

Tehran University of Medical Sciences Publication http://tums.ac.ir

Iran J Parasitol

Open access Journal at http://ijpa.tums.ac.ir



Iranian Society of Parasitology http://isp.tums.ac.ir

Original Article

Occurrence of *Acanthamoeba* Genotypes in Wastewater Samples in Tehran, Iran

Massoud BEHNIA ¹, Karim HATAM-NAHAVANDI ², Elham HAJIALILO ^{1,3}, Maryam NIYYATI ⁴, Fatemeh TARIGHI ¹, Azam BAKHTIAR AKRAM ¹, Mahboobeh SALIMI¹, *Mostafa REZAEIAN ^{1,5}

- 1. Dept. of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran
 - 2. Infection Disease and Tropical Medicine Research Center, Zahedan University of Medical Sciences, Zahedan, Iran
- 3. Dept. of Medical Parasitology and Mycology, School of Medicine, Qazvin University of Medical Sciences, Qazvin, Iran
- 4. Dept. of Medical Parasitology and Mycology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
 - 5. Center for Research of Endemic Parasites of Iran (CREPI), Tehran University of Medical Sciences, Tehran, Iran

Received 11 Dec 2016 Accepted 19 Apr 2017

Keywords:

Acanthamoeba, Genotype, Wastewater, Iran

*Correspondence Email:

rezaiian@sina.tums.ac.ir

Abstract

Background: Acanthamoeba spp. is potentially pathogenic free-living amoeba that can exist in various water sources. The presence of this amoeba in water sources could be a health hazard as Acanthamoeba could lead to severe diseases such as Acanthamoeba keratitis and encephalitis. This study aimed to determine the genotypes of isolated Acanthamoeba spp. in raw wastewater samples in Tehran, Iran.

Methods: Overall, 90 raw wastewater samples were collected from water treatment facilities in west and south of Tehran, Iran during 2014-2016. Water samples were filtered and cultured on non-nutrient agar (NNA) medium enriched with *Escherichia coli*. Morphological and molecular analyses were done on positive strains. The pathogenic ability of the isolated strains was determined using physical assays.

Results: Totally, 6 out of 90 (6.7%) samples were positive for *Acanthamoeba*, according to morphological characteristics of double-walled cysts. Genotyping and sequencing of the positive strains showed *Acanthamoeba* belonging to T4 (83%) and T11 (17%) genotypes. In vitro pathogenicity tests were revealed that five isolates were classified as non-pathogenic strains and one strain belonging to T4 genotype was classified as the highly pathogenic amoebae.

Conclusion: The current research reflected a low contamination of wastewater sources to *Acanthamoeba*. More studies regarding the contamination of wastewaters before and after treatment are required in different places of the country.

Introduction

canthamoeba spp. are one of the most prevalent protozoa with distribution, worldwide amphizoics amoebae were isolated from various environmental sources such as different kinds of waters including seawater, tap water, aquarium, fresh and saltwater lakes, hot spring resorts, bottled water and swimming pool (1, 2). To date, the classification of Acanthamoeba using molecular approaches revealed 20 genotypes (T1-T20) based on 18S rDNA gene region (3), most genotypes have been isolated from clinical cases, however, T4 genotype is the most common strain among the clinical and environmental specimens. The T4 genotype has greater pathogenic ability among the others (1, 4). Moreover, T4 genotype is the most common genotype isolated from corneal scrapes of keratitis patients in Iran (5, 6).

Acanthamoeba spp. acting as opportunistic amoebeae is able to develop two severe diseases including Granulomatous Amoebic Encephalitis (GAE) and sight-threatening corneal infection termed Acanthamoeba keratitis (AK) (1). The major risk factor for AK is the usage of contact lenses and exposure to contaminated water. Moreover, swimming or bathing in the contaminated aquatic environment is hazardous to contact lens wearers (1, 2). Since Acanthamoeba could act as a reservoir for pathogenic microorganism, this could double the threat for human health (1, 7). Several studies in different parts of Iran showed the occurrence of Acanthamoeba genotypes and other free-living amoebae (FLA) in the various sources of waters. The presence of Acanthamoeba was revealed in recreational water and tap water sources in northern and southern Iran, Gilan and Kish Island, respectively. The results showed the occurrence of T4 genotype in recreational waters and T3, T4, T5 and T11 genotypes in tap water sources (8, 9). Studies on river waters and hot springs in the country were also detected T3, T4, T5 and T15 genotypes of *Acanthamoeba* and *Naegleria* (10-13).

Despite numerous studies conducted on water sources in Iran, there were no data regarding the contamination of wastewater sources to *Acanthamoeba* spp. in the region and thus the main aim of the current study was to determine the occurrence of *Acanthamoeba* genotypes in wastewater of two treatment facilities in west and south of Tehran Province. The pathogenic potential of the isolated strains were also evaluated using thermos and osmotolerance assays.

This study aimed to determine the genotypes of isolated *Acanthamoeba* spp. in raw wastewater samples in Tehran, Iran.

Materials and Methods

Sampling

During 2014-2016, 90 raw wastewater samples were collected from water treatment facilities in west and south of Tehran Province. Fifty water samples were collected from the west of the Tehran, Shahrak-e Ekbātān, and 40 samples were obtained from the south of Tehran, Chahardonge region, both areas are located in the urban region. All of the specimens were transferred to Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Iran.

Sample processing and microscopic survey

Filtration of each sample was performed using nitrocellulose membranes (45-µm diameter). The filters were cultured on 1.5% non-nutrient agar (NNA) medium plates with heatinactivated *Escherichia coli* (14). The plates monitoring was performed daily using an inverted microscope (15). Cloning of the posi-

tive plates was done to purify the amoebae from bacterial and fungal contamination (5).

In vitro pathogenicity tests

Osmo-tolerance and thermo-tolerance assays were performed on the positive strains. For osmotolerance test, non-nutrient agar with 1 M and 0.5 M mannitol concentration was used to evaluate the outgrowth of amoebae (16). For thermo-tolerance assay, the growth of amoebae was surveyed by the inverted microscopy. The temperature was set at 37 and 42 °C for a week (17). All the plates were monitored for one week.

DNA extraction, PCR amplification, and Sequencing

Acanthamoeba spp. were harvested using sterile phosphate buffered saline (PBS), at pH 7.2, then the harvested amoebae were concentrated and lysed using lysozyme and glass beads treatment. High Pure polymerase chain reaction (PCR) Template Preparation Kit (Roche, Mannheim, Germany) was used for DNA extraction. PCR amplified the ASA.S1 region (500 bp) of 18S rRNA gene, along with primers IDP1 5'-GGCCCAGATCGTTTACCGTGAA-3' and JDP2 5'-TCTCACAAGCTGCTAGGGAGTCA-3' (16). A total of 30-µl volume of PCR reaction was fixed by using Amplicone (Taq DNA Polymerase Master Mix RED, Denmark), 0.1 µM of each primer, distilled water, and DNA template. The cycling conditions stared with 94 °C for 1 min; 35 cycles of 94 °C for 35 sec, annealing step was 56 °C for 45 sec and 72 °C for 1 min; final extension was 72 °C for 10 min. The presence of PCR product bands was confirmed with UV light after electrophoresis and staining of PCR product. Purification and sequencing were done using the ABI 3130X sequencer. The sequences adjusted with chromas (ver. 1.0.0.1), afterward compared with BLAST GenBank database.

Results

Of 50 collected samples from the raw wastewater treatment facility of Shahrak-e Ekbatan, 1 (2%) was positive for *Acanthamoeba* spp. From the other 40 specimens belonged to Chahardonge, 5 (12%) were positive (Table 1). Morphologically, all positive strains were belonged to group two morphology (Fig. 1). The result of sequencing showed that 1 (17%) strain labeled as a TWS2 belonged to T11 (corresponding to *A. hatchetii* with 100% homology to the genes available in the gene data bank) and 5 (83%) belonged to T4 genotype (TWS1, TWS3, TWS4, TWS5, TWS6).

In vitro pathogenicity tests including osmotolerance and thermo-tolerance assays were confirmed that only a single isolate (TW1) was highly pathogenic as the strain showed a good growth at high temperature and osmolarity, while the other strains were classified as non-pathogenic amoebae (Table 1).

Table 1: Data of genotyping and pathogenicity test from wastewater samples during 2014-2016, Tehran, Iran

| Code | Locality | NNA culture | PCR | Genera | Genotype | Pathogenicity |
|------|-------------|----------------|-----|--------------|----------|---------------|
| TWS1 | Ekbātān | + | + | Acanthamoeba | T4 | High |
| TWS2 | Ekbātān | + | + | Acanthamoeba | T11 | Low |
| TWS3 | Ekbātān | + | + | Acanthamoeba | T4 | Low |
| TWS4 | Ekbātān | + | + | Acanthamoeba | T4 | Low |
| TWS5 | chahardonge | + | + | Acanthamoeba | T4 | Low |
| TWS6 | Ekbātān | + | + | Acanthamoeba | T4 | Low |

TWS: Tehran wastewater sample

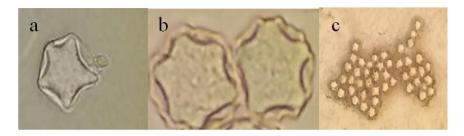


Fig. 1: Light microscopy photographs of *Acanthamoeba* cyst in non-nutrient agar. (a) T11 genotype. (b) T4 genotype x1000. (c) T4 genotype x100

Discussion

This study is the first survey of Acanthamoeba spp. isolated from the wastewater specimens in the country. Our study showed 6.7% of the samples were positive for Acanthamoeba spp., in which 83% belonged to T4 genotype. The data of this study reflected the occurrence of Acanthamoeba spp. in the wastewater of the capital in the country. However, despite the other environmental studies in Iran, the current research reflected a low contamination of wastewater sources to Acanthamoeba. This may be due to inhibitory effect of the contaminant in the polluted water. Only Acanthamoeba was detected in the studied samples and other freeliving amoebae were not observed. This can be explained by more resistance of Acanthamoeba cysts in harsh conditions and the fact that other free-living amoebae are more fragile (18). The result of the other studies is opposite to ours. A study conducted in wastewater treatment plant (WWTP) process and sludge in agriculture in the Navarra Community of Spain region showed Acanthamoeba spp. as the most common protozoa in outlet water and sludge (19). Moreover, another research in wastewater treatment plant of central Spain detected Acanthamoeba belonged to T16 and T7 genotypes. Moreover, phylogenetic analysis detected T19 genotype among the samples (20). Another research in South Africa showed the occurrence of free-living amoebae (87.2%) in wastewater treatment plant (21). The current study showed the presence of T4 and T11 genotypes in the contaminated water samples.

Acanthamoeba belonging to T3, T4, T7 and T9 genotypes, Hartmannella and Naegleria were isolated from the Spanish wastewater treatment plants (22). Moreover, the most frequent FLA was the genus Acanthamoeba (59%) from wastewater in the north of Mexico City (23). Despite the mentioned researchers in the world, there were no data regarding Acanthamoeba spp. distribution of wastewater samples in Iran.

Most of the wastewaters recovered after treatment and reused directly for irrigation systems, river flow augmentation and industrial consumption (13, 24). As expected, the present study reflected that the most common strain belonged to T4 genotype. T4 genotype was also the frequently isolated strain from clinical and environmental samples (5, 6, 10). However, the interesting point in the present research was the low pathogenicity of most strains. This is in agreement with study that revealed not all T4s have pathogenic potential (25). Moreover, nine strains from nasal swabs of immunocompromised patients belonged to T4 genotype were classified as non-pathogenic ones (16).

Acanthamoeba belonged to T11 genotype (A. hatchetii) was another isolated strain in the wastewater specimens, this genotype was reported from the keratitis patients in Iran and isolated from the environmental samples previously (6, 26). However, this strain showed low pathogenic potential. More studies regarding the pathogenic ability of the isolated

strains such as in vivo tests and animal-based studies are of importance.

Conclusion

The data of current study was reflected the low occurrence of *Acanthamoeba* genotypes in the raw wastewater samples. More studies about the contamination of wastewaters before and after treatment are required in different places of the country.

Conflict of Interests

The authors declare that there is no conflict of interest.

References

- Khan NA. Acanthamoeba. biology and increasing importance in human health. FEMS Microbiol Rev. 2006;30(4):564-95.
- 2. Schuster FL, Visvesvara GS. Free-living amoebae as opportunistic and non-opportunistic pathogens of humans and animals. Int J Parasitol. 2004;34(9):1001-27.
- 3. Corsaro D, Walochnik J, Köhsler M et al. Acanthamoeba misidentification and multiple labels: redefining genotypes T16, T19, and T20 and proposal for Acanthamoeba micheli sp. nov. (genotype T19). Parasitol Res. 2015; 114(7):2481-90.
- 4. Maghsood AH, Sissons J, Rezaian M et al. Acanthamoeba genotype T4 from the UK and Iran and isolation of the T2 genotype from clinical isolates. J Med Microbiol. 2005;54(Pt 8):755-9.
- 5. Niyyati M, Lorenzo-Morales J, Rezaie S et al. Genotyping of *Acanthamoeba* isolates from clinical and environmental specimens in Iran. Exp Parasitol. 2009;121(3):242-5.
- 6. Hajialilo E, Behnia M, Tarighi F et al. Isolation and genotyping of *Acanthamoeba* strains (T4, T9, and T11) from amoebic keratitis patients in Iran. Parasitol Res. 2016;115(8):3147-51.
- 7. Horn M, Wagner M. Bacterial endosymbionts of free-living amoebae. J Eukaryot Microbiol. 2004;51(5):509-14.
- 8. Niyyati M, Nazar M, Haghighi A et al. Reporting of T4 genotype of *Acanthamoeba* isolates in recreational water sources of Gilan Province, Northern Iran. Novel Biomed. 2015;3(1):20-4.

- Niyyati M, Lasgerdi Z, Lorenzo-Morales J. Detection and molecular characterization of potentially pathogenic free-living amoebae from water sources in Kish Island, Southern Iran. Microbiol Insights. 2015;8(Suppl 1):1-6.
- 10. Nazar M, Haghighi A, Niyyati M et al. Genotyping of *Acanthamoeba* isolated from water in recreational areas of Tehran, Iran. J Water Health. 2011;9(3):603-8.
- Solgi R, Niyyati M, Haghighi A et al. Thermotolerant *Acanthamoeba* spp. isolated from therapeutic hot springs in Northwestern Iran. J Water Health. 2012;10(4):650-6.
- 12. Niyyati M, Lasjerdi Z, Nazar M et al. Screening of recreational areas of rivers for potentially pathogenic free-living amoebae in the suburbs of Tehran, Iran. J Water Health. 2012;10(1):140-6.
- Levine AD, Asano T. Recovering sustainable water from wastewater. Environ Sci Technol. 2004;38(11):201A-208A.
- Lorenzo-Morales J, Ortega-Rivas A, Martínez E et al. *Acanthamoeba* isolates belonging to T1, T2, T3, T4 and T7 genotypes from environmental freshwater samples in the Nile Delta region, Egypt. Acta Trop. 2006;100(1-2):63-9.
- 15. Page FC. A new key to freshwater and soil *Gymnamoebae*: with instructions for culture. Ambleside, UK: Freshwater Biological Association; 1988. p. 122.
- Memari F, Niyyati M, Haghighi A et al. Occurrence of pathogenic *Acanthamoeba* genotypes in nasal swabs of cancer patients in Iran. Parasitol Res. 2015;114(5):1907-12.
- 17. Khan NA. Pathogenicity, morphology, and differentiation of *Acanthamoeba*. Curr Microbiol. 2001;43(6):391-5.
- 18. Ramirez E, Robles E, Martinez B et al. Distribution of free-living amoebae in a treatment system of textile industrial wastewater. Exp Parasitol. 2014;145 Suppl:S34-8.
- Marin I, Goni P, Lasheras A, Ormad M. Efficiency of a Spanish wastewater treatment plant for removal potentially pathogens: Characterization of bacteria and protozoa along water and sludge treatment lines. Ecol Eng. 2015;74:28-32.
- 20. Magnet A, Henriques-Gil N, Galván-Diaz AL et al. Novel *Acanthamoeba* 18S rRNA gene sequence type from an environmental isolate. Parasitol Res. 2014;113(8):2845-50.

- 21. Muchesa P, Mwamba O, Barnard TG et al. Detection of free-living amoebae using amoebal enrichment in a wastewater treatment plant of gauteng province, South Africa. Biomed Res Int. 2014; 2014:575297.
- 22. García A, Goñi P, Clavel A et al. Potentially pathogenic free-living amoebae (FLA) isolated in Spanish wastewater treatment plants. Environ Microbiol Rep. 2011;3(5):622-6.
- 23. Ramirez E, Robles E, Bonilla P et al. Occurrence of Pathogenic Free-Living Amoebae and Bacterial Indicators in a Constructed Wetland

- Treating Domestic Wastewater from a Single Household. Eng Life Sci. 2005;5(3):253-8.
- Asano T. Wastewater Reclamation and Reuse: Water Quality Management Library: CRC Press; 1998
- Mirjalali H, Niyyati M, Abedkhojasteh H et al. Pathogenic assays of *Acanthamoeba* belonging to the T4 genotype. Iran J Parasitol. 2013;8(4):530-5.
- 26. Astorga B, Lorenzo-Morales J, Martín-Navarro CM et al. *Acanthamoeba* Belonging to T3, T4, and T11: Genotypes Isolated from Air-Conditioning Units in Santiago, Chile. J Eukaryot Microbiol. 2011;58(6):542-4.

Available at: http://ijpa.tums.ac.ir 521