Review Article

Vanadium Inhalation in a Mouse Model for the Understanding of Air-Suspended Particle Systemic Repercussion

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There is an increased concern about the health effects that air-suspended particles have on human health which have been dissected in animal models. Using CD-1 mouse, we explore the effects that vanadium inhalation produce in different tissues and organs. Our findings support the systemic effects of air pollution. In this paper, we describe our findings in different organs in our conditions and contrast our results with the literature.

1. Vanadium as an Example of Chemical Components Adherent to Suspended Particles

Airborne suspended particles (PM) consist of a heterogeneous mixture of solid and liquid particles suspended in air, continually varying in size and chemical composition in time and space. Primary particles are emitted directly into the atmosphere, such as diesel, whereas secondary particles are generated through physicochemical transformation of gases, such as nitrate and sulfate formation from gaseous nitric acid and sulfur dioxide (SO₂), respectively.

The numerous natural and anthropogenic sources of PM include motor vehicle emissions, tire fragmentation, power generation and other industrial combustion, smelting and other metal processing industries, agriculture, construction and demolition activities, residential wood burning, windblown soil, pollens and molds, forest fires and combustion of agricultural debris, volcanic emissions, and sea spray. Although there are numerous chemicals that have been detected in PM in different locations, some of the most common components include nitrates, sulfates, elemental

and organic carbon, organic and biological compounds, and a variety of metals such as vanadium [1].

Vanadium is present in crude oil as an organic–metallic complex. The oil quantity of vanadium varies with the extraction source, finding important differences within countries; for example, Western Venezuelan and Maya Mexican oils have the highest concentrations of vanadium. Also in residual fuel oil, the concentrations are high (0.2 to $0.160 \mu g/g$). Coal is also rich in vanadium (14 to 56 mg/kg). Natural sources also contribute to vanadium particulate concentrations in the atmosphere. Continental dust, volcanic eruptions, sea salt spray, and forest fires are some of the most productive genesis of vanadium [2].

2. Generalities of Vanadium

Vanadium, an outcome of the combustion of oil products, is inhaled adhered to suspended particles. Literature indicated that one of the main mechanisms of vanadium action is through the generation of reactive oxygen species (ROS), by the activation of mitogen activated protein kinases (MAPKs) that will result in the phosphorylation of downstream



FIGURE 1: Comet classification. (a) Comet measurement, (b) no damage, (c) low damage, (d) medium damage, (e) high damage, and (f) total damage. The fluorochromization was performed with ethidium bromide and fluorescence microscope (1000x).

enzymes such as ERK1/2, resulting in the activation or deactivation of diverse signaling pathways.

Vanadium could also damage genetic material and induce hematotoxicity [3–5], immune toxicity [6], hepatotoxicity [7], lung toxicity [8–10], neurotoxicity [11–13], and reproductive toxicity [14–16] All these organs and systems are altered by vanadium inhalation, and in the next sections, we will describe the changes observed in our animal model, in an attempt to translate our findings to the actions that airsuspended particles could have on human health.

3. An Animal Model to Assess Systemic Effects of Air-Suspended Particle

Our model was evaluated in CD-1 mouse establishing a whole-body inhalation model [11, 17, 18]. The exposure schedule consisted of V₂O₅ 0.02 M in saline, one hour twice a week. An ultranebulizer (UtraNeb 99 De Vilbis) with the size of the particle emitted being less than $5 \,\mu$ m (range 0.5– $5 \,\mu$ m) at a flow rate of 10 L/min was attached to a transparent acrylic box reaching stable concentrations in the chamber of 1436 μ m/m³.

3.1. Genotoxicity. DNA damage is induced by exogenous agents such as environmental pollutants [19] or endogenous ones (free radicals) [20]. Genetic damage mechanisms include oxidation of nitrogenous bases or sugar residues; cross-links or DNA strand breaks [21, 22]. When DNA is altered, there are mechanisms for repairing the damage that could disrupt the strands and produce mutations or genetic material loss. Any agent that interacts with the genetic materials is dangerous [23] because the damage end

result could be mitotic arrest, transcription gene induction, replicating errors, and genetic instability that could result in cancer [22, 24].

The adverse effects of vanadium on genetic material are controversial, contradictory, and scarce *in vivo* [25], and because of the scarce information available in the literature, IARC classifies vanadium as a 2B category (possible human carcinogen).

Single-Strand Breaks. Alkaline comet assay was used for the evaluation of single-strand breaks in peripheral leukocytes and bone marrow cells of exposed animals. This early genotoxic biomarker evaluates single-strand breaks [26]. With our model, we identify genotoxic damage in CD-1 mice which correlates with time of exposure. The greater the exposure, the longer the comet tails. With the same model, bone marrow was explored comparing male with female mice, and a clear difference was noticed in male mice, whose cells had greater damage (Figure 1).

Micronucleus. Micronucleus (MN) comprises aneugenic and clastogenic events in the cell that leads to chromosomal instability. The result of this type of genetic damage can induce cancer [27]. Applying Krishna and Hayashi [28] method to quantify MN in reticulocytes, we identified sexand age- (pre- and post-puberty mice) specific differences at 30, 60, and 90 days after the exposure had ended (Figure 2). Our findings indicate that adult females were the least affected. We suggest that estrogens as antioxidant molecules could be protecting females form reactive species produced by the interaction with vanadium [29, 30], but more research about these issues should be encouraged. 3.2. Bone and Teeth Effects. Vanadium is concentrated in the bone inducing changes in its inorganic matrix density. Vanadium concentrations increased in the teeth with the time of exposure. The increased vanadium was associated with a decrease in the thickness of the teeth's enamel layer [31]. If the exposure is during teeth development, incisive histodifferentiation is altered resulting in ameloblast and predentin disorganization (Figures 3(a) and 3(b)).

These changes may explain the increased incidence of cavities in highly polluted areas as reported by Bowen [32].

3.3. Hematopoietic System. Reports of vanadium hematotoxicity are limited, and in many cases, contradictory. Some studies describe alterations in erythrocytes such as anemia, changes in hemoglobin and hematocrit concentrations [33– 35], an increment in osmotic red blood cells fragility [36, 37], contrasting with other reports of microcytic erythrocytosis [38], and the acceleration of maturation on erythropoiesis [39]. Other researchers have found modifications in white blood cells count [33–35], contrasting with other reports of negative results in all hematologic parameters [40, 41]. In the referred studies, the exposure route was oral or parenteral, with different vanadium compounds, but vanadium pentoxide inhalation, the principal source of exposure, has been poorly evaluated.

In subchronic vanadium exposure (eight-week inhalation), we found normochromic normocytic anemia, with a decrease in hemoglobin and hematocrit [3]. This anemia could be a consequence of hemolysis, as other researchers have proposed [36, 37], or might be a kind of erythrocyte suicide identified as eryptosis, specifically described for vanadium toxicity [42]; also an impairment of erythropoiesis could be assumed.

One of the most relevant findings in our model was the change on platelets. There was an increase in platelets counts (thrombocytosis) and morphological changes as megaplatelets [43]. This event was correlated with an increase of size and number of megakaryocytes (megakaryocytosis) in bone marrow and spleen [4, 44]. There are no previous reports of thrombocytosis, but spleen megakaryocytosis is reported as a collateral finding by Al-bayati et al. in 1992 [45]. We consider that anemia may be a consequence of a detour from the bipotential progenitor cell (PEM) differentiation toward megakaryocytes and platelets instead of erythrocytes lineage, but this explanation needs further investigation. Thrombocytosis might be found as a reactive process or as a clonal event. There are some findings in our model such as the morphological changes of platelets (presence of megaplatelets) and megakaryocytes, accompanied with platelets functional impairment (data not published) that might suggest the presence of a clonal event known as essential thrombocythemia, but this hypothesis needs further analysis. Platelet production as a consequence of megakaryocyte proliferation depends mainly on thrombopoietin (TPO), and signaling pathways like JAK/STAT are required, so we are analyzing this pathway activation in our model [5].

Leukocytosis, with neutrophilia and lymphocytosis were observed in our model during the whole exposure. The same findings were reported in a study published by Zaporowska and Wasilewsky in 1992 [35]; however, they explored oral exposure. Fibrinogen and total serum proteins were also elevated [3]. These findings support a chronic inflammation state, and this could be also the explanation for thrombocytosis, but it does not explain changes in platelets and megakaryocytes function and morphology, so further research should be encouraged to understand these changes and to test the possibility of a clonal disorder.

3.4. Hepatotoxicity. Liver is also an inhaled vanadium target reporting that increased lipid peroxidation, evaluated by TBARS, is present during the exposure to this element. Applying another biomarker, 4-hydroxynonenal demonstrates that the hepatocytes' cell membrane is altered by oxidative damage. The oxidative destruction of polyunsaturated fatty acids in the membranes is the alteration that is developed as a positive staining for 4-hydroxynonenal, and the increase of TBARS. The histological evaluation of the liver in our model showed spotlights of inflammatory infiltrate. Other findings to be noted were an increase in the number of binucleated cells with meganucleus and modifications that suggest liver regenerative activity (Figure 4).

3.5. Immune System

3.5.1. Thymus. The thymus provides a variety of specialized microenvironments that support and direct T-cell differentiation and selection. Thymic microenvironment is delimited by a variety of cells such as macrophages, epithelial and dendritic cells (DC), fibroblasts, and immature T cells (thymocytes).

Dendritic thymic cells (DCs) are regularly distributed into the thymus medulla; they are a heterogeneous population of bone-marrow-derived immune cells, and differ in surface markers, migratory patterns, localizations and cytokine production. DCs are responsible for the negative selection of autoreactive T cells as discussed by Wu and Liu [46].

There are some reports that indicate the changes in the cortex-medulla thymic distribution as a consequence of V_2O_5 exposure as well as changes in cell populations' arrangement in a mice model. These changes suggest a modification in the CD's morphophysiology with possible health repercussion as discussed by González-Villalva et al. [43]. In the same experimental model we have been using, by immunohistochemistry for CD11c, we identify a decrease in color density for CD11c as the time of exposure increases (Figures 5(a) and 5(b)). This could be explained because of the vanadium oxidative potential, which generates reactive oxygen species (ROS); that could modify transcriptional factors associated with the inflammatory response as discussed by Gusik et al. [47].

3.5.2. Spleen. Vanadium exposure produces different effects on spleen such as splenomegaly, increase in megacaryocytes number and size, and the enlargement of the size of the white pulp, mainly because of the increase of CD19+ cells



FIGURE 2: Micronucleus assay with mouse peripheral blood reticulocytes using acridine orange-coated slides. (a) Reticulocytes (white arrows) and erythrocytes (dark cells) (400x). (b) Micronuclei (head arrow) in reticulocyte (1000x).



FIGURE 3: (a) Control dental primordium section from upper incisor with trichrome Masson stain. (A): Ameloblast. (O): Odontoblast. Predentine (PD). (b) Exposed dental primordium section from upper incisor with trichrome Masson stain. (A): Ameloblast. (O): Odontoblast. Predentine (PD).



FIGURE 4: Meganucleated hepatocytes (arrow) were observed in the group of animals exposed to inhaled vanadium pentoxide. (400x).

(Figures 6(a) and 6(b)). Recently, it has been noted that these changes are due to an increment on the lymphocyte proliferation (Figures 7(a) and 7(b)).

Also we have observed nuclear changes in spleen lymphocytes and thymus after the first week of exposure and until three months of treatment. This could be due to a modification in the cytoskeleton and nucleoskeleton of these cells (Figures 8(a) and 8(b)). *3.5.3. Lymph Nodes.* In lymph nodes, we observed that after 4 weeks of inhalation, there was an evident increase in the presence of CD25, a late lymphocyte activation marker. This was particularly true in the lymph node medulla, which contains primarily B cells. These findings match with what was observed in the spleen since, once again, B cells appear to be specially affected by vanadium (Figures 9 and 10).

3.6. Lung Toxicity. Adverse effects of vanadium in the lung have been associated to suspended particles air pollution exposure in humans, and experimental studies have been carried out in primates and rodents. These studies report that inhalation exposure results in impaired lung function, influx of inflammatory cells in bronchiolar lavage, and fibrotic changes in the lungs [8, 9, 48].

In rats and mice exposed to V_2O_5 (whole-body), increases in inflammation and epithelial hyperplasia were observed at a 2 mg/m^3 dose. In rats exposed to 4 mg/m^3 , fibrosis and a restrictive lung disease were observed [38].

In humans, accidental or inadvertent exposure to this compound in the workplaces has been associated with inflammatory responses in the respiratory tract, including bronchitis (often called boilermaker's bronchitis), pneumonia, rhinitis, and pharyngitis [49].



FIGURE 5: (a) CD11c+ thymic cells (*) in control mice 2 week. Thymic cortex (c) and thymic medulla (m). (b) CD11c+ thymic cells (*) in exposed mice 2 week. Thymic cortex (c) and thymic medulla (m).



FIGURE 6: Enlargement of spleen lymphoid nodules due to V_2O_5 exposure. (a) Normal histology of spleen in control animals. (b) The growth of the white pulp mainly by lymphoid nodules (arrows) in the spleen from V_2O_5 that exposed animals is observed.



FIGURE 7: V_2O_5 effects on the lymphocyte proliferation index. In control spleen, a basal proliferation index is observed. (a) After vanadium exposure, the number of lymphocytes in division increased contrasting with control (b) (1000x).

In our model, the inhalation of vanadium caused an increase of cytokines such as IL-6 and TNF- α , located in the bronchiolar epithelium. This inflammatory response led to thickening of the bronchiolar muscular layer evaluated by alpha-actin immunohistochemistry [8]. This change was evidenced at 12-week exposure (Figure 11). The modifications found suggest airway remodeling similar to that observed in asthma [8].

Bonner and coworkers [8] showed that rat intratracheal instillation with vanadium caused lung inflammation, airway hyperreactivity, fibroproliferative changes, mucus cell metaplasia, smooth muscle layer thickening, and airway fibrosis. However, we did not observe fibrosis in our inhalation model.

What we observed were changes in the nonciliated bronchiolar cell (NCBC) and ciliated cells nucleus that



FIGURE 8: Spleen lymphocytes' nuclear changes after V_2O_5 exposure. (a) In control animal's nuclear morphology, regular ultrastructure in lymphocytes is observed (7000x) (arrow). (b) Vanadium exposure produces alterations in lymphocyte's nuclear morphology similar to the changes observed in malignant cells (8000x) (arrows).



FIGURE 9: Immunofluorescence staining for CD25 in a lymph node medulla in a control mouse.



FIGURE 10: Immunofluorescence staining for CD25 in lymph node medulla after 4-week inhalation.

resembled pseudoinclusions [10]; also a decrease in the number of the NCBC was noticed [50].

3.7. Neurotoxicity. The central nervous system (CNS) exhibits a great vulnerability to oxidative stress damage, because of its continuous production of reactive oxygen species as a consequence of its high aerobic metabolism, low levels of antioxidants, and its extended surface rich

in peroxidizable fatty acids which has been invoked as an excellent field for neuronal injury [51]. This damage might be expressed as neurodegenerative changes [52, 53].

In our mice model, vanadium induced functional, cytological, and ultrastructural alterations in the olfactory bulb, motor cortex (Figure 12(b)), hippocampus, striatum, substantia nigra, and ependyma [54]. In the olfactory bulb granule cells, spine density decreased at different times of exposure (4, 5, 6, 7, 8, 10, and 12 weeks). In addition, lipofuscin granules, swelling organelles, vacuolation, and cytoplasmic condensation with disrupted mitochondria were recorded; apoptosis and necrotic neuronal deaths were also identified [54]. In our hands, these alterations were associated with olfactory dysfunction [55].

Our model showed nigrostriatal dopaminergic system damage that resulted in a decrease in the number of tyrosine hydroxylase immunoreactive neurons in the *substantia nigra pars compacta* and dendritic spine loss in the medium-sized spiny neurons of the striatum in exposed mice [12]. These changes might be the consequence of dopamine autooxidation resulting in quinones and semiquinones that may lead to *substantia nigra* cell death [56]. Oxidative damage, in our model, was confirmed by 4-hydroxynonenal (4-HNE) a lipid peroxidation marker by immunohistochemistry (Figure 12(d)). The damage observed in motor cortex neurons caused cell death, possibly by excitotoxicity with motor impairment [57].

In hippocampal formation, we observed morphological and functional alterations in the pyramidal neurons to CA1 and granule cells to dentate gyrus. A decrease in dendritic spines and pyramidal and granule cell necrosis were detected. These morphological alterations might correlate with spatial memory impairment [13, 58].

Blood-brain-barrier was also affected observing cilia loss in the fourth ventricle, as well as cell sloughing and dissolution of tight junctions between ependymal cells and basal membrane causing an evident disruption of the BBB [11]. These modifications were extended to the choroid plexus in lateral ventricles [57]. Additionally, an increase in gelatinases MMP-9 and MMP-2 activation in the olfactory bulb, prefrontal cortex, hippocampus, and striatum were



FIGURE 11: Mouse bronchioles. (a) Normal bronchioles label with alpha-actin (arrows). (b) Thickening smooth muscle layer after 12-week exposure to vanadium. Original magnification 100x.



FIGURE 12: Photomicrograph of cortex pyramidal cells with Golgi stain. (a) Dendrite with spine dendrite in a control. (b) Exposure to vanadium, area with spine dendritic loss (arrow). (c) Image from a section of the cerebral cortex in control mice (confocal): the neuronal bodies (arrows). (d) Pyramidal cell neurons exposed to vanadium stained with 4-HNE (arrows). Scale bars: (a), (b) $10 \mu m$; (c), (d) $50 \mu m$.

observed [59], changes that could be associated to previously reported dendritic remodeling in the tissues [60] (Figure 12).

3.8. Reproductive Toxicity. The male reproductive system consists of the testes, their associated ducts, the accessory glands, and the penis. The testes produce sperm and testosterone. The seminiferous tubules are located in the testes [61], and the lining cells of the tubules comprise the seminiferous epithelium. Some of them are the source of sperm and some are there to sustain the production. The Sertoli or sustentacular cell helps the developing of sperm cells and defines the seminiferous tubule in two separate

and physiologically different compartments, the basal and the luminal by occluding junctions of adjacent Sertoli cells. The seminiferous epithelium contains the sperm cells; the spermatogonium (stem cell) originates the sperm cells. These cells undergo meiosis and become haploid spermatocytes and spermatids, which further transform its complete morphological differentiation into spermatozoa [62].

Animal evidence demonstrates that i.p. vanadium treatment has adverse impacts on male reproductive health [14, 15, 63]. We compared the fine structure of seminiferous tubules in two groups of CD1 mice (experimental and control) [16], the experimental group underwent V_2O_5 inhalation at 0.02 M for a 12-week time period. Our studies



FIGURE 13: Actin Immunohistochemistry stain in seminiferous tubules. (a) Control animal showing positive immunoreactivity in Sertoli (arrow), Leydig (*) and germinal cells (clasp). (b) After six weeks of treatment the immunoreactivity decreased in all the testicular cells.



FIGURE 14: Immunohistochemical analysis for Cx43 localization in mice testes exposed for four weeks to V_2O_5 inhalation. (a) Cx43 in control mice was localized between Sertoli cells and germinal cells (arrows). (b) Cx43 immunostaining was absent in exposed mice.

demonstrated that V_2O_5 causes ultrastructural changes in testis of exposed mice. Ultrastructure examination revealed spermatogonia, spermatocytes, and Sertoli cells necrosis; the cells had obvious pathological changes including vacuolation cytoplasm and mitochondrial swelling, as well as decrease or vanish of mitochondria crista. Compared with the control group, the intercellular membranes of various seminiferous cells, as well as spermatozoa and Sertoli cells, were detached in the V_2O_5 -exposed groups in a dose-dependent manner. In addition, other effects, such as pseudonuclear inclusions, were found.

Some toxic reproductive effects that were observed in this and other models, like alterations in spermatogenesis, low sperm count, decrease in sperm motility, and increase in the frequency of sperm morphologic abnormalities including morphological nuclear changes, suggest damage to the cytoskeleton [18].

Cytoskeletal proteins play an important role in the regulation of spermatogenesis and other process in the testis. Gamma-tubulin is a part of the cytoskeletal proteins and an important member of centrosome, which is the main microtubule-organizing center in animal cells. In a previous report, we observed that vanadium exposure decreased gamma-tubulin in Sertoli, Leydig and germinal cells in a time-dependent manner [18].

On the other hand, we have also observed that V is able to alter other cytoskeletal proteins like the actin in testicular cells. Actin is involved in different processes like cell division, cell morphological changes during differentiation, hematotesticular barrier integrity, and organization of testicular cells by junctional complex. Similar to tubulins, vanadium also alters actin in Sertoli, Leydig, and germinal cells in a time dependent manner. However, actin was less sensitive to the effect of vanadium than gamma-tubulin in testicular cells (Figure 13).

The damage to the cytoskeleton of testicular cells produced by vanadium implies changes in testicular functions leading to infertility, a phenomenon that has been implicated in vanadium exposure.

Metals also affect intercellular junctions by either reducing the amount of these or inducing aberrant intracellular localization of these membranous proteins [64]. Evidence indicates that these cell adhesion molecules are important as early targets for a variety of toxic substances including vanadium [65]. We used immunohistochemistry for localization of the gap-junctional proteins in Sertoli and spermatogenic cells, and Cx43 presence was observed essentially in the cell membrane of the basal compartment in control mice testis. Immunostained area revealed that there was a lack of membranous distribution of Cx43 and a shift from membranous immunoreactivity to predominantly cytoplasmic in exposed animals to V_2O_5 by four weeks [65] (Figure 14).

4. Conclusions

Atmospheric pollution repercussions by air-suspended particles are almost neglected. The use of animal models like the one described here helps to understand the interactions of the respiratory system, that is, the main entrance for air pollutants, and other organs such as the liver, reproductive system and nervous system. The increase in neurodegenerative diseases, infertility, and immune disorders needs a deeper evaluation for its possible link with air pollutants. These studies will support the reinforcement of atmospheric exposure limits worldwide.

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