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## Understanding cold bias: Variable response of skeletal Sr/Ca to seawater $p\text{CO}_2$ in acclimated massive *Porites* corals

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Coral skeletal Sr/Ca is a palaeothermometer commonly used to produce high resolution seasonal sea surface temperature (SST) records and to investigate the amplitude and frequency of ENSO and interdecadal climate events. The proxy relationship is typically calibrated by matching seasonal SST and skeletal Sr/Ca maxima and minima in modern corals. Applying these calibrations to fossil corals assumes that the temperature sensitivity of skeletal Sr/Ca is conserved, despite substantial changes in seawater carbonate chemistry between the modern and glacial ocean. We present Sr/Ca analyses of 3 genotypes of massive *Porites* spp. corals (the genus most commonly used for palaeoclimate reconstruction), cultured under seawater  $p\text{CO}_2$  reflecting modern, future (year 2100) and last glacial maximum (LGM) conditions. Skeletal Sr/Ca is indistinguishable between duplicate colonies of the same genotype cultured under the same conditions, but varies significantly in response to seawater  $p\text{CO}_2$  in two genotypes of *Porites lutea*, whilst *Porites murrayensis* is unaffected. Within *P. lutea*, the response is not systematic: skeletal Sr/Ca increases significantly (by 2–4%) at high seawater  $p\text{CO}_2$  relative to modern in both genotypes, and also increases significantly (by 4%) at low seawater  $p\text{CO}_2$  in one genotype. This magnitude of variation equates to errors in reconstructed SST of up to  $-5^\circ\text{C}$ .

Coral skeletal Sr/Ca is influenced by sea surface temperature (SST) and is a widely used palaeothermometer<sup>1</sup>. Strong correlations have been observed between instrumental SST records and skeletal Sr/Ca in modern corals over seasonal to decadal timescales<sup>2–4</sup>, providing a reliable proxy for the reconstruction of multi-centennial SST variations and valuably extending the record of many natural climate oscillations back to the 16<sup>th</sup> Century<sup>5–7</sup>. Over millennial timescales, the accuracy of SST estimates from Sr/Ca ratios in fossil corals has been questioned as they frequently yield improbably cool or spatially inconsistent values<sup>8–10</sup>. Whilst some of these inconsistencies have been explained by regional oceanographic dynamics<sup>11</sup>, a number of possible influences on skeletal Sr/Ca in coral remain unresolved. Skeletal Sr/Ca is correlated with skeletal growth rate in some<sup>12,13</sup>, but not all<sup>14,15</sup> corals and may be influenced by diagenesis<sup>16,17</sup>, skeletal ultrastructure<sup>14,18</sup>, metabolic cycles<sup>19</sup>, and potential variations in seawater Sr/Ca<sup>20</sup>. Calibration equations describing the temperature dependence of coral Sr/Ca also vary significantly, both between species and within a single species sampled from different reef locations<sup>21,22</sup>. However these factors appear insufficient to explain cold bias e.g. in an early Holocene Tahitian fossil coral<sup>10</sup>. Understanding the origin of sensitivity variations within the Sr/Ca-SST relationship is essential if we are to interpret accurately the high resolution climate records encoded in coral skeletons and benefit from their application in resolving the controls on past climates and predicting future change. Coral palaeoproxy relationships are derived by correlating annual SST variations with skeletal Sr/Ca in modern specimens<sup>21,22</sup>. In applying these empirical relationships to fossil specimens, it is implicitly assumed that the processes affecting skeletal geochemistry have not changed between the present and past. It is unlikely that this is the case: in particular, atmospheric  $\text{CO}_2$  has varied significantly over glacial-interglacial time scales<sup>23</sup> affecting both seawater pH and dissolved inorganic carbon (DIC) chemistry<sup>24</sup>, factors which influence skeletal trace element incorporation in some calcitic marine organisms<sup>25</sup>.

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The influence of pH and DIC on coral skeletal Sr/Ca is poorly constrained. Culture studies investigating the effect of reduced seawater pH across a range of ocean acidification scenarios yield mixed results: seawater pH and Sr/Ca were negatively correlated in *Acropora digitifera*<sup>26</sup> and newly settled recruits of *Favia fragum*<sup>27</sup> but were unrelated in *Montipora capitata*<sup>13</sup>. No systematic variation in skeletal Sr/Ca was found over a wider pH range (both lower and higher than present day) in *Stylophora pistillata*<sup>28</sup>. With the exception of *A. digitifera*<sup>26</sup>, these studies used addition of HCl or NaOH to manipulate seawater pH at constant seawater  $p\text{CO}_2$ , consequently shifting other carbonate system parameters to unrealistic values<sup>29</sup>. Additionally, the short acclimation times (5–35 days) of three<sup>26–28</sup> of the four studies are probably insufficient to enable coral physiological responses e.g. changes in zooxanthellae density, to adapt to changes in environment.

We test the direct impact of variations in seawater  $p\text{CO}_2$  on skeletal Sr/Ca in massive *Porites* spp. corals, the coral genus most commonly used for palaeoclimate reconstruction. Heads from 3 genetically distinct corals ( $2 \times$  *Porites lutea* and  $1 \times$  *Porites murrayensis*; Fig. 1) were divided into smaller sub-colonies (each  $>8$  cm diameter) and cultured in a mixture of natural and synthetic seawater. Corals were housed in a purpose-built large-volume aquarium system constructed of low  $\text{CO}_2$  permeability materials. The seawater in each treatment was bubbled with gas mixes set to reach the target seawater  $p\text{CO}_2$  compositions. Corals were maintained at ambient  $p\text{CO}_2$  conditions for 2 months, adjusted to  $p\text{CO}_2$  treatment conditions over another 2 months and then acclimated at the final treatment  $p\text{CO}_2$  for 5 months. Final seawater  $p\text{CO}_2$  target levels ranged from the last glacial maximum (LGM;  $\sim 180 \mu\text{atm}$ ), through present day ( $\sim 400 \mu\text{atm}$ ) to levels projected by the year 2100 ( $\sim 750 \mu\text{atm}$ ). Actual seawater  $p\text{CO}_2$  levels (198, 416 and  $750 \mu\text{atm}$ ) in the reservoirs were calculated from total alkalinity and DIC using the CO2SYS program<sup>30</sup>. Seawater Sr/Ca, temperature, salinity and DIC system parameters were maintained within narrow limits throughout the study (Table 1).

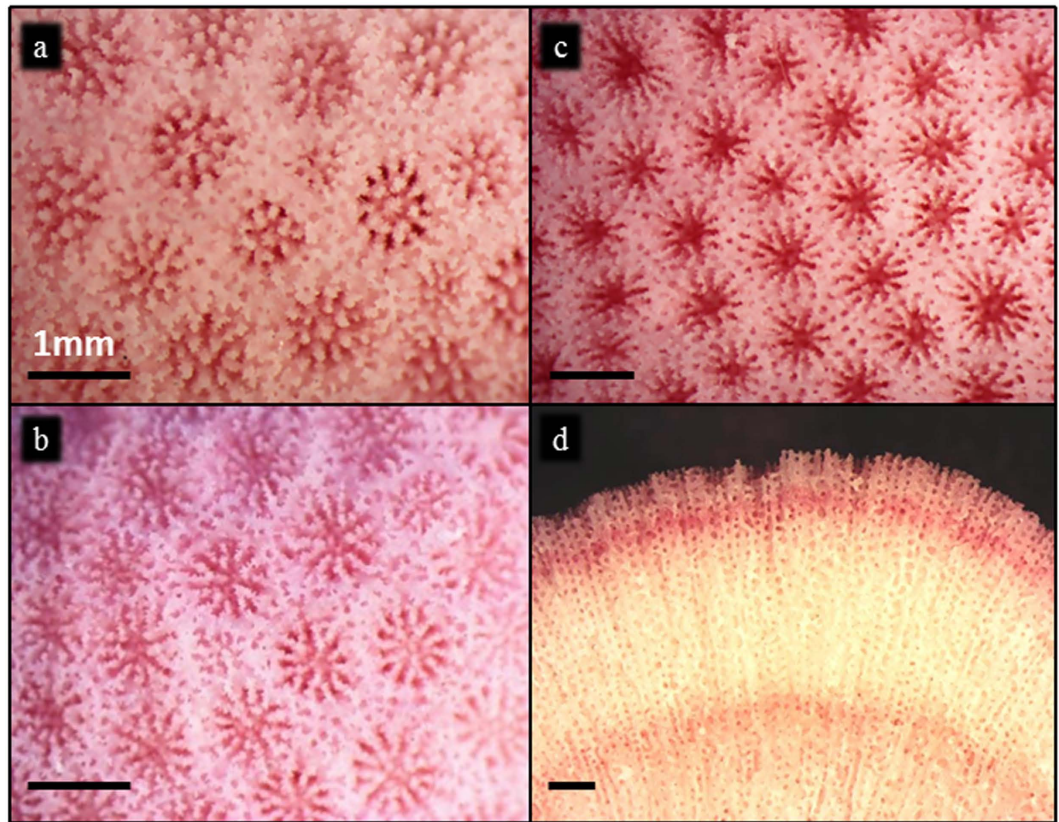
The skeleton deposited in the 5-week period, following the acclimation, was identified by alizarin red staining (Fig. 1), and skeletal Sr/Ca of this region was analysed by secondary ion mass spectrometry (SIMS). Skeletal deposition is concentrated at the skeleton surface in massive *Porites* corals, although subtle thickening of the trabeculae may occur over a period of weeks<sup>31</sup>. The high spatial resolution of SIMS allows the selective analysis of skeleton deposited at the skeletal surface, avoiding the centres of calcification where Sr/Ca is anomalously high, and any material which thickens the trabecula and may be deposited a considerable time later<sup>32</sup>. The accuracy of our SIMS estimates is hindered by uncertainty in the elemental composition of the NaHxby2 standard, but our analytical precision, which is critical to palaeothermometry, is good (within 0.6%, or  $0.018 \text{ mmol mol}^{-1}$ , ( $2\sigma$ ), equivalent to  $\pm 0.4^\circ\text{C}$ ).

## Results and Discussion

**Skeletal Sr/Ca varies with seawater  $p\text{CO}_2$  in some genotypes.** Coral aragonite Sr/Ca partition coefficients ( $K_D^{\text{Sr/Ca}}$  = skeletal Sr/Ca/seawater Sr/Ca) are estimated for each coral (Supplementary Table S1).  $K_D^{\text{Sr/Ca}}$  varies significantly in response to perturbations in seawater  $p\text{CO}_2$  in both genotypes of *P. lutea* but is unaffected in *P. murrayensis* (Fig. 2). Seawater temperatures do not vary significantly between the treatments (Table 1) and we observe excellent agreement (within 0.3%) in the  $K_D^{\text{Sr/Ca}}$  of duplicate sub colonies of the same coral genotype within each treatment (Fig. 2), indicating that minor differences in coral positioning and lighting in each tank do not affect skeletal Sr incorporation.  $K_D^{\text{Sr/Ca}}$  is significantly increased at high seawater  $p\text{CO}_2$  compared to ambient in both genotypes of *P. lutea* (by 4% in genotype 1, and 2% in genotype 2;  $p < 0.05$ , ANOVA and Tukey post-hoc).  $K_D^{\text{Sr/Ca}}$  is also significantly increased (by 4%;  $p < 0.05$ ) in *P. lutea* genotype 2 at low  $p\text{CO}_2$  compared to ambient. In contrast,  $K_D^{\text{Sr/Ca}}$  does not vary significantly between the *P. murrayensis* sub-colonies cultured over the full seawater  $p\text{CO}_2$  range.

Two hypotheses have been proposed to explain the impact of seawater  $p\text{CO}_2$  on coral skeletal Sr/Ca<sup>26</sup>. Coral aragonite precipitates from an extracellular calcifying fluid enclosed in a semi-isolated space between the coral tissue and underlying skeleton, and corals increase the pH of this fluid (upregulate pH) above that of ambient seawater to promote high fluid aragonite saturation states favourable for skeletal precipitation. The coral calcification fluid is derived from seawater which is transported paracellularly (between cells) to the calcification site and from additional Ca transported transcellularly (across cells) via L-type Ca channels<sup>33,34</sup> and the enzyme Ca-ATPase<sup>33,35</sup>. Enhancing transcellular Ca transport could decrease the Sr/Ca of the calcification fluid. This hypothesis is appealing as corals maintained at high seawater  $p\text{CO}_2$  are able to upregulate calcification fluid pH more to partially offset the effect of lowered seawater pH<sup>36</sup>. Ca-ATPase is a  $\text{Ca}^{2+}:\text{H}^+$  antiport which increases the pH of the calcification fluid<sup>37</sup> and may be expected to have a higher activity under high seawater  $p\text{CO}_2$ . This hypothesis suggests that coral skeletal Sr/Ca is decreased at high seawater  $p\text{CO}_2$ . However,  $\text{Sr}^{2+}$  has a similar ionic radius to  $\text{Ca}^{2+}$  and inhibition of Ca channels and Ca-ATPase in the branching coral, *Pocillopora damicornis*, did not affect skeletal Sr/Ca, suggesting that transmembrane Ca transport does not fractionate Sr/Ca in the calcification fluid<sup>38</sup>.

Alternatively, skeletal Sr/Ca variations may reflect elemental partitioning during aragonite precipitation (Rayleigh fractionation). Rayleigh fractionation occurs when variable volumes of an isolated fluid reservoir are used for solid precipitation, and a trace element ion (in this case Sr) is either discriminated against or preferentially incorporated compared to the major precipitating ion (in this case Ca). The Sr/Ca partition coefficient in aragonite is  $> 1$ <sup>39</sup>, so Sr is preferentially incorporated into the growing crystals over Ca<sup>40</sup>. As precipitation proceeds, the Sr/Ca of the fluid remaining in the reservoir, and of the carbonate subsequently precipitated from it, decreases. The final Sr/Ca of the aragonite reflects the proportion of the reservoir used in precipitation<sup>40</sup>. The deposition rate of coral aragonite is strongly dependent on the saturation state of the calcifying fluid<sup>41,42</sup>. At high calcification fluid saturation states, a higher proportion of the fluid reservoir may be used for calcification resulting in a low skeletal Sr/Ca. Although corals at high seawater  $p\text{CO}_2$  increase the pH of the calcification fluid more than their counterparts cultured at ambient  $p\text{CO}_2$ , they do not ultimately attain such a high fluid pH<sup>36</sup> and calcification rates in these corals are usually reduced<sup>43</sup>. This hypothesis suggests that coral skeletal Sr/Ca will be increased at high seawater  $p\text{CO}_2$ . Neither hypothesis immediately explains our observed pattern of skeletal Sr/Ca



**Figure 1. Multiple genotypes of massive *Porites* spp. stained with alizarin.** Representative colonies of (a) *Porites lutea* genotype 1; (b) *Porites lutea* genotype 2; (c) *Porites murrayensis*, cultured at ambient  $p\text{CO}_2$  within our aquarium system. (d) Alizarin stain lines mark skeletal extension during the 5-week experimental period. Scale bars are 1mm.

variations where skeletal Sr/Ca is increased in some corals at both low and high seawater  $p\text{CO}_2$ . However it is unlikely that the two hypotheses operate independently as the saturation state of the calcification fluid is probably highly responsive to Ca-ATPase activity.

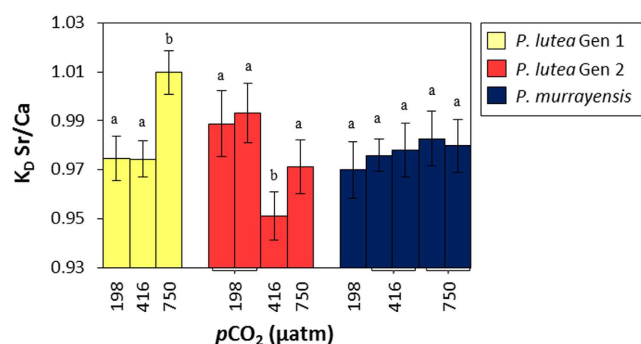
**Growth rate and skeletal Sr/Ca.** We explored the relationship between coral aragonite:seawater  $K_D^{\text{Sr/Ca}}$  and skeletal growth rate, inferring that Rayleigh fractionation is linked to skeletal precipitation rate. We observe significant differences in both the skeletal extension and calcification rate of some genotypes between treatments (Supplementary Table S2). However, correlation between  $K_D^{\text{Sr/Ca}}$  and either skeletal linear extension or calcification rate (Supplementary Fig. S1) are insignificant in all coral genotypes ( $p > 0.3$ ; Pearson's correlation). Our data suggest that variations in Sr incorporation between corals do not reflect Rayleigh fractionation.

**Alternative controls on skeletal Sr/Ca.** We observe significant variation in skeletal Sr/Ca between different coral genotypes cultured at the same seawater  $p\text{CO}_2$  (Fig. 3) demonstrating that other factors also influence Sr incorporation. The origin of this is currently unknown. The mucus layer at the coral surface behaves as a Donnan matrix, concentrating  $\text{Ca}^{2+}$  from seawater prior to transport across the coral ectoderm tissue<sup>44</sup> and matrix variations between corals could conceivably subtly influence the Sr/Ca of seawater transported to the calcification fluid. Organic molecules at the calcification site also play a major role in regulating aragonite nucleation and growth<sup>45</sup>. Skeletal organic materials differ between coral species<sup>46</sup>, and may influence skeletal trace element content either because biomolecules impact the incorporation of trace elements in the crystal lattice, or due to direct organic-metal complexing. Understanding the role of these organic templates in fractionating Sr/Ca during calcification, and quantifying differences between coral genotypes even within an individual species, will be a critical step forward in resolving biological influences on skeletal Sr/Ca from the thermal signature.

**Implications for SST reconstructions.** Our data show that seawater  $p\text{CO}_2$  can be a significant factor affecting the Sr/Ca of massive tropical coral skeletons frequently used for palaeotemperature reconstruction. Decreasing seawater  $p\text{CO}_2$  from ambient (416  $\mu\text{atm}$ ) to values consistent with the LGM (198  $\mu\text{atm}$ ) increases skeletal Sr/Ca by 4% in one of the three coral genotypes. The sensitivity of coral skeletal Sr/Ca to temperature is typically  $\sim 0.8\text{--}1\% \text{ } ^\circ\text{C}^{-1}$  and the effect of the decrease in seawater  $p\text{CO}_2$  implies a cooling of 4–5  $^\circ\text{C}$  in this single genotype, although all colonies were cultured at the same temperature. The influence of seawater  $p\text{CO}_2$  on skeletal

Treatment	Sr/Ca (mmol/mol)	Temperature (°C)	Salinity	TA (μeq/kg)	DIC (μmol/kg)	pH <sub>T</sub>	pCO <sub>2</sub> (μatm)	HCO <sub>3</sub> <sup>-</sup> (μmol/kg)	CO <sub>3</sub> <sup>2-</sup> (μmol/kg)	Ω <sub>Ar</sub>
Last Glacial Maximum	12.27 ± 0.09	25.3 ± 0.1	35.1 ± 0.04	2289 ± 14	1831 ± 11	8.28	198	1511	315	5.01
Modern Day	12.07 ± 0.03	25.2 ± 0.2	35.2 ± 0.08	2290 ± 8	1999 ± 8	8.03	416	1780	207	3.29
Future (~Year 2100)	11.85 ± 0.03	25.1 ± 0.2	35.2 ± 0.05	2293 ± 5	2113 ± 11	7.81	750	1954	138	2.19

**Table 1. Summary of the physical and chemical composition of seawater in each culture treatment, measured over the 5-week experimental period.** Values are the mean ± 1σ. Dissolved inorganic carbon (DIC) and Total Alkalinity (TA) were used to calculate seawater carbonate system components (pH<sub>T</sub>, pCO<sub>2</sub>, HCO<sub>3</sub><sup>-</sup>, CO<sub>3</sub><sup>2-</sup> and Ω<sub>Ar</sub>) using CO2SYS (Pierrot *et al.*<sup>30</sup>, whereby pH is Total scale; K<sub>1</sub> and K<sub>2</sub> are from Mehrbach *et al.*<sup>61</sup> refit by Dickson and Millero; K<sub>SO4</sub> as determined by Dickson.

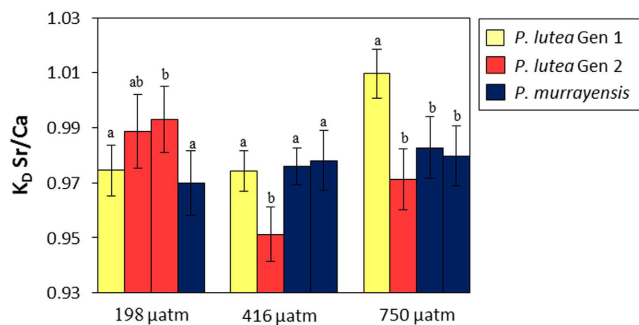


**Figure 2. Influence of seawater pCO<sub>2</sub> on coral aragonite:seawater Sr/Ca partition coefficients (K<sub>D</sub><sup>Sr/Ca</sup>) for 3 genotypes of *Porites* spp. following acclimation to 198, 416 and 750 μatm pCO<sub>2</sub> at 25 °C.** Bars represent the mean K<sub>D</sub><sup>Sr/Ca</sup> of multiple analyses (n = 12–41; see Table S1) across 2 or more skeletal units within individual colonies. Within each species/genotype, different letters indicate significant differences between treatments (p < 0.05; ANOVA and Tukey post-hoc). Error bars represent the combined 95% confidence limits of seawater and skeletal Sr/Ca measurements.

Sr/Ca may explain the ‘cold bias’ observed in some fossil corals, which suggest that tropical SSTs at the last or penultimate deglaciations in the Atlantic<sup>47</sup> and Pacific<sup>48</sup> Oceans were ~6 °C cooler than at present. These estimates contrast with other marine proxies which indicate that SSTs typically cooled by 2–3 °C at most at this time<sup>49,50</sup>. Proposed variations in seawater Sr/Ca over glacial-interglacial periods are insufficient to explain the ‘cold bias’ observed in coral SST reconstruction<sup>20</sup>.

We find that the influence of seawater pCO<sub>2</sub> on skeletal Sr/Ca is inconsistent between corals, even of the same species, and this prevents the calculation of a correction factor to compensate for past variations in seawater pCO<sub>2</sub>. Skeletal Sr/Ca is unaffected by pCO<sub>2</sub> over the modern-LGM range relevant to paleotemperature reconstruction in two of our cultured genotypes, yet one genotype shows a substantial, and significant response that is observed across duplicate sub-colonies. We are unable to relate the seawater pCO<sub>2</sub> effect to coral growth rate indicating that coral extension cannot be used to correct skeletal Sr/Ca for an indirect influence of pCO<sub>2</sub> in *Porites* spp., despite successful application of a growth correction in other coral genera<sup>51</sup>. Further culture work is required to resolve the relationship between skeletal Sr/Ca and seawater pCO<sub>2</sub> across a wider range of coral genotypes, and to identify how seawater pCO<sub>2</sub> contributes to ‘cold bias’. At present, the accuracy of absolute tropical past SST estimates from skeletal Sr/Ca of fossil corals is limited by this uncertainty. In contrast, relative changes in annual seawater temperature are usually well preserved in coral skeletal Sr/Ca<sup>52</sup> and fossil specimens may yet provide good records of Sr/Ca derived seasonal SST variations which can be used to infer the amplitude and frequency of past ENSO<sup>53</sup> and interdecadal climate events<sup>54</sup>. However, it will also be important to quantify in further studies whether skeletal Sr/Ca temperature sensitivity varies between different coral genotypes, and is affected by seawater pCO<sub>2</sub>.

In this study we hypothesise that inconsistencies in SST reconstructions of the LGM from coral Sr/Ca may be explained by a change in seawater pCO<sub>2</sub> between the modern and glacial ocean, equivalent to ~220 μatm. Approximately 50% of this increase in pCO<sub>2</sub> towards present day has occurred in the last 150 years as a result of rising greenhouse gas emissions, overlapping with multi-centennial coral Sr/Ca records. Anthropogenic CO<sub>2</sub> emissions are isotopically depleted in <sup>13</sup>C, and a centennial-scale decrease in the δ<sup>13</sup>C of surface ocean DIC attributable to rising atmospheric CO<sub>2</sub> concentrations<sup>55</sup> is also observed in the skeletal aragonite of >200 year-old corals collected at the end of the 20<sup>th</sup> century<sup>56,57</sup>. Between 1850 and 2000, the ~85 μatm increase in atmospheric CO<sub>2</sub> is clearly preserved in the carbon isotopic ratios of these corals, yet there is no discernible influence on skeletal Sr/Ca<sup>56</sup>. In our culture study, a significant increase in skeletal Sr/Ca was observed at low pCO<sub>2</sub> compared to ambient in one of three genotypes, and we may therefore expect some corals to show a decrease in Sr/Ca with rising CO<sub>2</sub> over the 20<sup>th</sup> century. In practice, any CO<sub>2</sub> forcing on coral Sr/Ca is likely to be obscured by significant interannual variations in SST caused by ENSO against a global trend of increasing SST<sup>55,6,58</sup>. It is also important to



**Figure 3. Genotypic variations in mean coral aragonite: seawater  $K_D^{Sr/Ca}$  in individual colonies of *Porites* spp. following > 5 months of acclimation to 198, 416 and 750  $\mu\text{atm}^3$   $p\text{CO}_2$  at 25 °C.** Within each  $p\text{CO}_2$  treatment, different letters indicate significant differences between genotypes ( $p < 0.05$ ; ANOVA and Tukey post-hoc). Error bars represent the combined 95% confidence limits of seawater and skeletal Sr/Ca measurements.

bear in mind that 70% of the increase in atmospheric  $\text{CO}_2$  between 1850 and 2000 occurred in the second half of the twentieth century, so the majority of these multi-centennial coral Sr/Ca records span very small (10–20  $\mu\text{atm}$ ) changes in  $p\text{CO}_2$ . Longer coral Sr/Ca datasets extending into the 21<sup>st</sup> century are required to investigate the effect of >100  $\mu\text{atm}$  changes in  $\text{CO}_2$  on *in situ* skeletal Sr/Ca.

Our study demonstrates that the variations in published skeletal Sr/Ca–SST calibrations between corals of the same species do not necessarily reflect differences in physical (e.g. temperature, light levels and ocean currents) and chemical (e.g. nutrient levels and composition) conditions<sup>22,52</sup>. Rather skeletal Sr/Ca varies significantly between coral genotypes even of the same species cultured under the same conditions. Resolving how genotypic variations affect coral biomineralisation and impact skeletal geochemistry is a key target for future research.

## Methods

**Aquarium System.** Culturing experiments were carried out in the marine culturing facility at the Department of Earth & Environmental Sciences, University of St Andrews, UK. Corals were maintained in 21 litre cast acrylic tanks, recirculated from high density polyethylene reservoirs containing ~900 litres of seawater. The reservoirs were bubbled (at 10 L  $\text{min}^{-1}$ ) with gas mixes set to reach the target seawater  $p\text{CO}_2$  compositions. The modern day treatment was aerated with untreated ambient air. The high and low  $\text{CO}_2$  treatments were aerated with ambient or low- $\text{CO}_2$  air, respectively, combined with high purity  $\text{CO}_2$  (Foodfresh, BOC, UK). Flow rates of air and  $\text{CO}_2$  were regulated by high-precision mass flow controllers (SmartTrak 50 Series, Sierra USA) controlled by purpose-written MATLAB<sup>®</sup> programs. Low- $\text{CO}_2$  air was produced by bubbling ambient air through a caustic solution (0.9 M NaOH and 0.1 M  $\text{Ca}(\text{OH})_2$ ) and rinsing it by bubbling through deionised water<sup>59</sup>. The  $[\text{CO}_2]$  of the low- $\text{CO}_2$  air (before mixing with  $\text{CO}_2$ ) was monitored every 2 hours by automated non-dispersive infra-red  $\text{CO}_2$  analysers (WMA04, PP systems, USA) and ranged from 20–100  $\mu\text{atm}$  depending on the age of the caustic solution. The  $[\text{CO}_2]$  of the low, ambient and high  $\text{CO}_2$  gas streams (after addition of any  $\text{CO}_2$ ) was monitored automatically 3–4 times per day and were  $180 \pm 3$ ,  $400 \pm 5$  and  $761 \pm 6$   $\mu\text{atm}$  (mean  $\pm 1\sigma$ ) over the experimental period. Corals were maintained under LED lighting (Maxspect R420R 160w–10000k) on a 12 h light: 12 h dark cycle, with wavelength settings of 100% A and 20% B such that light intensity at coral depth was ~300  $\mu\text{mol}$ . Corals were fed weekly with rotifers.

**Monitoring Seawater Composition.** Dissolved inorganic carbon (DIC) was measured weekly in each reservoir by LI-7000  $\text{CO}_2$  differential, non-dispersive, infrared gas analyser (Apollo SciTech; AS-C3). Samples were calibrated against a natural seawater certified reference material (CRM; A. Dickson, Scripps Institution of Oceanography). Internal reproducibility was calculated from the standard deviation of 8 replicate measurements of a single sample ( $\sigma/\sqrt{n}$ ), and was always <0.1%. Multiple measurements of the CRM were analysed as unknown samples over the 4 week period to check the calibration, and these were in good agreement with the certified value (unknown =  $2019 \pm 6$  ( $1\sigma$ )  $\mu\text{mol kg}^{-1}$ ,  $n = 4$ ; CRM =  $2014 \mu\text{mol kg}^{-1}$ ).

Total alkalinity (TA) was measured twice daily by automated Gran titration (Metrohm, 888 Titrando). Precision of duplicate ~30 ml TA analyses was typically  $\pm 2 \mu\text{eq kg}^{-1}$ . Between days, the precision of multiple measurements of synthetic  $\text{Na}_2\text{CO}_3$  standards (1015, 1190, and 1379  $\mu\text{mol kg}^{-1}$ ) was consistently  $\pm 3 \mu\text{eq kg}^{-1}$  ( $1\sigma$ ,  $n = 14$ , 10 and 8, respectively). The total alkalinity, [Ca] and [Sr] of the culture seawater was maintained by additions of 0.6 M  $\text{Na}_2\text{CO}_3$  and a mixture of 0.58 M  $\text{CaCl}_2$  + 0.02 M  $\text{SrCl}_2$  by 200  $\mu\text{l}$  volume solenoid diaphragm pumps, evenly spaced over a 24 hour period, controlled by a custom-written MATLAB<sup>®</sup> dosing control program. We adjusted addition rates to maintain total alkalinity within narrow limits ( $\pm \leq 14 \mu\text{mol kg}^{-1}$ ). Total alkalinity variations of this magnitude have little effect on seawater carbonate chemistry (~0.002 pH units, ~0.6% [DIC]) at constant seawater  $p\text{CO}_2$ .

Seawater samples were collected weekly during the experimental period for Sr and Ca analysis by quadrupole ICP-MS (Thermo Scientific X Series) at the National Oceanography Centre, Southampton. Samples were diluted 1000-fold in 5%  $\text{HNO}_3$  (with 5 ppb In as an internal standard) and calibrated against matrix-matched synthetic

standards prepared from 1000  $\mu\text{g ml}^{-1}$  single-element stock solutions (Inorganic Ventures) in 5%  $\text{HNO}_3$ . Based on replicate analyses ( $n = 4$ ) of IAPSO standard seawater, external reproducibility ( $\sigma/\sqrt{n}$ ) was 0.6% for Sr/Ca. Measured values of Sr/Ca in IAPSO ( $8.78 \pm 0.05$  ( $1\sigma$ )  $\text{mmol mol}^{-1}$ ) were in excellent agreement with certified values ( $8.77 \text{ mmol mol}^{-1}$ ).

Nutrients were measured in filtered (Whatman GFF;  $\sim 0.7 \mu\text{m}$ ) seawater samples from each reservoir by flow cell spectrophotometry (Lachat 8000; Scottish Association of Marine Science). Concentrations of  $\text{NH}_4^+$  ( $< 0.19 \pm 0.01 \mu\text{mol L}^{-1}$ ; mean  $\pm 1\sigma$ ),  $\text{PO}_4^{3-}$  ( $< 0.06 \pm 0.00 \mu\text{mol L}^{-1}$ ),  $\text{SiO}_4^{4-}$  ( $< 1.3 \pm 0.03 \mu\text{mol L}^{-1}$ ) and  $\text{NO}_3 + \text{NO}_2$  ( $< 0.95 \pm 0.02 \mu\text{mol L}^{-1}$ ) were within the range reported for pristine reef waters<sup>60</sup>.

**Coral sample processing and SIMS.** Skeletal samples were prepared from the maximum growth axis and fixed in epoxy resin (Epo thin, Struers UK) in 2.5 cm diameter circular moulds, under vacuum. Polished, gold coated sections were analysed using a Cameca imf-4f ion microprobe in the School of Geosciences at the University of Edinburgh ( $^{16}\text{O}^-$  ion beam accelerated at 10.8 kV; primary beam current = 8 nA; energy offset = 75 eV; field aperture 1; contrast aperture 2). Trace element data were collected over 8 days, and multiple analyses of a fossil coral aragonite standard, NaHaxby2 (Sr/Ca = 2.87  $\text{mmol mol}^{-1}$ ), were made each day. Calculation of relative ion yields (RIYs) and standardisation of data was performed as in Allison *et al.*<sup>16</sup>. Internal reproducibility was calculated from ten cycles of a single SIMS analysis ( $2\sigma/\sqrt{10}$ ) and was  $\sim 0.4\%$  for Sr/Ca. External reproducibility (the precision of  $\sim 15$  daily analyses on the standard) was 0.6%, or 0.018  $\text{mmol mol}^{-1}$ , ( $2\sigma$ ) for Sr/Ca. Where significant variations in the RIY for NaHaxby2 were observed between days, instrument drift was distinguished from heterogeneity in the composition of the standard through additional analyses of the same area on a subsequent day. Samples were calibrated against mean standard RIYs, either from daily or weekly analyses depending on this assessment of instrument performance. See Allison *et al.*<sup>32</sup> for representative images of skeletal ultrastructure and SIMS analyses pits<sup>32</sup>.

Daily calcification rates ( $\mu\text{mol CaCO}_3 \text{ cm}^{-2} \text{ d}^{-1}$ ) were estimated for each sub colony from the measured change in seawater TA over an isolation period in the light (5 hours), and the dark (7 hours), on three separate occasions over the experimental period. Linear extension ( $\mu\text{m}$ ) was measured as the distance between the alizarin stain lines in each analysed trabecula.

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### Author Contributions

N.A., A.F. and C.H. designed the study and built the culturing system with K.H. Coral culturing was performed by C.C. and N.A. SIMS was performed by C.C. and N.A. All authors contributed to the analysis of the results and to the writing of the paper.

### Additional Information

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