Ware LB, Matthay MA, Parsons PE, Thompson BT, Januzzi JL, Eisner MD. Pathogenetic and prognostic significance of altered coagulation and fibrinolysis in acute lung injury/acute respiratory distress syndrome. *Crit Care Med*. 2007;35:1821–8.
60. Gallagher DC, Parikh SM, Balonov K, Miller A, Gautam S, Talmor D. Circulating angiopoietin 2 correlates with mortality in a surgical population with acute lung injury/adult respiratory distress syndrome. *Shock*. 2008;29:656–61.
61. Ganter MT, Cohen MJ, Brohi K, Chesebro BB, Staudenmayer KL, Rahn P. Angiopoietin-2, marker and mediator of endothelial activation with prognostic significance early after trauma? *Ann Surg*. 2008;247:320–6.
62. Kurzius-Spencer M, Foster K, Littau S, Richey KJ, Clark BM, Sherrill D. Tracheobronchial markers of lung injury in smoke inhalation victims. *J Burn Care Res*. 2008;29:311–8.
63. Nathani N, Perkins GD, Tunnicliffe W, Murphy N, Manji M, Thickett DR. Kerbs von Lungren 6 antigen is a marker of alveolar inflammation but not of infection in patients with acute respiratory distress syndrome. *Crit Care*. 2008;12:R12.
64. van der Heijden M, van Nieuw Amerongen GP, Koolwijk P, van Hinsbergh VW, Groeneveld AB. Angiopoietin-2, permeability oedema, occurrence and severity of ALI/ARDS in septic and non-septic critically ill patients. *Thorax*. 2008;63:903–9.
65. Calfee CS, Eisner MD, Parsons PE, Thompson BT, Conner ER Jr, Matthay MA. Soluble intercellular adhesion molecule-1

- and clinical outcomes in patients with acute lung injury. *Intensive Care Med.* 2009;35:248–57.
66. Chi XJ, Cai J, Luo CF, Cheng N, Hei ZQ, Li SR. Relationship between the expression of Toll-like receptor 2 and 4 in mononuclear cells and postoperative acute lung injury in orthotopic liver transplantation. *Chin Med J (Engl).* 2009;122:895–9.
 67. Determann RM, Millo JL, Waddy S, Lutter R, Garrard CS, Schultz MJ. Plasma CC16 levels are associated with development of ALI/ARDS in patients with ventilator-associated pneumonia: a retrospective observational study. *BMC Pulm Med.* 2009;9:49.
 68. Kropski JA, Fremont RD, Calfee CS, Ware LB. Clara cell protein (CC16), a marker of lung epithelial injury, is decreased in plasma and pulmonary edema fluid from patients with acute lung injury. *Chest.* 2009;135:1440–7.
 69. Determann RM, Royakkers A, Wolthuis EK, Vlaar AP, Choi G, Paulus F. Ventilation with lower tidal volumes as compared with conventional tidal volumes for patients without acute lung injury: a preventive randomized controlled trial. *Crit Care.* 2010;14:R1.
 70. Determann RM, Royakkers AA, Haitsma JJ, Zhang H, Slutsky AS, Ranieri VM. Plasma levels of surfactant protein D and KL-6 for evaluation of lung injury in critically ill mechanically ventilated patients. *BMC Pulm Med.* 2010;10:6.
 71. Fremont RD, Koyama T, Calfee CS, Wu W, Dossett LA, Bossert FR. Acute lung injury in patients with traumatic injuries: utility of a panel of biomarkers for diagnosis and pathogenesis. *J Trauma.* 2010;68:1121–7.
 72. Aman J, van der Heijden M, van Lingen A, Girbes AR, van Nieuw Amerongen GP, van Hinsbergh VW. Plasma protein levels are markers of pulmonary vascular permeability and degree of lung injury in critically ill patients with or at risk for acute lung injury/acute respiratory distress syndrome. *Crit Care Med.* 2011;39:89–97.
 73. Guervilly C, Lacroix R, Forel JM, Roch A, Camoin-Jau L, Papazian L. High levels of circulating leukocyte microparticles are associated with better outcome in acute respiratory distress syndrome. *Crit Care.* 2011;15:R31.
 74. Jabaudon M, Futier E, Roszyk L, Chalus E, Guerin R, Petit A. Soluble form of the receptor for advanced glycation end products is a marker of acute lung injury but not of severe sepsis in critically ill patients. *Crit Care Med.* 2011;39:480–8.
 75. Kohno T, Anzai T, Shimizu H, Kaneko H, Sugano Y, Yamada S. Impact of serum high-mobility group box 1 protein elevation on oxygenation impairment after thoracic aortic aneurysm repair. *Heart Vessels.* 2011;26:306–12.
 76. Osaka D, Shibata Y, Kanouchi K, Nishiwaki M, Kimura T, Kishi H. Soluble endothelial selectin in acute lung injury complicated by severe pneumonia. *Int J Med Sci.* 2011;8:302–8.
 77. Quesnel C, Piednoir P, Gelly J, Nardelli L, Garnier M, Lecon V. Alveolar fibrocyte percentage is an independent predictor of poor outcome in patients with acute lung injury. *Crit Care Med.* 2012;40:21–8.
 78. Schultz MJ, Determann RM, Royakkers AA, Wolthuis EK, Korevaar JC, Levi MM. Bronchoalveolar activation of coagulation and inhibition of fibrinolysis during ventilator-associated lung injury. *Crit Care Res Pract.* 2012;2012:961784.
 79. Agrawal A, Matthay MA, Kangelaris KN, Stein J, Chu JC, Imp BM. Plasma angiopoietin-2 predicts the onset of acute lung injury in critically ill patients. *Am J Respir Crit Care Med.* 2013;187:736–42.
 80. Jones SW, Zhou H, Ortiz-Pujols SM, Maile R, Herbst M, Joyner BL Jr. Bronchoscopy-derived correlates of lung injury following inhalational injuries: a prospective observational study. *PLoS ONE.* 2013;8:e64250.
 81. Hoeboer SH, Groeneveld AB, van der Heijden M, Oudemans-van Straaten HM. Serial inflammatory biomarkers of the severity, course and outcome of late onset acute respiratory distress syndrome in critically ill patients with or at risk for the syndrome after new-onset fever. *Biomark Med.* 2015;9:605–16.
 82. Roubinian NH, Looney MR, Kor DJ, Lowell CA, Gajic O, Hubmayr RD. Cytokines and clinical predictors in distinguishing pulmonary transfusion reactions. *Transfusion.* 2015;55:1838–46.
 83. Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics.* 2000;56:455–63.
 84. Meduri GU, Yates CR. Systemic inflammation-associated glucocorticoid resistance and outcome of ARDS. *Ann N Y Acad Sci.* 2004;1024:24–53.
 85. Calfee CS, Gallagher D, Abbott J, Thompson BT, Matthay MA, Network NA. Plasma angiopoietin-2 in clinical acute lung injury: prognostic and pathogenetic significance. *Crit Care Med.* 2012;40:1731–7.
 86. Zhong M, Zhang L, Wang F, Peng S, Zhang J, Xuan G. The levels of angiopoietin-2 in patients with acute respiratory distress syndrome and its value on prognosis. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue.* 2014;26:804–9.
 87. Stiehl T, Thamm K, Kaufmann J, Schaeper U, Kirsch T, Haller H. Lung-targeted RNA interference against angiopoietin-2 ameliorates multiple organ dysfunction and death in sepsis. *Crit Care Med.* 2014;42:e654–e62.
 88. Bhatia M, Moochhala S. Role of inflammatory mediators in the pathophysiology of acute respiratory distress syndrome. *J Pathol.* 2004;202:145–56.
 89. Butt Y, Kurdowska A, Allen TC. Acute lung injury: a clinical and molecular review. *Arch Pathol Lab Med.* 2016;140:345–50.
 90. Baskar S, Klein AL, Zeff A. The use of IL-1 receptor antagonist (anakinra) in idiopathic recurrent pericarditis: a narrative review. *Cardiol Res Pract.* 2016;2016:7840724.
 91. Galani V, Tatsaki E, Bai M, Kitsoulis P, Lekka M, Nakos G. The role of apoptosis in the pathophysiology of acute respiratory distress syndrome (ARDS): an up-to-date cell-specific review. *Pathol Res Pract.* 2010;206:145–50.
 92. Gu Y, Hu X, Liu C, Qv X, Xu C. Interleukin (IL)-17 promotes macrophages to produce IL-8, IL-6 and tumour necrosis factor-alpha in aplastic anaemia. *Br J Haematol.* 2008;142:109–14.
 93. Strle K, McCusker RH, Tran L, King A, Johnson RW, Freund GG. Novel activity of an anti-inflammatory cytokine: IL-10 prevents TNFalpha-induced resistance to IGF-I in myoblasts. *J Neuroimmunol.* 2007;188:48–55.
 94. Daniel AE, Timmerman I, Kovacevic I, Hordijk PL, Adriane L, Paatero I. Plasminogen activator inhibitor-1 controls vascular integrity by regulating VE-cadherin trafficking. *PLoS ONE.* 2015;10:e145684.
 95. Azevedo ZM, Moore DB, Lima FC, Cardoso CC, Bougleux R, Matos GI. Tumor necrosis factor (TNF) and lymphotoxin-alpha (LTA) single nucleotide polymorphisms: importance in ARDS in septic pediatric critically ill patients. *Hum Immunol.* 2012;73:661–7.

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