

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Role of viral infections in the development and exacerbation of asthma in children



Tuomas Jartti, MD,^a and James E. Gern, MD^b Turku, Finland, and Madison, Wis

INFORMATION FOR CATEGORY 1 CME CREDIT

Credit can now be obtained, free for a limited time, by reading the review articles in this issue. Please note the following instructions.

Method of Physician Participation in Learning Process: The core material for these activities can be read in this issue of the Journal or online at the JACI Web site: www.jacionline.org. The accompanying tests may only be submitted online at www.jacionline.org. Fax or other copies will not be accepted.

Date of Original Release: October 2017. Credit may be obtained for these courses until September 30, 2018.

Copyright Statement: Copyright © 2017-2018. All rights reserved.

Overall Purpose/Goal: To provide excellent reviews on key aspects of allergic disease to those who research, treat, or manage allergic disease.

Target Audience: Physicians and researchers within the field of allergic disease.

Accreditation/Provider Statements and Credit Designation: The American Academy of Allergy, Asthma & Immunology (AAAAI) is accredited by the Accreditation Council for Continuing Medical Education (ACCME) to provide continuing medical education for physicians. The AAAAI designates this journal-based CME activity for a maximum of 1.00 *AMA PRA Category 1 Credit*TM. Physicians should claim only the credit commensurate with the extent of their participation in the activity.

List of Design Committee Members: Tuomas Jartti, MD, and James E. Gern, MD (authors); Cezmi A. Akdis, MD (editor)

Disclosure of Significant Relationships with Relevant Commercial

Companies/Organizations: T. Jartti has received a grant from the Sigrid Juselius Foundation. J. E. Gern has received a grant from the National Institutes of Health/National Institutes of Allergy and Infectious Disease; has consultant arrangements with Janssen, Regeneron, and PReP Biosciences; and has received travel support from Boehringer Ingelheim. C. A. Akdis disclosed no relevant financial relationships.

Activity Objectives:

- To become familiar with current evidence pertaining to the role of viral respiratory tract infections in the development and exacerbation of asthma.
- 2. To provide an overview of interactions between aeroallergen sensitization and viral infection in the development of asthma.
- 3. To highlight the gaps in existing knowledge regarding the role of viral infection in asthmatic patients.

Recognition of Commercial Support: This CME activity has not received external commercial support.

List of CME Exam Authors: Nicholas Klaiber, MD, and Wei Zhao, MD, PhD.

Disclosure of Significant Relationships with Relevant Commercial Companies/Organizations: The exam authors disclosed no relevant financial relationships.

Viral infections are closely linked to wheezing illnesses in children of all ages. Respiratory syncytial virus (RSV) is the main causative agent of bronchiolitis, whereas rhinovirus (RV) is most commonly detected in wheezing children thereafter. Severe respiratory illness induced by either of these viruses is associated with subsequent development of asthma, and the risk is greatest for young children who wheeze with RV infections. Whether viral illnesses actually cause asthma is the subject of intense debate. RSV-induced wheezing illnesses during infancy influence respiratory health for years. There is definitive evidence that RSV-induced bronchiolitis can damage the airways to promote airway obstruction and recurrent wheezing. RV likely causes less structural damage and yet is a significant contributor to wheezing illnesses in young children and in the context of asthma. For both viruses, interactions between viral virulence factors, personal risk factors (eg, genetics), and

From ^athe Department of Paediatrics, Turku University Hospital and University of Turku, and ^bthe Departments of Pediatrics and Medicine, University of Wisconsin School of Medicine and Public Health, Madison.

Supported by the Sigrid Jusélius Foundation, Helsinki, Finland, and National Institutes of Health grants UG3 OD023282, P01 HL070831, U19 AI104317, and UM1 AI114271.

Received for publication June 16, 2017; revised August 3, 2017; accepted for publication August 22, 2017.

environmental exposures (eg, airway microbiome) promote more severe wheezing illnesses and the risk for progression to asthma. In addition, allergy and asthma are major risk factors for more frequent and severe RV-related illnesses. Treatments that inhibit inflammation have efficacy for RV-induced wheezing, whereas the anti-RSV mAb palivizumab decreases the risk of severe RSV-induced illness and subsequent recurrent wheeze. Developing a greater understanding of personal and environmental factors that promote more severe viral illnesses might lead to new strategies for the prevention of viral wheezing illnesses and perhaps reduce the subsequent risk for asthma. (J Allergy Clin Immunol 2017;140:895-906.)

Key words: Asthma, bronchiolitis, child, exacerbation, respiratory syncytial virus, rhinovirus, virus, wheeze, wheezing

0091-6749/\$36.00

© 2017 Published by Elsevier Inc. on behalf of the American Academy of Allergy, Asthma & Immunology

http://dx.doi.org/10.1016/j.jaci.2017.08.003

Corresponding author: Tuomas Jartti, MD, Department of Paediatrics, Turku University Hospital, PO Box 52, FIN-20520 Turku, Finland. E-mail: tuomas.jartti@utu.fi.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections

Abbrevia	tions used			
CDHR3:	Cadherin-related family member 3			
nBreg:	Neonatal regulatory B			
NGF:	Nerve growth factor			
OR:	Odds ratio			
RSV:	Respiratory syncytial virus			
RV:	Rhinovirus			
TLR:	Toll-like receptor			

Discuss this article on the JACI Journal Club blog: www.jacionline.blogspot.com.

Bronchiolitis, acute wheezing illnesses, and asthma are a huge clinical burden. The prevalence of bronchiolitis is approximately 20% to 30% in the first year and 10% to 20% in the second year of life.^{1,2} Up to 30% to 50% of children have acute wheezing at least once before school age.¹ Of these, 30% to 40% will have recurrent wheezing.¹ Eventually, the prevalence of asthma is approximately 5% to 10% in children.³

The diagnostics of viral respiratory tract infections has improved markedly during the last 2 decades because of the development of PCR techniques. Several new respiratory viruses and their subgroups have been discovered, and especially rhinovirus (RV) diagnostics have markedly improved.⁴ We have learned that bronchiolitis and early wheezing episodes are almost always (90% to 100% of cases) associated with viral infections.^{5,6} The overall virus detection rates slightly decrease by age, being 80% to 95% in older children.⁷

Prediction of childhood asthma has been limited for many years to assessment of traditional risk factors, such as atopic characteristics (aeroallergen sensitization, increased blood eosinophil count, or atopic eczema), parental asthma, or factors related to parental atopy. Acute wheezing illnesses with RV and respiratory syncytial virus (RSV) are early markers for recurrent wheezing.^{6,8-14} In addition, RV-induced wheezing episodes in infancy are a major risk factor for later asthma, especially in children with atopic features. Once asthma is established, exposure to allergens and RV infections are important triggers of asthma exacerbations in children.¹⁵

This review will focus on the role of viral infections on the development and exacerbation of asthma in children. Understanding the mechanisms of these events could suggest novel insights into the pathogenesis of asthma and would help to identify novel strategies for the prevention and treatment of asthma.

CLINICAL DEFINITIONS

Bronchiolitis is a virus-induced infection with inflammation of the small bronchioles and their surrounding tissue. Clinically, it is characterized as the first expiratory breathing difficulty in children less than 2 years of age. Other lower respiratory tract symptoms include dry cough, tachypnea, hyperinflation, chest retraction, and widespread crackles or wheezing. In many studies wheezing is not a mandatory diagnostic criterion, and the upper age limit varies from 6 months to 2 years.¹⁶

Wheezing is defined as a whistling sound during expiration accompanied by dyspnea.¹⁷ Wheezing can be diagnosed if there is a reversible expiratory airway obstruction and the illness does not



FIG 1. RV and RSV interactions with airway epithelial cells. *ICAM-1*, Intercellular adhesion molecule 1; *LDL-R*, low-density lipoprotein receptor.

fulfill the diagnosis of bronchiolitis or asthma. Moreover, wheezing is divided into different phenotypes based on natural history, such as "transient early," "persistent," and "late-onset" wheezing. Typically, the 2 latter phenotypes are more closely associated with sensitization and asthma.¹

Asthma is a chronic disorder characterized by airway inflammation, increased mucus secretion, and bronchial hyperresponsiveness, all of which cause reversible airflow obstruction.¹⁷ The chronic inflammation, disrupted epithelium, and airway remodeling increase the susceptibility to many environmental factors, such as viral infections and allergens.

VIRUS CHARACTERISTICS RVs

RVs are nonenveloped positive-strand RNA viruses in the family Picornaviridae and genus *Enterovirus* and are classified into 3 species (RV-A, RV-B and RV-C; Fig 1).¹⁸ There are more than 160 distinct RV genotypes, including 80 RV-A and 32 RV-B serotypes and 65 newly identified RV-C serotypes. RV structural and genetic variability has inhibited efforts to develop antivirals. For example, small molecules ("capsid binding agents") that inhibit RV-A and RV-B binding and replication are not effective against RV-C because of differences in capsid structure.¹⁹ 3C protease inhibitors are effective *in vitro*, but results in clinical trials were disappointing.^{20,21} The large number of antigenically distinct RV types has been a barrier to vaccine development, although new approaches have identified some degree of cross-reactivity among RV types,²² and a highly multiplexed RV vaccine is immunogenic in animal models.²³

Detection and epidemiology. RV-C does not grow in conventional cell cultures, which delayed its discovery until 2006,²⁴ approximately 50 years after the first discovery of RVs. PCR is the method of choice for identifying RVs from nasal mucus samples.²⁵ Up to 35% of asymptomatic subjects have positive results for RV,²⁶ but the virus does not cause chronic infection or "colonization" in healthy subjects.²⁷ Both symptomatic and asymptomatic infections can induce systemic immune responses in young wheezing children.²⁸

RVs circulate year-round, with multiple coexisting genotypes,²⁹ and peak prevalence in temperate climates occurs in the early autumn and late spring. Most infections cause common cold symptoms.³⁰ The prevalence of RV-induced bronchiolitis/wheezing is age dependent. In children hospitalized for lower respiratory tract illness, RSV is detected most often until about 12 months of age, and RV is most common in older children.⁷ RV predominates as an etiologic agent in 50% to

						Prevalence of recurrent wheezing/	
Study site	Inclusion criteria	First author, year	No.	Outcome, age (y)	Virus risk factors, OR (95% CI)†	asthma in virus groups	Other risk factors, OR (95% CI)†
Kuopio, Finland	Bronchiolitis, 1-23 mo, hospitalized	Kotaniemi- Syrjänen, 2003	82	Asthma, 7.2	RV: 4.1 (1.0-17), other viruses NS	RV: 52%	_
		Hyvärinen, 2007	81	Asthma, 12.3	NS	_	_
		Ruotsalainen, 2013	67 patients with bronchiolitis, 155 control subjects	Asthma, 16.5	RV: 7.3 (2.1-26), RSV: 5.7 (1.6-20)	RV: 28% RSV: 24%	B-eos: 21.3 (1.1-430), atopic dermatitis: 6.0 (1.3-27), as-IgE: 6.0 (1.1-33)
Madison, Wisconsin	Wheezing, <12 mo, outpatients, high atopy risk, birth cohort	Lemanske, 2005	275	Recurrent wheezing, 3-4	Wheezing <12 mo — RV: 10 (4.1- 26), RSV: 3.5 (1.7-7.5), other: 4.6 (2.0-11)	Wheezing <12 mo — RV: 65%, RSV: 48%, other: 49%	Positive egg IgE level: 3.0 (1.1-7.8), older siblings: 2.6 (1.3-5.2)
		Jackson, 2008	259	Asthma, 6	Wheezing <12 mo — RV only: 2.9 (1.1-7.5), RSV only: 1.2 (0.4-3.2) Wheezing <24 mo — RV only: 5.6 (2.4-13), RSV only: 1.3 (0.4-3.8) Wheezing <36 mo — RV only: 43 (12-149), RSV only: 14 (3.4-54)	Wheezing <12 mo — RV only: 47%, RSV only: 27% Wheezing <24 mo — RV only: 61%, RSV only: 26% Wheezing <36 mo — RV only: 89% RSV only: 73%	Aeroallergen sensitization, first year: 4.3 (1.4-13)
		Rubner, 2017	217	Asthma, 13	Wheezing, <36 mo — RV: 3.3 (1.5-7.1), RSV: NS	Wheezing, <36 mo — RV: 40%	Aeroallergen sensitization, first year: 6.0 (2.5-14), aeroallergen sensitization, first 3 y of life: 21, with RV: 7.9
Turku, Finland	First wheezing, 3-23 mo, hospitalized	Lehtinen, 2007	118	Recurrent wheezing, 2.1	RV HR: 5.1 (1.0-25); non-RV/RSV HR: NS	RV: 50%	Atopy HR: 4.7 (1.9-11), eczema HR: 3.3 (1.3-8.4), age HR: 3.0 (1.4-6.6)
		Lukkarinen, 2017	127	Atopic and nonatopic asthma, 7.7	Atopic asthma — RV: 5.0 (1.1-22) Nonatopic asthma — non- RSV/RV: 8.0 (2.3-28)	RV without atopic characteristics: -, RV and sensitization: 50%, plus eczema: 71%, plus parental asthma: 100%	Atopic asthma — sensitization: 12 (3.0-44), eczema: 4.8 (1.4-17) Nonatopic asthma — age: 7.3 (1.7-31), parental smoking: 3.8 (1.2-13)

TABLE I. RV-induced wheezing illnesses during infancy and the risk of recurrent wheezing and asthma*

(Continued)

TABLE I. (Continued)

Study site	Inclusion criteria	First author, year	No.	Outcome, age (y)	Virus risk factors, OR (95% Cl)†	Prevalence of recurrent wheezing/ asthma in virus groups	Other risk factors, OR (95% Cl)†
Perth, Australia	Wheezing, <12 mo, outpatients, high atopy risk	Kusel, 2007	198	Recurrent wheezing, 5	RV: 2.5 (1.1-5.9) RSV: 2.5 (1.0-11.3)	_	Atopic by age 2 y and RSV: 4.1 (1.3-9.5), atopic by age 2 y and RV: 3.2 (1.1-9.5)
		Kusel, 2012	147	Asthma, 10	RV or RSV: NS	—	RV and atopic after 2 y: RR 3.4 (1.1-10)
Rome, Italy	First bronchiolitis, <12 mo, hospitalized	Midulla, 2012	262 patients with bronchiolitis, 39 control subjects	Recurrent wheezing, 14 mo	RV: 3.3 (1.0-11)	RV: 80%	Absence of lung consolidation: 2.6 (1.1-6.1), family history of asthma: 2.5 (1.2-4.9)
		Midulla, 2014	230	Recurrent wheezing, 3.2	RV: 3.2 (1.1-9.6)	RV: 64%	B-eos >400 cells/ μL: 8.4 (1.6-45)
Soma, Japan	Lower respiratory tract infection, ≤3 y, hospitalized	Takeyama, 2014	80 patients with wheezing and 136 patients without wheezing at admission	Recurrent wheezing, 4.2	Wheezing group: RV vs RSV, P = .035 Nonwheezing group: NS	Wheezing group — RV: 81%	(
Three centers, Finland	Bronchiolitis, <24 mo, hospitalized	Bergroth, 2016	365 total 177, <12 mo with first episode	Asthma, 1.7	All — RV: 9.1 (4.3-19) Non-RV/RSV: 2.7 (1.3-5.6) <12 mo with first episode — RV: 20.4 (4.9-86), non-RV/RSV: 3.8 (1.1-13)	All — RV: 61%, non-RV/RSV: 36% <12 mo with first episode, —	_

---, No data given; *as-IgE*, allergen-specific IgE; *B-eos*, blood eosinophil count; *HR*, hazard ratio; *NS*, nonsignificant; *RR*, risk ratio. *Including prospective studies that have used both RV and RSV detection.

†Unless otherwise stated.

80% of wheezing episodes and asthma exacerbations in children.^{6-8,31-33} In infants aged less than 12 months, RV causes approximately 20% to 40% of bronchiolitis or acute wheezing episodes in emergency department and hospital settings and is second only to RSV.^{34,35} RV is also the leading cause of bronchiolitis, leading to hospitalization outside the winter RSV-induced bronchiolitis season.³⁶ RV-A and RV-C species cause more severe respiratory illness than RV-B species.³⁷

School-age asthma after RV-induced bronchiolitis/ wheezing. RV-induced severe bronchiolitis/early wheezing is a more robust marker of asthma risk than wheezing episodes caused by RSV or other viruses (Table I).^{6,8,13,14,38,39} High-risk birth cohorts, which have included wheezing children with at least 1 atopic parent, have shown a close association between early-life RV-induced wheezing and school-age asthma.^{6,9} The Childhood Origins of Asthma (COAST) study demonstrated that the risk for asthma by age 6 years was increased if the children had wheezing with RV (odds ratio [OR], 9.8) versus RSV (OR, 2.6) during the first 3 years, and furthermore, 90% of the children with RV-induced wheezing in the third year of life had asthma by age 6 years (OR, 26).^{6,40} Although RV-induced wheezing was an independent asthma risk factor, aeroallergen sensitization markedly increases the RV-associated risk of asthma.^{6,9} An Australian birth cohort study showed that the risk for wheezing at age 5 years was increased if the wheezing at less than 1 year of age was associated with RV either alone (OR, 3.2) or with concomitant RSV (OR, 4.1) but only in children with sensitization at an age of less than 2 years.⁹ Therefore data from these high-risk birth cohorts suggest that atopic airways have an increased susceptibility to long-term dysfunction after RV-induced wheezing illnesses.

In addition, the subsequent asthma risk has also been demonstrated in population-based long-term follow-up studies in children hospitalized for the wheezing episode.^{8,12} In a Finnish study asthma at age 7 years was more common after RV-induced (52%; OR, 4.1) than after RSV-induced (15%) severe bronchiolitis.⁸ In prospective studies viral wheezing episodes in infancy are associated with increased asthma risk for as long as 15 to 18 years.¹¹ One study focused on the first episode of severe wheezing and demonstrated an association between RV and

school-age atopic asthma (84%; OR, 5.5) in an 8-year follow-up.¹³ When the upper age limit of bronchiolitis is set to 6 months, non–RSV-induced bronchiolitis (most of these cases are RV induced) has shown higher asthma risk (24%) at age 6.5 years than RSV-induced bronchiolitis (8%).⁴¹ The association between RV-induced wheezing and the development of childhood asthma has been confirmed in a recent meta-analysis including 15 original articles.⁴²

RSV

RSV is a pneumovirus in the Paramyxoviridae family and is a single-stranded enveloped RNA virus with 2 major antigenic groups, A and B (Fig 1). The genetic diversity of proteins among the A and B RSV groups forms several subgroups with 10 A genotypes and 13 B genotypes.⁴³ There is evidence that some strains are more likely to cause lower airway disease, and virulence factors have been identified.⁴⁴ Monoclonal antibodies to the RSV fusion (F) protein inhibit viral attachment and the severity of clinical illness. An RSV vaccine has been elusive, but several candidates are now in clinical development.^{45,46}

Detection and epidemiology. Rapid RSV antigen detection and PCR appear to have equal sensitivity in detecting the virus⁴⁷; however, the former is used more often in clinical decision making. RSV causes up to 80% of bronchiolitis cases, the peak incidence being in infants between 3 and 6 months of age. Epidemics occur typically at midwinter, and nearly all children have had RSV infection by age 2 years. During the first year of life, approximately 20% require outpatient medical care, whereas 2% to 3% with more severe illness need hospitalization caused by RSV-induced bronchiolitis are age less than 3 months, prematurity with the presence of chronic lung disease, congenital heart disease, immunodeficiency, and neuromuscular disorders.^{2,48-51}

Childhood asthma after RSV-induced bronchiolitis. Many prospective long-term follow-up studies have shown that RSV-induced bronchiolitis is associated with later development of asthma.^{11,52,53} For example, the Tucson Respiratory Study linked RSV-induced bronchiolitis to asthma up to age 11 years, and a Finnish study linked RSV-induced bronchiolitis to self-reported asthma up to age 15 to 18 years.^{11,52} A case-control study of Swedish children reported an association between RSV-induced bronchiolitis and subsequent allergic sensitization,^{10,54-56} but this finding has not been reproduced in birth cohort studies.⁵³

Whether this association is causal has been the subject of considerable debate. A retrospective cohort study including more than 95,000 infants showed that infants born 3 months before the RSV season had the greatest risk for hospitalization because of lower respiratory tract illness and the highest risk for having asthma between the ages of 4 and 5 years,⁵⁷ suggesting causality. A Danish study including more than 18,000 Danish twins reached a different conclusion: severe RSV-induced illnesses were associated with recurrent wheezing during early childhood,^{58,59} but this was attributable to genetic predisposition for both more severe RSV-induced illness and asthma.⁵⁹ Two studies of RSV immunoprophylaxis with palivizumab in preterm or high-risk infants demonstrated that prevention of more severe RSV-induced illness decreased recurrent wheezing but not atopic asthma.^{60,61}



FIG 2. Interacting factors that contribute to the severity of virus-induced wheezing illnesses and the risk for subsequent development of childhood asthma.

SIMILARITIES AND DIFFERENCES BETWEEN RV AND RSV

Clinical characteristics

There are several differences between RV- and RSV-induced illnesses with respect to at-risk populations and clinical characteristics.^{36,62} Children hospitalized for RV-induced wheezing tend to be older, are more likely to have wheezed previously, more often have allergic sensitization compared with those with RSV-induced wheezing,^{5,7,33,63-66} and also show a favorable response to oral corticosteroid treatment whereas those with RSV-induced wheezing do not.^{67,68} Although RSV can generally cause more severe infections in infants than RV, the inception of wheeze might be more rapid (and duration shorter) with RV compared with RSV infection.^{69,70} These observations are further supported by cluster analysis, which included 2615 children with bronchiolitis and identified showed 4 different distinct clinical profiles.⁷⁰

Pathophysiology

RV and RSV are both transmitted mainly through direct contacts and aerosol particles. Both viruses replicate in ciliated epithelial cells of the upper airways and in medium- to large-sized lower airways (Fig 1).⁷¹ RSV infections can extend into the small airways and can also infect type I pneumocytes. These viruses attach to unique cellular receptors: intercellular adhesion molecule 1 used by RV-B and most RV-As, low-density lipoprotein receptor used by some RV-As, cadherin-related family member 3 (CDHR3) used by RV-C, and CX3CR1 used by RSV.^{72,73} RSV induces epithelial cell apoptosis and necrosis and generally causes more damage to the airway epithelium compared with RV. Surfactant proteins have shown protective effects against RSV infection by regulating innate and adaptive immunity and participating in host defense pathways, such as regulation of proinflammatory cytokine production, chemotaxis, or tissue repair.

After RV attachment, infected cells recognize RV pathogen-associated molecular patterns through interaction with 2 different families of pattern recognition receptors: Toll-like receptor (TLR) 2, TLR3, TLR7, and TLR8 and retinoic acid–inducible gene I–like receptors.^{74,75} TLR4 is a key regulator

of both innate and adaptive immune responses in RSV infection.⁴³ These receptors activate transcription factors (eg, interferon regulatory transcription factor 7 and nuclear factor κB) that promote the expression of type 1 and type III interferons and several inflammatory cytokine genes.⁷⁶ Early innate immune responses, such as type 1 interferons, occur very rapidly after either RV or RSV infection of the epithelium.³⁶ Both viruses induce cytokines (IL-1, TNF, IL-6, IL-12, IL-18, and IFN- γ), chemokines (CCL3, CCL5, CXCL8, and CXCL10), and growth factors that activate and attract granulocytes, dendritic cells, and monocytes at the site of infection. Transcriptional profiling of PBMCs during RSV infections demonstrates stimulation of innate immune and cell-cycle pathways and downregulation of B and T cell-related genes. The latter finding was more pronounced in infants infected with RSV compared with RV, which is consistent with other studies that have demonstrated virus-specific patterns of gene expression.^{77,78} Interestingly, RSV can infect B cells,⁷⁹ whereas some RVs can bind to B cells and induce proliferation.⁸⁰ The combined effects of the virus and the inflammatory response lead to epithelial damage and sloughing, mucus production, and ultimately airway obstruction leading to wheezing. 36,51,81,82

Recent findings from cross-sectional and birth cohort studies indicate that environmental exposures modify the host response to respiratory viruses in early childhood. For example, children on European dairy farms are less likely to have transient wheezing illnesses, which are mostly of viral cause.⁸³ Similarly, children from Wisconsin dairy farm families have fewer medically attended respiratory illnesses compared with those from nonfarm families.⁸⁴ *Lactobacillus johnsonii*, an environmental microbe associated with pet ownership, can protect against RSV-induced pathology in a mouse model.⁸⁵ In urban areas early-life exposure to high levels of indoor allergens (cockroach, mouse, and cat) and diverse microbes in household dust are related to reduced risk of recurrent wheeze.⁸⁶

Finally, there is evidence that viruses and bacteria interact in patients with respiratory illnesses. Viral infections and illnesses are associated with transient detection of common bacterial pathogens (*Moraxella catarrhalis*, *Streptococcus pneumoniae*, and *Haemophlius influenzae*).^{87,88} Furthermore, airway microbiome communities dominated by these organisms were associated with increased risk of wheezing illnesses.⁸⁷ Finally, hospitalization for RSV-induced bronchiolitis is associated with airway microbiomes dominated by *H influenzae* or *Streptococcus* species, which were in turn associated with enhanced innate immune activation.⁸⁹

WHY IS VIRUS-INDUCED WHEEZING DURING INFANCY ASSOCIATED WITH CHILDHOOD ASTHMA?

There are several interacting factors that contribute to the strong linkage between virus-induced wheezing illnesses and the risk of childhood asthma (Fig 2). First, some host factors can predispose children to more severe RV or RSV infections and later to asthma. Second, viral wheezing illnesses might damage the airways to promote variable airway obstruction. Third, there might be interactions between risk factors and environmental exposures that promote asthma. Finally, antibiotic use, urbanization, and increased hygiene cause loss of microbial biodiversity (measures of richness and species distribution) and

TABLE II. Genes linked to both RSV-induced bronchiolitis and asthma

Cytokines	Receptors	Other
CCL5	CCR5	ADAM33
CXCL8	CX3CR1	MS4A2
IFNG	IL4RA	NOS2A
IL4	TLR4	TNX
IL6	TLR10	SPA1
IL10	VDR	SPA2
IL13		SFTPD
IL17		
IL18		
TGFB1		

eventually dysbiosis (imbalance in a microbial ecosystem) to promote allergic diseases.⁹⁰⁻⁹² In contrast, diverse microbial communities might help maintain normal airway physiology during viral infection and thereby moderate or prevent respiratory symptoms.⁹³

Mechanisms linking RV-induced wheezing to asthma development

The susceptibility to RV-induced bronchiolitis/early wheezing seems to be linked to predisposition because the prevalence of RV-induced bronchiolitis has been as high as 50% to 80% during the first year of life in infants with recurrent moderate-to-severe respiratory illnesses from atopic families.²⁹ Genetic variation at the 17q21 locus increases the risk for RV-induced wheezing in early childhood.⁹⁴ Although the mechanism for this effect is still unknown, it is notable that farm exposure also interacts with 17q21 polymorphisms to influence the risk for allergy and asthma.⁹⁵

In addition, low interferon responses secondary to young age, allergic sensitization, or both could increase susceptibility to viral infections and illnesses.^{96,97} Many studies have linked RV-induced wheezing in early life to other atopic biomarkers, allergen-specific sensitization, increased eosinophil counts in nasal mucus or blood, or the presence of atopic eczema, which have additive effects on asthma risk.^{5,7,9,96,98,99} Allergen exposure and high-affinity IgE receptor cross-linking have been shown to impair virus-induced type I and III interferon production in peripheral blood cells.¹⁰⁰ Also, type 2 inflammatory cytokines can inhibit antiviral responses in airway epithelial cells.¹⁰¹ The interaction between RV-induced wheezing and atopy is likely to become stronger by increasing age in children because the prevalence and intensity of respiratory allergy increases with age.^{7,39}

Interactions between allergic sensitization and RV-induced wheezing have been described. For example, both RV infections and allergens can enhance the airway epithelial cell production of IL-25 and IL-33, which promotes type 2 airway inflammation and remodeling.^{102,103} The IL-33 polymorphism is linked to intermediate and late-onset wheezing and allergic sensitization.¹⁰⁴ The first line of defense against RV infection is the airway epithelium, which is relatively resistant to infection when undamaged. Disrupted airway epithelium can favor RV replication by opening the way to deeper cell layers in which RV replicates most actively and by increasing the number of intercellular adhesion molecule 1 receptors as shown in

recent *in vitro* studies.¹⁰⁵ Damaged barrier function of the airway epithelium can also lead to enhanced absorption of aeroallergens or invasion of bacterial pathogens through the airway wall.¹⁰⁶ Finally, RVs can contribute to airway remodeling by inducing vascular endothelial growth factor, TGF- β , and chemoattractants for airway smooth muscle cells.^{107,108} These effects might be more pronounced for RV infections in early life.¹⁰⁹ Thus repeated RV infections that extend to the lower airways could damage the airways and lead to remodeling of airway structures.

RSV-induced wheezing and asthma

RSV-induced bronchiolitis is linked to an increased risk for asthma. The severity of the acute illness is related to the subsequent risk for asthma.¹¹⁰ Polymorphisms in a number of genes, mostly related to immune regulation, are associated with increased risk for both RSV-induced bronchiolitis and asthma (Table II).^{43,111} Other risk factors include young age and low lung function, but unlike RV-induced wheezing illnesses, atopic children are probably not at increased risk for RSV-induced bronchiolitis.⁶³

The risk of more severe RSV infections has been linked to the balance between type 1 and 2 immune responses. Infants with more severe versus mild RSV have shown reduced IFN- γ expression in PBMCs and airway cells, and IFN- γ expression has been correlated with subsequent asthma.¹¹²⁻¹¹⁴ TLR4 polymorphisms and LPS have been linked to induction of type 2 immune responses, and these factors also influence the severity of RSV-induced illness in children.¹¹⁴ Recently, it was shown in mice that aeroallergen-induced IL-33 predisposes to pneumovirus-induced asthma by dampening impaired IFN- α and IFN- λ production.¹¹⁵ Also, IFN- β has immune-modulating properties and can inhibit eosinophilic asthma–like pathophysiology in pneumovirus-infected mice.¹¹⁶

Overall, RSV-induced bronchiolitis typically affects neonates at the critical time window of lung development and therefore might have long-term influences. Murine models have shown that RSV infection causes more lung damage during the proliferative stage of lung growth compared with the equilibrated stage.⁴³ Neonatal regulatory B (nBreg) cells are highly permissive to RSV infection, and the frequency of nBreg cells has predicted the severity of acute bronchiolitis disease.79 Thus nBreg cell activity might have an important role in modulating an early-life host response to RSV. RSV infection increases the expression of nerve growth factor (NGF) and its receptors in the developing lungs.¹¹⁷ NGF controls the structural development of nervous system and its ability to respond to environmental changes. Neurotrophin levels have correlated with the intensity of allergic conditions, including asthma, and overexpression of NGF could lead to airway hyperreactivity during and after RSV infection.

Other viruses. Nearly all acute wheezing episodes are associated with viral infection.⁵ Although RV and RSV most commonly cause wheezing illnesses, other contributing viruses include enterovirus,^{7,118} bocavirus,¹¹⁹ parainfluenzavirus,^{6,8} coronaviruses,^{9,29,88} metapneumovirus,³² influenza virus,^{9,120} and adenoviruses.³² Polyomaviruses have also been detected in the lower airways but almost always with other viral pathogens, and it is uncertain whether they contribute to wheezing

illnesses.¹²¹ It should be noted that only RSV- or RV-induced wheezing illnesses are associated with increased risk for recurrent wheeze and asthma; infections of lesser severity with these viruses are ubiquitous. It remains to be determined whether less severe infections with these viruses, other viruses, or even certain bacteriophages could protect against the acquisition of asthma.

Prevention of asthma related to viral wheeze

Antiviral and anti-inflammatory treatments have been tested for long-term efficacy in prevention recurrent wheezing and asthma. Palivizumab, an mAb that reduces the rate of severe RSV infection, decreased recurrent wheezing in up to a 6-year follow-up in preterm infants but did not affect rates of atopic asthma.^{61,122} For infants with an initial RV-induced wheezing episode, a 3-day course of oral prednisolone decreased the time to initiation of asthma control therapy in the subgroup of children with high RV genome load.^{67,68} Interestingly, all children with high RV genome load who were treated with placebo required asthma control medication within just 14 months. These studies indicate that preventive strategies targeting either the virus or the inflammatory response could help prevent recurrent wheezing and perhaps some asthma phenotypes.

ASTHMA EXACERBATION

Several factors can contribute to asthma exacerbation, including infections, underuse of asthma control medications, or exposure to allergens or pollutants.¹²³ In most cases there is more than 1 contributing factor, and this is especially true for severe exacerbations. Viral infections are of special importance because they contribute to up to 90% of exacerbations, especially during the fall and spring in temperate climates, when viral respiratory tract infections are most common. Furthermore, viral infections play a major role in seasonal peaks of exacerbations that coincide with the return of children to schools after summer and spring breaks.¹²⁴

Among the plethora of respiratory viruses that can cause wheezing illnesses, RVs are most closely associated with exacerbations of childhood asthma.^{125,126} RVs can cause a spectrum of illnesses ranging from asymptomatic infections to severe lower respiratory tract illnesses. This is also true for children with asthma, and in fact, most RV infections in children with asthma do not cause exacerbations.¹²⁷⁻¹²⁹ There are a number of cofactors that are associated with a greater likelihood of more severe RV-induced illnesses, and the list of contributing factors is quite similar for infants (Fig 2) and children with asthma. It is the more severe RV-induced illnesses, that promote asthma exacerbations.¹³⁰

Viral factors. Just as RV-A and RV-C are associated with wheezing illnesses in early childhood, these viruses are also more often associated with exacerbations of asthma compared with RV-B. RV-C might be more strongly associated with more severe exacerbations, including those requiring hospitalization.¹³¹⁻¹³⁴ This could be due to faster replication rate and induction of more robust cellular responses, as demonstrated in cultures of differentiated airway epithelial cells.¹³⁵ In cohort studies RV-B

infections do not increase the risk for exacerbations,¹²⁹ but they might slightly increase the risk of exacerbation in children whose asthma is of greater severity.¹²⁷

Host factors. In children with established asthma, allergy and genetic factors can increase the risk of virus-induced wheezing illnesses and asthma exacerbations. Allergy is an important factor, and in fact, allergy and viral respiratory tract infections synergistically increase the risk for acute asthma exacerbations.33 The effects of allergy are dose related. For example, in a cohort of Costa Rican children, the quantity of dust mite IgE was positively associated with the risk for RV-induced asthma exacerbations.¹³⁶ In addition, in a Boston cohort of children with acute exacerbations of asthma, the severity of acute RV-induced exacerbation was positively related to IgE specific for either dust mite or mouse proteins.¹³⁷ Furthermore, in a study of children with asthma who were monitored for viral infections during the peak months (April and September) for RV infections, allergic sensitization was associated with a greater severity of RV-induced respiratory illnesses and RV-induced symptoms of asthma.¹²⁸

Recent studies with omalizumab, which prevents IgE binding to its receptor, have established that neutralizing allergic inflammation can enhance interferon responses and reduce RV-induced illnesses and asthma exacerbations. For example, moderate-to-severe asthma urban children with were randomized to treatment with either standard asthma controller therapy or standard therapy plus omalizumab. In 2 separate studies both year-round and preseasonal treatment with omalizumab eliminated the seasonal peaks in exacerbations, most of which are associated with viral infection.^{138,13} Analysis of weekly samples of nasal secretions confirmed that omalizumab reduced virus-induced exacerbations, and this effect was most pronounced in children with more severe asthma.¹³⁸ Furthermore, omalizumab increased IFN-α responses of blood cells stimulated with RV ex vivo. This finding suggested that omalizumab, by neutralizing IgE, indirectly improved antiviral responses. This theory is further supported by the findings that omalizumab reduced the frequency, quantity, and duration of RV detection in nasal secretions sampled weekly and also reduced the frequency of RV-induced colds by about one third.¹²⁷ In a study of children with acute asthma exacerbations, RV-induced exacerbations were less severe in those children who had been treated with omalizumab compared with other controllers.¹⁴⁰

Collectively, these findings suggest that allergic inflammation impairs antiviral responses, and in fact, there is considerable evidence that childhood asthma is associated with reduced interferon responses of blood cells and the airway epithelium (see review in this issue by Edwards et al¹⁴¹). In adults with allergic asthma, inhaled IFN- β was administered during colds in an attempt to prevent viral exacerbations. Although the primary end point was not achieved, IFN- β improved lung function in the study population and also improved asthma control in a subset of adults with more severe asthma.¹⁴²

Several genetic factors have been linked to the risk of asthma exacerbations.¹⁴³ Of these factors, a polymorphism (rs6967330) in CDHR3 is associated with acute severe exacerbations in young children, including asthma-related hospitalizations.¹⁴⁴ This polymorphism is in the coding



FIG 3. Opportunities for treatment or prevention of virus-induced wheezing illnesses. Potential interventions are shown in red. *OMZ*, Omalizumab.

region (C529Y) and leads to increased CDHR3 expression on the cell surface.¹⁴⁴ Interestingly, CDHR3 is the cellsurface receptor for RV-C, and the rs6967330 polymorphism leads to increased RV-C cell binding and replication.⁷³ These findings suggest that the linkage between rs6967330 and exacerbations of asthma is due to greater severity of RV-C infections.

Environmental factors

Several environmental factors can influence the severity of illness during RV infections and increase the probability of RV-induced exacerbation in children with asthma. Observational studies demonstrated that exposure to pollutants, such as NO₂, can increase the severity of virus-induced lower respiratory tract symptoms and reductions in lung function in children with asthma.¹⁴⁵ In addition, high exposure to allergens in children with allergic asthma increases the risk for virus-induced exacerbations.¹⁴⁶ Maternal stress and depression have been associated with acute wheezing illnesses (which are predominantly viral) in young children; the mechanism of this association is unknown but was not due to enhanced T_H2 responses or impaired antiviral responses as measured in PBMCs.¹⁴⁷ Vitamin D levels were inversely related to measures of asthma severity, including hospitalization for severe exacerbations.¹⁴⁸ However, in a randomized clinical trial vitamin D supplementation of adults with low vitamin D levels did not reduce rates of clinical colds or exacerbation rates.^{149,150}

Viral infections are generally recognized as important contributors to acute asthma exacerbations. However, prospective monitoring of nasal secretions during the peak season for RV infections and asthma exacerbations provides evidence of a close relationship between viral, bacterial, and respiratory symptoms. RV infections increase the frequency and quantity of *S pneumoniae*, *M catarrhalis*, and *H* influenzae detected in airway secretions.¹²⁹ In addition, viral infections associated with detection of these common bacterial pathogens are more likely to be symptomatic, and in children with asthma, they are more likely to be associated with asthma exacerbations.¹²⁹

Additional studies were conducted to identify differences in the microbiome composition of asymptomatic viral infections compared with those associated with asthma exacerbations.¹⁵¹ Compared with baseline samples before RV infection, samples during acute RV-induced exacerbations had increased detection of *Moraxella* species. In contrast, asymptomatic infections were associated with increases in *Corynebacterium* species compared with the baseline samples. These findings suggest that RV infections modify the upper airway microbiome and that quantitative and qualitative changes in the airway microbiome modify the probability that RV infection will lead to an exacerbation of asthma.

CONCLUSIONS AND THERAPEUTIC OPPORTUNITIES

Respiratory viruses interact with host and environmental factors to increase the risk of wheezing illnesses in infants and to increase the risk of exacerbations in asthmatic children. These findings suggest that there are a number of therapeutic opportunities to reduce the frequency and severity of viral respiratory illnesses and hopefully secondary effects on the incidence and exacerbation of asthma (Fig 3). The search goes on for effective antivirals and vaccines for RV and RSV, and several candidates are currently in clinical trials (reviewed by Edwards et al¹⁵²). Antiviral approaches also include strategies to enhance resistance to multiple respiratory viruses through administration of interferons or other immunostimulatory molecules. Furthermore, the search is on to identify biologic exposures (eg, microbes and proteins) that might help to promote the development of healthy mucosal immune responses that resist viral infection. Research into the airway microbiome has identified a number of intriguing associations with bacteria that might either protect against viral illnesses or add to the problem. These findings suggest possibilities for 2 new therapeutic approaches: (1) identify a new class of probiotics selected to promote resistance to viral illnesses and (2) develop strategies to inhibit pathogenic bacteria that synergize with viruses and add to illness severity. The latter approach could involve antibacterial vaccines or probiotics. Targeted antimicrobial therapy could also be considered, although potential benefits must be weighed against further selection of antimicrobial-resistant organisms and possible short- and long-term detrimental effects on commensal or beneficial airway or gut bacteria. Finally, treatments that can either prevent allergy or moderate its severity could secondarily boost antiviral responses and also reduce inflammatory responses that lead to airway obstruction and remodeling.

Collectively, these new approaches provide hope that new insights into personal risk factors (genetics, allergy, and antiviral immunity), environmental exposures (farm, urban, and microbes), and viral virulence can be harnessed to reduce the morbidity of viral respiratory illnesses and childhood asthma.

What do we know?

- Risk factors for RV-induced wheezing in early childhood include low lung function, genetic predisposition, and atopic characteristics, and the combination of RV-induced wheezing and atopy predicts a high likelihood of subsequent childhood asthma.
- Risk factors for RSV-induced bronchiolitis include genetics, low lung function, and young age but not atopy. RSV-induced bronchiolitis also predicts recurrent wheezing and childhood asthma.
- The airway inflammation that predisposes to early RV-induced wheezing appears to be responsive for corticosteroid medication.
- Prevention of RSV-induced bronchiolitis with the anti-RSV mAb palivizumab decreases risk of severe RSV-induced illness, recurrent wheezing, and perhaps nonatopic asthma.
- In children with asthma, infections with respiratory viruses, most commonly RV, contribute to most acute exacerbations.
- Virus-induced exacerbations are usually multifactorial, and other contributing factors include allergy, airway bacteria, genetics, medication adherence, and exposure to allergens and pollutants.

What is still unknown?

- Do RV infections cause asthma, unmask asthma, or both?
- Will prevention of bronchiolitis (either RV or RSV induced) and other viral wheezing illnesses in early life lead to preservation of lung function and reduced rates of persistent childhood asthma?
- Biomarkers are needed to serve as early clinical markers for susceptibility to viral illnesses and the progression to asthma; these biomarkers could be used to identify patients for secondary prevention trials of asthma.
- Which lifestyle factors and environmental exposures (microbial and others) lead to the development of robust antiviral immune and lung development to reduce the risk for virus-induced wheezing illnesses?
- During viral infections, what are the microbial mechanisms that modify airway inflammation and influence airway physiology?
- What is the role of the host antiviral immune response in causing airway obstruction and respiratory symptoms?

REFERENCES

- Taussig LM, Wright AL, Holberg CJ, Halonen M, Morgan WJ, Martinez FD. Tucson children's respiratory study: 1980 to present. J Allergy Clin Immunol 2003;111:661-75.
- 2. Meissner HC. Viral bronchiolitis in children. N Engl J Med 2016;374:1793-4.
- Lai CK, Beasley R, Crane J, Foliaki S, Shah J, Weiland S, et al. Global variation in the prevalence and severity of asthma symptoms: phase three of the International Study of Asthma and Allergies in Childhood (ISAAC). Thorax 2009;64:476-83.
- Jartti T, Soderlund-Venermo M, Hedman K, Ruuskanen O, Makela MJ. New molecular virus detection methods and their clinical value in lower respiratory tract infections in children. Paediatr Respir Rev 2013;14:38-45.
- Turunen R, Koistinen A, Vuorinen T, Arku B, Söderlund-Venermo M, Ruuskanen O, et al. The first wheezing episode: respiratory virus etiology, atopic characteristics, and illness severity. Pediatr Allergy Immunol 2014;25:796-803.

- Jackson DJ, Gangnon RE, Evans MD, Roberg KA, Anderson EL, Pappas TE, et al. Wheezing rhinovirus illnesses in early life predict asthma development in high-risk children. Am J Respir Crit Care Med 2008;178:667-72.
- Jartti T, Lehtinen P, Vuorinen T, Ruuskanen O. Bronchiolitis: age and previous wheezing episodes are linked to viral etiology and atopic characteristics. Pediatr Infect Dis J 2009;28:311-7.
- Kotaniemi-Syrjänen A, Vainionpää R, Reijonen TM, Waris M, Korhonen K, Korppi M. Rhinovirus-induced wheezing in infancy—the first sign of childhood asthma? J Allergy Clin Immunol 2003;111:66-71.
- Kusel MM, de Klerk NH, Kebadze T, Vohma V, Holt PG, Johnston SL, et al. Early-life respiratory viral infections, atopic sensitization, and risk of subsequent development of persistent asthma. J Allergy Clin Immunol 2007;119:1105-10.
- Sigurs N, Aljassim F, Kjellman B, Robinson PD, Sigurbergsson F, Bjarnason R, et al. Asthma and allergy patterns over 18 years after severe RSV bronchiolitis in the first year of life. Thorax 2010;65:1045-52.
- Ruotsalainen M, Hyvarinen MK, Piippo-Savolainen E, Korppi M. Adolescent asthma after rhinovirus and respiratory syncytial virus bronchiolitis. Pediatr Pulmonol 2013;48:633-9.
- Midulla F, Pierangeli A, Cangiano G, Bonci E, Salvadei S, Scagnolari C, et al. Rhinovirus bronchiolitis and recurrent wheezing: one year follow-up. Eur Respir J 2012;39:396-402.
- Lukkarinen M, Koistinen A, Turunen R, Lehtinen P, Vuorinen T, Jartti T. Rhinovirus-induced first wheezing episode predicts atopic but not nonatopic asthma at school age. J Allergy Clin Immunol 2017 [Epub ahead of print].
- Midulla F, Nicolai A, Ferrara M, Gentile F, Pierangeli A, Bonci E, et al. Recurrent wheezing 36 months after bronchiolitis is associated with rhinovirus infections and blood eosinophilia. Acta Paediatr 2014;103:1094-9.
- Khetsuriani N, Lu X, Teague WG, Kazerouni N, Anderson LJ, Erdman DD. Novel human rhinoviruses and exacerbation of asthma in children. Emerg Infect Dis 2008;14:1793-6.
- Korppi M, Koponen P, Nuolivirta K. Upper age limit for bronchiolitis: 12 months or 6 months? Eur Respir J 2012;39:787-8, author reply 788-9.
- National Asthma Education and Prevention Program. National Heart, Lung, and Blood Institute, National Institutes of Health. Expert panel report 3: guidelines for the diagnosis and management of asthma. Bethesda: US Department of Health and Human Services; 2007.
- McIntyre CL, Knowles NJ, Simmonds P. Proposals for the classification of human rhinovirus species A, B and C into genotypically assigned types. J Gen Virol 2013;94:1791-806.
- Basta HA, Ashraf S, Sgro JY, Bochkov YA, Gern JE, Palmenberg AC. Modeling of the human rhinovirus C capsid suggests possible causes for antiviral drug resistance. Virology 2014;448:82-90.
- Hao W, Bernard K, Patel N, Ulbrandt N, Feng H, Svabek C, et al. Infection and propagation of human rhinovirus C in human airway epithelial cells. J Virol 2012; 86:13524-32.
- 21. Hayden FG, Turner RB, Gwaltney JM, Chi-Burris K, Gersten M, Hsyu P, et al. Phase II, randomized, double-blind, placebo-controlled studies of ruprintrivir nasal spray 2-percent suspension for prevention and treatment of experimentally induced rhinovirus colds in healthy volunteers. Antimicrob Agents Chemother 2003;47:3907-16.
- 22. Glanville N, McLean GR, Guy B, Lecouturier V, Berry C, Girerd Y, et al. Cross-serotype immunity induced by immunization with a conserved rhinovirus capsid protein. PLoS Pathog 2013;9:e1003669.
- Lee S, Nguyen MT, Currier MG, Jenkins JB, Strobert EA, Kajon AE, et al. A polyvalent inactivated rhinovirus vaccine is broadly immunogenic in rhesus macaques. Nat Commun 2016;7:12838.
- Arden KE, McErlean P, Nissen MD, Sloots TP, Mackay IM. Frequent detection of human rhinoviruses, paramyxoviruses, coronaviruses, and bocavirus during acute respiratory tract infections. J Med Virol 2006;78:1232-40.
- Bochkov YA, Palmenberg AC, Lee WM, Rathe JA, Amineva SP, Sun X, et al. Molecular modeling, organ culture and reverse genetics for a newly identified human rhinovirus C. Nat Med 2011;17:627-32.
- Turner RB, Lee WM. Rhinovirus. In: Richman DD, Whitley RJ, Hayden FG, editors. Clinical virology. 4th ed. Washington (DC): ASM Press; 2017: 1143-64.
- Jartti T, Gern JE. Rhinovirus-associated wheeze during infancy and asthma development. Curr Respir Med Rev 2011;7:160-6.
- Heinonen S, Jartti T, Garcia C, Oliva S, Smitherman C, Anguiano E, et al. Rhinovirus detection in symptomatic and asymptomatic children: value of host transcriptome analysis. Am J Respir Crit Care Med 2016;193:772-82.
- Jartti T, Lee WM, Pappas T, Evans M, Lemanske RF Jr, Gern JE. Serial viral infections in infants with recurrent respiratory illnesses. Eur Respir J 2008;32:314-20.
- **30.** Toivonen L, Schuez-Havupalo L, Karppinen S, Teros-Jaakkola T, Rulli M, Mertsola J, et al. Rhinovirus infections in the first 2 years of life. Pediatrics 2016;138:e20161309.

- Kusel MM, de Klerk NH, Holt PG, Kebadze T, Johnston SL, Sly PD. Role of respiratory viruses in acute upper and lower respiratory tract illness in the first year of life: a birth cohort study. Pediatr Infect Dis J 2006;25:680-6.
- 32. Jartti T, Lehtinen P, Vuorinen T, Österback R, van den Hoogen B, Osterhaus AD, et al. Respiratory picornaviruses and respiratory syncytial virus as causative agents of acute expiratory wheezing in children. Emerg Infect Dis 2004;10:1095-101.
- 33. Rakes GP, Arruda E, Ingram JM, Hoover GE, Zambrano JC, Hayden FG, et al. Rhinovirus and respiratory syncytial virus in wheezing children requiring emergency care. IgE and eosinophil analyses. Am J Respir Crit Care Med 1999;159:785-90.
- Marguet C, Lubrano M, Gueudin M, Le Roux P, Deschildre A, Forget C, et al. In very young infants severity of acute bronchiolitis depends on carried viruses. PLoS One 2009;4:e4596.
- Midulla F, Scagnolari C, Bonci E, Pierangeli A, Antonelli G, De Angelis D, et al. Respiratory syncytial virus, human bocavirus and rhinovirus bronchiolitis in infants. Arch Dis Child 2010;95:35-41.
- Rossi GA, Colin AA. Infantile respiratory syncytial virus and human rhinovirus infections: respective role in inception and persistence of wheezing. Eur Respir J 2015;45:774-89.
- 37. Lee WM, Lemanske RF Jr, Evans MD, Vang F, Pappas T, Gangnon R, et al. Human rhinovirus species and season of infection determine illness severity. Am J Respir Crit Care Med 2012;186:886-91.
- Bergroth E, Aakula M, Korppi M, Remes S, Kivisto JE, Piedra PA, et al. Post-bronchiolitis use of asthma medication: a prospective 1-year follow-up study. Pediatr Infect Dis J 2016;35:363-8.
- 39. Rubner FJ, Jackson DJ, Evans MD, Gangnon RE, Tisler CJ, Pappas TE, et al. Early life rhinovirus wheezing, allergic sensitization, and asthma risk at adolescence. J Allergy Clin Immunol 2017;139:501-7.
- Lemanske RF Jr, Jackson DJ, Gangnon RE, Evans MD, Li Z, Shult PA, et al. Rhinovirus illnesses during infancy predict subsequent childhood wheezing. J Allergy Clin Immunol 2005;116:571-7.
- Koponen P, Helminen M, Paassilta M, Luukkaala T, Korppi M. Preschool asthma after bronchiolitis in infancy. Eur Respir J 2012;39:76-80.
- 42. Liu L, Pan Y, Zhu Y, Song Y, Su X, Yang L, et al. Association between rhinovirus wheezing illness and the development of childhood asthma: a meta-analysis. BMJ Open 2017;7:e013034.
- Wu P, Hartert TV. Evidence for a causal relationship between respiratory syncytial virus infection and asthma. Expert Rev Anti Infect Ther 2011;9:731-45.
- Hotard AL, Laikhter E, Brooks K, Hartert TV, Moore ML. Functional analysis of the 60-nucleotide duplication in the respiratory syncytial virus buenos aires strain attachment glycoprotein. J Virol 2015;89:8258-66.
- Mazur NI, Martinon-Torres F, Baraldi E, Fauroux B, Greenough A, Heikkinen T, et al. Lower respiratory tract infection caused by respiratory syncytial virus: current management and new therapeutics. Lancet Respir Med 2015;3:888-900.
- Graham BS. Vaccine development for respiratory syncytial virus. Curr Opin Virol 2017;23:107-12.
- Griffiths C, Drews SJ, Marchant DJ. Respiratory syncytial virus: infection, detection, and new options for prevention and treatment. Clin Microbiol Rev 2017;30:277-319.
- Ralston SL, Lieberthal AS, Meissner HC, Alverson BK, Baley JE, Gadomski AM, et al. Clinical practice guideline: the diagnosis, management, and prevention of bronchiolitis. Pediatrics 2014;134:e1474-502.
- American Academy of Pediatrics Subcommittee on Diagnosis and management of bronchiolitis. Pediatrics 2006;118:1774-93.
- Scottish Intercollegiate Guidelines Network. Bronchiolitis in children. A national clinical guideline.
- Williams JV, Piedra PA, Englund JA. Respiratory syncytial virus, human metapneumovirus and parainfluenza viruses. In: Richman DD, Whitley RJ, Hayden FG, editors. Clinical virology. 4th ed. Washington (DC): ASM Press; 2017: 873-902.
- 52. Stein RT, Sherrill D, Morgan WJ, Holberg CJ, Halonen M, Taussig LM, et al. Respiratory syncytial virus in early life and risk of wheeze and allergy by age 13 years. Lancet 1999;354:541-5.
- 53. Henderson J, Hilliard TN, Sherriff A, Stalker D, Al Shammari N, Thomas HM. Hospitalization for RSV bronchiolitis before 12 months of age and subsequent asthma, atopy and wheeze: a longitudinal birth cohort study. Pediatr Allergy Immunol 2005;16:386-92.
- 54. Sigurs N, Bjarnason R, Sigurbergsson F, Kjellman B, Bjorksten B. Asthma and immunoglobulin E antibodies after respiratory syncytial virus bronchiolitis: a prospective cohort study with matched controls. Pediatrics 1995;95:500-5.
- 55. Sigurs N, Bjarnason R, Sigurbergsson F, Kjellman B. Respiratory syncytial virus bronchiolitis in infancy is an important risk factor for asthma and allergy at age 7. Am J Respir Crit Care Med 2000;161:1501-7.
- 56. Sigurs N, Gustafsson PM, Bjarnason R, Lundberg F, Schmidt S, Sigurbergsson F, et al. Severe respiratory syncytial virus bronchiolitis in infancy and asthma and allergy at age 13. Am J Respir Crit Care Med 2005;171:137-41.

- 57. Wu P, Dupont WD, Griffin MR, Carroll KN, Mitchel EF, Gebretsadik T, et al. Evidence of a causal role of winter virus infection during infancy in early childhood asthma. Am J Respir Crit Care Med 2008;178:1123-9.
- 58. Stensballe LG, Simonsen JB, Thomsen SF, Larsen AM, Lysdal SH, Aaby P, et al. The causal direction in the association between respiratory syncytial virus hospitalization and asthma. J Allergy Clin Immunol 2009;123:131-7.e1.
- 59. Thomsen SF, van der Sluis S, Stensballe LG, Posthuma D, Skytthe A, Kyvik KO, et al. Exploring the association between severe respiratory syncytial virus infection and asthma: a registry-based twin study. Am J Respir Crit Care Med 2009;179:1091-7.
- 60. Carroll KN, Gebretsadik T, Escobar GJ, Wu P, Li SX, Walsh EM, et al. Respiratory syncytial virus immunoprophylaxis in high-risk infants and development of childhood asthma. J Allergy Clin Immunol 2017;139:66-71.e3.
- **61.** Mochizuki H, Kusuda S, Okada K, Yoshihara S, Furuya H, Simoes EA. Palivizumab prophylaxis in preterm infants and subsequent recurrent wheezing: 6 year follow up study. Am J Respir Crit Care Med 2017;196:29-38.
- **62.** Vandini S, Calamelli E, Faldella G, Lanari M. Immune and inflammatory response in bronchiolitis due to respiratory syncytial virus and rhinovirus infections in infants. Paediatr Respir Rev 2017 [Epub ahead of print].
- 63. Jartti T, Kuusipalo H, Vuorinen T, Söderlund-Venermo M, Allander T, Waris M, et al. Allergic sensitization is associated with rhinovirus-, but not other virus-, induced wheezing in children. Pediatr Allergy Immunol 2010;21:1008-14.
- 64. Mansbach JM, Clark S, Teach SJ, Gern JE, Piedra PA, Sullivan AF, et al. Children hospitalized with rhinovirus bronchiolitis have asthma-like characteristics. J Pediatr 2016;172:202-4.e1.
- 65. Korppi M, Kotaniemi-Syrjänen A, Waris M, Vainionpää R, Reijonen TM. Rhinovirus-associated wheezing in infancy: comparison with respiratory syncytial virus bronchiolitis. Pediatr Infect Dis J 2004;23:995-9.
- 66. Turunen R, Vuorinen T, Bochkov Y, Gern J, Jartti T. Clinical and virus surveillance after the first wheezing episode: special reference to rhinovirus A and C species. Pediatr Infect Dis J 2017;36:539-44.
- Jartti T, Nieminen R, Vuorinen T, Lehtinen P, Vahlberg T, Gern J, et al. Short- and long-term efficacy of prednisolone for first acute rhinovirus-induced wheezing episode. J Allergy Clin Immunol 2015;135:691-8.e9.
- 68. Koistinen A, Lukkarinen M, Turunen R, Vuorinen T, Vahlberg T, Camargo CA Jr, et al. Prednisolone for the first rhinovirus-induced wheezing and 4-year asthma risk: a randomized trial. Pediatr Allergy Immunol 2017 [Epub ahead of print].
- **69.** Mansbach JM, McAdam AJ, Clark S, Hain PD, Flood RG, Acholonu U, et al. Prospective multicenter study of the viral etiology of bronchiolitis in the emergency department. Acad Emerg Med 2008;15:111-8.
- Dumas O, Mansbach JM, Jartti T, Hasegawa K, Sullivan AF, Piedra PA, et al. A clustering approach to identify severe bronchiolitis profiles in children. Thorax 2016;71:712-8.
- Mosser AG, Vrtis R, Burchell L, Lee WM, Dick CR, Weisshaar E, et al. Quantitative and qualitative analysis of rhinovirus infection in bronchial tissues. Am J Respir Crit Care Med 2005;171:645-51.
- Johnson SM, McNally BA, Ioannidis I, Flano E, Teng MN, Oomens AG, et al. Respiratory syncytial virus uses CX3CR1 as a receptor on primary human airway epithelial cultures. PLoS Pathog 2015;11:e1005318.
- 73. Bochkov YA, Watters K, Ashraf S, Griggs TF, Devries MK, Jackson DJ, et al. Cadherinrelated family member 3, a childhood asthma susceptibility gene product, mediates rhinovirus C binding and replication. Proc Natl Acad Sci U S A 2015;112:5485-90.
- 74. Han M, Chung Y, Young Hong J, Rajput C, Lei J, Hinde JL, et al. Toll-like receptor 2-expressing macrophages are required and sufficient for rhinovirusinduced airway inflammation. J Allergy Clin Immunol 2016;138:1619-30.
- Slater L, Bartlett NW, Haas JJ, Zhu J, Message SD, Walton RP, et al. Co-ordinated role of TLR3, RIG-I and MDA5 in the innate response to rhinovirus in bronchial epithelium. PLoS Pathog 2010;6:e1001178.
- Bosco A, Wiehler S, Proud D. Interferon regulatory factor 7 regulates airway epithelial cell responses to human rhinovirus infection. BMC Genomics 2016;17:76.
- Mejias A, Dimo B, Suarez NM, Garcia C, Suarez-Arrabal MC, Jartti T, et al. Whole blood gene expression profiles to assess pathogenesis and disease severity in infants with respiratory syncytial virus infection. PLoS Med 2013;10:e1001549.
- Mejias A, Suarez NM, Ramilo O. Detecting specific infections in children through host responses: a paradigm shift. Curr Opin Infect Dis 2014;27:228-35.
- Zhivaki D, Lemoine S, Lim A, Morva A, Vidalain PO, Schandene L, et al. Respiratory syncytial virus infects regulatory B cells in human neonates via chemokine receptor CX3CR1 and promotes lung disease severity. Immunity 2017;46:301-14.
- Aab A, Wirz O, van de Veen W, Sollner S, Stanic B, Ruckert B, et al. Human rhinoviruses enter and induce proliferation of B lymphocytes. Allergy 2017;72:232-43.
- Tregoning JS, Schwarze J. Respiratory viral infections in infants: causes, clinical symptoms, virology, and immunology. Clin Microbiol Rev 2010;23:74-98.
- 82. Guo-Parke H, Canning P, Douglas I, Villenave R, Heaney LG, Coyle PV, et al. Relative respiratory syncytial virus cytopathogenesis in upper and lower respiratory tract epithelium. Am J Respir Crit Care Med 2013;188:842-51.

- 83. Fuchs O, Genuneit J, Latzin P, Buchele G, Horak E, Loss G, et al. Farming environments and childhood atopy, wheeze, lung function, and exhaled nitric oxide. J Allergy Clin Immunol 2012;130:382-8.e6.
- 84. Ludka-Gaulke T, Ghera P, Waring SC, Keifer M, Seroogy C, Gern JE, et al. Farm exposure in early childhood is associated with a lower risk of severe respiratory illnesses. J Allergy Clin Immunol 2017 [Epub ahead of print].
- 85. Fujimura KE, Demoor T, Rauch M, Faruqi AA, Jang S, Johnson CC, et al. House dust exposure mediates gut microbiome *Lactobacillus* enrichment and airway immune defense against allergens and virus infection. Proc Natl Acad Sci U S A 2014;111:805-10.
- 86. Lynch SV, Wood RA, Boushey H, Bacharier LB, Bloomberg GR, Kattan M, et al. Effects of early-life exposure to allergens and bacteria on recurrent wheeze and atopy in urban children. J Allergy Clin Immunol 2014;134:593-601.e12.
- 87. Teo SM, Mok D, Pham K, Kusel M, Serralha M, Troy N, et al. The infant nasopharyngeal microbiome impacts severity of lower respiratory infection and risk of asthma development. Cell Host Microbe 2015;17:704-15.
- Bisgaard H, Hermansen MN, Bonnelykke K, Stokholm J, Baty F, Skytt NL, et al. Association of bacteria and viruses with wheezy episodes in young children: prospective birth cohort study. BMJ 2010;341:c4978.
- 89. de Steenhuijsen Piters WA, Heinonen S, Hasrat R, Bunsow E, Smith B, Suarez-Arrabal MC, et al. Nasopharyngeal microbiota, host transcriptome, and disease severity in children with respiratory syncytial virus infection. Am J Respir Crit Care Med 2016;194:1104-15.
- 90. Haahtela T, Laatikainen T, Alenius H, Auvinen P, Fyhrquist N, Hanski I, et al. Hunt for the origin of allergy—comparing the Finnish and Russian Karelia. Clin Exp Allergy 2015;45:891-901.
- 91. Huang YJ, Marsland BJ, Bunyavanich S, O'Mahony L, Leung DY, Muraro A, et al. The microbiome in allergic disease: current understanding and future opportunities—2017 PRACTALL document of the American Academy of Allergy, Asthma & Immunology and the European Academy of Allergy and Clinical Immunology. J Allergy Clin Immunol 2017;139:1099-110.
- Blaser MJ. Antibiotic use and its consequences for the normal microbiome. Science 2016;352:544-5.
- 93. Hasegawa K, Mansbach JM, Ajami NJ, Espinola JA, Henke DM, Petrosino JF, et al. Association of nasopharyngeal microbiota profiles with bronchiolitis severity in infants hospitalised for bronchiolitis. Eur Respir J 2016;48: 1329-39.
- 94. Caliskan M, Bochkov YA, Kreiner-Moller E, Bonnelykke K, Stein MM, Du G, et al. Rhinovirus wheezing illness and genetic risk of childhood-onset asthma. N Engl J Med 2013;368:1398-407.
- 95. Loss GJ, Depner M, Hose AJ, Genuneit J, Karvonen AM, Hyvarinen A, et al. The early development of wheeze. environmental determinants and genetic susceptibility at 17q21. Am J Respir Crit Care Med 2016;193:889-97.
- 96. Jackson DJ, Evans MD, Gangnon RE, Tisler CJ, Pappas TE, Lee WM, et al. Evidence for a causal relationship between allergic sensitization and rhinovirus wheezing in early life. Am J Respir Crit Care Med 2012;185:281-5.
- Gern JE, Brooks GD, Meyer P, Chang A, Shen K, Evans MD, et al. Bidirectional interactions between viral respiratory illnesses and cytokine responses in the first year of life. J Allergy Clin Immunol 2006;117:72-8.
- 98. Singh AM, Evans MD, Gangnon R, Roberg KA, Tisler C, DaSilva D, et al. Expression patterns of atopic eczema and respiratory illnesses in a high-risk birth cohort. J Allergy Clin Immunol 2010;125:491-3.
- **99.** Nicolai A, Frassanito A, Nenna R, Cangiano G, Petrarca L, Papoff P, et al. Risk factors for virus-induced acute respiratory tract infections in children younger than 3 years and recurrent wheezing at 36 months follow-up after discharge. Pediatr Infect Dis J 2017;36:179-83.
- 100. Durrani SR, Montville DJ, Pratt AS, Sahu S, DeVries MK, Rajamanickam V, et al. Innate immune responses to rhinovirus are reduced by the high-affinity IgE receptor in allergic asthmatic children. J Allergy Clin Immunol 2012;130:489-95.
- 101. Contoli M, Ito K, Padovani A, Poletti D, Marku B, Edwards MR, et al. Th2 cytokines impair innate immune responses to rhinovirus in respiratory epithelial cells. Allergy 2015;70:910-20.
- 102. Saglani S. Viral infections and the development of asthma in children. Ther Adv Infect Dis 2013;1:139-50.
- 103. Beale J, Jayaraman A, Jackson DJ, Macintyre JD, Edwards MR, Walton RP, et al. Rhinovirus-induced IL-25 in asthma exacerbation drives type 2 immunity and allergic pulmonary inflammation. Sci Transl Med 2014;6:256ra134.
- 104. Savenije OE, Mahachie John JM, Granell R, Kerkhof M, Dijk FN, de Jongste JC, et al. Association of IL33-IL-1 receptor-like 1 (IL1RL1) pathway polymorphisms with wheezing phenotypes and asthma in childhood. J Allergy Clin Immunol 2014;134:170-7.
- 105. Jakiela B, Brockman-Schneider R, Amineva S, Lee WM, Gern JE. Basal cells of differentiated bronchial epithelium are more susceptible to rhinovirus infection. Am J Respir Cell Mol Biol 2008;38:517-23.

- 106. Sajjan U, Wang Q, Zhao Y, Gruenert DC, Hershenson MB. Rhinovirus disrupts the barrier function of polarized airway epithelial cells. Am J Respir Crit Care Med 2008;178:1271-81.
- 107. Shariff S, Shelfoon C, Holden NS, Traves SL, Wiehler S, Kooi C, et al. Human rhinovirus infection of epithelial cells modulates airway smooth muscle migration. Am J Respir Cell Mol Biol 2017;56:796-803.
- 108. Leigh R, Oyelusi W, Wiehler S, Koetzler R, Zaheer RS, Newton R, et al. Human rhinovirus infection enhances airway epithelial cell production of growth factors involved in airway remodeling. J Allergy Clin Immunol 2008;121:1238-45.
- 109. Hong JY, Bentley JK, Chung Y, Lei J, Steenrod JM, Chen Q, et al. Neonatal rhinovirus induces mucous metaplasia and airways hyperresponsiveness through IL-25 and type 2 innate lymphoid cells. J Allergy Clin Immunol 2014;134:429-39.
- 110. Carroll KN, Wu P, Gebretsadik T, Griffin MR, Dupont WD, Mitchel EF, et al. The severity-dependent relationship of infant bronchiolitis on the risk and morbidity of early childhood asthma. J Allergy Clin Immunol 2009;123:1055-61.e1.
- 111. Singh AM, Moore PE, Gern JE, Lemanske RF Jr, Hartert TV. Bronchiolitis to asthma: a review and call for studies of gene-virus interactions in asthma causation. Am J Respir Crit Care Med 2007;175:108-19.
- 112. Aberle JH, Aberle SW, Dworzak MN, Mandl CW, Rebhandl W, Vollnhofer G, et al. Reduced interferon-gamma expression in peripheral blood mononuclear cells of infants with severe respiratory syncytial virus disease. Am J Respir Crit Care Med 1999;160:1263-8.
- 113. Renzi PM, Turgeon JP, Marcotte JE, Drblik SP, Berube D, Gagnon MF, et al. Reduced interferon-gamma production in infants with bronchiolitis and asthma. Am J Respir Crit Care Med 1999;159:1417-22.
- 114. Caballero MT, Serra ME, Acosta PL, Marzec J, Gibbons L, Salim M, et al. TLR4 genotype and environmental LPS mediate RSV bronchiolitis through Th2 polarization. J Clin Invest 2015;125:571-82.
- 115. Lynch JP, Werder RB, Simpson J, Loh Z, Zhang V, Haque A, et al. Aeroallergen-induced IL-33 predisposes to respiratory virus-induced asthma by dampening antiviral immunity. J Allergy Clin Immunol 2016;138:1326-37.
- 116. Simpson J, Lynch JP, Loh Z, Zhang V, Werder RB, Spann K, et al. The absence of interferon-beta promotor stimulator-1 (IPS-1) predisposes to bronchiolitis and asthma-like pathology in response to pneumoviral infection in mice. Sci Rep 2017;7:2353.
- 117. Hu C, Wedde-Beer K, Auais A, Rodriguez MM, Piedimonte G. Nerve growth factor and nerve growth factor receptors in respiratory syncytial virus-infected lungs. Am J Physiol Lung Cell Mol Physiol 2002;283:L494-502.
- 118. Andréoletti L, Lesay M, Deschildre A, Lambert V, Dewilde A, Wattré P. Differential detection of rhinoviruses and enteroviruses RNA sequences associated with classical immunofluorescence assay detection of respiratory virus antigens in nasopharyngeal swabs from infants with bronchiolitis. J Med Virol 2000;61:341-6.
- 119. Soderlund-Venermo M, Lahtinen A, Jartti T, Hedman L, Kemppainen K, Lehtinen P, et al. Clinical assessment and improved diagnosis of bocavirus-induced wheezing in children. Finland. Emerg Infect Dis 2009;15:1423-30.
- 120. Miller EK, Gebretsadik T, Carroll KN, Dupont WD, Mohamed YA, Morin LL, et al. Viral etiologies of infant bronchiolitis, croup and upper respiratory illness during 4 consecutive years. Pediatr Infect Dis J 2013;32:950-5.
- 121. Ren L, Gonzalez R, Xie Z, Zhang J, Liu C, Li J, et al. WU and KI polyomavirus present in the respiratory tract of children, but not in immunocompetent adults. J Clin Virol 2008;43:330-3.
- 122. Blanken MO, Rovers MM, Molenaar JM, Winkler-Seinstra PL, Meijer A, Kimpen JL, et al. Respiratory syncytial virus and recurrent wheeze in healthy preterm infants. N Engl J Med 2013;368:1791-9.
- 123. National Asthma Education and Prevention Program. Expert panel report 3 (EPR-3): guidelines for the diagnosis and management of asthma—summary report 2007. J Allergy Clin Immunol 2007;120(suppl):S94-138.
- 124. Johnston NW, Johnston SL, Norman GR, Dai J, Sears MR. The September epidemic of asthma hospitalization: school children as disease vectors. J Allergy Clin Immunol 2006;117:557-62.
- 125. Heymann PW, Carper HT, Murphy DD, Platts-Mills TA, Patrie J, McLaughlin AP, et al. Viral infections in relation to age, atopy, and season of admission among children hospitalized for wheezing. J Allergy Clin Immunol 2004;114:239-47.
- 126. Johnston SL, Pattemore PK, Sanderson G, Smith S, Lampe F, Josephs L, et al. Community study of role of viral infections in exacerbations of asthma in 9-11 year old children. BMJ 1995;310:1225-9.
- 127. Esquivel A, Busse WW, Calatroni A, Togias AG, Grindle KA, Bochkov YA, et al. Effects of omalizumab on rhinovirus infections, illnesses and exacerbations of asthma. Am J Respir Crit Care Med 2017 [Epub ahead of print].
- 128. Olenec JP, Kim WK, Lee WM, Vang F, Pappas TE, Salazar LE, et al. Weekly monitoring of children with asthma for infections and illness during common cold seasons. J Allergy Clin Immunol 2010;125:1001-6.
- 129. Kloepfer KM, Lee WM, Pappas TE, Kang TJ, Vrtis RF, Evans MD, et al. Detection of pathogenic bacteria during rhinovirus infection is associated with

increased respiratory symptoms and asthma exacerbations. J Allergy Clin Immunol 2014;133:1301-7.e3.

- 130. Zhu J, Message SD, Qiu Y, Mallia P, Kebadze T, Contoli M, et al. Airway inflammation and illness severity in response to experimental rhinovirus infection in asthma. Chest 2014;145:1219-29.
- 131. Zheng SY, Wang LL, Ren L, Luo J, Liao W, Liu EM. Epidemiological analysis and follow-up of human rhinovirus infection in children with asthma exacerbation. J Med Virol 2017 [Epub ahead of print].
- 132. Bizzintino J, Lee WM, Laing IA, Vang F, Pappas T, Zhang G, et al. Association between human rhinovirus C and severity of acute asthma in children. Eur Respir J 2011;37:1037-42.
- 133. Annamalay AA, Jroundi I, Bizzintino J, Khoo SK, Zhang G, Lehmann D, et al. Rhinovirus C is associated with wheezing and rhinovirus A is associated with pneumonia in hospitalized children in Morocco. J Med Virol 2017;89:582-8.
- 134. Cox DW, Bizzintino J, Ferrari G, Khoo SK, Zhang G, Whelan S, et al. Human rhinovirus species C infection in young children with acute wheeze is associated with increased acute respiratory hospital admissions. Am J Respir Crit Care Med 2013;188:1358-64.
- 135. Nakagome K, Bochkov YA, Ashraf S, Brockman-Schneider RA, Evans MD, Pasic TR, et al. Effects of rhinovirus species on viral replication and cytokine production. J Allergy Clin Immunol 2014;134:332-41.
- 136. Soto-Quiros M, Avila L, Platts-Mills TA, Hunt JF, Erdman DD, Carper H, et al. High titers of IgE antibody to dust mite allergen and risk for wheezing among asthmatic children infected with rhinovirus. J Allergy Clin Immunol 2012;129:1499-505.e5.
- 137. Kantor DB, Stenquist N, McDonald MC, Schultz BJ, Hauptman M, Smallwood CD, et al. Rhinovirus and serum IgE are associated with acute asthma exacerbation severity in children. J Allergy Clin Immunol 2016;138:1467-71.e9.
- 138. Teach SJ, Gill MA, Togias A, Sorkness CA, Arbes SJ Jr, Calatroni A, et al. Pre-seasonal treatment with either omalizumab an inhaled corticosteroid boost to prevent fall asthma exacerbations. J Allergy Clin Immunol 2015;136:1476-85.
- 139. Busse WW, Morgan WJ, Gergen PJ, Mitchell HE, Gern JE, Liu AH, et al. Randomized trial of omalizumab (anti-IgE) for asthma in inner-city children. N Engl J Med 2011;364:1005-15.
- 140. Kantor DB, McDonald MC, Stenquist N, Schultz BJ, Smallwood CD, Nelson KA, et al. Omalizumab is associated with reduced acute severity of rhinovirustriggered asthma exacerbation. Am J Respir Crit Care Med 2016;194:1552-5.
- 141. Edwards MR, Strong K, Cameron A, Walton RP, Jackson DJ, Johnston SL. Viral infections in allergy and immunology: How allergic inflammation influences viral infections and illness. J Allergy Clin Immunol 2017;140:909-20.
- 142. Djukanovic R, Harrison T, Johnston SL, Gabbay F, Wark P, Thomson NC, et al. The effect of inhaled IFN-beta on worsening of asthma symptoms caused by viral infections. A randomized trial. Am J Respir Crit Care Med 2014;190:145-54.
- 143. Park HW, Tantisira KG. Genetic signatures of asthma exacerbation. Allergy Asthma Immunol Res 2017;9:191-9.
- 144. Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Moller E, Mercader JM, Belgrave D, et al. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. Nat Genet 2014;46:51-5.
- 145. Chauhan AJ, Inskip HM, Linaker CH, Smith S, Schreiber J, Johnston SL, et al. Personal exposure to nitrogen dioxide (NO2) and the severity of virus-induced asthma in children. Lancet 2003;361:1939-44.
- 146. Murray CS, Poletti G, Kebadze T, Morris J, Woodcock A, Johnston SL, et al. Study of modifiable risk factors for asthma exacerbations: virus infection and allergen exposure increase the risk of asthma hospital admissions in children. Thorax 2006;61:376-82.
- 147. Ramratnam SK, Visness CM, Jaffee KF, Bloomberg GR, Kattan M, Sandel MT, et al. Relationships among maternal stress and depression, type 2 responses, and recurrent wheezing at age 3 years in low-income urban families. Am J Respir Crit Care Med 2017;195:674-81.
- 148. Brehm JM, Celedon JC, Soto-Quiros ME, Avila L, Hunninghake GM, Forno E, et al. Serum vitamin D levels and markers of severity of childhood asthma in Costa Rica. Am J Respir Crit Care Med 2009;179:765-71.
- 149. Castro M, King TS, Kunselman SJ, Cabana MD, Denlinger L, Holguin F, et al. Effect of vitamin D3 on asthma treatment failures in adults with symptomatic asthma and lower vitamin D levels: the VIDA randomized clinical trial. JAMA 2014;311:2083-91.
- 150. Denlinger LC, King TS, Cardet JC, Craig T, Holguin F, Jackson DJ, et al. Vitamin D supplementation and the risk of colds in patients with asthma. Am J Respir Crit Care Med 2016;193:634-41.
- 151. Kloepfer KM, Sarsani VK, Poroyko V, Lee WM, Pappas TE, Kang T, et al. Community acquired rhinovirus infection is associated with changes in the airway microbiome. J Allergy Clin Immunol 2017;140:312-5.e8.
- 152. Edwards MR, Walton RP, Jackson DJ, Feleszko W, Skevaki C, Jartti T, et al. The potential of anti-infectives and immunomodulators as therapies for asthma and asthma exacerbations. Allergy 2017 [Epub ahead of print].