

Establishment of a rat model with ageing insomnia induced by D-galactose and para-chlorophenylalanine

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Abstract. The current study aimed to establish a rat model of ageing insomnia induced by D-galactose and/or para-chlorophenylalanine. Following establishment of the model, body weights were measured, and Morris water maze and pentobarbital-induced sleep tests were performed. The serum levels of inflammatory mediators and the neural levels of neurotransmitters were detected. The results demonstrated that the body weights of PCPA+D-gal-induced ageing insomnia rats decreased significantly. Ageing insomnia rats exhibited longer latencies to the platform in the Morris water maze tests and fewer target crossings. The sleep latency of the model rats was longer and sleep time was shorter by contrast. The relative expression of hippocampal IL-6, TNF- α , NF- κ B and mGluR2 mRNA of the PCPA+D-gal-induced ageing insomnia group was higher, while the relative expression of 5-HT_{1A}R and GABA_AR _{α 1} mRNA were lower. The serum levels of IL-1 β , IL-6, TNF- α and brain level of glutamate increased in the PCPA+D-gal-induced ageing insomnia group, while the levels of 5-HT and GABA decreased. In conclusion, memory function, sleep time, expression of inflammatory factors and neurotransmitters are altered in ageing insomnia rats induced by D-galactose and para-chlorophenylalanine, indicating the successful establishment of a murine model of ageing insomnia.

Introduction

Insomnia is a subjective experience characterized by difficulty in falling asleep and/or staying asleep, which results in poor sleep quality and/or quantity (1). The sleep characteristics of the elderly are different from those of young people (2). In China, the prevalence of insomnia is 17% and the percentage of the people whose sleep time is <6 h is 23% (3), with increasing prevalence of insomnia among the elderly (4). The prevalence of elderly insomnia in China is 47.2% (5) and 57% in America (6). Insomnia severely affects the physical health and life quality of the elderly (7-11) and increases social and economic burden (6).

The main clinical characteristic of ageing insomnia is difficulty initiating sleep (12). Ageing individuals spend more time in the lighter stages of sleep than in deep sleep in non-rapid eye movement sleep (NREM), as demonstrated by electroencephalography (13). The main treatments of ageing insomnia are psychological/behavioral therapies, pharmacological treatment or a combination of both (6). Pharmacological treatments are initially recommended when non-pharmacological options do not attain satisfying sleep (14). Currently, the commonly used ageing animal models include: D-galactose (D-gal)-induced subacute (15), β -amyloid induced (16,17), thymic senescence (18), rapid (19) and natural (20) ageing models. The ageing model induced by D-gal has been widely used (21,22) and has been evaluated in behavioral, biochemical and neurochemical aspects (23). The modeling methods of animal models of insomnia mainly include chemical reagent stimulation using intraperitoneal injection of para-chlorophenylalanine (PCPA) (24), the horizontal platform environmental deprivation method (25), the stress stimulation sleep deprivation method (26), the gentle stimulation deprivation method and the forced exercise deprivation method (27). Rat and mouse models can be used; however, rat models are optimal due to model stability in the establishment of the ageing model induced by D-gal (28). Therefore, the current study established rat models.

Inflammatory factors are closely related to sleep and ageing (29,30). Sleep has a regulatory effect on immune

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function, with pro-inflammatory cytokines reaching their peak levels during early night sleep and anti-inflammatory cytokines reaching their peak levels during daytime waking hours (31). Inflammatory cytokines interleukin (IL)-1 and tumor necrosis factor (TNF)- α participate the regulation of physiological sleep in the central nervous system (32). Inflammatory cytokines are also involved in oxidative stress, and inflammation is associated with aging (33). IL-1 β , IL-6 and TNF- α are the key inflammatory cytokines involved in oxidative stress. They are closely related to reactive oxygen species (ROS) and promote the activation of NF- κ B, as well as the expression of proinflammatory factor-induced nitric oxide synthase and cyclooxygenase-2 (34). Sleep is closely related to neurotransmitters (35). Glutamate, an important excitatory amino acid transmitter in brain tissue, can stimulate the activity of neurons (36). Gamma-aminobutyric acid (GABA), an important inhibitory neurotransmitter in the central nervous system, can inhibit neuronal excitability and exert sedative and hypnotic effects (37). Glutamate and GABA play an important role in maintaining the balance of nerve cell inhibition and excitation function (38).

In the current study, the ageing insomnia rat model was induced by continuous subcutaneous injection of D-gal and intraperitoneal injection of PCPA. The levels of senility and sleep characteristics of ageing insomnia rat model were evaluated, including memory function, sleep duration, inflammation factors and neurotransmitters. The results of the current study may provide experimental evidence for further research on ageing insomnia.

Materials and methods

Reagents. D-gal (purity, 99%) was purchased from Beijing Solarbio Science and Technology Co., Ltd. (cat no. 1013G051). PCPA and pentobarbital were purchased from Sigma-Aldrich, Merck KGaA (cat no. SHBJ7057). IL-1 β (cat no. JL20884), IL-6 (cat no. JL20896), TNF- α (cat no. JL29364), 5-hydroxytryptamine (5-HT; cat no. JL13043), glutamate (cat no. JL13664) and gamma-aminobutyric acid (GABA; cat no. JL47835) kits were purchased from Shanghai JiangLai Biotech. Co. Ltd. TRIzol[®] reagent was obtained from Invitrogen (Invitrogen; Thermo Fisher Scientific, Inc.). RevertAid First Strand cDNA Synthesis kit was purchased from Thermo Fisher Scientific, Inc. TB Green Premix Ex Taq II (Tli RNase Plus) was purchased from Takara Bio, Inc.

Animals. A total of 40 male Sprague Dawley rats (weight, 200 \pm 20 g; age, 2 months) were obtained from the Xinjiang Medical University Animal experiment center [approval protocol no. SYXK (X) 2018-003]. The rats were housed in a specific pathogen-free environment with room temperature of 25 \pm 2 $^{\circ}$ C, 12-h light/dark cycles and free access to water and food. Their health and behavior were monitored daily. The primary humane endpoints used to determine when animals should be euthanized were reduced heart and respiration rate. 'Guidelines for euthanasia of experimental animals' was followed to minimize suffering and distress of the animals (39).

Experimental procedures were conducted in accordance with the China Experimental Animals Administration

Legislation and were approved by the Ethics Committee of Xinjiang Medical University.

Establishment of rat models. Ageing rat model induced by D-gal or by PCPA are two classical rat models used in ageing and insomnia research, respectively (21,23). D-gal and PCPA were used to establish ageing insomnia rat model in the current study, which was a composite rat model. A total of 40 rats were randomly divided into 4 groups: Controls, PCPA group (insomnia group), D-gal group (ageing group) and PCPA+D-gal group (ageing insomnia group) with 10 rats in each group.

Controls were subcutaneously injected with normal saline (120 mg/kg) for 42 days, then intraperitoneally injected with normal saline (300 mg/kg) for 3 days. Rats in the PCPA group were subcutaneously injected with normal saline (120 mg/kg) for 42 days, then intraperitoneally injected with PCPA (300 mg/kg) for 3 days. Rats in the D-gal group were subcutaneously injected with D-gal (120 mg/kg) for 42 days and then intraperitoneally injected with normal saline (300 mg/kg) for 3 days. Rats in the PCPA+D-gal group were subcutaneously injected with D-gal (120 mg/kg) for 42 days and then intraperitoneally injected with PCPA (300 mg/kg) once a day for 3 days. Acute and/or adverse reactions of D-gal and PCPA were not observed during the experiment. Weight gain was measured every week and for the final time on the day 46.

Morris water maze test. The spatial memory of the rats was assessed via the Morris water maze test on day 46. The Morris Water Maze (Chengdu Taimeng Co., Ltd.) consisted of a tank (radius, 120 cm; height, 50 cm) containing water at a height of 30 cm and a temperature of \sim 25 $^{\circ}$ C. The water maze was divided into 4 quadrants and the escape platform (diameter, 12 cm) was placed at a fixed position in the 3rd quadrant, 2.5 cm under the water. The midpoint was selected as the fixed entry point in each quadrant. The rats were placed into the water with their backs to the wall of the pool and detained on the platform for 10 sec as a sign of success in finding the platform. A water maze device was used to record the time required for the rats to find the platform from the entry point, which was defined as escape latency. Rats stayed on the platform for 10 sec and then the next quadrant experiment was conducted. When the rats took >2 min to find the target platform, they were directed to the platform, stayed there for 10 sec and escape latency was recorded as 2 min. For spatial navigation training, the experiment was conducted once a day at each water entry point in each quadrant for a total of 4 times for 5 days. It is hypothesized that the time of each rat crossing the original platform within 2 min reflects the spatial memory ability, and the platform was removed only on the 6th day. Latency time to the platform and the number of target crossings were recorded and processed by a computer equipped with a TaiMeng Behavior Analysis System (Chengdu Taimeng Co., Ltd.).

Measurement of pentobarbital-induced sleeping behavior. Experiments were carried out on day 46. Pentobarbital was diluted with 0.9% physiological saline and intraperitoneally injected into the rats (35 mg/kg). The rats were then

placed into another cage. Sleep latency was recorded as time elapsed between the pentobarbital injection and the time that rats could maintain 60 sec without flipping. Sleep time was recorded as the time between the elapse and the time that rats could not continue to remain in the supine position within 30 sec.

Sample collection. Following behavioral tests, the rats were anesthetized with 10% chloral hydrate (Chengdu Kelong Chemical Co., Ltd.) by intraperitoneal injection into the abdominal cavity with 300 mg/kg (40). No sign of peritonitis was observed following the injection. No rats died during the experiment and all 40 rats were anesthetized. Blood samples (8-10 ml) were collected from the abdominal aorta. The rats died of hemorrhagic shock following blood sample collection. Death was determined by non-spontaneous breathing, lack of heartbeat and cold limbs. Cardiac and respiratory arrest were observed for 3-5 min to confirm death. Body weight at the time of sacrifice (on day 46) are shown in Table SI. Brain tissue was dissected on ice for further analysis. Experiments, including anesthesia of rats, abdominal aortic blood collection, brain tissue separation, plasma centrifugation (2,000 x g for 15 min at 4°C), liquid nitrogen quick-freezing tissue, labeling and preservation of tissue and body disposal lasted ~5 h.

ELISA. Neural serum levels of IL-1 β , IL-6, TNF- α and neurotransmitter levels of 5-HT, glutamate and in brain tissue were detected using ELISA kits, according to the manufacturer's protocol.

Reverse transcription quantitative (RT-q) PCR. The hippocampi were isolated and collected to examine the mRNA level of inflammatory cytokines and neurotransmitter receptors. RNA extraction was conducted using TRIzol[®] reagent. cDNA was synthesized using a RevertAid First Strand cDNA Synthesis kit, according to the manufacturers' instructions. RT-qPCR reactions were conducted by a CFX96 RT-qPCR system (Bio-Rad Laboratories, Inc.) using TB Green[™] Premix Ex Taq[™] II (Tli RNase Plus). The thermocycling conditions included pre-denaturation at 95°C for 10 min, denaturation at 95°C for 10 sec, annealing for 10 sec (annealing temperatures: 60°C for nuclear factor κ -light-chain-enhancer of activated B cells (NF- κ B), 5-hydroxytryptamine 1A receptor (5-HT_{1A}R) and metabotropic glutamate receptor 2 (mGluR2); 62°C for IL-6 and GABA receptor α 1 subtype (GABA_AR _{α 1}); 62.5°C for TNF- α), extension at 72°C for 10 sec, 40 cycles of extension at 95°C for 15 sec and extension at 60°C for 1 min. Measurements were performed in triplicate. The mRNA level of inflammatory cytokines and neurotransmitters was normalized to reference *GAPDH* gene and quantified as relative expression of mRNA using the 2^{- $\Delta\Delta$ C_q} method (41). The primer sequences are listed in Table I.

Statistical analysis. Each experiment was repeated three times. Data are presented as the mean \pm standard deviation or standard error. SPSS software (version 21.0; IBM Corp.) was used for statistical analysis. One-way ANOVA and Tukey's post-hoc test were performed to compare differences between groups. P<0.05 was considered to indicate a statistically significant difference.

Results

Insomnia rats and ageing insomnia rats lost body weight rapidly on day 45 due to sleep deprivation induced by PCPA injection. The body weight of the rats in different groups were initially observed following model establishment. The results demonstrated that rats in the D-gal and PCPA+D-gal groups appeared to shed hair and move slower from the 5th week, while rats in the PCPA and the PCPA+D-gal groups appeared to be excited and irascible from day 43. Weight gain in the D-gal and PCPA+D groups was significantly lower compared with controls on days 7-42 (Fig. 1). Furthermore, weight gain in the PCPA group was significantly higher compared with PCPA+D group on days 7-42. Weight gain in the PCPA and PCPA+D-gal groups was significantly lower compared with controls on day 45. Moreover, weight gain in the D-gal group was significantly higher compared with the PCPA+D-gal group on day 45.

Spatial memory ability decreases in ageing insomnia PCPA+D-gal rats. A Morris water maze was performed to assess the spatial memory ability of the rats. Rats in the PCPA+D-gal groups suffered significant impairment in spatial learning ability on account of the longer latency time compared with controls from days 2-5 (Fig. 2A). Furthermore, escape latency in the PCPA group was significantly shorter compared with the PCPA+D-gal group from days 2-5 (Fig. 2A). No significant difference of escape latency was found between the ageing D-gal and insomnia PCPA groups (Fig. 2A).

Subsequently, the spatial memory was evaluated on the 6th day. The escape latency of rats in PCPA+D-gal groups was longer compared with the control group (Fig. 2B). Furthermore, escape latency in the D-gal and PCPA groups was shorter compared with the PCPA+D-gal group (Fig. 2B).

Rats in D-gal and PCPA+D-gal groups exhibited lower target crossing numbers compared with the control group (Fig. 2C). Additionally, there were significantly more target crossings in the PCPA group compared with the PCPA+D-gal group (Fig. 2C). No significant difference was found between the PCPA and D-gal groups (Fig. 2C).

According to the results of Morris water maze tests, the current study concluded that the spatial memory ability of the PCPA+D-gal ageing insomnia rats significantly decreased compared with the control group.

Ageing insomnia PCPA+D-gal rats exhibit shorter sleep latency and sleep time. Sleep latency and sleep time through pentobarbital injections was investigated. The results demonstrated that sleep latency of PCPA and PCPA+D-gal rats was significantly longer compared with the control group (Fig. 3A). Furthermore, D-gal rats exhibited shorter sleep latency compared with the PCPA+D-gal group (Fig. 3A).

The sleep time of D-gal and PCPA+D-gal rats was significantly shorter compared with the control group (Fig. 3B). Additionally, PCPA rats exhibited longer sleep time compared with the PCPA+D-gal group (Fig. 3B). There was no significant difference between the PCPA and D-gal groups (Fig. 3B).

Table I. Forward and reverse primers used for reverse transcription quantitative PCR analysis.

Primer	Forward	Reverse
IL-6	5'-AGGAGTGGCTAAGGACCAAGACC-3'	5'-TGCCGAGTAGACCTCATAGTGACC-3'
TNF- α	5'-CCAATGGCGTGGAGCTGAGAG-3'	5'-TCTGGTAGGAGACGGCGATGC-3'
NF- κ b	5'-TGTGGTGGAGGACTTGTCTGAG G-3'	5'-AGTGCTGCCTTGTCTGTTCTTGAG-3'
5-HT _{1A} R	5'-AGGACCACGGCTACACCATCTAC-3'	5'-CTGACAGTCTTGC GGATTCCGGAAG-3'
mGluR2	5'-ATC ACTGGTGGTATTGGCGGTTCC-3'	5'-TGGCACTGGTAGAGGCGTAGC-3'
GABA _A R $_{\alpha 1}$	5'-TGAGCACACTGACTGGAAGAAGC-3'	5'-TGGTCTCAGGCGATTGTCATAACC-3'
GAPDH	5'-GACATGCCGCCTGGAGAAAC-3'	5'-AGCCCAGGATGCCCTTTAGT-3'

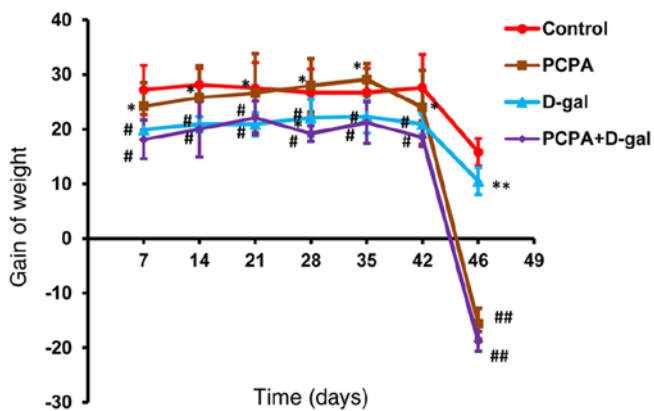


Figure 1. Weight gain in rats was measured each week and for the final time on day 46. Data are presented as mean \pm standard deviation. * $P < 0.05$ and ** $P < 0.01$ vs. control group. # $P < 0.05$ and ## $P < 0.01$ vs. PCPA+D-gal group. PCPA, para-chlorophenylalanine; D-gal, D-galactose.

Expression of pro-inflammatory cytokines and neurotransmitter receptors at mRNA level are also influenced by D-gal and PCPA injections. Hippocampal IL-6, TNF- α , NF- κ B, 5-HT_{1A}R, mGluR2 and GABA_AR $_{\alpha 1}$ mRNA expression was determined using RT-qPCR to establish whether inflammatory cytokines and neurotransmitter receptors were involved in cognitive impairment and insomnia. The results demonstrated that IL-6, TNF- α and NF- κ B mRNA expression were upregulated in the hippocampus of rats in the PCPA, D-gal and PCPA+D-gal groups compared with the control group (Fig. 4A-C). However, there was no significant difference in the IL-6, TNF- α and NF- κ B mRNA expression levels in the PCPA and D-gal groups compared with the PCPA+D-gal group. Additionally, the PCPA and PCPA+D-gal groups demonstrated downregulated 5-HT_{1A}R mRNA expression compared with the control group (Fig. 4D). The PCPA, D-gal and PCPA+D-gal groups demonstrated downregulated GABA_AR $_{\alpha 1}$ mRNA expression compared with the control group (Fig. 4F). No significant difference was discovered between D-gal, PCPA and PCPA+D-gal groups. Furthermore, the PCPA and PCPA+D-gal groups exhibited upregulated mGluR2 mRNA expression compared with the control group (Fig. 4E). The D-gal group also demonstrated downregulated mGluR2 mRNA expression compared with the PCPA+D-gal group. The results indicate that the subcutaneous injection of D-gal, intraperitoneal injection of PCPA and combined

injection of D-gal and PCPA altered inflammatory cytokine and neurotransmitter receptor mRNA expression in the rat hippocampus.

Expression levels of pro-inflammatory cytokines in neural serum and neurotransmitters in neural tissue. Subsequently, ELISA assays were used to investigate the expression of pro-inflammatory cytokines and neurotransmitters. Serum expression of IL-1 β , IL-6 and TNF- α in the PCPA+D-gal rats were significantly increased compared with the control group (Fig. 5A-C). Furthermore, the levels of these cytokines in the PCPA and D-gal rats were significantly decreased compared with the PCPA+D-gal group. No significant difference of IL-1 β , IL-6 and TNF- α was found between the PCPA and D-gal groups. The results indicated that the subcutaneous injection of D-gal, intraperitoneal injection of PCPA and combined injection of D-gal and PCPA all induced significant chronic inflammation in the rats.

Furthermore, neural levels of 5-HT, glutamate and GABA were measured. The results demonstrated a significant decrease in the levels of 5-HT in the PCPA+D-gal groups compared with the control group (Fig. 5D). The levels of 5-HT in brains of D-gal and PCPA rats were significantly higher compared with the PCPA+D-gal group. The PCPA+D-gal groups exhibited a significant increase in the levels of glutamate compared with the control group (Fig. 5E). Furthermore, neural glutamate levels in the D-gal and PCPA rats were significantly decreased compared with the PCPA+D-gal group (Fig. 5E). The PCPA and PCPA+D-gal groups exhibited a significant decrease in GABA levels compared with the control group (Fig. 5F). No significant difference was found in the levels of 5-HT, glutamate and GABA between the D-gal and PCPA groups. These results demonstrated that the subcutaneous injection of D-gal, intraperitoneal injection of PCPA and the combined injection of D-gal and PCPA induced expression changes of the neurotransmitters in brains of the rats.

Discussion

Oxidative damage induced by ROS serves a crucial role in the pathophysiology of ageing (42,43). High doses of D-gal induce ROS overexpression via the metabolism of D-gal (44). The injection of D-gal results in the increase of oxygen free radicals in the rat brain (21). Additionally, ageing alterations induced by D-gal are similar to natural ageing processes (45). A previous study demonstrated that the levels of neurotransmitters in

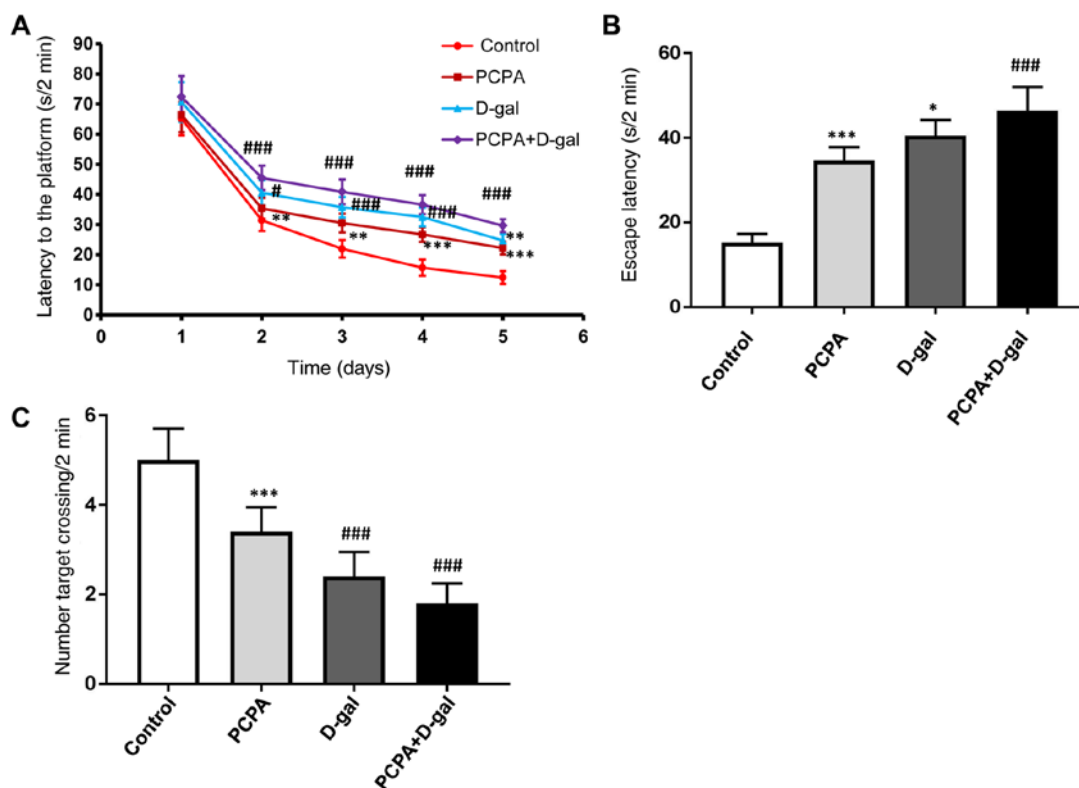


Figure 2. The spatial memory ability of rats was observed using a Morris water maze test. (A) Latency to the platform within 2 min during 5 days spatial navigation training (s/2 min). (B) Escape latency to the platform within 2 min on day 6 (s/2 min). (C) The number of target crossings within 2 min. Data are presented as mean \pm SEM. # P <0.05 and ### P <0.001 vs. control group. * P <0.05, ** P <0.01 and *** P <0.001 vs. PCPA+D-gal group. PCPA, para-chlorophenylalanine; D-gal, D-galactose.

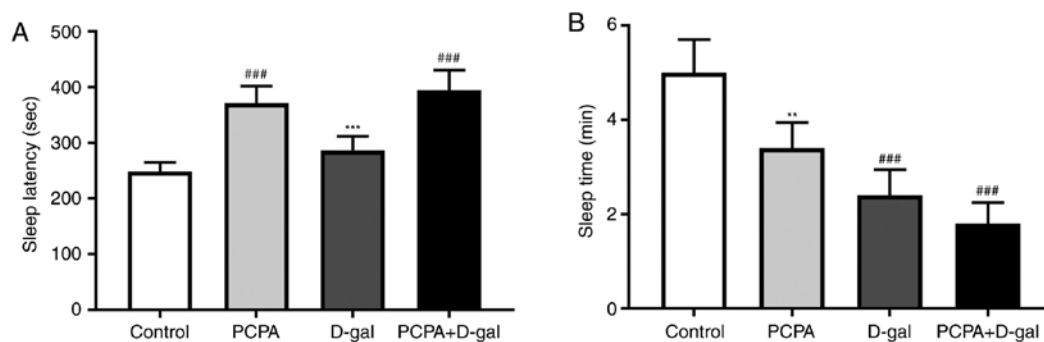


Figure 3. Pentobarbital (35 mg/kg) was intraperitoneally injected into rats to observe sleep latency and sleep time. The (A) sleep latency and (B) sleep time were recorded. Data are presented as mean \pm SEM. ### P <0.001 vs. control group. ** P <0.01 and *** P <0.001 vs. PCPA+D-gal group. PCPA, para-chlorophenylalanine; D-gal, D-galactose.

the brain of PCPA-induced insomnia rats were altered (23). Therefore, D-gal and PCPA were used in this study to establish rat models of ageing and insomnia, respectively. Furthermore, another previous study reported that D-gal-induced ageing rats lose weight rapidly compared with the control group (46). In the current study, the results indicated that rats in both D-gal and PCPA+D-gal groups lost weight rapidly compared with control rats.

Cognitive decline increases with age during natural ageing (47,48). It has been reported that in the Morris water maze, the latency time of D-gal rats was longer, the number of target crossings was lower and that cognitive function decreased (49). Sleep disorder is closely related to

cognition (50) and patients with primary insomnia have been reported to suffer from subjective memory impairment in virtual water mazes (51). Sleep duration is negatively correlated with age in older adults (52) and chronic insomnia in the elderly is likely to exacerbate cognitive impairment (53). In the current study, D-gal-induced ageing rats, PCPA-induced the insomnia rats and D-gal- and PCPA-induced ageing insomnia rats demonstrated cognitive decline in the Morris water maze. Therefore, the ageing and insomnia model exhibited decreased cognitive function.

Chronic inflammation is closely related to ageing (54). Inflammatory factors, such as IL-1 β , IL-6 and TNF- α , serve roles in D-gal-induced oxidative stress to simulate the

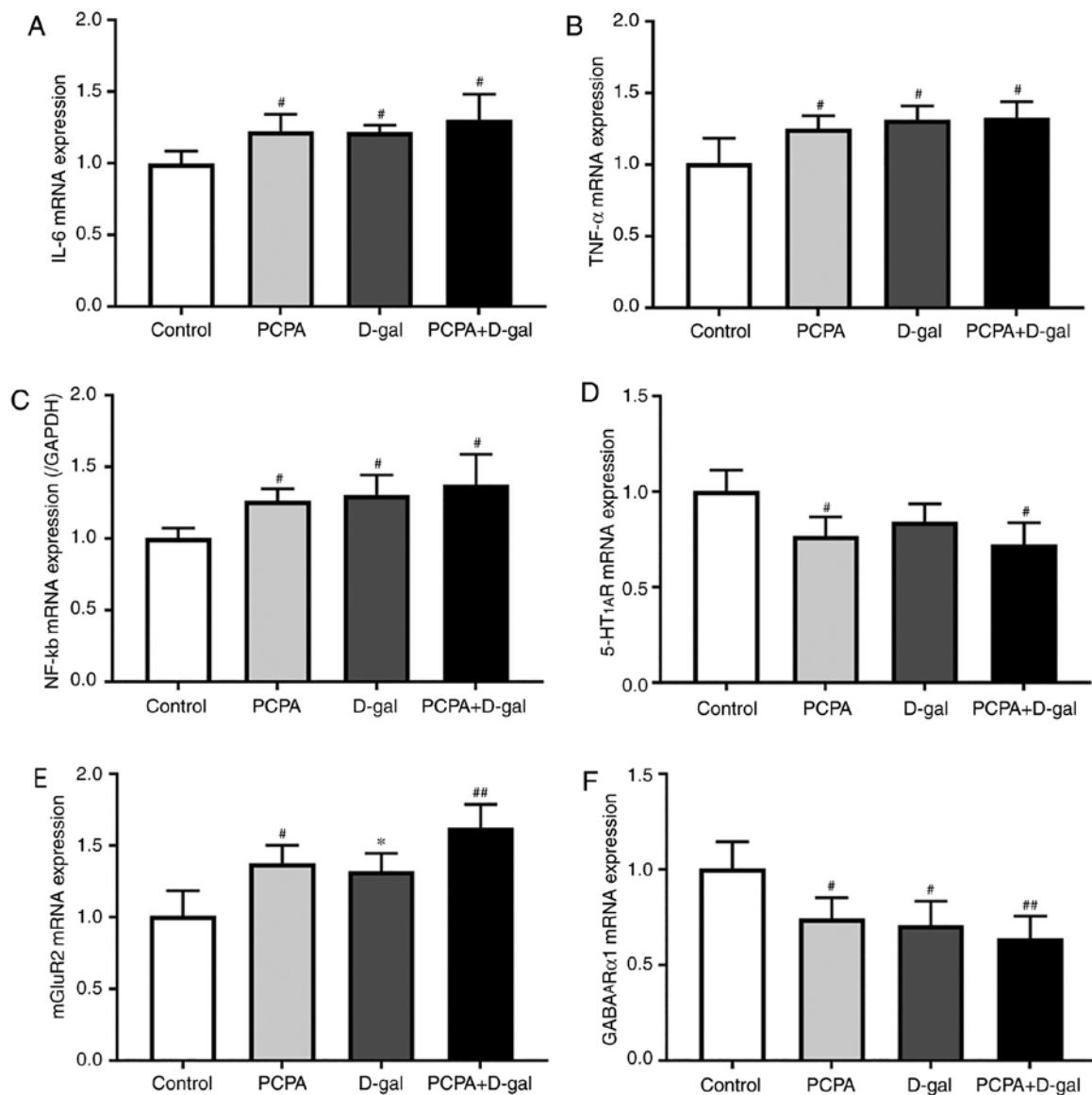


Figure 4. Gene expression of inflammatory factors IL-6, TNF- α , NF- κ B and neurotransmitter receptors of 5-HT $_{1A}$ R, mGluR2, Gamma-aminobutyric acid A receptor α 1 subtype in the hippocampus were assayed using RT-PCR. mRNA expression of (A) IL-6, (B) TNF- α , (C) NF- κ B, (D) 5-HT $_{1A}$ R, (E) mGluR2 mRNA, and (F) GABA $_{A\alpha 1}$. Data are presented as mean \pm SEM. # P <0.05, ## P <0.01 vs. control group. * P <0.05 vs. PCPA+D-gal group. IL-6, interleukin 6; TNF- α , tumor necrosis factor α ; NF- κ B, nuclear factor κ -light-chain-enhancer of activated B cells; 5-HT $_{1A}$ R, 5-hydroxytryptamine 1A receptor; mGluR2, metabotropic glutamate receptor 2.

ageing process (46). It has been reported that the plasma concentrations of IL-1 β , IL-6 and TNF- α in the D-gal and PCPA induced ageing insomnia rats were significantly higher compared with the control group (55). Furthermore, inflammatory cytokines affect neurotransmitters related to sleep, such as norepinephrine and 5-HT, which are transmitted to the central nervous system (56,57). IL-1 β , IL-6 and TNF- α are the key inflammatory factors related to sleep regulation (58), and IL-1 β increases non-rapid eye movement sleep in electroencephalography (59). The I- κ /NF- κ B signaling pathway is associated with sleep regulation and the immune system and hippocampus play a central role in insomnia (60). A previous study demonstrated that plasma levels of IL-6 and TNF- α in PCPA-induced insomnia rats were significantly higher compared with the control group (25). In the current study, the results demonstrated that the serum content of IL-1 β , IL-6 and

TNF- α and mRNA expression of IL-6, TNF- α and NF- κ B in neural tissues in the PCPA+D-gal group were higher compared with control rats.

Glutamate is an important excitatory amino acid transmitter in brain tissue (36). mGluR2 is distributed in both pre- and post-synaptic neurons, inhibits adenylate cyclization and regulates ion channel receptors by coupling with Gi/o, and negatively regulates neurotransmitter release (61). mGluR2 is involved in the physiology of sleep (62). Additionally, GABA is a major inhibitory neurotransmitter in the central nervous system (37). GABA inhibits neuronal excitation in the nervous system, and GABA receptors are widely used in the treatment of anxiety disorder, insomnia, epilepsy (62). Glutamate and GABA serve important roles in maintaining the stability of the balance between the inhibitory and excitatory functions of nerve cells (63). It has been demonstrated the amount of GABA

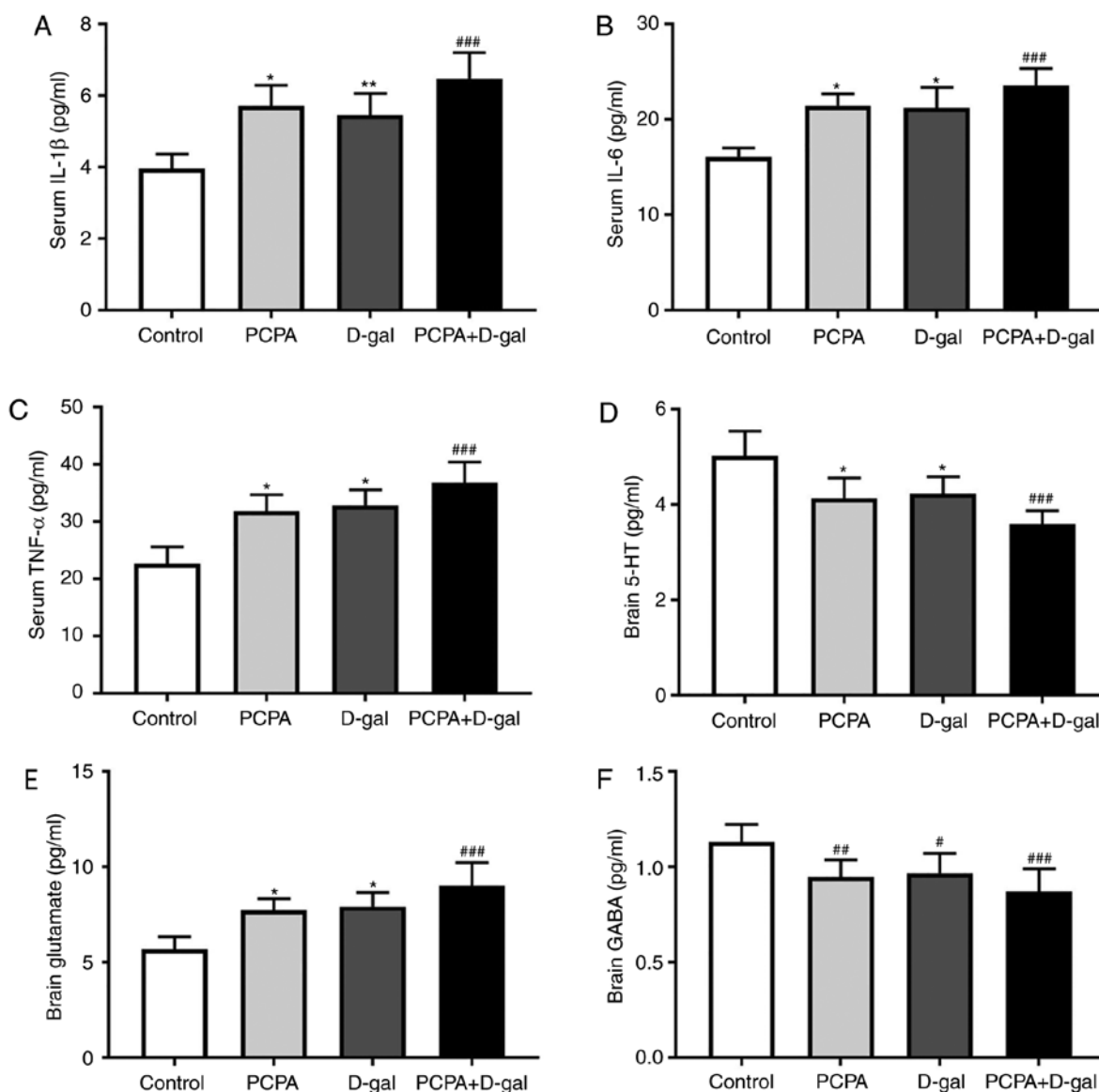


Figure 5. Protein expression of inflammatory factors IL-1 β , IL-6, TNF- α in serum and expression of 5-HT, glutamate, GABA in neural tissue were assayed via ELISA. Serum (A) IL-1 β , (B) IL-6 (C), TNF- α and neural (D) 5-HT, (E) glutamate and (F) GABA. Data are presented as mean \pm SEM. * P <0.05, ** P <0.01 and *** P <0.001 vs. control group. * P <0.05, ** P <0.01 vs. PCPA+D-gal group. IL, interleukin, TNF- α , tumor necrosis factor α ; 5-HT, 5-hydroxytryptamine; GABA, gamma-aminobutyric acid; PCPA, para-chlorophenylalanine; D-gal, D-galactose.

in the brain tissue is positively associated with the changes in sleep-wake depth (64). Furthermore, GABA serves inhibitory roles by binding to its receptor (65) and GABA_AR $_{\alpha 1}$ has a sedative effect (66). It has been reported that the expression of glutamate in cerebral cortex, hypothalamus and hippocampus of in PCPA-induced insomnia rats increased, while the content of 5-HT and GABA decreased compared with the control group (23). Woo *et al* (67) found that the expression of GABA_AR $_{\alpha 1}$ protein was upregulated in the hypothalamus of mice with improved sleep time. In the current study, the results demonstrated that expression of 5-HT and GABA decreased while glutamate increased in the D-gal- and PCPA-induced ageing insomnia rats compared with the control group. This indicated that ageing insomnia may be associated with 5-HT, glutamate and GABA neurotransmitters.

The present study established an ageing insomnia rat model by injection of D-galactose and PCPA. Changes in

memory ability, inflammatory factors and neurotransmitters of model rats were observed. Future studies should include more behavioral tests, age-related redox marker detections (oxidative stress), apoptotic proteins and body weight/5-HT measurements in order to verify the results of the present study and elucidate ageing insomnia in the rat model.

In conclusion, the current study established an ageing insomnia rat model by an injection combination of D-gal with PCPA, and evaluated the changes in cognitive behavior, sleep duration, inflammation factors and neurotransmitters in PCPA+D-gal ageing insomnia rats. This ageing insomnia rat model induced by D-gal and PCPA may provide experimental model for further research on ageing insomnia.

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Availability of data and materials

The datasets used and/or analysed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

XR, QW, XZ and ND designed the study; XZ supplied the samples for this study; XR, GW, TL and DY analyzed the data; XZ supervised the whole study; XR and XZ wrote the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

All experimental procedures were conducted in accordance with China Experimental Animals Administration Legislation and were approved by the Ethics Committee of Xinjiang Medical University (Urumqi, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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