# REVIEW



# Global status of Azithromycin and Erythromycin Resistance Rates in *Neisseria* gonorrhoeae: A Systematic Review and Metaanalysis

Zhiwei Lu<sup>a,b</sup>, Danyal Abbasi Tadi<sup>c</sup>, Jinchao Fu<sup>d,\*</sup>, Khalil Azizian<sup>e</sup>, and Ebrahim Kouhsari<sup>f,g,\*</sup>

<sup>a</sup>Graduate School, Zhejiang Chinese Medical University, Hangzhou, Zhejiang, China; <sup>b</sup>Hangzhou Heyunjia Hospital, Hangzhou, Zhejiang, China; <sup>c</sup>Department of Veterinary, Azad University of Shahr-e Kord, Shahr-e Kord, Iran; <sup>d</sup>Department of General Practice, Shulan (Hangzhou) Hospital Affiliated to Zhejiang Shuren University Shulan International Medical College, Hangzhou, Zhejiang, China; <sup>e</sup>Department of Microbiology, Faculty of Medicine, Kurdistan University of Medical Sciences, Sanandaj, Iran; <sup>f</sup>Laboratory Sciences Research Center, Golestan University of Medical Sciences, Gorgan, Iran; <sup>a</sup>Department of Laboratory Sciences, Faculty of Paramedicine, Golestan University of Medical Sciences, Gorgan, Iran

**Background**: The widespread development of antibiotic resistance or decreased susceptibility in *Neisseria gonorrhoeae* (NG) infection is a global and significant human public health issue. **Objectives**: Therefore, this meta-analysis aimed to estimate worldwide resistance rates of NG to the azithromycin and erythromycin according to years, regions, and antimicrobial susceptibility testing (AST). **Methods**: We systematically searched the published studies in PubMed, Scopus, and Embase from 1988 to 2021. All analyses were conducted using Stata software. **Results**: The 134 reports included in the meta-analysis were performed in 51 countries and examined 165,172 NG isolates. Most of the included studies were from Asia (50 studies) and Europe (46 studies). In the metadata, the global prevalence over the past 30 years were 6% for azithromycin and 48% for erythromycin. There was substantial change in the prevalence of macrolides NG resistance over time (P < 0.01). In this metadata, among 58 countries reporting resistance data for azithromycin, 17 (29.3%) countries reported that >5% of specimens had azithromycin resistance. **Conclusions**: The implications of this study emphasize the rigorous or improved antimicrobial stewardship, early diagnosis, contact tracing, and enhanced intensive global surveillance system are crucial for control of further spreading of gonococcal emergence of antimicrobial resistance (AMR).

\*To whom all correspondence should be addressed: Jinchao Fu, Email: Jinchaofu0307@163.com; Ebrahim Kouhsari, Email: Ekouhsari1987@gmail.com; ORCID: 0000-0001-5893-6483.

Abbreviations: NG, *Neisseria gonorrhoeae*; AST, antimicrobial susceptibility testing; AMR, antimicrobial resistance; WPR, weighted pooled resistance rate; CLSI, Clinical and Laboratory Standards Institute; EUCAST, European Committee on Antimicrobial Susceptibility Testing.

Keywords: Animicrobial Resistance, *Neisseria gonorrhoeae*, antibiotic resistance, azithromycin, erythromycin, systematic review and meta-analysis

Author Contributions: ZL, DAT, JF contributed to the conception and design of the work. KA and EK contributed to design of the work, and final approval of the version to be published. ZL and JF contributed in drafting the work and revising it critically for important intellectual content. JF and EK contributed in revising the article and final approval of the version to be published.

# INTRODUCTION

Sexually transmitted infections (STIs) are a significant human health issue around the world [1,2]. Gonorrhea, as one of the most frequent bacterial STIs, continues to be a major threat to public health [3]. In 2020, the WHO estimated that there were 82.4 million (95% CI 47.7 million-130.4 million) new cases of gonorrhea among adolescents and adults. Antimicrobial resistance (AMR) in Neisseria gonorrhoeae (NG) is a developing clinical and public health challenge [3]. Emerging azithromycin-resistance among the NG reported in most countries which would lead to an increase in serious problems, including infertility, ectopic pregnancy, and increased transmission of HIV [4]. Thus, an effective and inexpensive anti-infective agent is a necessity to control gonorrhea. The aim of this meta-analysis is: (1) to assess the weighted pooled resistance rate (WPR: proportion of strains resistant to specific antibiotic agent) rates of NG to azithromycin and erythromycin according to years, regions, and antimicrobial susceptibility testing (AST).

#### Methods

This review is reported accordant with the Preferred Reporting Items for Systematic Reviews and Meta Analyses guidelines (PRISMA) [5].

#### Search Strategy and Study Selection

We systematically searched for studies in four international databases including PubMed, Scopus, and Embase (until August 2021) by utilizing the related keywords: ("*Neisseria gonorrhoeae*" OR "gonorrhoea" OR "Gonococcus") AND ("antimicrobial resistance" OR "antibiotic resistance" OR "macrolides resistance" OR "azithromycin resistance" "erythromycin resistance") in the Title/Abstract/Keywords fields. The search strategy was designed and conducted by study investigators.

Reference lists of all studies were also reviewed for any other related publication. The records found through database searching were merged and the duplicates were removed using EndNote X8 (Thomson Reuters, New York, NY, USA). One of the team researchers randomly assessed the search results and confirmed that no relevant study had been ignored. All these steps were done by the three authors and any disagreements about article selection were resolved through discussion, and a fourth author acted as arbiter.

# Inclusion and Exclusion Criteria

The included articles met the following criteria: (1) original study that investigated NG AMR in human clinical isolates; (2) peer-reviewed articles published in English between January 1980 and August 2021; (3) sample

size of NG isolates; (4) the methods used for resistance testing (MIC-based methods, disk diffusion, mix methods (both methods)); (5) reported the AMR rate in NG isolates following the criteria by the Clinical and Laboratory Standards Institute (CLSI) standards [6], European Committee on Antimicrobial Susceptibility Testing (EUCAST) [7], and/or the WHO WPR Resistance Surveillance Programme guidelines [8]. The clinical azithromycin resistant breakpoints were interpreted using the EUCAST (>0.5 µg/mL) and CLSI (epidemiological cutoff value  $\geq 2.0 \ \mu g/mL$ ). In 2019, EUCAST and CLSI excluded their clinical breakpoints for azithromycin and an epidemiological cutoff value (MIC >1 mg/L) has been used for the identification of non-susceptible isolates with determinants of azithromycin resistance [9]. Studies were excluded if they contained duplicate data or were overlapping articles, (2) animal research, reviews, meta-analvsis and/or systematic review, conference abstracts, and macrolides resistance were not evidently described.

### Data Extraction and Assessment of Study Quality

Data extracted from each included study was; author, study period, publication year, regions, sample size of NG isolates, sample size of macrolides resistant NG isolates, and AST (disk diffusion, agar dilution, microbroth dilution, E-test). The Newcastle-Ottawa scale (NOS) was adapted for cross-sectional studies for quality assessment [10]. NOS is based on a star scoring system, which a maximum of 8 (for cross-sectional studies) can be awarded to each study. Quality assessment was checked independently by two authors, and any disagreements were solved by the third. Studies that received a score of 6 or above were considered to be high quality.

#### Statistical Analysis

Cross-sectional articles presenting eligible information on macrolides susceptibility in NG isolates were pooled and analyzed based on random-effects model with Stata/SE software, v.17.1 (StataCorp, College Station, TX, USA). Freeman-Tukey double arcsine transformation was performed to estimate the WPR. The inconsistency across studies was examined by the forest plot as well as the I<sup>2</sup> statistic. Values of I<sup>2</sup> (25%, 50%, and 75%) were interpreted as the presence of low, medium, or high heterogeneity, respectively. So, the DerSimonian and Laird random effects models were used [11]. Macrolides resistance rate was expressed as percentage and 95% confidence interval (CI) basis. Publication bias was calculated using Egger's test.

# Study Outcomes

The primary outcome was the WPR rate of NG to azithromycin and erythromycin resistance. Subgroup



Figure 1. Flow chart of study selection.

analyses were then utilized by year publication (1988-2013, 2014-2018, and 2019-2021), (2) regions, (3) AST, and (4) interpretation of resistance (CLSI, EUCAST, and WHO).

# RESULTS

# Results of the Systematic Literature Search

Initially, a total of 2,350 studies were identified. The full texts of the remaining 505 articles were reviewed. Ultimately, 134 studies were included [12-145] based on their irrelevance/duplication and the inclusion and exclusion criteria (Figure 1). The 134 reports included in the analysis were conducted in 51 countries, examined 165,172 NG isolates, and were published between 1988 and 2021. The main group of included articles (50 studies) were conducted from Asia and Europe (46 studies) (Table 1). The macrolides WPR rates and the subgroup analyses by year, continents, AST, and interpretation of resistance are shown in Table 1.

#### Azithromycin Resistance

The susceptibility to azithromycin was determined in 133 studies included 165,172 NG isolates; the WPR was 6% (95% CI 5%-7%) with substantial heterogeneity (I<sup>2</sup>=98.76%; P=0.01) was observed between included studies (Table 1). Also, significant publication bias was detected (Egger rank correlation test, P < 0.01). To analyze the trends for changes in the prevalence of azithromycin resistance in more recent years, we performed a subgroup analysis for three periods (1988-2013; 2014-2018; and 2019-2021) (Table 1, Figure 2). The subgroup analysis that compared the data from 1988-2013 (WPR 2.31%; 95% CI 1.40%-3.40%), 2014-2018 (WPR 6.85%; 95% CI 4.62%-9.46%), and 2019-2021 (WPR 7.25%; 95% CI 5.62%-9.06%) indicated an increase in the resistance rate. However, there was significant variation in the proportion of azithromycin resistance isolates over time (P < 0.01). Among 51 countries reporting resistance data for azithromycin, 22 (43.13%) countries (China, Bangladesh, Brazil, Guyana, Cuba, Israel, Korea, Germany, Vietnam, Italy, Belarus, Norway, Hungary, Slovenia, Estonia, Poland, Spain, Iran, Argentina, Ethiopia, Ireland, and Côte d'Ivoire) reported that >5% of specimens had azithromycin resistance (Figures 3 and 4). There was a statistically significant difference in the azithromycin resistance rates between different continents (P < 0.01); and this rate was higher in South America than Asia (9.2% vs. 6.3%), Europe (9.2% vs. 5.8%), Africa (9.2% vs. 5.3%), North America (9.2% vs. 2.1%), and Oceania (9.2% vs.1.7%) (Figure 4). No significant difference was found in the method used for AST and the interpretation of resistance (P=0.36).

Variables	Number of Studies (n, N)	Prevalence (%) of Resistance (95% Cl)	Heterogeneity (I <sup>2</sup> ) (%)	Egger test
Azithromycin	133 (6104, 165172)	6.00 (5.00-7.00)	98.76	<0.01
Publication date				
1988-2013	34 (409, 52191)	2.31 (1.40-3.40)	94.85	
2014-2018	46 (2018, 68318)	6.85 (4.62-9.46)	98.82	
2019-2021	53 (3677, 44013)	7.25 (5.62-9.06)	97.22	
Continent				
North America	15 (819, 96842)	2.13 (1.10-3.43)	98.71	
Asia	49 (1945, 17500)	6.36 (4.07-9.07)	97.48	
Africa	15 (307, 2995)	5.28 (0.35-14.15)	98.32	
Europe	45 (2887, 45035)	5.88 (4.63-7.26)	96.19	
South America	8 (139, 1752)	9.27 (4.89-14.79)	90.68	
Oceania	1 (7, 398)	1.76 (0.71-3.59)	0.00	
Interpretation of resistance				
CLSI	65 (1823, 112801)	4.00 (3.00-5.00)	98.20	
EUCAST	60 (3237, 44863)	7.00 (6.00-9.00)	96.64	
WHO	8 (1044, 6858)	8.00 (3.00-14.00)	98.23	
AST				
MIC-based methods	116 (5772, 160099)	5.64 (4.46-6.93)	98.86	
Disk diffusion	10 (185, 2005)	9.24 (1.79-20.78)	94.38	
Mix methods	7 (147, 2418)	2.76 (0.376.80)	97.65	
Erythromycin	16 (3010, 13161)	21 (13-29)	98.69	0.68
Publication date				
1988-2013	11 (1423, 8186)	17.16 (6.08-32.12)	98.37	
2014-2018	1 (18, 25)	72.00 (50.61-87.93)	0.00	
2019-2021	4 (1569, 4950)	21.50 (7.17-40.61)	97.74	
Continent				
North America	4 (2713, 12123)	13.61 (6.01-23.62)	99.26	
Asia	3 (58, 176)	36.31 (0.00-93.16)	-	
Africa	4 (62, 223)	23.74 (0.00-88.05)	99.00	
Europe	5 (177, 639)	17.52 (0.45-49.43)	98.68	
South America		-	-	
Interpretation of resistance				
CLSI	10 (2791, 12527)	17.00 (9-26)	98.97	
EUCAST	5 (219, 558)	37.00 (13-64)	97.42	
WHO	1 (0, 76)	0.00 (0.00-2)	-	
AST				
MIC-based methods	1 (2, 12974)	24.11 (15.46-33.95)	98.92	
Disk diffusion	3 (18, 156)	12.58 (0.00-64.08)	0	
Mix methods	12 (2990, 31)	6.45 (0.79-21.42)	0	

 Table 1. Subgroup Analysis for Azithromycin and Erythromycin Resistance Rates

CI, confidence interval; n, number of events (drug resistance); N, total number of isolates from the included studies



Figure 2. The prevalence of antibiotics weighted pooled resistance over years.

#### Erythromycin Resistance

The susceptibility to erythromycin was determined in 16 studies that included 13,161 NG isolates; the WPR was 21% (95% CI 13%-29%) with substantial heterogeneity (I<sup>2</sup>=98.69%, P < 0.01) was observed between included studies (Table 1). Also, no significant publication bias was detected (Egger rank correlation test, P=0.68). As shown in the Table 1 and Figure 2, the prevalence of erythromycin resistance notably increased from 17.16% (95% CI 6.08-32.12) of 18 strains 1988-2013 reaching 72% (95% CI 50.61-87.93) of 1,569 strains in 2014-2018. Regarding erythromycin resistance, a huge increase was noted from the earlier time period of 1988-2013 to 2014-2018, but this was based on a single study [146] with very few isolates. The frequency of erythromycin resistance during the years 2019-2021 represents a gradual increase over the years 1988-2013. However, there was significant variation in the proportion of erythromycin resistance isolates over time (P < 0.01). Among 11 countries reporting resistance data for erythromycin, 4 (~22%) countries (Bangladesh, Nigeria, Denmark, and Belarus) reported that >25% of specimens had resistance. There was a no statistically significant difference in the erythromycin resistance rates between different continents (P=0.83). No significant difference was found in the method used for AST (P=0.08). No significant difference was found in the interpretation of resistance based on CLSI, EUCAST, and WHO (P=0.79).

#### DISCUSSION

This metadata was conducted to study the prevalence of macrolides resistance in NG isolates around the world. It is proposed for the implication of continuous and robust monitoring of AMR of clinical NG isolates. NG AMR has become an important issue around the world in the past few decades [1,2]. In our metadata, the global WPRs over the past 30 years are higher than the 5% for azithromycin (6%) and erythromycin (21%). The WHO recommends that alternative agents should be used when NG AMR rate reaches a 5% threshold [147,148].

Ceftriaxone is the last choice for empirical first-line gonorrhea monotherapy in many countries [9]. However, emergence of decreased susceptibility or resistance to ceftriaxone have been stated globally. Currently most countries commonly suggest dual antimicrobial therapy (ceftriaxone 250-500mg plus azithromycin 1-2g) as empirical first-line gonorrhea treatment to prevent the emergence/accumulation of AMR [147]. However, as documented in the WHO's global gonococcal AMR surveillance 2017-18, there are major gaps in NG AMR surveillance [149]. The emergence of azithromycin and/or ceftriaxone resistance has been developing rapidly, which raises significant global challenges over the continued efficacy of current gonorrhea treatments [150-153]. During 2018, the WHO Gonococcal AMR Surveillance Programme (WHO-GASP) reported data on azithromycin resistance in NG among 61 countries in WHO regions; 44 (72%) countries reported that > 5% of specimens had resistance [1,2,9]. In this metadata, among 58 countries reporting resistance data for azithromycin, 17 (29.3%) countries (China, Bangladesh, Japan, Korea, Italy, Slovenia, Portugal, Slovenia, Iran, Denmark, Spain, and Ethiopia) reported that > 5% of specimens had azithromycin resistance. The area discrepancies in azithromycin-resistance rate of NG isolates may result from gonorrhea control policy, antibiotic stewardship, the lack of an



Figure 3. Global map of reported weighted pooled resistance rates for (A) azithromycin and (B) erythromycin in the study.

				Effect size		
Study				with 95% CI		
Azithromycin						
North America				2.13 [ 1.10, 3.43]		
Asia				6.36 [ 4.07, 9.07]		
Africa				5.28 [ 0.35, 14.15]		
Europe				5.88 [ 4.63, 7.26]		
South America				9.27 [ 4.89, 14.79]		
Oceania				1.76 [ 0.71, 3.59]		
		٠		5.63 [ 4.51, 6.86]		
Erythromycin						
North America				13.61 [ 6.01, 23.62]		
Asia				36.31 [ 0.00, 93.16]		
Africa				23.74 [ 0.00, 88.05]		
Europe			-	17.52 [ 0.45, 49.43]		
		٠		20.59 [ 13.09, 29.21]		
-5	0	0	50	100		

Figure 4. Forest plot for the summary of azithromycin and erythromycin WPRs by regions.

implementing response plans, antimicrobial misuse, and several AST methods. The selection and spread of azithromycin resistance are driven by unsuitable treatment of patients with suboptimal doses of azithromycin for gonococcal infection. Gonorrhea and AMR in NG are significant public health concerns globally, especially in Asian and European countries. In our metadata, the most reports were from Asia (50 reports) and Europe (46 studies) than on the other continents. Current evidence of AMR NG in Asian and European data supports rigorous monitoring of definite antibiotic policy, and active surveillance of infections. Furthermore, there is an alarm for the high prevalence (>10%) of azithromycin resistance in NG strains in China, Iran, Bangladesh, Myanmar, Vietnam, Norway, Ireland, Estonia, Hungary, and Slovenia. It is significant to appreciate the epidemiology of AMR in NG to inform treatment guidelines and prevention and control measures, in addition to wider gonorrhea prevention efforts.

This meta-analysis revealed a significant increase in the proportion of azithromycin resistance isolates over time, which is consistent with other reports [154,155]. Over the period, our findings showed that the trend of azithromycin resistance had increased from 1988-2013 to 2019-2021 (2.3% and 7.2%, respectively). Azithromycin resistance rate over times was increased as compared to erythromycin. The previous clinical and in vitro AMR data displayed early that erythromycin is not sufficiently effective for the treatment of gonorrhea [156,157]. So, due to the limited clinical effects of drugs (like; erythromycin) for the treatment of gonorrhea, currently most countries commonly suggest dual antimicrobial therapy (ceftriaxone 250-500mg plus azithromycin 1-2g) as empirical first-line gonorrhea treatment to prevent the emergence/accumulation of AMR [147]. The reason for the decrease in resistance to erythromycin in the past years is the lack of use of this drug in the treatment of gonorrhea.

The prevalence of azithromycin and erythromycin resistant NG isolates in South America (9.2%) and Asia (36.3%) is higher than in other continents. These results indicate that macrolides resistance patterns differed among geographic regions and in different periods.

Despite the significance of AST for clinical management of infection and AMR surveillance, the methodologies and breakpoints of the CLSI and EUCAST are used worldwide. Divergences in clinical breakpoints between CLSI and EUCAST considerably impact susceptibility interpretation of clinical isolates. This has implications not only for antibiograms at institutions switching between the two AST systems, but for broader AMR surveillance initiatives comparing data within and between countries using different systems or over the time period during which a shift in methodology is applied.

The creeping increase in the emergence of NG isolates with resistance to ceftriaxone plus azithromycin is a cause for a rising global challenge that necessitates further monitoring and investigation. The increase in emergence of reduced susceptibility or low- to high-level azithromycin resistance of NG strains has previously been reported to be related to mutational changes in the specific residues (the peptidyl-transferase loop) in domain V of the 23S rRNA, namely A2059G, C2611T, A2143G, A2058G, and multiple transferable resistance (Mtr) system as a tripartite efflux pump [158,159]. The main limitation of our review that is that we used collection date for data samples, rather than the publication date for analysis. The correct date of samples was not available or mentioned in all the included studies, so the authors performed the subgroup analysis by year published instead of the year of collection data of samples.

However, there were also several limitations of our meta-analysis to be addressed: (1) most of the included studies in this meta-analysis were conducted in European and Asian countries; (2); not including the strains identification code of NG in the included studies.

# CONCLUSIONS

In this metadata, the global resistance prevalence was 6% for azithromycin and 48% for erythromycin. These are major insights into the epidemiological macrolides resistance pattern of NG over time and has highlighted the distribution of macrolides-resistant strains in continents/countries. The creeping increase in the emergence of NG isolates with resistance to ceftriaxone and/or azithromycin is a cause for a rising global challenge that necessitates robust monitoring and more investigation. The implications of this study emphasize the rigorous or improved antimicrobial stewardship, early diagnosis, contact tracing, and enhanced intensive global surveillance system are crucial for control of further spreading of gonococcal AMR.

Acknowledgments: This study (Grant number: 14-113102) was supported and approved by Golestan University of Medical Sciences (Gorgan, Iran) for which we are very grateful.

**Competing Interests**: The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript.

**Data availability statement**: All the data in this review are included in the manuscript.

#### REFERENCES

 Kirkcaldy RD, Weston E, Segurado AC, Hughes G. Epidemiology of gonorrhoea: a global perspective. Sex Health. 2019 Sep;16(5):401-11.

- Unemo M, Golparian D, Eyre DW. Antimicrobial Resistance in *Neisseria gonorrhoeae* and Treatment of Gonorrhea. Methods Mol Biol. 2019;1997:37–58.
- Multi-drug resistant gonorrhea: World Health Organization; 10 November 2021. Available from: https://www.who.int/ news-room/fact-sheets/detail/multi-drug-resistant-gonorrhoea
- Derbie A, Mekonnen D, Woldeamanuel Y, Abebe T. Azithromycin resistant gonococci: a literature review. Antimicrob Resist Infect Control. 2020 Aug;9(1):138.
- Moher D, Liberati A, Tetzlaff J, Altman DG, med PGJP. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. 2009;6(7):e1000097.
- Performance Standards for Antimicrobial Susceptibility Testing, CLSI supplement M100. CLSI; Clinical and Laboratory Standards Institute; 2022.
- The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 12.0, 2022. http://www.eucast.org
- Ison CA, Town K, Obi C, Chisholm S, Hughes G, Livermore DM, et al. Decreased susceptibility to cephalosporins among gonococci: data from the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) in England and Wales, 2007–2011. Lancet Infect Dis. 2013;13(9):762-8.
- Unemo M, Lahra MM, Escher M, Eremin S, Cole MJ, Galarza P, et al. WHO global antimicrobial resistance surveillance for *Neisseria gonorrhoeae* 2017–18: a retrospective observational study. 2021;2(11):e627-e36.
- Newcastle O. Newcastle-Ottawa: Scale customized for cross-sectional studies. 2018.
- DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials. 1986 Sep;7(3):177–88.
- Adamson PC, Van Le H, Le HH, Le GM, Nguyen TV, Klausner JD. Trends in antimicrobial resistance in *Neisseria gonorrhoeae* in Hanoi, Vietnam, 2017-2019. BMC Infect Dis. 2020 Nov;20(1):809.
- Ahmed MU, Chawdhury FA, Hossain M, Sultan SZ, Alam M, Salahuddin G, et al. Monitoring antimicrobial susceptibility of *Neisseria gonorrhoeae* isolated from Bangladesh during 1997-2006: emergence and pattern of drug-resistant isolates. J Health Popul Nutr. 2010 Oct;28(5):443–9.
- Alfsnes K, Eldholm V, Olsen AO, Brynildsrud OB, Bohlin J, Steinbakk M, et al. Genomic epidemiology and population structure of *Neisseria gonorrhoeae* in Norway, 2016-2017. Microb Genom. 2020 Apr;6(4). https://doi. org/10.1099/mgen.0.000359.
- 15. Aniskevich A, Shimanskaya I, Boiko I, Golubovskaya T, Golparian D, Stanislavova I, et al. Antimicrobial resistance in *Neisseria gonorrhoeae* isolates and gonorrhea treatment in the Republic of Belarus, Eastern Europe, 2009–2019. BMC Infect Dis. 2021;21(1):1–9.
- Attram N, Agbodzi B, Dela H, Behene E, Nyarko EO, Kyei NN, et al. Antimicrobial resistance (AMR) and molecular characterization of *Neisseria gonorrhoeae* in Ghana, 2012-2015. PLoS One. 2019 Oct;14(10):e0223598.
- Azizmohammadi S, Azizmohammadi S. Antimicrobial susceptibility pattern of *Neisseria gonorrhoeae* isolated from fertile and infertile women. Trop J Pharm Res.

2016;15(12):2653-7.

- Bailey AL, Potter RF, Wallace MA, Johnson C, Dantas G, Burnham CA. Genotypic and Phenotypic Characterization of Antimicrobial Resistance in *Neisseria gonorrhoeae*: a Cross-Sectional Study of Isolates Recovered from Routine Urine Cultures in a High-Incidence Setting. MSphere. 2019 Jul;4(4):e00373–19.
- Bala M, Ray K, Gupta SM. Antimicrobial resistance pattern of *Neisseria gonorrhoeae* isolates from peripheral health centres and STD clinic attendees of a tertiary care centre in India. Int J STD AIDS. 2008 Jun;19(6):378–80.
- Barbee LA, Soge OO, Katz DA, Dombrowski JC, Holmes KK, Golden MR. Increases in *Neisseria gonorrhoeae* with reduced susceptibility to azithromycin among men who have sex with men in Seattle, King County, Washington, 2012–2016. Clin Infect Dis. 2018 Feb;66(5):712–8.
- Barros Dos Santos KT, Skaf LB, Justo-da-Silva LH, Medeiros RC, Francisco Junior RD, Caniné MC, et al. Evidence for Clonally Associated Increasing Rates of Azithromycin Resistant *Neisseria gonorrhoeae* in Rio de Janeiro, Brazil. BioMed Res Int. 2019 Jan;2019:3180580.
- 22. Bazzo ML, Golfetto L, Gaspar PC, Pires AF, Ramos MC, Franchini M, et al.; Brazilian-GASP Network. First nationwide antimicrobial susceptibility surveillance for *Neisseria gonorrhoeae* in Brazil, 2015-16. J Antimicrob Chemother. 2018 Jul;73(7):1854–61.
- 23. Bharara T, Bhalla P, Rawat D, Garg VK, Sardana K, Chakravarti A. Rising trend of antimicrobial resistance among *Neisseria gonorrhoeae* isolates and the emergence of N. gonorrhoeae isolate with decreased susceptibility to ceftriaxone. Indian J Med Microbiol. 2015 Jan-Mar;33(1):39–42.
- 24. Boiko I, Golparian D, Jacobsson S, Krynytska I, Frankenberg A, Shevchenko T, et al. Genomic epidemiology and antimicrobial resistance determinants of *Neisseria gonorrhoeae* isolates from Ukraine, 2013-2018. APMIS. 2020 Jul;128(7):465–75.
- 25. Boiko I, Golparian D, Krynytska I, Bezkorovaina H, Frankenberg A, Onuchyna M, et al. Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates and treatment of gonorrhoea patients in Ternopil and Dnipropetrovsk regions of Ukraine, 2013-2018. APMIS. 2019 Jul;127(7):503–9.
- 26. Brunner A, Nemes-Nikodem E, Jeney C, Szabo D, Marschalko M, Karpati S, et al. Emerging azithromycin-resistance among the *Neisseria gonorrhoeae* strains isolated in Hungary. Ann Clin Microbiol Antimicrob. 2016 Sep;15(1):53.
- 27. Brunner A, Nemes-Nikodem E, Mihalik N, Marschalko M, Karpati S, Ostorhazi E. Incidence and antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from patients attending the national *Neisseria gonorrhoeae* reference laboratory of Hungary. BMC Infect Dis. 2014 Aug;14(1):433.
- 28. Buder S, Dudareva S, Jansen K, Loenenbach A, Nikisins S, Sailer A, et al.; GORENET study group. Antimicrobial resistance of *Neisseria gonorrhoeae* in Germany: low levels of cephalosporin resistance, but high azithromycin resistance. BMC Infect Dis. 2018 Jan;18(1):44.
- Calado J, Castro R, Lopes A, Campos MJ, Rocha M, Pereira F. Antimicrobial resistance and molecular characteristics

of *Neisseria gonorrhoeae* isolates from men who have sex with men. Int J Infect Dis. 2019 Feb;79:116–22.

- Carannante A, Ciammaruconi A, Vacca P, Anselmo A, Fillo S, Palozzi AM, et al. Genomic characterization of gonococci from different anatomic sites, Italy, 2007–2014. Microb Drug Resist. 2019 Nov;25(9):1316–24.
- Cobo F, Cabezas-Fernández MT, Avivar C. Typing and antimicrobial susceptibility of 134 *Neisseria gonorrhoeae* strains from Southern Spain. Rev Esp Quimioter. 2019 Apr;32(2):114–20.
- 32. Cobo F, Cabezas-Fernández MT, Cabeza-Barrera MI. Antimicrobial susceptibility and typing of *Neisseria gonorrhoeae* strains from Southern Spain, 2012-2014. Enferm Infecc Microbiol Clin. 2016 Jan;34(1):3–7.
- 33. Cole MJ, Spiteri G, Jacobsson S, Pitt R, Grigorjev V, Unemo M; Euro-GASP Network. Is the tide turning again for cephalosporin resistance in *Neisseria gonorrhoeae* in Europe? Results from the 2013 European surveillance. BMC Infect Dis. 2015 Aug;15(1):321.
- 34. Costa LM, Pedroso ER, Vieira Neto V, Souza VC, Teixeira MJ. Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from patients attending a public referral center for sexually transmitted diseases in Belo Horizonte, State of Minas Gerais, Brazil. Rev Soc Bras Med Trop. 2013 May-Jun;46(3):304–9.
- 35. Crucitti T, Belinga S, Fonkoua MC, Abanda M, Mbanzouen W, Sokeng E, et al. Sharp increase in ciprofloxacin resistance of *Neisseria gonorrhoeae* in Yaounde, Cameroon: analyses of a laboratory database period 2012-2018. Int J STD AIDS. 2020 May;31(6):579–86.
- 36. Costa-Lourenço AP, Abrams AJ, Dos Santos KT, Argentino IC, Coelho-Souza T, Caniné MC, et al. Phylogeny and antimicrobial resistance in *Neisseria gonorrhoeae* isolates from Rio de Janeiro, Brazil. Infect Genet Evol. 2018 Mar;58:157–63.
- 37. Dan M, Mor Z, Gottliev S, Sheinberg B, Shohat T. Trends in antimicrobial susceptibility of *Neisseria gonorrhoeae* in Israel, 2002 to 2007, with special reference to fluoroquinolone resistance. Sex Transm Dis. 2010 Jul;37(7):451–3.
- 38. de Korne-Elenbaas J, Bruisten SM, de Vries HJ, Van Dam AP. Emergence of a *Neisseria gonorrhoeae* clone with reduced cephalosporin susceptibility between 2014 and 2019 in Amsterdam, The Netherlands, revealed by genomic population analysis. J Antimicrob Chemother. 2021 Jun;76(7):1759–68.
- 39. Fuertes de Vega I, Baliu-Piqué C, Bosch Mestres J, Vergara Gómez A, Vallés X, Alsina Gibert M. Risk factors for antimicrobial-resistant *Neisseria gonorrhoeae* and characteristics of patients infected with gonorrhea. Enferm Infecc Microbiol Clin (Engl Ed). 2018 Mar;36(3):165–8.
- 40. Dillon JA, Rubabaza JP, Benzaken AS, Sardinha JC, Li H, Bandeira MG, et al. Reduced susceptibility to azithromycin and high percentages of penicillin and tetracycline resistance in *Neisseria gonorrhoeae* isolates from Manaus, Brazil, 1998. Sex Transm Dis. 2001 Sep;28(9):521–6.
- 41. Donegan EA, Wirawan DN, Muliawan P, Schachter J, Moncada J, Parekh M, et al. Fluoroquinolone-resistant *Neisseria gonorrhoeae* in Bali, Indonesia: 2004. Sex Transm Dis. 2006 Oct;33(10):625–9.
- 42. Dong Y, Yang Y, Wang Y, Martin I, Demczuk W, Gu W.

Shanghai *Neisseria gonorrhoeae* Isolates Exhibit Resistance to Extended-Spectrum Cephalosporins and Clonal Distribution. Front Microbiol. 2020 Oct;11:580399.

- 43. Ehinmidu J, Bolaji R, Adegboye E. Isolation and antibiotic susceptibility profile of *Neisseria gonorrhoeae* isolated from urine samples in Zara, Northern Nigeria. Journal of Phytomedicine and Therapeutics. 2004;9.
- 44. Enders M, Turnwald-Maschler A, Regnath T. Antimicrobial resistance of *Neisseria gonorrhoeae* isolates from the Stuttgart and Heidelberg areas of southern Germany. Eur J Clin Microbiol Infect Dis. 2006 May;25(5):318–22.
- 45. Fentaw S, Abubeker R, Asamene N, Assefa M, Bekele Y, Tigabu E. Antimicrobial susceptibility profile of Gonococcal isolates obtained from men presenting with urethral discharge in Addis Ababa, Ethiopia: implications for national syndromic treatment guideline. PLoS One. 2020 Jun;15(6):e0233753.
- 46. Ibargoyen García U, Nieto Toboso MC, Azpeitia EM, Imaz Perez M, Hernandez Ragpa L, Álava Menica JA, et al. Epidemiological surveillance study of gonococcal infection in Northern Spain. Enferm Infecc Microbiol Clin (Engl Ed). 2020 Feb;38(2):59–64.
- 47. Gianecini R, Romero ML, Oviedo C, Vacchino M, Galarza P, Gonococcal Antimicrobial Susceptibility Surveillance Programme-Argentina Working G; Gonococcal Antimicrobial Susceptibility Surveillance Programme-Argentina (GASSP-AR) Working Group. Emergence and Spread of *Neisseria gonorrhoeae* Isolates With Decreased Susceptibility to Extended-Spectrum Cephalosporins in Argentina, 2009 to 2013. Sex Transm Dis. 2017 Jun;44(6):351–5.
- Glazkova S, Golparian D, Titov L, Pankratova N, Suhabokava N, Shimanskaya I, et al. Antimicrobial susceptibility/resistance and molecular epidemiological characteristics of *Neisseria gonorrhoeae* in 2009 in Belarus. APMIS. 2011 Aug;119(8):537–42.
- 49. Golparian D, Bazzo ML, Golfetto L, Gaspar PC, Schörner MA, Schwartz Benzaken A, et al.; Brazilian-GASP Network. Genomic epidemiology of *Neisseria gonorrhoeae* elucidating the gonococcal antimicrobial resistance and lineages/sublineages across Brazil, 2015-16. J Antimicrob Chemother. 2020 Nov;75(11):3163–72.
- 50. Golparian D, Brilene T, Laaring Y, Viktorova E, Johansson E, Domeika M, et al. First antimicrobial resistance data and genetic characteristics of *Neisseria gonorrhoeae* isolates from Estonia, 2009-2013. New Microbes New Infect. 2014 Sep;2(5):150–3.
- 51. Golparian D, Harris SR, Sánchez-Busó L, Hoffmann S, Shafer WM, Bentley SD, et al. Genomic evolution of *Neisseria gonorrhoeae* since the preantibiotic era (1928-2013): antimicrobial use/misuse selects for resistance and drives evolution. BMC Genomics. 2020 Feb;21(1):116.
- 52. Guerrero-Torres MD, Menéndez MB, Guerras CS, Tello E, Ballesteros J, Clavo P, et al. Epidemiology, molecular characterisation and antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates in Madrid, Spain, in 2016. Epidemiol Infect. 2019 Sep;147:e274.
- 53. Hamasuna R, Yasuda M, Ishikawa K, Uehara S, Hayami H, Takahashi S, et al. The second nationwide surveillance of the antimicrobial susceptibility of *Neisseria gonorrhoeae* from male urethritis in Japan, 2012-2013. J Infect Chemo-

ther. 2015 May;21(5):340-5.

- 54. Hamasuna R, Yasuda M, Ishikawa K, Uehara S, Takahashi S, Hayami H, et al. Nationwide surveillance of the antimicrobial susceptibility of *Neisseria gonorrhoeae* from male urethritis in Japan. J Infect Chemother. 2013 Aug;19(4):571–8.
- 55. Harris SR, Cole MJ, Spiteri G, Sánchez-Busó L, Golparian D, Jacobsson S, et al.; Euro-GASP study group. Public health surveillance of multidrug-resistant clones of *Neisseria gonorrhoeae* in Europe: a genomic survey. Lancet Infect Dis. 2018 Jul;18(7):758–68.
- 56. Hjelmevoll SO, Golparian D, Dedi L, Skutlaberg DH, Haarr E, Christensen A, et al. Phenotypic and genotypic properties of *Neisseria gonorrhoeae* isolates in Norway in 2009: antimicrobial resistance warrants an immediate change in national management guidelines. Eur J Clin Microbiol Infect Dis. 2012 Jun;31(6):1181–6.
- 57. Hofstraat SH, Götz HM, van Dam AP, van der Sande MA, van Benthem BH. Trends and determinants of antimicrobial susceptibility of *Neisseria gonorrhoeae* in the Netherlands, 2007 to 2015. Euro Surveill. 2018 Sep;23(36):1700565.
- Holderman JL, Thomas JC 4th, Schlanger K, Black JM, Town K, St Cyr SB, et al. Sustained transmission of *Neis-seria gonorrhoeae* with high-level resistance to azithromycin, in Indianapolis, Indiana, 2017–2018. Clin Infect Dis. 2021 Sep;73(5):808–15.
- 59. Horn NN, Kresken M, Körber-Irrgang B, Göttig S, Wichelhaus C, Wichelhaus TA; Working Party Antimicrobial Resistance of the Paul Ehrlich Society for Chemotherapy. Antimicrobial susceptibility and molecular epidemiology of *Neisseria gonorrhoeae* in Germany. Int J Med Microbiol. 2014 Jul;304(5-6):586–91.
- 60. Hovhannisyan G, von Schoen-Angerer T, Babayan K, Fenichiu O, Gaboulaud V. Antimicrobial susceptibility of Neisseria gonorrheae strains in three regions of Armenia. Sex Transm Dis. 2007 Sep;34(9):686–8.
- 61. Ieven M, Van Looveren M, Sudigdoadi S, Rosana Y, Goossens W, Lammens C, et al. Antimicrobial susceptibilities of *Neisseria gonorrhoeae* strains isolated in Java, Indonesia. Sex Transm Dis. 2003 Jan;30(1):25–9.
- 62. Jacobsson S, Cole MJ, Spiteri G, Day M, Unemo M; Euro-GASP Network. Associations between antimicrobial susceptibility/resistance of *Neisseria gonorrhoeae* isolates in European Union/European Economic Area and patients' gender, sexual orientation and anatomical site of infection, 2009-2016. BMC Infect Dis. 2021 Mar;21(1):273.
- 63. Jean SS, Lu MC, Shi ZY, Tseng SH, Wu TS, Lu PL, et al. In vitro activity of ceftazidime-avibactam, ceftolozane-tazobactam, and other comparable agents against clinically important Gram-negative bacilli: results from the 2017 Surveillance of Multicenter Antimicrobial Resistance in Taiwan (SMART). Infect Drug Resist. 2018 Oct;11:1983– 92.
- 64. Jeverica S, Golparian D, Matičič M, Potočnik M, Mlakar B, Unemo M. Phenotypic and molecular characterization of *Neisseria gonorrhoeae* isolates from Slovenia, 2006-12: rise and fall of the multidrug-resistant NG-MAST genogroup 1407 clone? J Antimicrob Chemother. 2014 Jun;69(6):1517–25.

- 65. Jiang FX, Lan Q, Le WJ, Su XH. Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from Hefei (2014-2015): genetic characteristics of antimicrobial resistance. BMC Infect Dis. 2017 May;17(1):366.
- 66. Joesoef MR, Knapp JS, Idajadi A, Linnan M, Barakbah Y, Kamboji A, et al. Antimicrobial susceptibilities of *Neisseria gonorrhoeae* strains isolated in Surabaya, Indonesia. Antimicrob Agents Chemother. 1994 Nov;38(11):2530–3.
- Kam KM. WONG PW, Cheung MM, HO NKY, Lo KK. Quinolone-Resistant *Neisseria gonorrhoeae* in Hong Kong. Sex Transm Dis. 1996;23(2):103–8.
- 68. Karymbaeva S, Boiko I, Jacobsson S, Mamaeva G, Ibraeva A, Usupova D, et al. Antimicrobial resistance and molecular epidemiological typing of *Neisseria gonorrhoeae* isolates from Kyrgyzstan in Central Asia, 2012 and 2017. BMC Infect Dis. 2021 Jun;21(1):559.
- Khaki P, Bhalla P, Sharma P, Chawla R, Bhalla K. Epidemilogical analysis of *Neisseria gonorrhoeae* isolates by antimicrobial susceptibility testing, auxotyping and serotyping. Indian J Med Microbiol. 2007 Jul;25(3):225–9.
- 70. Khanam R, Ahmed D, Rahman M, Alam MS, Amin M, Khan SI, et al. Antimicrobial Susceptibility of *Neisseria* gonorrhoeae in Bangladesh (2014 Update). Antimicrob Agents Chemother. 2016 Jun;60(7):4418–9.
- 71. Kirkcaldy RD, Soge O, Papp JR, Hook EW 3rd, del Rio C, Kubin G, et al. Analysis of *Neisseria gonorrhoeae* azithromycin susceptibility in the United States by the Gonococcal Isolate Surveillance Project, 2005 to 2013. Antimicrob Agents Chemother. 2015 Feb;59(2):998–1003.
- 72. Kirkcaldy RD, Zaidi A, Hook EW 3rd, Holmes KK, Soge O, del Rio C, et al. *Neisseria gonorrhoeae* antimicrobial resistance among men who have sex with men and men who have sex exclusively with women: the Gonococcal Isolate Surveillance Project, 2005-2010. Ann Intern Med. 2013 Mar;158(5 Pt 1 5 Part 1):321–8.
- 73. Kivata MW, Mbuchi M, Eyase FL, Bulimo WD, Kyanya CK, Oundo V, et al. gyrA and parC mutations in fluoroquinolone-resistant *Neisseria gonorrhoeae* isolates from Kenya. BMC Microbiol. 2019 Apr;19(1):76.
- Kubanov A, Vorobyev D, Chestkov A, Leinsoo A, Shaskolskiy B, Dementieva E, et al. Molecular epidemiology of drug-resistant *Neisseria gonorrhoeae* in Russia (Current Status, 2015). BMC Infect Dis. 2016 Aug;16(1):389.
- 75. Kubanova A, Frigo N, Kubanov A, Sidorenko S, Lesnaya I, Polevshikova S, et al. The Russian gonococcal antimicrobial susceptibility programme (RU-GASP)—national resistance prevalence in 2007 and 2008, and trends during 2005-2008. Euro Surveill. 2010 Apr;15(14):19533.
- Kularatne R, Maseko V, Gumede L, Kufa T. Trends in Neisseria gonorrhoeae antimicrobial resistance over a ten-year surveillance period, Johannesburg, South Africa, 2008–2017. Antibiotics (Basel). 2018 Jul;7(3):58.
- 77. Kulkarni SV, Bala M, Muqeeth SA, Sasikala G, Nirmalkar AP, Thorat R, et al. Antibiotic susceptibility pattern of *Neisseria gonorrhoeae* strains isolated from five cities in India during 2013-2016. J Med Microbiol. 2018 Jan;67(1):22–8.
- 78. Lagace-Wiens PR, Duncan S, Kimani J, Thiong'o A, Shafi J, McClelland S, et al. Emergence of fluoroquinolone resistance in *Neisseria gonorrhoeae* isolates from four

clinics in three regions of Kenya. Sex Transm Dis. 2012 May;39(5):332–4.

- 79. Lan PT, Golparian D, Ringlander J, Van Hung L, Van Thuong N, Unemo M. Genomic analysis and antimicrobial resistance of *Neisseria gonorrhoeae* isolates from Vietnam in 2011 and 2015-16. J Antimicrob Chemother. 2020 Jun;75(6):1432–8.
- Le W, Su X, Lou X, Li X, Gong X, Wang B, et al. Susceptibility Trends of Zoliflodacin against Multidrug-Resistant *Neisseria gonorrhoeae* Clinical Isolates in Nanjing, China, 2014 to 2018. Antimicrob Agents Chemother. 2021 Feb;65(3):e00863–20.
- Lebedzeu F, Golparian D, Titov L, Pankratava N, Glazkova S, Shimanskaya I, et al. Antimicrobial susceptibility/ resistance and NG-MAST characterisation of *Neisseria* gonorrhoeae in Belarus, Eastern Europe, 2010-2013. BMC Infect Dis. 2015 Jan;15(1):29.
- 82. Lee H, Unemo M, Kim HJ, Seo Y, Lee K, Chong Y. Emergence of decreased susceptibility and resistance to extended-spectrum cephalosporins in *Neisseria gonorrhoeae* in Korea. J Antimicrob Chemother. 2015 Sep;70(9):2536–42.
- Lee RS, Seemann T, Heffernan H, Kwong JC, Gonçalves da Silva A, Carter GP, et al. Genomic epidemiology and antimicrobial resistance of *Neisseria gonorrhoeae* in New Zealand. J Antimicrob Chemother. 2018 Feb;73(2):353–64.
- 84. Li W, Zhu By, Qin Sq, Yang Mc, Liang M, He S, et al. Surveillance of antimicrobial susceptibilities of *Neisseria gonorrhoeae* from 2013 to 2015 in Guangxi Province, China. Japanese Journal of Infectious Diseases. JJID. 2017;2017:169.
- Liu JW, Xu WQ, Zhu XY, Dai XQ, Chen SC, Han Y, et al. Gentamicin susceptibility of *Neisseria gonorrhoeae* isolates from 7 provinces in China. Infect Drug Resist. 2019 Aug;12:2471–6.
- 86. Liu YH, Huang YT, Liao CH, Hsueh PR. Antimicrobial susceptibilities and molecular typing of *neisseria gonorrhoeae* isolates at a medical centre in Taiwan, 2001-2013 with an emphasis on high rate of azithromycin resistance among the isolates. Int J Antimicrob Agents. 2018 May;51(5):768–74.
- Liu YH, Wang YH, Liao CH, Hsueh PR. Emergence and Spread of *Neisseria gonorrhoeae* Strains with High-Level Resistance to Azithromycin in Taiwan from 2001 to 2018. Antimicrob Agents Chemother. 2019 Aug;63(9):e00773–19.
- Lo JY, Ho KM, Lo AC. Surveillance of gonococcal antimicrobial susceptibility resulting in early detection of emerging resistance. J Antimicrob Chemother. 2012 Jun;67(6):1422–6.
- Loo VG, Simor AE, Jaeger R, Low DE. Survey of *Neisseria gonorrhoeae* antimicrobial susceptibility in Ontario. Can J Infect Dis. 1990;1(4):136–8.
- 90. Mabonga E, Parkes-Ratanshi R, Riedel S, Nabweyambo S, Mbabazi O, Taylor C, et al. Complete ciprofloxacin resistance in gonococcal isolates in an urban Ugandan clinic: findings from a cross-sectional study. Int J STD AIDS. 2019 Mar;30(3):256–63.
- 91. Maduna LD, Kock MM, van der Veer BM, Radebe O, McIntyre J, van Alphen LB, et al. Antimicrobial Resistance of *Neisseria gonorrhoeae* Isolates from High-Risk Men in

Johannesburg, South Africa. Antimicrob Agents Chemother. 2020 Oct;64(11):e00906–20.

- 92. Martin I, Sawatzky P, Allen V, Hoang L, Lefebvre B, Mina N, et al. Emergence and characterization of *Neisseria gonorrhoeae* isolates with decreased susceptibilities to ceftriaxone and cefixime in Canada: 2001-2010. Sex Transm Dis. 2012 Apr;39(4):316–23.
- 93. Martin I, Sawatzky P, Allen V, Lefebvre B, Hoang L, Naidu P, et al. Multidrug-resistant and extensively drug-resistant *Neisseria gonorrhoeae* in Canada, 2012-2016. Can Commun Dis Rep. 2019 Feb;45(2-3):45–53.
- 94. Martin I, Sawatzky P, Liu G, Allen V, Lefebvre B, Hoang L, et al. Antimicrobial susceptibilities and distribution of sequence types of *Neisseria gonorrhoeae* isolates in Canada: 2010. Can J Microbiol. 2013 Oct;59(10):671–8.
- 95. Martins RA, Cassu-Corsi D, Nodari CS, Cayô R, Natsumeda L, Streling AP, et al. Temporal evolution of antimicrobial resistance among *Neisseria gonorrhoeae* clinical isolates in the most populated South American Metropolitan Region. Mem Inst Oswaldo Cruz. 2019;114:e190079.
- 96. Mehta SD, Maclean I, Ndinya-Achola JO, Moses S, Martin I, Ronald A, et al. Emergence of quinolone resistance and cephalosporin MIC creep in *Neisseria gonorrhoeae* isolates from a cohort of young men in Kisumu, Ken-ya, 2002 to 2009. Antimicrob Agents Chemother. 2011 Aug;55(8):3882–8.
- 97. Melendez JH, Hardick J, Barnes M, Page KR, Gaydos CA. Antimicrobial Susceptibility of *Neisseria gonorrhoeae* Isolates in Baltimore, Maryland, 2016: The Importance of Sentinel Surveillance in the Era of Multi-Drug-Resistant Gonorrhea. Antibiotics (Basel). 2018 Aug;7(3):77.
- 98. Mlynarczyk-Bonikowska B, Serwin AB, Golparian D, Walter de Walthoffen S, Majewski S, Koper M, et al. Antimicrobial susceptibility/resistance and genetic characteristics of *Neisseria gonorrhoeae* isolates from Poland, 2010-2012. BMC Infect Dis. 2014 Feb;14(1):65.
- 99. Morris SR, Moore DF, Hannah PB, Wang SA, Wolfe J, Trees DL, et al. Strain typing and antimicrobial resistance of fluoroquinolone-resistant *Neisseria gonorrhoeae* causing a California infection outbreak. J Clin Microbiol. 2009 Sep;47(9):2944–9.
- 100. Nacht C, Agingu W, Otieno F, Odhiambo F, Mehta SD. Antimicrobial resistance patterns in *Neisseria gonor-rhoeae* among male clients of a sexually transmitted infections clinic in Kisumu, Kenya. Int J STD AIDS. 2020 Jan;31(1):46–52.
- 101. Naznin M, Salam MA, Hossain MZ, Alam MS. Current status of gonococcal antimicrobial susceptibility with special reference to Azithromycin and Ceftriaxone: report from a tertiary care hospital in Bangladesh. Pak J Med Sci. 2018 Nov-Dec;34(6):1397–401.
- 102. Nemes-Nikodém É, Brunner A, Pintér D, Mihalik N, Lengyel G, Marschalkó M, et al. Antimicrobial susceptibility and genotyping analysis of Hungarian *Neisseria* gonorrhoeae strains in 2013. Acta Microbiol Immunol Hung. 2014 Dec;61(4):435–45.
- 103. Olsen B, Pham TL, Golparian D, Johansson E, Tran HK, Unemo M. Antimicrobial susceptibility and genetic characteristics of *Neisseria gonorrhoeae* isolates from Vietnam, 2011. BMC Infect Dis. 2013 Jan;13(1):40.

- 104. Olsen B, Månsson F, Camara C, Monteiro M, Biai A, Alves A, et al. Phenotypic and genetic characterisation of bacterial sexually transmitted infections in Bissau, Guinea-Bissau, West Africa: a prospective cohort study. BMJ Open. 2012 Mar;2(2):e000636.
- 105. Pinto M, Rodrigues JC, Matias R, Água-Doce I, Cordeiro D, Correia C, et al.; PTGonoNet. Fifteen years of a nationwide culture collection of *Neisseria gonorrhoeae* antimicrobial resistance in Portugal. Eur J Clin Microbiol Infect Dis. 2020 Sep;39(9):1761–70.
- 106. Qin X, Zhao Y, Chen W, Wu X, Tang S, Li G, et al. Changing antimicrobial susceptibility and molecular characterisation of *Neisseria gonorrhoeae* isolates in Guangdong, China: in a background of rapidly rising epidemic. Int J Antimicrob Agents. 2019 Dec;54(6):757–65.
- 107. Queirós C, Borges da Costa J, Lito L, Filipe P, Melo Cristino J. Estudio retrospectivo acerca de la evolución y el desarrollo de resistencias antimicrobianas en casos diagnosticados de gonorrea en un hospital de nivel terciario en Portugal durante 10 años. Actas Dermosifiliogr (Engl Ed). 2020 Nov;111(9):761–7.
- 108. Rambaran S, Naidoo K, Dookie N, Moodley P, Sturm AW. Resistance Profile of *Neisseria gonorrhoeae* in KwaZulu-Natal, South Africa Questioning the Effect of the Currently Advocated Dual Therapy. Sex Transm Dis. 2019 Apr;46(4):266–70.
- 109. Regnath T, Mertes T, Ignatius R. Antimicrobial resistance of *Neisseria gonorrhoeae* isolates in south-west Germany, 2004 to 2015: increasing minimal inhibitory concentrations of tetracycline but no resistance to third-generation cephalosporins. Euro Surveill. 2016 Sep;21(36):30335.
- 110. Rice RJ, Knapp JS. Susceptibility of *Neisseria gonor-rhoeae* associated with pelvic inflammatory disease to ce-foxitin, ceftriaxone, clindamycin, gentamicin, doxycycline, azithromycin, and other antimicrobial agents. Antimicrob Agents Chemother. 1994 Jul;38(7):1688–91.
- 111. Ryan L, Golparian D, Fennelly N, Rose L, Walsh P, Lawlor B, et al. Antimicrobial resistance and molecular epidemiology using whole-genome sequencing of *Neisseria gonorrhoeae* in Ireland, 2014-2016: focus on extended-spectrum cephalosporins and azithromycin. Eur J Clin Microbiol Infect Dis. 2018 Sep;37(9):1661–72.
- 112. Salmerón P, Viñado B, Arando M, Alcoceba E, Romero B, Menéndez B, et al. *Neisseria gonorrhoeae* antimicrobial resistance in Spain: a prospective multicentre study. J Antimicrob Chemother. 2021 May;76(6):1523–31.
- 113. Salmerón P, Viñado B, El Ouazzani R, Hernández M, Barbera MJ, Alberny M, et al. Antimicrobial susceptibility of *Neisseria gonorrhoeae* in Barcelona during a five-year period, 2013 to 2017. Euro Surveill. 2020 Oct;25(42):1900576.
- 114. Serra-Pladevall J, Barberá MJ, Rodriguez S, Bartolomé-Comas R, Roig G, Juvé R, et al. *Neisseria* gonorrhoeae antimicrobial susceptibility in Barcelona: penA, ponA, mtrR, and porB mutations and NG-MAST sequence types associated with decreased susceptibility to cephalosporins. Eur J Clin Microbiol Infect Dis. 2016 Sep;35(9):1549–56.
- 115. Sethi S, Golparian D, Bala M, Dorji D, Ibrahim M, Jabeen K, et al. Antimicrobial susceptibility and genetic

characteristics of *Neisseria gonorrhoeae* isolates from India, Pakistan and Bhutan in 2007-2011. BMC Infect Dis. 2013 Jan;13(1):35.

- 116. Shimuta K, Unemo M, Nakayama S, Morita-Ishihara T, Dorin M, Kawahata T, et al.; Antibiotic-Resistant Gonorrhea Study Group. Antimicrobial resistance and molecular typing of *Neisseria gonorrhoeae* isolates in Kyoto and Osaka, Japan, 2010 to 2012: intensified surveillance after identification of the first strain (H041) with high-level ceftriaxone resistance. Antimicrob Agents Chemother. 2013 Nov;57(11):5225–32.
- 117. Sood S, Mahajan N, Verma R, Kar HK, Sharma VK. Emergence of decreased susceptibility to extended-spectrum cephalosporins in *Neisseria gonorrhoeae* in India. Natl Med J India. 2013 Jan-Feb;26(1):26–8.
- 118. Sosa J, Ramirez-Arcos S, Ruben M, Li H, Llanes R, Llop A, et al. High percentages of resistance to tetracycline and penicillin and reduced susceptibility to azithromycin characterize the majority of strain types of *Neisseria gonorrhoeae* isolates in Cuba, 1995-1998. Sex Transm Dis. 2003 May;30(5):443–8.
- 119. Starnino S, Suligoi B, Regine V, Bilek N, Stefanelli P, Dal Conte I, et al.; *Neisseria gonorrhoeae* Italian Study Group. Phenotypic and genotypic characterization of *Neisseria* gonorrhoeae in parts of Italy: detection of a multiresistant cluster circulating in a heterosexual network. Clin Microbiol Infect. 2008 Oct;14(10):949–54.
- 120. Takayama Y, Nakayama S, Shimuta K, Morita-Ishihara T, Ohnishi M. Characterization of azithromycin-resistant *Neisseria gonorrhoeae* isolated in Tokyo in 2005-2011. J Infect Chemother. 2014 May;20(5):339–41.
- 121. Takuva S, Mugurungi O, Mutsvangwa J, Machiha A, Mupambo AC, Maseko V, et al. Etiology and antimicrobial susceptibility of pathogens responsible for urethral discharge among men in Harare, Zimbabwe. Sex Transm Dis. 2014 Dec;41(12):713–7.
- 122. Tanaka M, Furuya R, Irie S, Kanayama A, Kobayashi I. High Prevalence of Azithromycin-Resistant *Neisseria gonorrhoeae* Isolates With a Multidrug Resistance Phenotype in Fukuoka, Japan. Sex Transm Dis. 2015 Jun;42(6):337– 41.
- 123. Tanaka M, Furuya R, Kobayashi I, Ohno A, Kanesaka I. Molecular characteristics and antimicrobial susceptibility of penicillinase-producing *Neisseria gonorrhoeae* isolates in Fukuoka, Japan, 1996-2018. J Glob Antimicrob Resist. 2021 Sep;26:45–51.
- 124. Tanaka M, Koga Y, Nakayama H, Kanayama A, Kobayashi I, Saika T, et al. Antibiotic-resistant phenotypes and genotypes of *Neisseria gonorrhoeae* isolates in Japan: identification of strain clusters with multidrug-resistant phenotypes. Sex Transm Dis. 2011 Sep;38(9):871–5.
- 125. Tayimetha CY, Unemo M. Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates in Yaoundé, Cameroon from 2009 to 2014. Sex Transm Dis. 2018 Dec;45(12):e101–3.
- 126. Thakur SD, Levett PN, Horsman GB, Dillon JR. High levels of susceptibility to new and older antibiotics in *Neisseria gonorrhoeae* isolates from Saskatchewan (2003-15): time to consider point-of-care or molecular testing for precision treatment? J Antimicrob Chemother. 2018

Jan;73(1):118-25.

- 127. Town K, Harris S, Sánchez-Busó L, Cole MJ, Pitt R, Fifer H, et al. Genomic and Phenotypic Variability in *Neisseria gonorrhoeae* Antimicrobial Susceptibility, England. Emerg Infect Dis. 2020 Mar;26(3):505–15.
- 128. Tribuddharat C, Pongpech P, Charoenwatanachokchai A, Lokpichart S, Srifuengfung S, Sonprasert S. Gonococcal Antimicrobial Susceptibility and the Prevalence of blaTEM-1 and blaTEM-135 Genes in *Neisseria gonorrhoeae* Isolates from Thailand. Jpn J Infect Dis. 2017 Mar;70(2):213–5.
- 129. Tshokey T, Tshering T, Pradhan AR, Adhikari D, Sharma R, Gurung K, et al. Antibiotic resistance in Neisseria gonorrhoea and treatment outcomes of gonococcal urethritis suspected patients in two large hospitals in Bhutan, 2015. PLoS One. 2018 Aug;13(8):e0201721.
- 130. Unemo M, Ahlstrand J, Sánchez-Busó L, Day M, Aanensen D, Golparian D, et al.; European Collaborative Group. High susceptibility to zoliflodacin and conserved target (GyrB) for zoliflodacin among 1209 consecutive clinical *Neisseria gonorrhoeae* isolates from 25 European countries, 2018. J Antimicrob Chemother. 2021 Apr;76(5):1221–8.
- 131. Vandepitte J, Hughes P, Matovu G, Bukenya J, Grosskurth H, Lewis DA. High prevalence of ciprofloxacin-resistant gonorrhea among female sex workers in Kampala, Uganda (2008-2009). Sex Transm Dis. 2014 Apr;41(4):233–7.
- 132. Visser M, van Westreenen M, van Bergen J, van Benthem BH. Low gonorrhoea antimicrobial resistance and culture positivity rates in general practice: a pilot study. Sex Transm Infect. 2020 May;96(3):220–2.
- 133. Vorobieva V, Firsova N, Ababkova T, Leniv I, Haldorsen BC, Unemo M, et al. Antibiotic susceptibility of *Neisseria gonorrhoeae* in Arkhangelsk, Russia. Sex Transm Infect. 2007 Apr;83(2):133–5.
- 134. Wang F, Liu JW, Li YZ, Zhang LJ, Huang J, Chen XS, et al. Surveillance and molecular epidemiology of *Neisseria gonorrhoeae* isolates in Shenzhen, China, 2010-2017. J Glob Antimicrob Resist. 2020 Dec;23:269–74.
- 135. West B, Changalucha J, Grosskurth H, Mayaud P, Gabone RM, Ka-Gina G, et al. Antimicrobial susceptibility, auxotype and plasmid content of *Neisseria gonorrhoeae* in northern Tanzania: emergence of high level plasmid mediated tetracycline resistance. Genitourin Med. 1995 Feb;71(1):9–12.
- 136. Wind CM, Schim van der Loeff MF, van Dam AP, de Vries HJ, van der Helm JJ. Trends in antimicrobial susceptibility for azithromycin and ceftriaxone in *Neisseria gonorrhoeae* isolates in Amsterdam, the Netherlands, between 2012 and 2015. Euro Surveill. 2017 Jan;22(1):30431.
- 137. Yan J, Chen Y, Yang F, Ling X, Jiang S, Zhao F, et al. High percentage of the ceftriaxone-resistant *Neisseria* gonorrhoeae FC428 clone among isolates from a single hospital in Hangzhou, China. J Antimicrob Chemother. 2021 Mar;76(4):936–9.
- 138. Yan J, Xue J, Chen Y, Chen S, Wang Q, Zhang C, et al. Increasing prevalence of *Neisseria gonorrhoeae* with decreased susceptibility to ceftriaxone and resistance to azithromycin in Hangzhou, China (2015-17). J Antimicrob Chemother. 2019 Jan;74(1):29–37.

- 139. Yang Y, Yang Y, Martin I, Dong Y, Diao N, Wang Y, et al. NG-STAR genotypes are associated with MDR in *Neisseria gonorrhoeae* isolates collected in 2017 in Shanghai. J Antimicrob Chemother. 2020 Mar;75(3):566–70.
- 140. Yéo A, Kouamé-Blavo B, Kouamé CE, Ouattara A, Yao AC, Gbedé BD, et al. Establishment of a Gonococcal Antimicrobial Surveillance Programme, in Accordance With World Health Organization Standards, in Côte d'Ivoire, Western Africa, 2014-2017. Sex Transm Dis. 2019 Mar;46(3):179–84.
- 141. Yin YP, Han Y, Dai XQ, Zheng HP, Chen SC, Zhu BY, et al. Susceptibility of *Neisseria gonorrhoeae* to azithromycin and ceftriaxone in China: A retrospective study of national surveillance data from 2013 to 2016. PLoS Med. 2018 Feb;15(2):e1002499.
- 142. Yuan LF, Yin YP, Dai XQ, Pearline RV, Xiang Z, Unemo M, et al. Resistance to azithromycin of *Neisseria gonorrhoeae* isolates from 2 cities in China. Sex Transm Dis. 2011 Aug;38(8):764–8.
- 143. Zheng XL, Xu WQ, Liu JW, Zhu XY, Chen SC, Han Y, et al. Evaluation of Drugs with Therapeutic Potential for Susceptibility of *Neisseria Gonorrhoeae* Isolates from 8 Provinces in China from 2018. Infect Drug Resist. 2020 Dec;13:4475–86.
- 144. Zheng Z, Liu L, Shen X, Yu J, Chen L, Zhan L, et al. Antimicrobial resistance and molecular characteristics among *Neisseria gonorrhoeae* clinical isolates in a Chinese tertiary hospital. Infect Drug Resist. 2019 Oct;12:3301–9.
- 145. Zhu B, Hu Y, Zhou X, Liu K, Wen W, Hu Y. Retrospective Analysis of Drug Sensitivity of *Neisseria gonorrhoeae* in Teaching Hospitals of South China. Infect Drug Resist. 2021 Jun;14:2087–90.
- 146. Naznin M, Salam MA, Hossain MZ, Alam MS. Current status of gonococcal antimicrobial susceptibility with special reference to Azithromycin and Ceftriaxone: Report from a tertiary care hospital in Bangladesh. Pak J Med Sci. 2018 Nov-Dec;34(6):1397-1401.
- 147. Unemo M, Lahra MM, Cole M, Galarza P, Ndowa F, Martin I, et al. World Health Organization Global Gonococcal Antimicrobial Surveillance Program (WHO GASP): review of new data and evidence to inform international collaborative actions and research efforts. Sex Health. 2019 Sep;16(5):412–25.
- 148. WHO. Global action plan to control the spread and impact of antimicrobial resistance in *Neisseria gonorrhoeae*. World Health Organization; 2012.
- 149. WHO. Antimicrobial resistance global report on surveillance: 2014 summary. World Health Organization; 2014.
- 150. Liang JY, Cao WL, Li XD, Bi C, Yang RD, Liang YH, Li P, Ye XD, Chen XX, Zhang XB. Azithromycin-resistant *Neisseria gonorrhoeae* isolates in Guangzhou, China (2009-2013): coevolution with decreased susceptibilities to ceftriaxone and genetic characteristics. BMC Infect Dis. 2016 Apr 14;16:152.
- 151. Unemo M. Current and future antimicrobial treatment of gonorrhoea - the rapidly evolving *Neisseria gonorrhoeae* continues to challenge. BMC Infect Dis. 2015 Aug 21;15:364.
- 152. Radovanovic M, Kekic D, Jovicevic M, Kabic J, Gajic I, Opavski N, Ranin L. Current Susceptibility Surveil-

lance and Distribution of Antimicrobial Resistance in N. gonorrheae within WHO Regions. Pathogens. 2022 Oct 25;11(11):1230.

- 153. Shaskolskiy B, Kandinov I, Dementieva E, Gryadunov D. Antibiotic Resistance in *Neisseria gonorrhoeae*: Challenges in Research and Treatment. Microorganisms. 2022 Aug 24;10(9):1699.
- 154. Fletcher-Lartey S, Dronavalli M, Alexander K, Ghosh S, Boonwaat L, Thomas J, et al. Trends in Antimicrobial Resistance Patterns in *Neisseria Gonorrhoeae* in Australia and New Zealand: A Meta-analysis and Systematic Review. Antibiotics (Basel). 2019 Oct;8(4):191.
- 155. George CRR, Enriquez RP, Gatus BJ, Whiley DM, Lo YR, Ishikawa N, Wi T, Lahra MM. Systematic review and survey of *Neisseria gonorrhoeae* ceftriaxone and azithromycin susceptibility data in the Asia Pacific, 2011 to 2016. PLoS One. 2019 Apr 3;14(4):e0213312.
- 156. Brown ST, Pedersen HB, Holmes KK. Comparison of erythromycin base and estolate in gonococcal urethritis. JAMA. 1977 Sep 26;238(13):1371-3. https://doi. org/10.1001/jama.1977.03280140049015.
- 157. Lewis DA. The Gonococcus fights back: is this time a knock out? Sex Transm Infect. 2010 Nov;86(6):415-21. 2010;86(6):415-21.
- 158. Laumen JG, Manoharan-Basil SS, Verhoeven E, Abdellati S, De Baetselier I, Crucitti T, et al. Molecular pathways to high-level azithromycin resistance in *Neisseria gonorrhoeae.* J Antimicrob Chemother. 2021 Jun;76(7):1752–8.
- 159. Młynarczyk-Bonikowska B, Majewska A, Malejczyk M, Młynarczyk G, Majewski S. Multiresistant *Neisseria gonorrhoeae*: a new threat in second decade of the XXI century. Med Microbiol Immunol. 2020 Apr;209(2):95–108.